

Harnessing autophagy to fight SARS-CoV-2: An update in view of recent drug development efforts

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Abstract

Drug repurposing is an attractive option for identifying new treatment strategies, in particular in extraordinary situations of urgent need such as the current coronavirus disease 2019 (Covid-19) pandemic. Recently, the World Health Organization announced testing of three drugs as potential Covid-19 therapeutics that are known for their dampening effect on the immune system. Thus, the underlying concept of selecting these drugs is to temper the potentially life-threatening overshooting of the immune system reacting to severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) infection. This viewpoint discusses the possibility that the impact of these and other drugs on autophagy contributes to their therapeutic effect by hampering the SARS-CoV-2 life cycle.

KEYWORDS

autophagy, Covid-19, drug repurposing, pharmacology, SARS-CoV2, virophagy

1 | INTRODUCTION

Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) has gained notoriety for causing the currently raging coronavirus disease 2019 (Covid-19) pandemic.¹ Humongous efforts are ongoing worldwide to cope with the impact on health and society. Although vaccines could be developed and marketed with unprecedented swiftness, drug development will take significantly longer. In light of the obvious exigency, drug repurposing is a promising strategy that is being followed by many scientists in preclinical and clinical research.² For example, the World Health Organization (WHO) launched the research program “Solidarity” in 2020 to test four compounds as options for antiviral treatment, namely remdesivir (originally developed as an inhibitor

of viral RNA polymerase to treat hepatitis C, Ebola, or Marburg virus infection), interferon β 1a (boosting the host response to viral infection), hydroxychloroquine (a malaria drug), and a combination of lopinavir and ritonavir (both HIV drugs). Unfortunately, an interim report of the study, including 11 330 in-patients with Covid-19 at 405 hospitals in 30 countries, revealed little or no effect.³

A more recent initiative in the Solidarity program evaluates three established immune-modulatory drugs for Covid-19 treatment.⁴ The selection of these drugs was based on a different rationale, that is, instead of trying to fight the virus directly, the aim is to confine the damage of an exaggerated immune response to the own body. A previous study showed that limiting the host defense can have beneficial effects in critically ill patients with Covid-19.⁵ The selected drugs are

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infiximab, imatinib, and artesunate. The aim of this short review is to point to a potential involvement of autophagy in the action of these drugs, which may play a more prominent role than generally acknowledged. The review additionally includes the drug ivermectin, which received media attention as its promising results were reported in clinical trials,⁶ and also covers antidepressants.

2 | AUTOPHAGY

In general, autophagy is an evolutionary conserved intracellular degradation process pivotal for cellular protein, energy, and organelle homeostasis.⁷ It is active under the basic condition at a low level ensuring continuous turnover and can be activated under certain stress conditions such as proteotoxicity or starvation.⁸ Material destined for degradation or recycling is engulfed by or transported into a double-membrane structure called “autophagosome.” Through additional membrane remodeling processes, these autophagosomes fuse with lysosomes producing autolysosomes, with prior fusion with late endosomes as a potential intermediate step.⁹ As detailed in excellent reviews, this process is tightly controlled and executed by a vast array of proteins, ATG proteins in particular, but also EPG proteins required for the more complex autophagy in multicellular organisms.^{7–11} Autophagic flux refers to the activity through all consecutive steps of autophagy and typically is defined as a measure of autophagic degradation activity.¹² Analytical tools assessing autophagic flux need to be chosen with great care to avoid erroneous conclusions.¹³ Several compounds currently are being developed, targeting different proteins in the autophagic cascade, given its involvement in various physiological and pathophysiological conditions, including viral infection.^{14,15}

3 | AUTOPHAGY AND CORONAVIRUSES

The link between autophagy and invading pathogens is anything but new and both pro- and antiviral roles of autophagy were identified.¹⁶ For example, evidence suggests that double-membrane structures derived from the endoplasmic reticulum both are required for the initial steps of autophagy and serve as replication sites for coronaviruses^{17–22} (see also Figure 1). Later on, several coronavirus proteins were shown to induce the formation of double-membrane structures, such as the nonstructural proteins 2, 3, 4, and 6.²³ The broad activity against coronavirus replication of compounds that interfere with the generation of these structures further corroborates their importance.

