

Whole-Genome Sequences of Four Strains Closely Related to Members of the *Mycobacterium chelonae* **Group, Isolated from Biofilms in a Drinking Water Distribution System Simulator**

Vicente Gomez-Alvarez, Randy P. Revetta

U.S. Environmental Protection Agency, Office of Research and Development, Cincinnati, Ohio, USA

We report here the draft genome sequences of four *Mycobacterium chelonae* **strains from biofilms subjected to a "chlorine burn" in a chloraminated drinking water distribution system simulator. These opportunistic pathogens have been detected in hospital and municipal water distribution systems, in which biofilms have been recognized as an important factor for their persistence.**

Received 9 November 2015 **Accepted** 7 December 2015 **Published** 21 January 2016

Citation Gomez-Alvarez V, Revetta RP. 2016. Whole-genome sequences of four strains closely related to members of the *Mycobacterium chelonae* group, isolated from biofilms in a drinking water distribution system simulator. Genome Announc 4(1):e01539-15. doi:10.1128/genomeA.01539-15.

Copyright © 2016 Gomez-Alvarez and Revetta. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Vicente Gomez-Alvarez, gomez-alvarez.vicente@epa.gov.

espite the use of disinfectants in drinking water distribution systems (DWDS) to mitigate the presence of pathogens, a diverse and complex microbial community has been shown to inhabit DWDS [\(1\)](#page-1-0). In chloraminated DWDS, the microbial community is often dominated by members of the *Mycobacterium chelonae-abscessus* complex [\(2\)](#page-1-1). These rapidly growing nontuberculous mycobacteria (NTM) are opportunistic pathogens with the ability to form biofilms on the surface of DWDS [\(3\)](#page-1-2). Furthermore, this complex has been implicated in invasive infections in immunocompromised hosts and is represented by *M. chelonae*, *M. abscessus*, *M. immunogenum*, *M. salmoniphilum*, *M. franklinii*, and *M. saopaulense* [\(4](#page-1-3)[–](#page-1-4)[10\)](#page-1-5). Little information is available about the resilience of NTM in DWDS and the potential to cause public health problems [\(11,](#page-1-6) [12\)](#page-1-7).

Strains from this study were isolated from biofilms obtained from a chloraminated DWDS simulator [\(3\)](#page-1-2). Samples were collected from two distinct operational schemes [\(Table 1\)](#page-0-0). Colonies were recovered from R2A plates after 7 days at 27°C. DNA was extracted using the UltraClean DNA microbial isolation kit, according to the manufacturer's instructions (Mo Bio Laboratories, Solana Beach, CA). Paired-end 125-bp libraries were prepared using the Nextera XT DNA library kit, followed by rapid mode sequencing on the HiSeq 2500 platform (Illumina, Inc., San Diego, CA). Prior to assembly, libraries were (i) cleaned from contaminants (adapters, phiX, artifacts, and human), (ii) error corrected via Tadpole, (iii) normalized to \leq 100 \times , (iv) removed of low-coverage (\leq 6 \times) reads, and (v) filtered to a minimum length

read of 125 nucleotide (nt). The reads were processed using the software package BBMap version 35.34 (http://sourceforge.net /projects/bbmap) and *de novo* assembly with SPAdes version 3.5.0 [\(13\)](#page-1-8). The final assembly attributes are listed in [Table 1.](#page-0-0)

The average nucleotide identity (ANI), a similarity index between two genomes [\(14\)](#page-1-9), grouped the four strains into two subclusters (subclusters 2a and 2b). The genome similarity between the two subclusters is 97.245% to 97.553%, with an ANI within all strains of 99.960%. The proposed cutoff for species is 95% to 96% [\(15\)](#page-1-10). Coincidentally, strains of subclusters 2a and 2b were obtained from biofilms attached to polyvinyl chloride (PVC) and copper (Cu) surfaces, respectively. The isolates share an overall 95.086% ANI with *M. chelonae* ATCC 35752 [\(6\)](#page-1-11) and 83.008% to 83.275% ANI with *M. abscessus* ATCC 19977 [\(7\)](#page-1-12) and *M. immunogenum* SMUC14 [\(8\)](#page-1-13), respectively. ANI calculations were performed using the online calculator available from EzGenome (http://www.ezbiocloud.net/ezgenome/ani). Furthermore, a comparative analysis of the *rpoB*, *recA*, and *sodA* genes revealed \leq 95.04% \pm 0.05% sequence homology with representatives of the *M. chelonae-M. abscessus* complex [\(6](#page-1-11)[–](#page-1-4)[10\)](#page-1-5). Genomic comparison confirmed that these isolates belong to the *M. chelonae* group but may constitute a different subspecies.

