Review Article

Label-free electrical detection of DNA hybridization using carbon nanotubes and graphene

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Abstract

The interface between biosystems and nanomaterials is emerging for detection of various biomolecules and subtle cellular activities. In particular, the development of costeffective and sequence-selective DNA detection is urgent for the diagnosis of genetic or pathogenic diseases. Graphenebased nanocarbon materials, such as carbon nanotubes and thin graphene layers, have been employed as biosensors because they are biocompatible, extraordinarily sensitive, and promising for large-area detection. Electrical and labelfree detection of DNA can be achieved by monitoring the conductance change of devices fabricated from these carbon materials. Here, the recent advances in this research area are briefly reviewed. The key issues and perspectives of future development are also discussed.

Keywords: carbon nanotubes; graphene; DNA hybridization; biosensors; label-free detection; transistors

he interface between biosystems and nanomaterials is emerging as one of the most diverse and dynamic areas of science and technology. The research of nano/bio interfaces, comprising the dynamic physical or chemical interactions, kinetics, and thermodynamic exchanges between nanomaterial surfaces and the surfaces of biological components such as proteins, membranes, phospholipids, endocytic vesicles, organelles, and DNA is rapidly growing. In particular, the development of sequence-selective DNA sensors for diagnosis of genetic or pathogenic diseases has attracted much attention. Many methods have been adopted to detect the DNA hybridization process including the detection relying on optical (1-10), piezoelectric (11-16), and electrochemical transductions (17-27). However, fluorescent or electrochemical tags are required for these detection methods. Alternative approaches based on the resistance change of semiconductor nanomaterials (e.g.



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and graphene-based sensors.

silicon nanowires and carbon nanotubes) have been demonstrated as a potential for label-free electrical detection (28-35). The nanomaterials of graphene (aromatic sp²)-based carbon have been widely used for DNA sensing because they are biocompatible and highly sensitive to environmental perturbations such as electronic doping (36-40) and molecular adsorption (41-46). In this article, we briefly review the recent developments of label-free bioelectronic sensors for detecting DNA hybridization using the field-effect transistors (FETs) based on single-walled carbon nanotubes (SWNTs) and graphenerelated materials. The detection mechanisms for various devices are also discussed in detail.

Field-effect transistors (FETs) for signal transduction

Graphene, a one-atom-thick planar sheet of aromatic sp² carbon crystal, holds great promise for replacing conventional Si semiconductors in applications, including high-speed computer chips and biochemical sensors. It has been recently demonstrated that its intrinsic carrier mobility ($\sim 200,000 \text{ cm}^2/\text{V-s}$) is higher than other known materials at room temperature (47, 48). The twodimensional (2-D) graphene is the basic structural element of some carbon allotropes including graphite (3-D), SWNTs (1-D), and fullerenes. Among these allotropes, SWNTs can be easily adopted for device fabrication due to having a high length-to-diameter ratio. The SWNTs can be considered as a cylindrical roll-up of the planar graphene sheet with a sp² bonding of carbon atoms. These cylindrical carbon molecules have novel properties that make them potentially useful in many applications in nanotechnology, electronics, optics, and other fields of materials science, as well as a potential use in architectural fields. Similar to graphene, they exhibit extraordinary strength, high thermal conductivity, unique electrical properties, and are biocompatible. The diameter size of SWNTs (~1 nm) is comparable to the size of DNA molecules, which is suitable for revealing the interactions between biomolecules and nanomaterials.

The electronic components, such as resistors and FETs based on 2-D graphene thin layers and 1-D SWNTs, have been successfully demonstrated using nano-/ microlithographic fabrication. It is noteworthy that the transport carriers in both 2-D graphene thin layers and 1-D SWNTs flow plentifully on their surfaces as their π electrons are delocalized on the surfaces. Consequently, their conductance is highly sensitive to the environmental perturbations occurring proximate to the carbon surfaces.