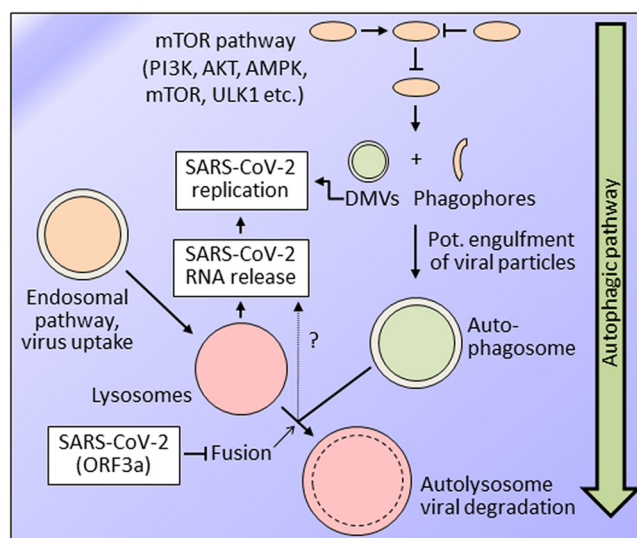


FIGURE 1 Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) replication and endosomal/autophagic pathways, simplified scheme. Current knowledge supports both beneficial and detrimental effects of the autophagic pathway for SARS-CoV-2 replication. A major entry route for the virus is endocytic uptake, which requires lysosomal acidification for viral RNA release. The autophagic pathway is a multifactorial and multistep pathway with a vast range of possibilities for pharmacological targeting. In the more initial phases, phagophores, and double-membrane vesicles (DMVs) are formed, most likely from the endoplasmic reticulum, possibly also promoted by some coronavirus proteins. SARS-CoV-2 replication takes place at the endoplasmic reticulum as well, at very similar, if not identical, membrane structures. SARS-CoV-2 inhibits the last step of autophagy leading to viral degradation, that is, the fusion of autophagosomes with lysosomes to form autolysosomes, thus inhibiting autophagic flux. Accordingly, compounds impacting autophagy are expected to be efficient in fighting SARS-CoV-2 only if they enhance autophagic flux

In contrast to this appearing congruence of viral mechanisms and early steps of autophagy, there is also firm evidence that coronaviruses interfere with late steps of autophagy to evade degradation. Very recently, for example, ORF3a (the protein derived from open reading frame 3a) of SARS-CoV-2 has been demonstrated to inhibit the fusion of autophagosomes with lysosomes, thereby increasing the number of autophagosomes but decreasing autophagic flux,^{24,25} in line with the effect of coronavirus infection.^{26,27} However, the exact details of how coronaviruses in general, and SARS-CoV-2 in particular, are intertwined with autophagy await further elucidation.^{23,28,29} Nevertheless, it appears plausible that coronaviruses may benefit from earlier steps of the autophagic pathway, but are vulnerable to the increased autophagic flux that clears out viral particles. This is supported by reports showing that induction of autophagy has the potential to fight coronavirus infection.^{26,27}

Chloroquine inhibits autophagy by interfering with autophagosome-lysosome fusion.³⁰ However, chloroquine and hydroxychloroquine exert additional effects like disorganizing the endo-lysosomal system that might have been the basis for the initial hope put on this drug for Covid-19 treatment.^{31,32} However, with more studies coming up, no overall beneficial effect of this drug on Covid-19 was apparent,^{33–35} and the drug now is abandoned in the WHO Solidarity program. Therefore, it appears likely that autophagy-targeting drugs need to promote autophagy flux rather than other aspects of autophagy. Although COVID-19 primarily is a respiratory disease, multiple organs are affected, either through cytokines or directly upon invasion of SARS-CoV-2.^{36–38} As autophagy is a conserved mechanism operative in most cells, pharmacological induction of autophagy has the potential to fight SARS-CoV-2 in all organs that are reached by the compound. However, the effect on overall health may depend on existing comorbidities, such as cancer, for example, where the effect of autophagy depends on the circumstances.^{39,40}

