

RESEARCH

Open Access

Cytokine profiles in pregnant gilts experimentally infected with porcine reproductive and respiratory syndrome virus and relationships with viral load and fetal outcome

Andrea Ladinig^{1*}, Joan K Lunney², Carlos JH Souza^{2,3}, Carolyn Ashley¹, Graham Plastow⁴ and John CS Harding¹

Abstract

In spite of extensive research, immunologic control mechanisms against Porcine Reproductive and Respiratory Syndrome virus (PRRSv) remain poorly understood. Cytokine responses have been exhaustively studied in nursery pigs and show contradictory results. Since no detailed reports on cytokine responses to PRRSv in pregnant females exist, the objectives of this study were to compare host cytokine responses between PRRSv-infected and non-infected pregnant gilts, and to investigate relationships between cytokine levels in infected gilts and viral load or fetal mortality rate. Serum samples and supernatants of peripheral blood mononuclear cells (PBMC) either stimulated with PRRSv or phorbol myristate acetate/Ionomycin (PMA/Iono) were analyzed for cytokines/chemokines: interleukins (IL) 1-beta (IL1 β), IL4, IL8, IL10, IL12, chemokine ligand 2 (CCL2), interferon alpha (IFN α) and interferon gamma (IFN γ). Three cytokines (IFN α , CCL2, IFN γ) in gilt serum differed significantly in inoculated versus control gilts over time. In supernatants of PRRSv stimulated PBMC from PRRSv-infected gilts, levels of IFN α were significantly decreased, while IL8 secretion was significantly increased. PRRSv infection altered the secretion of all measured cytokines, with the exception of IFN α , from PBMC after mitogen stimulation, indicating a possible immunomodulatory effect of PRRSv. IFN α , CCL2, and IFN γ in serum, and IFN γ in supernatants of PMA/Iono stimulated PBMC were significantly associated with viral load in tissues, serum or both. However, only IFN α in supernatants of PRRSv stimulated PBMC was significantly associated with fetal mortality rate. We conclude that of the eight cytokines tested in this study IFN α was the best indicator of viral load and severity of reproductive PRRSv infection.

Introduction

Cytokines and chemokines play a key role in the regulation of the innate, humoral (T-helper 2 [Th2]) and cellular (T-helper 1 [Th1]) immune responses [1]. *Early cytokines* including the type I interferons and pro-inflammatory cytokines (interleukins 1 (IL1), IL6 and tumor necrosis factor-alpha (TNF α)), and *late cytokines* such as interferon-gamma (IFN γ), are important regulators of adaptive immune responses [2]. Two important chemokines are interleukin 8 (IL8 or CXCL8), a potent recruiter of neutrophils to sites of infection, and chemokine ligand 2 (CCL2), which induces the migration of monocytes

from blood to become tissue macrophages [3]. Antiviral or type I interferons are produced by a variety of cells, with plasmacytoid dendritic cells (pDC) or interferon producing cells (IPCs) being specialists in this task [3]. Type II interferon, IFN γ , and IL12 are key inducers of Th1 immune responses [2,3]. The functions of IL10 are diverse, but principally aimed at immune regulation [3,4]. Unlike in human or mouse, in which IL4 is the major Th2 cytokine [5-7], the role of IL4 in pigs is not completely clear and its expression in vivo following viral infection is usually low or undetectable [8-10].

Recently, bead-based multiplex assays, also known as Fluorescent Microsphere Immunoassays (FMIA), became available for measurement of cytokines in porcine specimens. FMIA allows high throughput, simultaneous detection and quantification of multiple analytes and

* Correspondence: andrea.ladinig@usask.ca

¹Department of Large Animal Clinical Sciences, Western College of Veterinary Medicine, University of Saskatchewan, Saskatoon, SK, Canada
Full list of author information is available at the end of the article

significantly reduced time and sample volume requirements [11,12]. For detection of cytokines, FMIA technology relies on the availability of capture and detection antibodies (Abs) enabling specific and sensitive measurement of the respective analytes. Because a limited number of swine antibodies are available and not all work well in multiplex FMIA the use of FMIA to detect swine cytokines is presently limited [13].

Cytokine responses to Porcine Reproductive and Respiratory Syndrome virus (PRRSv) infection have been exhaustively studied using both *in vivo* and *in vitro* models. A thorough review is beyond the scope of the present paper. However, reports on cytokine responses to PRRSv infection *in vivo* contain contradictory results and were mainly performed in nursery pigs using respiratory models. Rowland et al. [14] used a reproductive model to investigate cytokine responses in PRRSv-infected fetuses but not in dams. To our knowledge, no detailed reports of cytokine responses to PRRSv infection in pregnant sows or gilts exist. Therefore, the objectives of the present study were: 1) to compare host cytokine responses between PRRSv-infected and non-infected gilts following experimental infection in the third trimester of gestation; 2) to investigate relationships between cytokine levels and viral load in gilt serum and gilt tissues; and 3) to investigate relationships between cytokine levels and fetal mortality rate defined at the level of the gilt as percent dead fetuses per litter. Three specific host responses were evaluated over 19 days post inoculation (dpi): 1) cytokine production in gilt serum, 2) cytokine production in supernatants of PRRSv-stimulated peripheral blood mononuclear cells (PBMC), and 3) cytokine production in supernatants of phorbol myristate acetate/Ionomycin (PMA/Iono) stimulated PBMC.

Materials and methods

Animal experiment and sample collection

The experimental infection protocol is described in detail in Ladinig et al. [15]. Briefly, on experimental day 0 (D0), 114 pregnant Landrace gilts (gestation day 85 ± 1 over 12 biweekly replicates) were inoculated with PRRSv isolate NVSL 97-7895 (1×10^5 TCID₅₀; 2 mL intramuscular and 1 mL into each nostril) (INOC), while 19 control gilts were similarly sham inoculated (CTRL). PRRSv negativity of gilts was confirmed before delivery to the University of Saskatchewan by ELISA (IDEXX PRRS X3 Ab test, IDEXX laboratories, Inc., Maine, USA) and PCR (Tetracore PRRS real-time PCR kit, Tetracore, Inc., Rockville, USA). Furthermore, all gilts were negative for PRRSv RNA by a strain-specific in-house quantitative reverse transcription polymerase chain reaction (qRT-PCR) in serum on D0 [15]. Serum and whole blood samples were collected on D0, D2, D6, D19 and D21. On D21 (gestation day 106 ± 1), gilts were humanely euthanized and

necropsy examinations were performed on gilts and their fetuses. Fetal preservation status was recorded and the percent dead fetuses were calculated for each litter. PRRSv RNA concentrations were measured in gilt serum and gilt tissue samples (lung, tonsil, reproductive and tracheobronchial lymph node) collected at termination using qRT-PCR [15]. The experiment was approved by the University of Saskatchewan's Animal Research Ethics Board, and adhered to the Canadian Council on Animal Care guidelines for humane animal use (permit #20110102).