DNA molecules consist of various nucleobases, cytosine, guanine, adenine, and thymine, which can interact with graphene or SWNT surfaces. Theoretical calculations and experiments have suggested that an interaction between nucleobases and graphene (or SWNT) surface can be explained by the van der Waals interaction (π - π interaction) and solvation energy contributed by solvent molecules (49). The interaction between recognition DNA (probe-DNA) and carbon surfaces or the binding of analyte DNA with preconjugated probe-DNA may cause significant changes in the conductivity of devices through different mechanisms such as electrostatic gating (caused by the charges on DNA) (50-53), electronic doping (from DNA to carbon materials) (54), or modification of the junction between electrodes and carbon materials (55, 56). Taking advantage of the sensitive electrical responses from these devices, SWNTs or graphene-related materials configured

into FETs have been successfully utilized to electrically differentiate the DNA molecules with single-base specificity. The physical mechanism underlying the sensing varies significantly with the device configuration and operation modes. Fig. 1 shows one of these detection strategies reported by Star et al. (54), and this example illustrates how the label-free electrical detection works for biomolecular detection. The source-drain conductance of the device was recorded before and after incubation with 12-mer oligonucleotide capture probes (5'-CCT AAT AAC AAT-3'), as well as after incubation with the complementary DNA targets. In this experiment, each electrical measurement was performed in dry state, after the device was immersed in desired solutions followed by rinsing and drying. The conductance decreases with the addition of probe- or target-DNA. Typically the percentage decrease in conductance caused by the addition of target-DNA can be correlated to the concentration of the target-DNAs. Also, the addition of mismatched DNA does not result in significant conductance drop. Therefore, the differentiation between the target and mismatched DNAs can be achieved by such a detection strategy.

Single-walled carbon nanotubes (SWNTs)

Kong et al. (36) have first successfully demonstrated the electrical detection of gases and chemicals using the FET based on individual SWNTs. Several biosensors based on nanotube FETs have been used for the electronic detection of the following interactions: biotin–streptavidin (57), human immunoglobin (IgG) (58–62), various monoclonal antibodies (63–66), and pig serum albumin (67). The utilization of SWNT networks for DNA detection has been reported by Star et al. (54) and Gui et al. (56). Inspired by these works, Heller et al. (52) have recently discussed the



Fig. 1. Transfer characteristics, conductance (*G*) as function of gate voltage (V_g), and schematic drawings of the SWNT FETs used for DNA assays before and after incubation with 12-mer oligonucleotide capture probes (5'–CCT AAT AAC AAT – 3'), as well as after incubation with the complementary DNA targets. (Reproduced with permission from Ref. (54), copyright 2006 National Academy of Sciences of the United States of America.)



Fig. 2. (a) Measurement setup, where a source-drain bias potential is applied and the device is gated through a Ag/AgCl reference electrode inserted in the electrolyte. (b, c) Changes in liquid gate sweeps of ambipolar devices measured during protein adsorption experiments. (b) Example of strong electrostatic gating (adsorption of 185 nM poly-L-lysine on an ambipolar SWNT device). (c) Example of a strong Schottky barrier effect in the case of adsorption of 1 μ M horse heart cytochrome-*c* on a short (40 nm) SWNT device. (Reproduced with permission from Ref. (52), copyright 2008 American Chemical Society).

possible sensing mechanisms for the individual SWNT device operated by liquid-gating as schematically illustrated in Fig. 2a, where a source-drain bias potential is applied and the device is gated through a Ag/AgCl reference electrode inserted in the electrolyte. Their study concludes that the electrostatic gating and changes of the Schottky barrier between electrodes and SWNTs are the two competitive detection mechanisms. Fig. 2b and c show two typical changes in liquid gate sweeps of ambipolar devices measured during protein adsorption experiments. Fig. 2b is an example of a strong electrostatic gating (adsorption of 185 nM poly-L-lysine on an ambipolar SWNT device) and Fig. 2c displays the result of a strong Schottky barrier effect in the case of adsorption of 1 μ M horse heart cytochrome-c on a short (40 nm) SWNT device. It is noted that these sensors, however, involve a high production cost because

nanolithographic facilities are required. Another concern is that these transistors may have significant deviceto-device variation due to the difficulty in obtaining desired single-chirality of SWNT species for device fabrication.