4 | ARTESUNATE

Like the other two drugs infliximab and imatinib, artesunate was added to the WHO Solidarity program because of its effects on the immune system.⁴ Artesunate is a derivate of artemisinin with established antimalaria features, but also potent anticancer effects. For considering its potential effects on autophagy, it is important to differentiate general effects on autophagy from effects on autophagy flux, given the complex interaction of SARS-CoV-2 with autophagy. The vast majority of publications assessing artesunate for its effects on autophagy report induction^{41–44} not all publications, however, assess autophagic flux following the established guidelines.¹³ Nevertheless, some flux assays have been performed such as the use of the late autophagy blockers chloroquine or bafilomycin A, where artesunate still enhances the autophagy marker LC3BII/I.^{45–47} Although all these studies support autophagy promoting function of artesunate, an inhibitory effect of artesunate has been observed using the tandem fluorescence tagged LC3B stably transfected into HeLa cells,⁴⁸ which is recognized as a valid method to determine autophagic flux.¹³ The reason for this seeming discrepancy is not known, which makes further studies mandatory.

5 | INFlixIMAB

Infliximab is a chimeric antibody targeting TNF- α used in clinical practice to treat autoimmune diseases such as Crohn's disease. Recently, it has been put forward that a

range of drugs, including infliximab, that are either approved or in a clinical trial with great promise to treat Crohn's disease induces autophagy as a relevant mechanism at least contributing to their effect.^{49,50} At least for infliximab, however, there is a scarcity of studies investigating the effect on autophagy directly,^{51,52} and no reports were found presenting autophagic flux assays for infliximab.

6 | IMATINIB

Imatinib is an ABL tyrosine kinase inhibitor used to treat chronic myeloid leukemia. Like chloroquine, it is a cationic amphiphilic drug and thus should have the potential to inhibit autophagy by accumulating in lysosomes and disturbing their function.⁵³ However, several studies report an autophagy-inducing effect of imatinib, including the assessment of autophagic flux using chloroquine as an inhibitor.^{54–57} Nevertheless, more studies are needed applying a broader range of autophagic flux assessments to solidify the conclusion that imatinib induces autophagy. Furthermore, it has been argued that compounds prone to induce phospholipidosis such as cationic amphiphilic drugs should be excluded from drug repurposing for SARS-CoV-2 treatment.⁵⁸ Nevertheless, it should be noted, that several other cationic amphiphilic drugs such as some antidepressants also induce autophagic flux.⁵⁹ Thus, this compound class may as well elicit more specific effects.

7 | ANTIDEPRESSANTS

Evidence is accumulating that patients with Covid-19 benefit from antidepressant treatment: A multicentric observational retrospective study with 7230 adults hospitalized for Covid-19 reported that those receiving antidepressant treatment had a reduced risk of intubation or death.⁶⁰ Similarly, a study with 3238 Covid-19 patients revealed a beneficial effect of the antidepressant fluvoxamine, reducing the need for emergency room observation or hospitalization.⁶¹ A small randomized clinical trial with 152 COVID-19 outpatients revealed a lower likelihood of clinical deterioration for patients receiving fluvoxamine.⁶² Furthermore, a recent preclinical study found the antidepressant fluoxetine as an inhibitor of SARS-CoV-2 in human lung tissue.⁶³ The beneficial effects of antidepressants frequently are conceptualized as cytokine effects⁶⁴ thus reducing the risk of a fatal cytokine storm.⁶⁵ However, antidepressants are known to induce autophagy as well.^{59,66} Thus, their effect on autophagy might not only be important for treating

depression but also to fight SARS-CoV-2. In fact, tricyclic antidepressants inhibit lysosomal acidic sphingomyelinase, thereby not only enhancing autophagy but also reducing SARS-CoV-2 entry into epithelial cells.^{67,68}