PBMC stimulation

PBMC were isolated from whole blood collected on D0, D2, D6 and D19 by gradient centrifugation using lymphocyte separation medium (Ficoll-Paque™ PLUS, GE Healthcare). Cells were re-suspended in RPMI medium 1640 (RPMI) (Life Technologies, Burlington, ON, Canada) supplemented with antibiotics (1% Pen/Strep) and 10% fetal bovine serum (FBS) (Life Technologies, Burlington, ON, Canada). On the day of isolation, freshly isolated PBMC were seeded in 48-well tissues culture plates (1×10^6 cells/ well, total volume 500 μ L). Duplicate wells for each stimulation time point were stimulated with either 10 ng/mL PMA (Sigma-Aldrich, Oakville, ON, Canada) and 250 ng/mL ionomycin (Sigma-Aldrich, Oakville, ON, Canada) (PMA/Iono) or with PRRSv isolate NVSL 97-7895 (multiplicity of infection =1). Unstimulated cells were seeded in duplicate wells containing cell culture medium only. Plates were incubated at 37 °C in 5% CO₂ for either 36 h (PMA/Iono stimulation) or 60 h (PRRSv stimulation). Cells and supernatants from duplicate wells were pooled and centrifuged (400 g, 10 min). Supernatant aliquots were stored frozen at -80 °C until further testing.

Cytokine testing

Serum samples (D0, D2, D6, D21) and PBMC supernatants (D0, D2, D6, D19) were analysed for the following innate, regulatory, Th1 and Th2 cytokines/chemokines by FMIA: IL1 β , IL8, CCL2, IFN α , IL10, IL12, and IL4. Due to the lack of availability of antibodies suitable for FMIA, an enzyme-linked immunosorbent assay (ELISA) was used to measure IFN γ .

Multiplex fluorescent microsphere immunoassay (FMIA)

For serum samples, a 7-plex in-house FMIA assay was developed as previously described with several modifications [12]. Beads, standards and Ab pairs used for the different cytokines are listed in Table 1. Briefly, capture antibodies were covalently coupled to magnetic beads (Biorad, Mississauga, ON, Canada). Coupling efficiency was tested by staining newly coupled beads with goat anti-mouse IgG-Phycoerythrin labeled Ab (Life Technologies, Burlington, ON, Canada) and comparing fluorescence

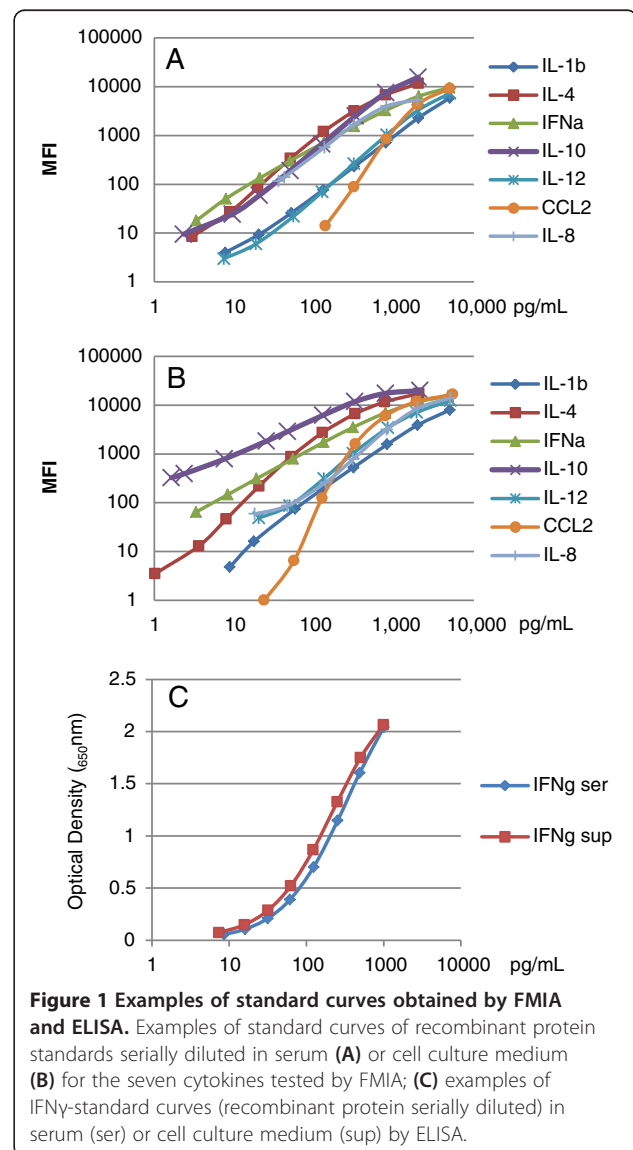
Table 1 Standards, capture and detection antibodies used by FMIA and ELISA

Cytokine (bead region)	Standard (source)	Capture Ab (source)	Detection Ab (source)
IL1 β (26)	DY681, part 841042 (RD)	DY681, part 841040 (RD)	BAF 681 (RD)
IL8 (27)	SD061 (L)	MCA1660 (C)	MAB5351 (RD)
CCL2 (53)	RP00175-025 (C)	Anti-poCCL2 clone 5-2 (Lu)	Anti-poCCL2 clone 18-1 (Lu)
IFN α (45)	17105-1 (C)	GTX11408 (GT)	27105-1 (C)
IL10 (28)	CSC0103, part SD064 (L)	ASC0104 (L)	ASC9109 (L)
IL12 (36)	DY912, part 841099 (RD)	MA0413S-500 (C)	BAM9122 (RD)
IL4 (34)	CSC1283, part 5S.128.10 (L)	CSC1283, part 5S.128.09 (L)	ASC0849 (L)
IFN γ (ELISA)	CSC4033, part SD066 (L)	CSC4033, part ASC4934D (L)	CSC4033, part ASC4839D (L)

(RD) R&D Systems, Minneapolis, MN, USA; (L) Life technologies, Burlington, ON, Canada; (C) Cedarlane, Burlington, ON, Canada; (GT) GeneTex, Irvine, CA, USA; (Lu) Lunney lab, Beltsville, MD, USA (Lunney et al., manuscript in preparation).