Genome assemblies were annotated with Prokka version 1.10 [\(16\)](#page-1-14), available as an application in Illumina BaseSpace Labs. The genome sequence of strain H002 contains 5,378 genes, 5,333 coding sequences (CDSs), 3 rRNAs, and 41 tRNAs; strain H003 contains 5,635 genes, 5,551 CDSs, 3 rRNAs, and 81 tRNAs; strain

TABLE 1 Summary statistics of whole-genome assemblies

Strain	Source (surface)	Operational scheme ^a	Fold coverage `X`	No. of contigs	Contig N_{50}	Assembly size (bp)	$G + C$ content $(\%)$	Accession no.
H ₀₀₂	Biofilm (Cu)	Standard I	74	60	192,652	5,421,590	64.00	LIYI00000000
H ₀₀₃	Biofilm (PVC)	Standard I	91	35	515,028	5,588,459	63.94	LJYL00000000
H ₀₇₂	Biofilm (Cu)	Standard II	79	54	210,836	5,449,380	64.00	LIYK00000000
H ₀₇₉	Biofilm (PVC)	Standard II	50	94	170,224	5,726,565	63.90	LJYO00000000

a Standard I, stable chloramine residual; Standard II, stable chloramine residual after a "chlorine-burn."

H072 contains 5,410 genes, 5,362 CDSs, 3 rRNAs, and 45 tRNAs; and strain H079 contains 5,824 genes, 5,362 CDSs, 3 rRNAs, and 76 tRNAs.

Nucleotide sequence accession numbers. The whole-genome shotgun project has been deposited in DDBJ/ENA/GenBank under the accession numbers listed in [Table 1.](#page-0-0) The versions described in this paper are the first versions.

ACKNOWLEDGMENTS

We thank Stacy Pfaller, Jonathan Pressman, and David Wahman for valuable discussions and assistance with this project.

The opinions expressed are those of the authors and do not necessarily reflect the official positions and policies of the U.S. EPA. Any mention of product or trade names does not constitute recommendation for use by the U.S. EPA.

FUNDING INFORMATION

The U.S. EPA through the Office of Research and Development funded and managed this research.

REFERENCES

- 1. **Roeselers G, Coolen J, van der Wielen PWJJ, Jaspers MC, Atsma A, de Graaf B, Schuren F**. 2015. Microbial biogeography of drinking water: patterns in phylogenetic diversity across space and time. Environ Microbiol **17:**2505–2514. http://dx.doi.org/10.1111/1462-2920.12739.
- 2. **Gomez-Alvarez V, Revetta RP, Santo Domingo JW**. 2012. Metagenomic analyses of drinking water receiving different disinfection treatments. Appl Environ Microbiol **78:**6095–6102. http://dx.doi.org/10.1128/ AEM.01018-12.
- 3. **Revetta RP, Gomez-Alvarez V, Gerke TL, Curioso C, Santo Domingo JW, Ashbolt NJ**. 2013. Establishment and early succession of bacterial communities in monochloramine-treated drinking water biofilms. FEMS Microbiol Ecol **86:**404 –414. http://dx.doi.org/10.1111/1574-6941.12170.
- 4. **Iroh Tam PY, Kline S, Ward G, Ferrieri P**. 2015. Non-tuberculous mycobacterial infection in hospitalized children: a case series. Epidemiol Infect **12:**1–9. http://dx.doi.org/10.1017/S0950268815000333.
- 5. **Simmon K, Brown-Elliott BA, Ridge PG, Durtschi JD, Mann LB, Slechta ES, Steigerwalt AG, Moser BD, Whitney AM, Brown JM, Voelkerding KV, McGowan KL, Reilly AF, Kirn TJ, Butler WR, Edelstein PH, Wallace RJ, Jr, Petti CA**. 2011. *Mycobacterium chelonaeabscessus* complex associated with sinopulmonary disease, northeastern USA. Emerg Infect Dis **17:**1692–1700. http://dx.doi.org/10.3201/ eid1709.101667.
- 6. **Hasan NA. Davidson RM, de Moura VCN, Garcia BJ, Reynolds PR, Epperson LE, Farias-Hesson E, DeGroote MA, Jackson M, Strong M**.