In contrast, SWNT network-based transistors are practically simpler in fabrication and involve lower production cost. Their electrical properties depend on the SWNT percolative path in the conduction channel and thus the device-to-device variations are expected to be small. Label-free electrical detection of DNA hybridization and antibody–antigen binding using FETs fabricated from SWNT networks have been successfully achieved recently (54). We have verified that the sensing of DNA hybridization for the devices with an apparent on/off ratio, the ratio of an on current and off current measured during the gate voltage sweeping, is dominated



Fig. 3. Transfer curves (drain current I_d vs gate voltage V_g) for photoresist capped Au-contacted SWNT FETs with (a) one junction exposed, (b) a channel exposed before immobilization, after immobilization, and upon hybridization with its complementary DNA, where the drain voltage was fixed at -0.5V. The inset for each graph shows the photoresist pattern as imaged in an optical microscope. (Reproduced with permission from Ref. (56), copyright 2007 American Chemical Society).

by the change in electrode-SWNT junctions rather than the conductance change of channel SWNTs (56). Fig. 3 compares the sensing performance for the SWNT FETs devices with (a) one junction exposed, and (b) only channel exposed before immobilization, after immobilization, and upon hybridization with its complementary DNA, where the unexposed area was covered with photoresist. The inset for each graph shows the photoresist pattern as imaged in an optical microscope. The results verify that the SWNT-electrode junction is more sensitive than SWNTs themselves. This can be reasoned that this FET is governed by the electrode-SWNT network junction when the contact resistance is nontrivial, where the contact barrier shall be sensitive to the charges brought in by DNA molecules. Separately, if a metallic tube network is used, the electrode-SWNT contact resistance becomes insignificantly small and therefore the DNA detection will be governed by the electronic doping (from DNA molecules to SWNTs) occurring at the channel area (68).

The reported detection limit of distinguishing complementary and one-base mismatched DNA for the FETs based on networks is on the order of ca. 1 nM of DNA. It has been demonstrated that the sensitivity can be improved by using a threading intercalacter attached with a redox-active functional group, where the intercalacter selectively inserts into hybridized DNA strands and the redox functional groups enhance the changes in device conductance (69). Another more efficient method is to bring more electrostatic charges to the proximity of electrode-SWNT contacts through introducing reporter-DNA-Au nanoparticle conjugates (70) as schematically illustrated in Fig. 4a. Each target DNA can bind to a Au-nanoparticle that has been previously attached with many reporter DNA molecules and consequently the change of conductance upon each hybridization event can be greatly magnified. Fig. 4b clearly demonstrates that the sensitivity of reporter DNA-Au nanoparticle assisted detection is significantly higher than those without the cooperation of reporter DNA. Fig. 4c shows that the sensitivity of this approach allows us to differentiate the complementary and one-base mismatched DNA with the sensitivity as good as ~ 100 fM of DNA.

An alternative method, by monitoring the shift of threshold voltage during the gate voltage sweep, reflecting on the increased density of charges trapped around SWNTs upon DNA addition or hybridization (71–74), has also been adopted to perform label-free detection of DNA hybridization (75). Due to that, the change in threshold voltage involves the trapping of moisture and strongly depends on the substrate surface qualities, the



Fig. 4. (a) Schematic illustration of DNA detection enhancement by reporter DNA–Au nanoparticle conjugates. The bottom right panel illustrates the possible molecular binding on SWNTs. (b) Statistics showing the percentage decrease of I_d in SWNT FETs for various sensing experiments. (c) Percentage I_d decrease versus DNA concentration in the sensing of complementary DNA, enhanced by 6A and 11A reporter DNA–AuNP conjugates. The dashed line shows the limit of selective detection, which is based on the I_d response to the mismatched DNA (1 nM) + 11A reporter DNA–AuNP conjugates. (Reproduced with permission from Ref. 70, copyright 2008 Wiley-VCH).

detection reliability, and sensitivity for this approach still require more investigations.