intensity of the newly coupled beads with existing batches (generously provided by Dr C. Souza, USDA, Beltsville, MD, USA). Serum from a healthy, non-infected gilt with no measurable levels of relevant cytokines by FMIA was used to prepare serial dilutions of cytokine standards (Figure 1A). The same serum was used as negative control on each FMIA plate. All serum samples were tested at a 1:3 dilution in phosphate buffered saline (PBS, pH 7.4) supplemented with 1% bovine serum albumin (BSA) and 0.05% sodium azide (PBS-BN). Samples and standards were incubated with magnetic beads in duplicate wells of a 96-well plate for 2 h. All incubations were performed in the dark (aluminum foil cover) at room temperature on a rotating plate shaker (500 rpm). A 96-well plate washer (Bio-Plex Pro™ II Wash Station, Biorad, Mississauga, ON, Canada) was used to perform 3 washes with PBS +0.05% Tween 20 (PBST) after each incubation step. Fifty μ L of anti-cytokine, biotinylated, secondary antibody (Ab) diluted in PBS-BN were added to each well and plates were incubated for 90 min. The optimal concentration of each biotinylated detection Ab was determined by titration. Finally, beads in each well were incubated in 50 μ L of a solution containing 10 μ g/mL of Streptavidin-R-Phycoerythrin (SAPE, PJRS20 Prozyme, Hayward, CA, USA) for 30 min, washed, and re-suspended in 100 μ L PBST. Coupled microspheres were analyzed on a Bio-Plex® 200 system (Biorad, Mississauga, ON, Canada) and analyzed with the Bio-Plex Manager software version 6.1 (Biorad, Mississauga, ON, Canada). All mean fluorescence intensity (MFI) measurements were background corrected by subtracting the MFI of the negative control (control serum diluted 1:3 in PBS-BN) from the MFI for the relevant analyte in each sample.

PBMC supernatants were tested in a similar manner. However, due to high cytokine concentrations present after either PRRSv or PMA/Iono stimulation, a higher sample dilution was required for certain analytes and use of a 7-plex assay was therefore not feasible. For supernatants of PRRSv stimulated PBMC, CCL2 was analyzed as a single-plex assay at a 1:100 dilution, while the remaining



analytes were tested in a 6-plex assay at a 1:5 dilution. For supernatants of PMA/Iono stimulated PBMC, IL-8 was analyzed at a 1:20 dilution, while the remaining analytes were tested in a 6-plex assay at a 1:5 dilution. Cell culture medium (RPMI +10% FBS +1% Pen/Strep) at the relevant dilution in PBS-BN was used as negative control and to prepare standard curves (Figure 1B). Biotinylated detection Abs were titrated in order to determine the optimal concentration to use in testing PBMC supernatants. Results from supernatants of PRRSv or PMA/Iono stimulated PBMC were adjusted by subtracting the cytokine levels in supernatants of unstimulated cells cultured for the same time point (36 or 60 h). These are reported as “adjusted” cytokine values herein.

IFN- γ ELISA

The assay was carried out using the Novex Swine IFN γ antibody duoset kit (Life Technologies Inc., Burlington, ON, Canada) according to the manufacturer's instructions. Low coefficient of variation (CV) Immulon 4 HBX 96 well plates (VWR International, Mississauga, ON, Canada) were coated with capture Ab diluted in Coating Buffer B (Life Technologies Inc., Burlington, ON, Canada). 5X Assay Buffer (Novex Antibody Pair Buffer Set, Life Technologies Inc., Burlington, ON, Canada) diluted in ddH₂O was used as a blocking solution and for dilution of detection and Streptavidin-HRP Abs and samples as appropriate. Plates were washed with PBST using a Bio-Plex Pro II Wash Station. Peroxidase-labeled conjugates were detected by the addition of SureBlue Reserve TMB Microwell Substrate (Mandel Scientific Company Inc, Guelph, ON, Canada). Color was developed in the dark until the optical density of the highest standard reached approximately 2.0 at a wavelength of 650 nm without addition of Stop Solution. Plates were read in a Vmax Microplate Reader (Molecular Devices Corporation, Downingtown, PA, USA) using SoftMaxPro software Version 1.1 (Sunnyvale, CA, USA). The standard curve (1000 pg/mL to 8 pg/mL) was constructed using doubling dilutions (Figure 1C). Concentrations of IFN γ in samples were estimated by comparison with a 4-parameter standard curve generated by the software. Serum samples were diluted 1:3 in assay buffer whereas the standard diluent was a 1:3 solution of control gilt serum in assay buffer. Supernatants of PRRSv and PMA/Iono stimulated PBMC were tested at 1:3 and 1:100 dilutions, respectively, in assay buffer. The standard diluent was a corresponding dilution of cell culture medium in assay buffer. As above, IFN γ levels in PBMC supernatants were adjusted by subtracting the IFN γ levels in supernatants of unstimulated cells; adjusted IFN γ values are compared and reported herein.

Cytokine quality control (QC)

Protein standards (Table 1) were used to prepare high and low concentration quality controls in either control

serum or cell culture media at the relevant dilution in PBS-BN. Both high and low quality controls were run in duplicate on each FMIA and ELISA plate. Process behavior control charts were used to monitor the inter-plate variation of each cytokine concentration in the QC samples, as well as the MFI of the highest and lowest standard on each FMIA plate. Plates were repeated if values fluctuated above or below the 3-sigma natural process limits on the appropriate control charts. Individual samples were repeated if the replicate CV was >20% as calculated by the instrument's software.

Statistical analysis

Separate statistical analyses were conducted using Stata 13 (STATA Corp, College Station, Texas, USA) to address each of three objectives. To meet the key model assumptions, data were transformed (logarithm base₁₀ or zero-skewness (lnskew0 function in STATA)) as appropriate. First, to determine if cytokine levels differed between INOC and CTRL gilts, multilevel mixed-effects linear regression models were developed. These models used gilt as a random effect and accounted for repeated measures by day. All remaining analyses used data from INOC gilts only and specifically focused on the cytokines that differed statistically between INOC and CTRL gilts. Our second objective was to determine potential relationships between cytokine levels (serum and supernatants) and PRRS viral load in serum and tissues. For these analyses, multilevel mixed-effects linear regression models controlling for experimental replicate were used. The area under the curve (AUC) for PRRSv RNA (target copies/ μ L, D0-D21) and cytokine protein (pg/mL) in serum (D0-D21) and supernatants (D0-D19) over time was calculated using the formula $AUC = (t_1 - t_0)(a_1 + a_0)/2 + (t_2 - t_1)(a_1 + a_2)/2 + \dots + (t_n - t_{n-1})(a_{n-1} + a_n)/2$. Our third objective was to determine potential associations between cytokine levels in serum (AUC) and fetal survival rate (percentage of dead fetuses per litter). For all analyses, cytokine levels in sera or adjusted cytokine levels in supernatants (as described above) were used.