8 | IVERMECTIN

Ivermectin is an antihelmintic macrolide of the avermectin group.⁶⁹ It is investigated as a potential anti-SARS-CoV-2 treatment with promising initial results, but also very recent dispute.^{6,71} Several mechanisms are discussed for its apparent antiviral activity^{70,71} and this viewpoint argues for adding autophagy to this panel. A number of publications report an autophagy-inducing effect of ivermectin,^{72–74} including a study carefully determining autophagic flux.⁷⁵ Therefore, autophagy should be considered as a mediator of the manifold effects of ivermectin in general,⁷⁰ and of its antiviral activity in particular. Of note, another antihelmintic drug, niclosamide, not only is known for its autophagy-inducing action but also has been demonstrated to reduce replication of the Middle East Respiratory Syndrome Coronavirus²⁶ as well as of SARS-CoV-2.²⁷

9 | CONCLUSION

The point of this article is to draw attention to autophagy as a potential contributing mechanism of selected drugs currently under investigation for repurposing to Covid-19 treatment. In other words, it is possible that the three drugs recently added to the WHO Solidarity program may not just prevent a life-threatening overreaction of the body during a SARS-CoV-2 infection, but also actually limit SARS-CoV-2 replication through activating autophagy. The interaction of SARS-CoV-2 with the autophagic pathway is complex (Figure 1), with evidence for both the virus taking advantage of the autophagic pathway and trying to tame the full activity of this pathway to prevent its degradation. It is obvious from this scenario that it will be essential to learn how exactly the SARS-CoV-2 life cycle is intertwined with the autophagic pathway. Future research should include all known forms of autophagy, such as macroautophagy, microautophagy, chaperone-mediated autophagy, secretory autophagy, and so forth. This also applies to better understanding the action of to be repositioned or new drugs at each level of the autophagic pathway.

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REFERENCES

- Zhou P, Yang XL, Wang XG, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature*. 2020;579(7798):270–273. doi:10.1038/s41586-020-2012-7
- Alam S, Kamal TB, Sarker MMR, Zhou JR, Rahman SMA, Mohamed IN. Therapeutic effectiveness and safety of repurposing drugs for the treatment of COVID-19: position standing in 2021. *Front Pharmacol*. 2021;12:659577. doi:10.3389/fphar.2021.659577
- WHO Solidarity Trial Consortium, Pan H, Peto R, et al. Repurposed antiviral drugs for Covid-19—Interim WHO Solidarity Trial Results. *N Engl J Med*. 2021;384(6):497–511. doi:10.1056/NEJMoa2023184
- Ledford H. International COVID-19 trial to restart with focus on immune responses Published online May 7, 2021. *Nature*. doi:10.1038/d41586-021-01090-z
- Investigators R-C, Gordon AC, Mouncey PR, et al. Interleukin-6 receptor antagonists in critically ill patients with Covid-19. *N Engl J Med*. 2021;384(16):1491–1502. doi:10.1056/NEJMoa2100433
- Zein A, Sulistiyana CS, Raffaelo WM, Pranata R. Ivermectin and mortality in patients with COVID-19: a systematic review, meta-analysis, and meta-regression of randomized controlled trials. *Diabetes Metab Syndr*. 2021;15(4):102186. doi:10.1016/j.dsx.2021.102186
- Nakatogawa H. Mechanisms governing autophagosome biogenesis. *Nat Rev Mol Cell Biol*. 2020;21(8):439–458. doi:10.1038/s41580-020-0241-0
- Dikic I, Elazar Z. Mechanism and medical implications of mammalian autophagy. *Nat Rev Mol Cell Biol*. 2018;19(6):349–364. doi:10.1038/s41580-018-0003-4
- Zhao YG, Codogno P, Zhang H. Machinery, regulation and pathophysiological implications of autophagosome maturation. *Nat Rev Mol Cell Biol*. 2021. doi:10.1038/s41580-021-00392-4
- Galluzzi L, Baehrecke EH, Ballabio A, et al. Molecular definitions of autophagy and related processes. *EMBO J*. 2017;36(13):1811–1836. doi:10.15252/embj.201796697
- Mizushima N, Yoshimori T, Ohsumi Y. The role of Atg proteins in autophagosome formation. *Annu Rev Cell Dev Biol*. 2011;27:107–132. doi:10.1146/annurev-cellbio-092910-154005
- Loos B, du Toit A, Hofmeyr JH. Defining and measuring autophagosome flux-concept and reality. *Autophagy*. 2014;10(11):2087–2096. doi:10.4161/15548627.2014.973338
- Klionsky DJ, Abdel-Aziz AK, Abdelfatah S, et al. Guidelines for the use and interpretation of assays for monitoring autophagy (4th edition)(1). *Autophagy*. 2021;17(1):1–382. doi:10.1080/15548627.2020.1797280
- Levine B, Kroemer G. Biological functions of autophagy genes: a disease perspective. *Cell*. 2019;176(1–2):11–42. doi:10.1016/j.cell.2018.09.048
- Mizushima N, Levine B. Autophagy in human diseases. *N Engl J Med*. 2020;383(16):1564–1576. doi:10.1056/NEJMra2022774
- Dong X, Levine B. Autophagy and viruses: adversaries or allies? *J Innate Immun*. 2013;5(5):480–493. doi:10.1159/000346388
- Knoops K, Kikkert M, Worm SH, et al. SARS-coronavirus replication is supported by a reticulovesicular network of