Graphene and related materials

Graphene and graphene derivatives, such as graphene oxide (GO), reduced graphene oxide (rGO) and n-doped graphene, are also potentially useful for the biosensing application. The GO is an oxidized form of graphene produced in solution and has negative charges when dispersed in water (76). It is strongly hydrophilic and has a brown/dark-brown color in aqueous solution. The GO can be reduced by chemical methods (77-81), thermal methods (82, 83), and ultraviolet-assisted methods (84) to form rGO. The conductivity can be increased by up to 4 orders of magnitude by reduction of GO to rGO (85-87). However, the conductivity of rGO is still less than that of the pristine graphene by a factor of 10-100 (88, 89). The rGO has been found to contain a considerable amount of topological defects (90) and this is probably one of the main reasons for the limited conductivity. Graphene could also be n-doped by introducing n-dopants (40, 43). Wang's group (91) has also recently reported that graphene nanoribbons could be covalently functionalized by nitrogen species on ribbon edges through highpower electrical joule heating in ammonia gas. All these approaches open up the routes for tailoring electrical properties of graphene sheets for various applications.

For the DNA hybridization study, GO has been used as a template to host a fluorescence quencher for the detection of DNA hybridization based on the fluorescence from the fluorophore-conjugated probe DNAs (92, 93). Electrical detection of DNA hybridization by



Fig. 5. Single-stranded (ss)–DNA tethering on GO (G-DNA) increases the conductivity of the device. Successive hybridization and dehybridization of DNA on the G-DNA device results in completely reversible increase and restoration of conductivity. Inset shows a fluorescent image for the GO device hybridized G-DNA (ds) sheet with wrinkles and folds clearly visible. (Reproduced with permission from Ref. 78, copyright 2008 American Chemical Society).

monitoring the conductance change of GO sheets has also been reported (94) (shown in Fig. 5), where the authors observe that the single-stranded (ss)-DNA tethering on the GO device (termed as G-DNA) increases its conductivity. Successive hybridization and dehybridization of DNA on the G-DNA device result in a completely reversible increase and restoration of conductivity. The conductance increase (restoration) was explained by the increased (decreased) electrostatic gating induced by the negative charges from DNA molecules. The electrical properties of GO films largely vary with the fabrication process of GO (oxidation processes of graphite); hence, the sensitivity of the devices produced from GO sheets shall strongly depend on the size and the shape of GO, the presence of wrinkles on a GO surface, the degree of oxidation in GO, and the defect density of the GO sheet. Therefore, to use GO as the sensing component, more efforts are needed to control the quality of GO sheets.

In contrast to GO, large-size graphene films are considered more reliable and achievable from the device fabrication viewpoint. Developing graphene-based biosensors becomes practical with the recent advances of chemical vapor deposition (CVD) of large-sized graphene film (up to wafer size) (95-97). We have fabricated largesized graphene transistors by transferring the as-grown CVD graphene films from Ni (or Cu) to glass substrates. These films can be configured as liquid-gated FETs (Fig. 6a) and they are able to detect the DNA hybridization with a detection sensitivity of 0.01 nM and the capability to distinguish single-base mismatch (98). As shown in Fig. 6b and c, the conductance of graphene devices exhibited amipolar behaviors subjecting to the gate voltage applied to the bath solution. The $V_{\rm g, min}$ that gives the minimum graphene conductance can be identified from the transfer curve and be used to monitor the Fermi energy state of the graphene film. Fig. 6b shows that the $V_{g,min}$ is sensitive to the immobilization of probe DNAs and hybridization of the complementary target DNAs. Specifically, $V_{g,min}$ is significantly left-shifted with the addition of DNA molecules, suggesting that the electrostatic gating from the DNA is not dominating and that DNA molecules *n*-dopes the graphene film. The interaction between DNA molecules and graphitic structure has been studied theoretically and experimentally (92, 93, 99-101). It has been demonstrated that the binding between graphene and nucleotides is dominated by the non-electrostatic interaction (99, 100, 102–104).