To account for multiple comparisons, all associations were considered statistically significant if $P < 0.01$. All final models were evaluated to ensure normality and homoscedasticity of residuals.

Results

Cytokine differences between inoculated and control gilts

Examples of standard curves of recombinant protein standards serially diluted in either serum or cell culture medium for the seven cytokines tested by FMIA are presented in Figures 1A and 1B. Examples of standard curves for the IFN γ ELISA are presented in Figure 1C.

Cytokine levels showed a high degree of variation between individual gilts in all sample types (Additional

file 1, Additional file 2 and Additional file 3). Serum levels of CCL2, IFN α and IFN γ differed significantly between INOC and CTRL gilts over time ($P < 0.001$ for all). INOC gilts showed a significant increase in CCL2 and IFN α in sera collected on D2 and D6, whereas IFN γ was increased significantly on D2 only (Figure 2). IFN γ levels measured in serum of INOC gilts on D21 were lower than CTRL gilts; however, differences were not statistically significant. High serum levels of IL1 β and IL8 at all 4 time points were measured in individual INOC and CTRL gilts, but PRRSv inoculation had no effect on these analytes over time (Additional file 1). Levels of IL12, IL10 and IL4 were very low in serum of both INOC and CTRL gilts and did not significantly differ over time (Additional file 1).

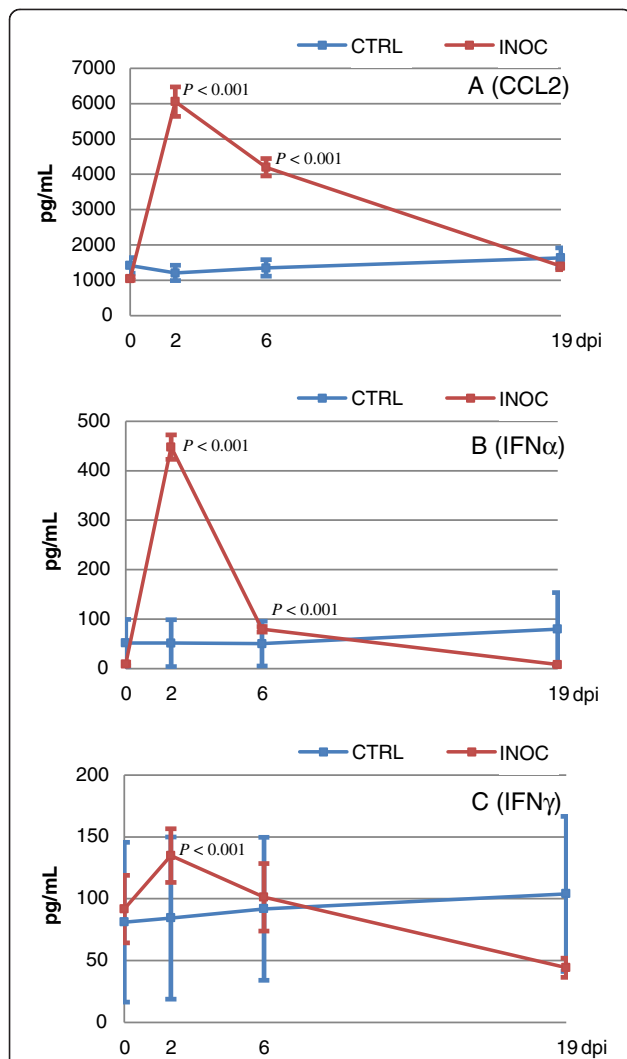


Figure 2 CCL2, IFN α and IFN γ levels in gilt serum over time. Mean (\pm SEM) CCL2 (A), IFN α (B) and IFN γ (C) levels in serum from 111 INOC and 19 CTRL gilts across respective study days; P -values indicate significant differences between INOC and CTRL gilts on individual days.

In supernatants of PRRSv stimulated PBMC, levels of IFN α and IL8 differed significantly between INOC and CTRL gilts over time ($P \leq 0.001$). PRRSv stimulation induced high levels of IFN α in PBMC from CTRL gilts and from INOC gilts before inoculation. Levels of IFN α were highly variable, ranging from 62 to 17025 pg/mL. A significant reduction in IFN α was detected in stimulated PBMC from INOC gilts compared to CTRL on D2, D6, and D19 (Figure 3A). Decreased levels were detected in all INOC gilts but to varying degrees. IL8 levels in supernatants of PRRSv stimulated PBMC were significantly increased on D2, D6, and D19 in INOC versus CTRL gilts (Figure 3B). Levels of IL1 β , CCL2, IFN γ , IL4, IL10 and IL12 in supernatants of PRRSv stimulated PBMC were either not detectable or low, and did not significantly differ between INOC and CTRL gilts over time (Additional file 2).

IFN α was not induced by PMA/Iono stimulation. All other cytokines from PMA/Iono stimulated PBMC differed between INOC and CTRL gilts over time ($P < 0.001$ for all except IFN γ where only a trend was reported ($P = 0.039$)) (Additional file 3). Among the 7 cytokines measured, four discrete patterns were observed reflecting the potential effects of gilt PRRSv infection on PBMC PMA/Iono