- modified endoplasmic reticulum. *PLoS Biol.* 2008;6(9):e226. doi:10.1371/journal.pbio.0060226
18. Axe, EL, Walker SA, Manifava M, et al. Autophagosome formation from membrane compartments enriched in phosphatidylinositol 3-phosphate and dynamically connected to the endoplasmic reticulum. *J Cell Biol.* 2008;182(4):685-701. doi:10.1083/jcb.200803137PM:18725538
 19. Prentice E, Jerome WG, Yoshimori T, Mizushima N, Denison MR. Coronavirus replication complex formation utilizes components of cellular autophagy. *J Biol Chem.* 2004;279(11):10136-10141. doi:10.1074/jbc.M306124200
 20. Reggiori F, Monastyrska I, Verheije MH, et al. Coronaviruses Hijack the LC3-I-positive EDEMosomes, ER-derived vesicles exporting short-lived ERAD regulators, for replication. *Cell Host Microbe.* 2010;7(6):500-508. doi:10.1016/j.chom.2010.05.013
 21. Ulasli M, Verheije MH, de Haan CA, Reggiori F. Qualitative and quantitative ultrastructural analysis of the membrane rearrangements induced by coronavirus. *Cell Microbiol.* 2010;12(6):844-861. doi:10.1111/j.1462-5822.2010.01437.x
 22. Cottam EM, Maier HJ, Manifava M, et al. Coronavirus nsp6 proteins generate autophagosomes from the endoplasmic reticulum via an omegasome intermediate. *Autophagy.* 2011;7(11):1335-1347. doi:10.4161/autophagy.7.11.16642
 23. Miller K, McGrath ME, Hu Z, et al. Coronavirus interactions with the cellular autophagy machinery. *Autophagy.* 2020;16(12):2131-2139. doi:10.1080/15548627.2020.1817280
 24. Miao G, Zhao H, Li Y, et al. ORF3a of the COVID-19 virus SARS-CoV-2 blocks HOPS complex-mediated assembly of the SNARE complex required for autolysosome formation. *Dev Cell.* 2021;56(4):427-442. doi:10.1016/j.devcel.2020.12.010
 25. Zhang Y, Sun H, Pei R, et al. The SARS-CoV-2 protein ORF3a inhibits fusion of autophagosomes with lysosomes. *Cell Discov.* 2021;7(1):31. doi:10.1038/s41421-021-00268-z
 26. Gassen NC, Niemeyer D, Muth D, et al. SKP2 attenuates autophagy through Beclin1-ubiquitination and its inhibition reduces MERS-Coronavirus infection. *Nat Commun.* 2019;10(1):5770. doi:10.1038/s41467-019-13659-4
 27. Gassen NC, Papies J, Bajaj T, et al. SARS-CoV-2-mediated dysregulation of metabolism and autophagy uncovers host-targeting antivirals. *Nat Commun.* 2021;12(1):3818. doi:10.1038/s41467-021-24007-w
 28. Delorme-Axford E, Klionsky DJ. Highlights in the fight against COVID-19: does autophagy play a role in SARS-CoV-2 infection? *Autophagy.* 2020;16(12):2123-2127. doi:10.1080/15548627.2020.1844940
 29. Garcia-Perez BE, Gonzalez-Rojas JA, Salazar MI, Torres-Torres C, Castrejon-Jimenez NS. Taming the autophagy as a strategy for treating COVID-19. *Cells.* 2020;9(12):2679. doi:10.3390/cells9122679
 30. Mauthe M, Orhon I, Rocchi C, et al. Chloroquine inhibits autophagic flux by decreasing autophagosome-lysosome fusion. *Autophagy.* 2018;14(8):1435-1455. doi:10.1080/15548627.2018.1474314
 31. Gao J, Tian Z, Yang X. Breakthrough: Chloroquine phosphate has shown apparent efficacy in treatment of COVID-19 associated pneumonia in clinical studies. *Biosci Trends.* 2020;14(1):72-73. doi:10.5582/bst.2020.01047