Meanwhile, the left-shift of $V_{g,min}$ after DNA hybridization suggests that the complementary DNAs can also effectively interact with graphene and impose the *n*-doping effect based on the graphene–nucleotide interaction. The shift in $V_{g,min}$ increased with the increasing concentration of the complementary DNA, specifically, 0.01 nM and 10 nM DNA solutions caused the >10 meV



Fig. 6. (a) Schematic illustration of the graphene device operated by liquid gating. The middle is an optical microscopy image of the graphene films. The bottom shows the DNA sequences used in the experiments. (b, c) Transfer characteristics for the graphene transistors before adding DNA, after immobilization with probe DNA, and after reaction with (b) complementary or (c) one-base mismatched DNA molecules with the concentration ranging from 0.01 to 500 nM. (Reproduced with permission from Ref 82, copyright 2010 Wiley-VCH).

and >55 meV shift, respectively. The $V_{g,min}$ did not shift further at the higher complementary DNA concentrations (>10 nM) likely due to saturation in hybridization with the limited number of probe DNAs. Furthermore, Fig. 6 demonstrates that the complementary DNA and the one-base mismatched DNA can be easily differentiated because $V_{g,min}$ is much less sensitive to the mismatched DNA, which only caused a ~ 20 meV shift at a high concentration (500 nM). The electrostatic gating effect by adsorbed charge species has been adopted to explain the shift of the $I_{\rm d}$ - $V_{\rm g}$ curve of carbon nanotube transistors (52, 53). If the same mechanism applies to our graphene devices, a positive shift of $V_{g,min}$ would be expected because the highly negative charges of the adsorbed DNAs can only be balanced by positive gate voltage. Thus, electrostatic gating is not dominating the electrical characteristics in our testing conditions. It is noted that the electrostatic gating by electrolytes may sometimes become a competing mechanism when the electrolyte concentration is too high or when the gate voltage scanning range is too large.

Future of carbon-based electrical detection of DNA hybridization

Label-free electrical detection of DNA hybridization using carbon-based materials has been carried out for years. High sensitivity and high specificity have been achieved. However, there are still numerous challenges for detecting low concentration of DNA analytes (< femto mole). Along with the decrease of DNA concentration, improvement of signal-to-noise ratio of devices shall become an important issue. More efforts are required to optimize operation conditions of carbon-based devices. It is also attractive to seek the device that allows the detection of a single hybridization event. In practical cases, detection of a longer DNA chain will also be necessary. The electrical detection of DNA hybridization largely depends on the device structure and the forms of these carbon materials. For example, an increase of semito-metallic ratio in the SWNT ensemble, controlling chirality of carbon nanotubes, and proper modification of graphene or graphene nanoribbon could potentially enhance the DNA detection sensitivity. The rapid, specific, and low-cost electrical detection of DNA hybridization could speed up the realization of the next generation smart homecare sensor system.

Conclusions

In this review, we have discussed the recent advances and key issues for the development of label-free detection of DNA hybridization using carbon nanotubes and graphene. Sensing mechanisms are also discussed for each type of device. The detection of a single-base polymorphorism or mutation is thought to be the key for diagnosis of genetic diseases and realization of personalized medicine. Carbon nanotube and graphenebased devices have shown great potential for the future application of DNA biosensors in terms of the high sensitivity and selectivity. From the fundamental viewpoint, more research efforts are needed to understand the interface (contact) effects, including nanotube–nanotube and graphene–electrode contacts, on sensing behaviors. In addition, intensive studies to bridge the knowledge to clinical detection of biomarkers are emergent.

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Conflict of interest and funding

There is no conflict of interest in the present study for any of the authors.

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