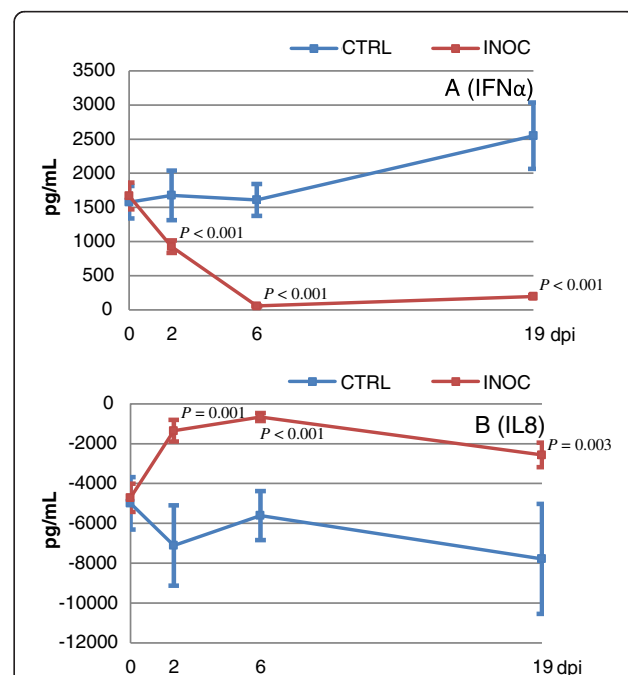


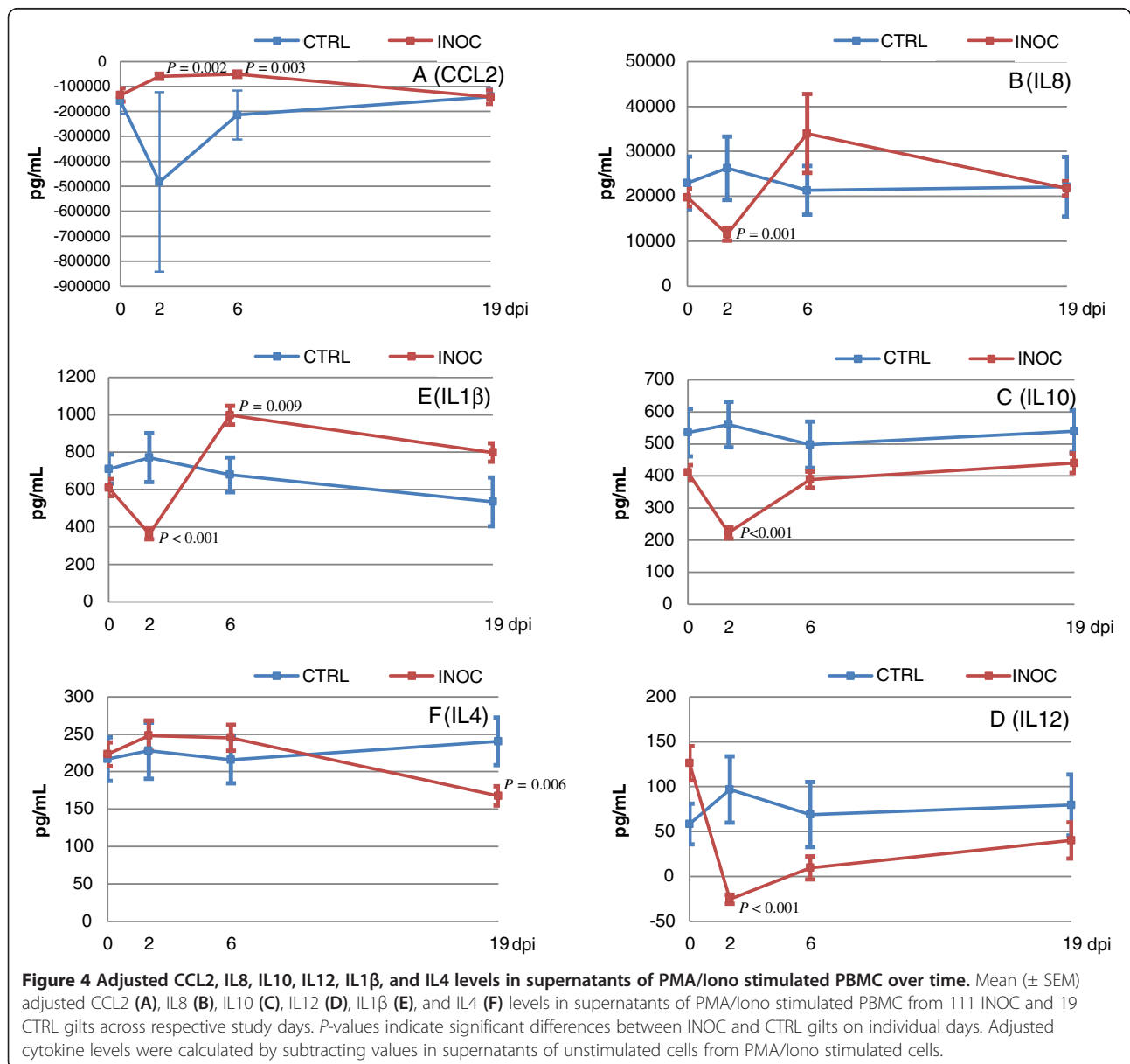
Figure 3 Adjusted IFN α and IL8 levels in supernatants of PRRSv stimulated PBMC over time. Mean (\pm SEM) adjusted IFN α (A) and IL8 (B) levels in supernatants of PRRSv stimulated PBMC from 111 INOC and 19 CTRL gilts across respective study days. P -values indicate significant differences between INOC and CTRL gilts on individual days. Adjusted cytokine levels were calculated by subtracting values in supernatants of unstimulated cells from PRRSv stimulated cells.

responsiveness. First, PRRSV infection resulted in a significantly increased CCL2 response to PMA/Iono stimulation on D2 and D6 but not on D19 (early induction; Figure 4A). These statistical differences however, are the result of a single outlier CTRL gilt with massive spontaneous production of CCL2 from unstimulated PBMC on day 2. With this outlier removed, CCL2 production significantly differed between INOC and CTRL over time ($P = 0.01$) with levels in INOC increased on D2 and D6 compared to CTRL. After adjusting for multiple comparison this was considered a trend (D2 $P = 0.027$, D6 $P = 0.019$). By contrast, levels of IL8, IL10, and IL12 were significantly decreased on D2 in INOC compared to CTRL gilts (early suppression; Figures 4B-D). Following PMA/Iono stimulation, levels of IL1 β decreased similarly on D2, but were

significantly increased in PBMC from INOC versus CTRL gilts on D6 (early suppression with rebound; Figure 4E). Finally, IL4 demonstrated late suppression with levels from PMA/Iono stimulated PBMC significantly decreased in INOC versus CTRL on D19. Prior to D19, IL4 levels numerically increased in INOC gilts (Figure 4F). Similarly, IFN γ levels decreased in INOC gilts on D19; however, differences were not significant but rather a trend ($P = 0.023$; Additional file 3).

Relationships between cytokines levels and PRRS viral load

Detailed results on viral loads in gilt serum and tissues can be found in Ladinig et al. [15]. Briefly, all INOC gilts were viremic on D2 and D6, and 94/111 (84.7%) remained viremic until termination. The percentage of tissues that



tested positive by PRRSv qRT-PCR at termination (D21) from INOC gilts were 90.1 (lung), 99.1 (tonsil), 100 (reproductive lymph node), and 99.1 (tracheobronchial lymph node), respectively. The mean viral loads of positive samples were $3.5 (\pm 1.2) \log_{10}$ RNA copies/mg in lung, $5.6 (\pm 0.8) \log_{10}$ RNA copies/mg in tonsil, $5.8 (\pm 0.8) \log_{10}$ RNA copies/mg in reproductive lymph node, and $4.8 (\pm 0.9) \log_{10}$ RNA copies/mg in tracheobronchial lymph node. AUC of IFN α and CCL2 in serum were positively associated with the AUC for viral load in serum (Table 2, Figures 5A and B). As shown in Figure 5, this association was mainly driven by gilts with high cytokine levels and high viral load over time, represented in the upper right quadrant. Importantly, no gilts had high cytokine levels but low viral loads over time (lower right quadrant). A positive association was detected between the AUC for CCL2 in serum and the viral load in lung. IFN α levels in serum trended to be positively associated with viral load in tonsil, while IFN γ levels showed a significant, but negative association (Table 2). No associations could be found for cytokine levels in serum with viral load in tracheobronchial lymph node. Although none of the cytokines measured in supernatants of PRRSv stimulated PBMC were associated with viral load in serum or tissues, levels of IFN γ in supernatants of PMA/Iono stimulated PBMC showed a negative association with viral load in reproductive lymph node (Table 2).