 32. Wang M, Cao R, Zhang L, et al. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res.* 2020;30(3):269-271. doi:10.1038/s41422-020-0282-0
 33. Group RC, Horby P, Mafham M, et al. Effect of hydroxychloroquine in hospitalized patients with Covid-19. *N Engl J Med.* 2020;383(21):2030-2040. doi:10.1056/NEJMoa2022926
 34. Boulware DR, Pullen MF, Bangdiwala AS, et al. A randomized trial of hydroxychloroquine as postexposure prophylaxis for Covid-19. *N Engl J Med.* 2020;383(6):517-525. doi:10.1056/NEJMoa2016638
 35. Singh AK, Singh A, Singh R, Misra A. Hydroxychloroquine in patients with COVID-19: a systematic review and meta-analysis. *Diabetes Metab Syndr.* 2020;14(4):589-596. doi:10.1016/j.dsx.2020.05.017
 36. Liu J, Li Y, Liu Q, et al. SARS-CoV-2 cell tropism and multi-organ infection. *Cell Discov.* 2021;7(1):17. doi:10.1038/s41421-021-00249-2
 37. Song E, Zhang C, Israelow B, et al. Neuroinvasion of SARS-CoV-2 in human and mouse brain. *J Exp Med.* 2021;218(3):e20202135. doi:10.1084/jem.20202135
 38. Wang X-M, Mannan R, Xiao L, et al. Characterization of SARS-CoV-2 and host entry factors distribution in a COVID-19 autopsy series. *Commun Med.* 2021;1(1):24. doi:10.1038/s43856-021-00025-z
 39. Alvarez-Meythaler JG, Garcia-Mayea Y, Mir C, Kondoh H, LLeonart ME. Autophagy Takes center stage as a possible cancer hallmark. *Front Oncol.* 2020;10(2178):586069. doi:10.3389/fonc.2020.586069
 40. Sun K, Deng W, Zhang S, et al. Paradoxical roles of autophagy in different stages of tumorigenesis: protector for normal or cancer cells. *Cell Biosci.* 2013;3(1):35. doi:10.1186/2045-3701-3-35
 41. Zhang J, Zhou L, Xiang JD, Jin CS, Li MQ, He YY. Artesunate-induced ATG5-related autophagy enhances the cytotoxicity of NK92 cells on endometrial cancer cells via interactions between CD155 and CD226/TIGIT. *Int Immunopharmacol.* 2021;97:107705. doi:10.1016/j.intimp.2021.107705
 42. Li L, Chen J, Zhou Y, Zhang J, Chen L. Artesunate alleviates diabetic retinopathy by activating autophagy via the regulation of AMPK/SIRT1 pathway. Published online Mar 4, 2021. *Arch Physiol Biochem.* doi:10.1080/13813455.2021.1887266
 43. Mancuso RI, Foglio MA, Olalla Saad ST. Artemisinin-type drugs for the treatment of hematological malignancies. *Cancer Chemother Pharmacol.* 2021;87(1):1-22. doi:10.1007/s00280-020-04170-5
 44. Sun X, Yan P, Zou C, et al. Targeting autophagy enhances the anticancer effect of artemisinin and its derivatives. *Med Res Rev.* 2019;39(6):2172-2193. doi:10.1002/med.21580
 45. Kong Z, Liu R, Cheng Y. Artesunate alleviates liver fibrosis by regulating ferroptosis signaling pathway. *Biomed Pharmacother.* 2019;109:2043-2053. doi:10.1016/j.biopha.2018.11.030
 46. Lee YS, Kalimuthu K, Seok Park Y, et al. Ferroptotic agent-induced endoplasmic reticulum stress response plays a pivotal role in the autophagic process outcome. *J Cell Physiol.* 2020;235(10):6767-6778. doi:10.1002/jcp.29571
 47. Zhao F, Vakhrusheva O, Markowitsch SD, et al. Artesunate impairs growth in cisplatin-resistant bladder cancer cells by cell cycle arrest, apoptosis and autophagy induction. *Cells.* 2020;9(12):2643. doi:10.3390/cells9122643