2015. Draft genome sequence of *Mycobacterium chelonae* type strain ATCC 35752. Genome Announc **3**(3):e00536-15.

- 7. **Ripoll F, Pasek S, Schenowitz C, Dossat C, Barbe V, Rottman M, Macheras E, Heym B, Herrmann J, Daffé M, Brosch R, Risler J, Gaillard J**. 2009. Non mycobacterial virulence genes in the genome of the emerging pathogen *Mycobacterium abscessus*. PLoS One **4:**e5660. http://dx.doi.org/ 10.1371/journal.pone.0005660.
- 8. **Greninger AL, Langelier C, Cunningham G, Keh C, Melgar M, Chiu CY, Miller S**. 2015. Two rapidly growing mycobacterial species isolated from a brain abscess: first whole-genome sequences of *Mycobacterium immunogenum* and *Mycobacterium llatzerense*. J Clin Microbiol **53:** 2374 –2377. http://dx.doi.org/10.1128/JCM.00402-15.
- 9. **Lourenço Nogueira CL, Simmon KE, Chimaera E, Cnockaert M, Carlos Palomino J, Martin A, Vandamme P, Brown-Elliott BA, Wallace RJ, Jr, Cardoso Leão S**. 2015. *Mycobacterium franklinii* sp. nov., a species closely related to members of the *Mycobacterium chelonae*-*Mycobacterium abscessus* group. Int J Syst Evol Microbiol **65:**2148 –2153. http://dx.doi.org/ 10.1099/ijs.0.000234.
- 10. **Nogueira CL, Whipps CM, Matsumoto CK, Chimaera E, Droz S, Tortoli E, de Freitas D, Cnockaert M, Palomino JC, Martin A, Vandamme P, Leão SC**. 2015. Description of *Mycobacterium saopaulense* sp. nov., a rapidly growing mycobacterium closely related with members of the *Mycobacterium chelonae*-*M*. *abscessus* group. Int J Syst Evol Microbiol http://dx.doi.org/10.1099/ijsem.0.000590.
- 11. **Van Ingen J, Boeree MJ, Dekhuijzen PNR, van Soolingen D**. 2009. Environmental sources of rapid growing nontuberculous mycobacteria causing disease in humans. Clin Microbiol Infect **15:**888 –893. http:// dx.doi.org/10.1111/j.1469-0691.2009.03013.x.
- 12. **Ashbolt NJ**. 2015. Microbial contamination of drinking water and human health from community water systems. Curr environ. Health Rep **2:**95–106.
- 13. **Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA**. 2012. SPAdes: a new genome assembly algorithm and its applications to singlecell sequencing. J Comput Biol **19:**455–477. http://dx.doi.org/10.1089/ cmb.2012.0021.
- 14. **Goris JA, Konstantinidis KT, Klappenbach JA, Coenye T, Vandamme P, Tiedje JM**. 2007. DNA-DNA hybridization values and their relationship to whole-genome sequence similarities. Int J Syst Evol Microbiol **57:**81–91. http://dx.doi.org/10.1099/ijs.0.64483-0.
- 15. **Richter M, Rosselló-Mora R**. 2009. Shifting the genomic gold standard for the prokaryotic species definition. Proc Natl Acad Sci USA **106:** 19126 –19131. http://dx.doi.org/10.1073/pnas.0906412106.
- 16. **Seemann T**. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics **30:**2068 –2069. http://dx.doi.org/10.1093/bioinformatics/ btu153.