Relationships between cytokines and fetal mortality rate

In INOC gilts, the percentage of dead fetuses 21 days post inoculation varied from 0% to 94.4% (mean $41.0 \pm 22.8\%$). AUC of IFN α in supernatants of PRRSv stimulated PBMC was significantly associated with percent dead ($\beta = 0.001$, $P = 0.006$) (Figure 5C), similarly AUC of IFN α in serum trended to be positively associated with fetal death ($\beta = 0.0035$, $P = 0.060$). For cytokine levels in

supernatants, a negative trend was detected between the AUC of IL10 in PMA/Iono stimulated PBMC and percent dead fetuses ($\beta = -0.001$, $P = 0.045$). For the remaining analytes, levels in serum or in supernatants were not associated with fetal death.

Discussion

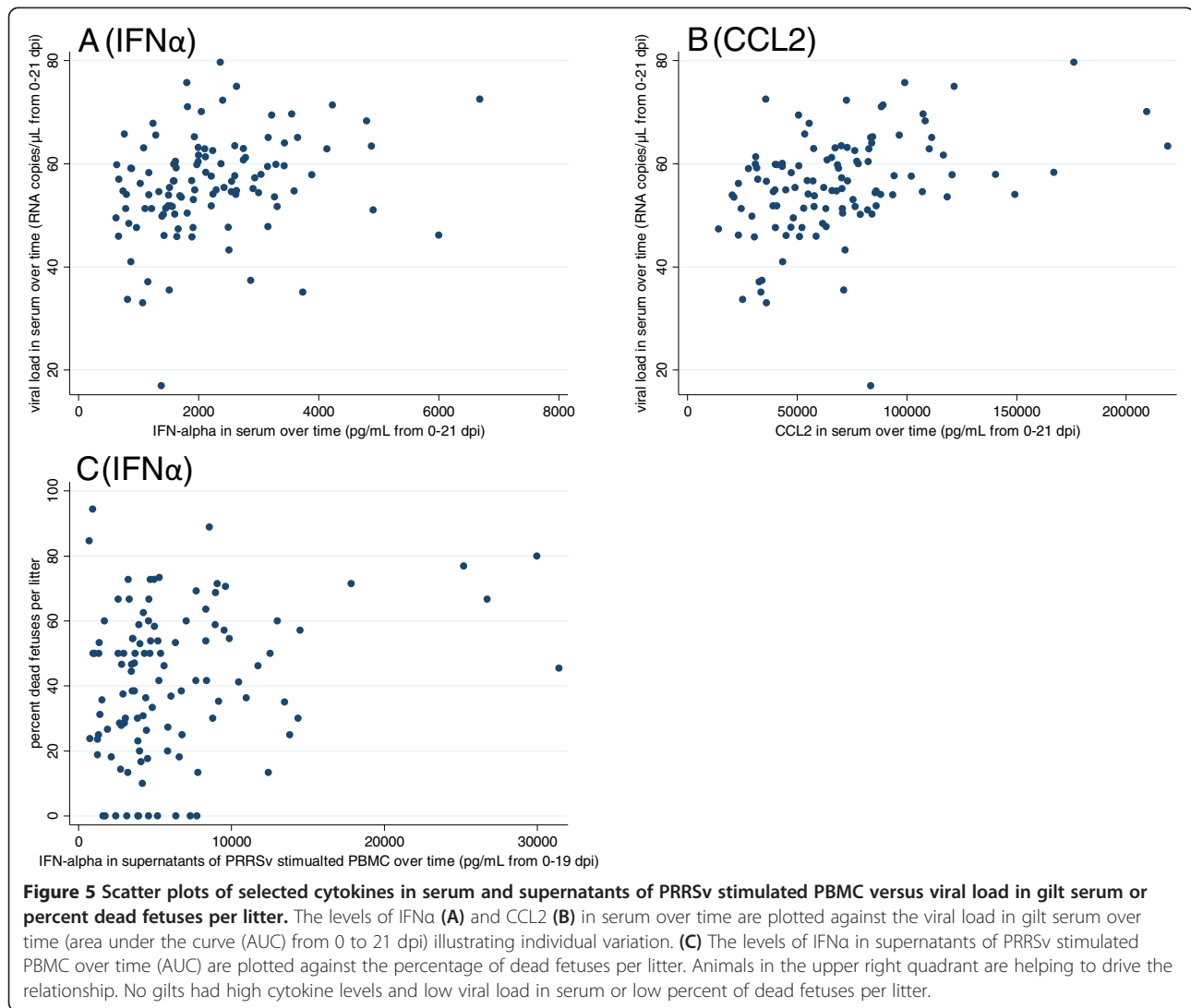
The results presented herein are part of an extensive and complex dataset obtained in a large scale, multi-institutional project aimed at finding phenotypic and genotypic predictors of PRRSv resistance in pregnant gilts. The objectives of this present study were to investigate cytokine responses in serum and supernatants of PBMC, either stimulated with PRRSv or a nonspecific mitogen (PMA/Iono), and to determine possible associations with the outcome of infection in a reproductive model. In order to investigate genotypic variation in the severity of reproductive PRRS, a large number of gilts were experimentally inoculated in the third trimester of gestation, while the number of CTRL gilts was reduced to the minimum needed to provide baseline data.