48. Button RW, Lin F, Ercolano E, et al. Artesunate induces necrotic cell death in schwannoma cells. *Cell Death Dis.* 2014;5:1466. doi:10.1038/cddis.2014.434
49. Azzman N. Crohn's disease: potential drugs for modulation of autophagy. *Medicina (Kaunas).* 2019;55(6):224. doi:10.3390/medicina55060224
50. Nys K, Agostinis P, Vermeire S. Autophagy: a new target or an old strategy for the treatment of Crohn's disease? *Nat Rev Gastroenterol Hepatol.* 2013;10(7):395-401. doi:10.1038/nrgastro.2013.66
51. Levin AD, Koelink PJ, Bloemendaal FM, et al. Autophagy contributes to the induction of anti-TNF induced macrophages. *J Crohns Colitis.* 2016;10(3):323-329. doi:10.1093/ecco-jcc/jjv174
52. Xie J, Zhu R, Peng Y, et al. Tumor necrosis factor-alpha regulates photoreceptor cell autophagy after retinal detachment. *Sci Rep.* 2017;7(1):17108. doi:10.1038/s41598-017-17400-3
53. Fu D, Zhou J, Zhu WS, et al. Imaging the intracellular distribution of tyrosine kinase inhibitors in living cells with quantitative hyperspectral stimulated Raman scattering. *Nat Chem.* 2014;6(7):614-622. doi:10.1038/nchem.1961
54. Elzinga BM, Nyhan MJ, Crowley LC, O'Donovan TR, Cahill MR, McKenna SL. Induction of autophagy by Imatinib sequesters Bcr-Abl in autophagosomes and down-regulates Bcr-Abl protein. *Am J Hematol.* 2013;88(6):455-462. doi:10.1002/ajh.23428
55. Ertmer A, Huber V, Gilch S, et al. The anticancer drug imatinib induces cellular autophagy. *Leukemia.* 2007;21(5):936-942. doi:10.1038/sj.leu.2404606
56. Guo S, Liang Y, Murphy SF, et al. A rapid and high content assay that measures cyto-ID-stained autophagic compartments and estimates autophagy flux with potential clinical applications. *Autophagy.* 2015;11(3):560-572. doi:10.1080/15548627.2015.1017181
57. Xie Q, Lin Q, Li D, Chen J. Imatinib induces autophagy via upregulating XIAP in GIST882 cells. *Biochem Biophys Res Commun.* 2017;488(4):584-589. doi:10.1016/j.bbrc.2017.05.096
58. Tummino TA, Rezelj VV, Fischer B, et al. Drug-induced phospholipidosis confounds drug repurposing for SARS-CoV-2. *Science.* 2021;373(6554):541-547. doi:10.1126/science.abi4708
59. Rein T. Is autophagy involved in the diverse effects of antidepressants? *Cells.* 2019;8(1):44. doi:10.3390/cells8010044
60. Hoertel N, Sanchez-Rico M, Vernet R, et al. Association between antidepressant use and reduced risk of intubation or death in hospitalized patients with COVID-19: results from an observational study. *Mol Psychiatry.* 2021. doi:10.1038/s41380-021-01021-4
61. Reis G, dos Santos Moreira Silva EA, Medeiros Silva DC, et al. Effect of early treatment with fluvoxamine on risk of emergency care and hospitalization among patients with COVID-19: the TOGETHER randomized platform clinical trial. *medRxiv.* 2021. preprint. doi:10.1101/2021.08.19.21262323