PMA, a diester of phorbol and potent tumor promoter which activates the signal transduction enzyme protein kinase C, is often used in conjunction with ionomycin, an ionophore inducing calcium transport into the cell, in order to stimulate immune responses in vitro [3,16]. In the present experiment, PMA/Iono stimulation of PBMC induced the secretion of all cytokines tested with the exception of IFN α . Four distinct patterns were observed in the way PRRSv infection altered PBMC cytokine secretions after PMA/Iono stimulation. The altered secretion of cytokines by PMA/Iono stimulated PBMC from INOC gilts after infection indicates a PRRSv immunomodulatory effect. However, the biological meaning of these findings is difficult to interpret. When testing the associations between cytokine levels in supernatants of PMA/Iono

Table 2 Associations between cytokine levels and viral load in serum and tissues following PRRSv infection in pregnant gilts

PRRSv RNA concentration ($\log_{10}/\mu\text{L}$ or mg)	Serum cytokine levels (AUC)			PMA/Iono supernatant (AUC)
	IFN α	CCL2	IFN γ	IFN γ
Serum (AUC)	$P = 0.003$ $\beta = 0.002134$	$P < 0.001$ $\beta = 0.000104$	ns	ns
Lung	ns	$P = 0.010$ $\beta = 0.000010$	ns	ns
Tonsil	$P = 0.016$ $\beta = 0.000185$	ns	$P < 0.001$ $\beta = -0.000115$	ns
Repro LN	ns	ns	ns	$P = 0.002$ $\beta = -0.0000004$

The associations between cytokine levels over time (represented as area under curve (AUC)) and viral load in gilt serum and tissues are presented. *P*-values (regression coefficients, β) were obtained by univariate, multilevel mixed-effects regression models. Only analytes that were found to be significantly associated with viral load in either serum or any of the tested tissues are included. No association was found among cytokines in supernatants of PRRSv stimulated PBMC and viral load. Repro LN = reproductive lymph node. PMA/Iono supernatant AUC = AUC of IFN γ levels secreted from PMA/Iono stimulated PBMC from PRRSv infected gilts. ns = not significant ($P > 0.01$).



stimulated PBMC and viral load or fetal mortality rate, the only significant association was between IFN γ and viral load in reproductive lymph node, where higher levels of IFN γ were associated with a lower viral burden.

Serum IFN γ levels were significantly increased in INOC gilts early in infection. This finding agrees with several reports of either numeric or significant increases in the number of IFN γ secreting cells by enzyme-linked immunospot (ELISPOT) assay, or in the protein levels measured in serum of pigs after PRRSv inoculation [12,17-24]. IFN γ levels in supernatants of PRRSv stimulated PBMC also showed a slight increase on D2, which was not statistically significant. Unlike IFN α and CCL2, no association was found between IFN γ levels in serum and viral load in gilt serum over time. On the other hand, viral loads in tonsil were negatively associated with IFN γ levels in serum

indicating a possible protective effect of IFN γ . However, IFN γ levels over time were not associated with fetal death. In contrast, Lowe et al. [25] showed that a strong cell mediated immune response determined by use of an IFN γ ELISPOT was correlated with protection against reproductive PRRS in 3 of 4 investigated farms.

The present study found IFN α to be one of the most relevant cytokines in PRRSv infection. High levels of IFN α were produced by PRRSv stimulated PBMC from CTRL gilts and INOC gilts before inoculation, which is in contrast to previous reports [26,27]. IFN α production, however, was significantly decreased in PBMC collected at all time points following PRRSv inoculation. The levels of IFN α in supernatants of PRRSv stimulated PBMC were positively associated with fetal mortality. Interestingly, it was previously demonstrated that North American type

PRRS viruses are potent inhibitors of type 1 interferon production in pDC [28], which are a major source of IFN α and play a key role in the early control of virus replication and the development of adaptive antiviral immune responses [29]. By contrast, a recent study showed that various type 1 and type 2 PRRSv strains induced IFN α secretion by pDC and showed either no or only weak suppression of IFN α secretion [27]. These recent data corroborate our findings of increased levels of IFN α in serum from INOC gilts. However, results of IFN α measurements in serum from PRRSv infected pigs are quite inconsistent. While some studies could not detect significant IFN α increases in serum of PRRSv inoculated pigs within one week of infection [12,18], others did [19,30]. Increased levels of IFN α might have negative effects in regards to PRRSv infection, since it was previously shown that IFN α up-regulated the expression of sialoadhesin and therefore enhanced PRRSv infection of monocytes [31].

In agreement with our results, Souza et al. [32] found significant increases in serum IFN α and CCL2 levels of nursery piglets within the first 2 weeks after inoculation with PRRSv 97–7895, the same virus strain as used for this study. IFN α and CCL2 levels in serum were also associated with PRRSv RNA concentration and growth performance where piglets with low concentration in serum showed a faster return to basal levels of IFN α and CCL2 in serum compared to highly viremic animals. Although we demonstrate the biological importance of IFN α in the reproductive PRRSv model herein, the relevance of CCL2 responses in reproductive PRRS is questionable. Although serum CCL2 levels were positively associated with viral burden in serum and lung tissue in the present study, no association was detected between CCL2 levels and fetal death. We propose the role of CCL2 in PRRSv infection primarily involves monocyte recruitment in lung, and is unrelated to mechanisms of fetal death or survival.

Although IL8 levels significantly increased in supernatants of PRRSv stimulated PBMC from INOC gilts after infection, this finding did not have any biological relevance as IL8 levels were not associated with any measured outcome, including PRRSv viral levels or fetal mortality rate. In agreement with previous reports [12,19], no significant increase in serum IL8 levels was detected in INOC compared to CTRL gilts.

In conclusion, PRRSv infection increased IFN α , CCL2, and IFN γ levels in serum but only before 7 dpi. Viral load in gilt serum was positively associated with serum IFN α and CCL2 levels. Although CCL2 and IFN γ were not associated with fetal outcome, IFN α levels in serum trended to be positively associated with fetal mortality rate. The importance of IFN α in regards to PRRSv infection was demonstrated by the significant inhibition of IFN α secretion by PRRSv stimulated PBMC from INOC

gilts after infection, and a significant positive association between fetal mortality rate and levels of IFN α in supernatants from PRRSv stimulated PBMC. Therefore, IFN α secretion could be used as a possible indicator of viral load and severity of reproductive PRRS. By contrast, IL8 was not associated with viral load or fetal death even though IL8 secretion by PRRSv stimulated PBMC from INOC gilts was significantly increased at all investigated time points after infection. Finally, levels of IFN γ in supernatants of PMA/Iono stimulated PBMC were suppressed by 21 days post-inoculation, and were negatively associated with viral load in reproductive lymph node. Together with the fact that IFN γ levels in serum were negatively associated with viral load in tonsil, this suggests a protective effect of IFN γ in PRRSv pathogenesis.

Additional files

Additional file 1: Mean cytokine levels (SD) in serum. Mean cytokine (SD) levels in serum are presented for the 8 analysed cytokines from 111 INOC and 19 CTRL gilts on respective study days post inoculation. If the repeated measures, multilevel mixed-effects regression model demonstrated values in INOC significantly differed from CTRL gilts over all experimental days (DAY*INOC), group differences on individual days were compared (INOC_CTRL). Due to multiple comparisons, $P < 0.01$ was considered statistically significant. ns = not significant.