62. Lenze EJ, Mattar C, Zorumski CF, et al. Fluvoxamine vs placebo and clinical deterioration in outpatients with symptomatic COVID-19: a randomized clinical trial. *JAMA.* 2020;324(22):2292-2300. doi:10.1001/jama.2020.22760
63. Zimniak M, Kirschner L, Hilpert H, et al. The serotonin reuptake inhibitor Fluoxetine inhibits SARS-CoV-2 in human lung tissue. *Sci Rep.* 2021;11(1):5890. doi:10.1038/s41598-021-85049-0
64. Kohler CA, Freitas TH, Stubbs B, et al. Peripheral alterations in cytokine and chemokine levels after antidepressant drug treatment for major depressive disorder: systematic review and meta-analysis. *Mol Neurobiol.* 2018;55(5):4195-4206. doi:10.1007/s12035-017-0632-1
65. Ye Q, Wang B, Mao J. The pathogenesis and treatment of the 'Cytokine Storm' in COVID-19. *J Infect.* 2020;80(6):607-613. doi:10.1016/j.jinf.2020.03.037
66. Zschocke J, Rein T. Antidepressants encounter autophagy in neural cells. *Autophagy.* 2011;7(10):1247-1248.
67. Gulbins A, Schumacher F, Becker KA, et al. Antidepressants act by inducing autophagy controlled by sphingomyelin-ceramide. *Mol Psychiatry.* 2018;23:2324-2346. doi:10.1038/s41380-018-0090-9
68. Carpinteiro A, Gripp B, Hoffmann M, et al. Inhibition of acid sphingomyelinase by ambroxol prevents SARS-CoV-2 entry into epithelial cells. *J Biol Chem.* 2021;296:100701. doi:10.1016/j.jbc.2021.100701
69. Crump A, Omura S. Ivermectin, 'wonder drug' from Japan: the human use perspective. *Proc Jpn Acad Ser B Phys Biol Sci.* 2011;87(2):13-28. doi:10.2183/pjab.87.13
70. Martin RJ, Robertson AP, Choudhary S. Ivermectin: an anthelmintic, an insecticide, and much more. *Trends Parasitol.* 2021;37(1):48-64. doi:10.1016/j.pt.2020.10.005
71. Zaidi AK, Dehgani-Mobaraki P. RETRACTED ARTICLE: The mechanisms of action of ivermectin against SARS-CoV-2: an evidence-based clinical review article. *J Antibiot (Tokyo).* 2021. doi:10.1038/s41429-021-00430-5
72. Liu J, Zhang K, Cheng L, Zhu H, Xu T. Progress in understanding the molecular mechanisms underlying the anti-tumour effects of ivermectin. *Drug Des Devel Ther.* 2020;14:285-296. doi:10.2147/DDDT.S237393
73. Zhang P, Ni H, Zhang Y, et al. Ivermectin confers its cytotoxic effects by inducing AMPK/mTOR-mediated autophagy and DNA damage. *Chemosphere.* 2020;259:127448. doi:10.1016/j.chemosphere.2020.127448
74. Zhu, S, Zhou J, Sun X, Zhou Z, Zhu Q. ROS accumulation contributes to abamectin-induced apoptosis and autophagy via the inactivation of PI3K/AKT/mTOR pathway in TM3 Leydig cells. *J Biochem Mol Toxicol.* 2020;34(8):22505. doi:10.1002/jbt.22505
75. Dou, Q, Chen HN, Wang K, et al. Ivermectin induces cyto-static autophagy by blocking the PAK1/Akt axis in breast cancer. *Cancer Res.* 2016;76(15):4457-4469. doi:10.1158/0008-5472.CAN-15-2887

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