Additional file 2: Mean cytokine levels (SD) in supernatants of unstimulated and PRRSv stimulated PBMC. Mean (SD) cytokine levels in supernatants of unstimulated and PRRSv stimulated PBMC are presented for the 8 analysed cytokines from 111 INOC and 19 CTRL gilts on the respective study days post inoculation. Adjusted values were calculated by subtracting values in supernatants of unstimulated cells from PRRSv stimulated cells and used in statistical analyses. Statistics determined whether values in INOC significantly differed from CTRL gilts over all experimental days (DAY*INOC), or on individual days (INOC_CTRL). Due to multiple comparisons, $P < 0.01$ was considered statistically significant; ns = not significant.

Additional file 3: Mean cytokine levels (SD) in supernatants of unstimulated and PMA/Iono stimulated PBMC. Mean (SD) cytokine levels in supernatants of unstimulated and phorbol myristate acetate/lonomycin (PMA/Iono) stimulated PBMC are presented for the 8 analysed cytokines from 111 INOC and 19 CTRL gilts on the respective study days post inoculation. Adjusted values were calculated by subtracting values in supernatants of unstimulated cells from PMA/Iono stimulated cells and used in statistical analyses. Statistics determined whether values in INOC significantly differed from CTRL gilts over all experimental days (DAY*INOC) or on individual days (INOC_CTRL). Due to multiple comparisons, $P < 0.01$ was considered statistically significant. ns = not significant.

Abbreviations

(Ab): Antibody; (APC): Antigen-presenting cells; (AUC): Area under the curve; (BSA): Bovine serum albumin; (CCL2): Chemokine ligand 2; (CV): Coefficient of variation; (DC): Dendritic cells; (ELISA): Enzyme-linked immunosorbent assay; (ELISPOT): Enzyme-linked immunospot assay; (FBS): Fetal bovine serum; (FMIA): Fluorescent Microsphere Immunoassays; (INOC): Gilts inoculated with PRRSv; (CTRL): Gilts that were sham inoculated; (IFN α): Interferon alpha; (IFN γ): Interferon gamma; (IPCs): Interferon producing cells; (IL): Interleukins; (MHC): Major histocompatibility complex; (MFI): Mean fluorescence intensity; (NK cells): Natural killer cells; (PBMC): Peripheral blood mononuclear cells; (PMA/Iono): Phorbol myristate acetate/Ionomycin; (PBS): Phosphate buffered saline; (PBS-BN): PBS with 0.05% sodium azide; (PBST): PBS +0.05% Tween 20; (pDC): Plasmacytoid dendritic cells; (PRRSv): Porcine Reproductive and Respiratory Syndrome virus; (QC): Quality control; (qRT-PCR): Quantitative reverse transcription polymerase chain reaction; (Th1): T-helper 1; (Th2): T-helper 2; (SAPE): Streptavidin-R-Phycoerythrin; (TNF α): Tumor necrosis factor-alpha.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

AL and JH conducted the research, led the interpretation of results and drafted the manuscript. GP and JKL assisted in the experimental design and interpretation. JH, AL and CA led the animal infection trials including animal care, collection of samples and PCR assays. JKL and CS helped develop the cytokine assays. JH and AL coordinated and oversaw statistical analysis of the data. All co-authors reviewed and contributed to writing of the manuscript. All authors read and approved the final manuscript.

Acknowledgements

The authors wish to acknowledge the numerous technicians and students from the Western College of Veterinary Medicine, Vaccine and Infectious Disease Organization, Prairie Diagnostic Services, Inc. and the University of Alberta who assisted with this project. We offer special thanks to Samuel Abrams for the help in the development of the FMIA assay, and to Ian Dohoo for his guidance with statistical analyses. Pregnant gilts were provided and bred by Fast Genetics Inc., Spiritwood, Canada, with management support of Dawn Friesen, Connie Heisler, Donell Wingerter and Benny Mote. Funding for the project was generously provided by grants from Genome Canada and Genome Prairie (grant number 2209-F), with administrative support from Genome Alberta.

Author details

¹Department of Large Animal Clinical Sciences, Western College of Veterinary Medicine, University of Saskatchewan, Saskatoon, SK, Canada. ²U.S. Department of Agriculture, Animal Parasitic Diseases Laboratory, Beltsville Agricultural Research Center, Agricultural Research Service, Beltsville, MD, USA. ³EMBRAPA Pesca e Aquicultura, Palmas, TO, Brazil. ⁴Department of Agricultural, Food, and Nutritional Science, Faculty of Agricultural, Life and Environmental Sciences, University of Alberta, Edmonton, AB, Canada.

Received: 3 April 2014 Accepted: 20 October 2014

Published online: 06 December 2014

References

- Dinarello CA (2000) Proinflammatory cytokines. *Chest* 118:503–508
- van Reeth K, Nauwynck H (2000) Proinflammatory cytokines and viral respiratory disease in pigs. *Vet Res* 31:187–213
- Murphy KP (2012) *Janeway's Immunobiology*. Garland Science, Taylor & Francis Group, LLC, New York, USA
- Moore KW, de Waal MR, Coffman RL, O'Garra A (2001) Interleukin-10 and the interleukin-10 receptor. *Annu Rev Immunol* 19:683–765
- Howard M, Farrar J, Hilfiker M, Johnson B, Takatsu K, Hamaoka T, Paul WE (1982) Identification of a T cell-derived B cell growth factor distinct from interleukin 2. *J Exp Med* 155:914–923
- Zhou Y, Lin G, Baarsch MJ, Scamurra RW, Murtaugh MP (1994) Interleukin-4 suppresses inflammatory cytokine gene transcription in porcine macrophages. *J Leukocyte Biol* 56:507–513
- Zhou Y, Lin G, Murtaugh MP (1995) Interleukin-4 suppresses the expression of macrophage NADPH oxidase heavy chain subunit (gp91-phox). *Biochim Biophys Acta* 1265:40–48
- Diaz I, Mateu E (2005) Use of ELISPOT and ELISA to evaluate IFN-gamma, IL-10 and IL-4 responses in conventional pigs. *Vet Immunol Immunopathol* 106:107–112
- Murtaugh MP, Baarsch MJ, Zhou Y, Scamurra RW, Lin G (1996) Inflammatory cytokines in animal health and disease. *Vet Immunol Immunopathol* 54:45–55
- Murtaugh MP, Johnson CR, Xiao Z, Scamurra RW, Zhou Y (2009) Species specialization in cytokine biology: is interleukin-4 central to the T(H)1-T(H)2 paradigm in swine? *Dev Comp Immunol* 33:344–352
- Christopher-Hennings J, Araujo KP, Souza CJ, Fang Y, Lawson S, Nelson S, Clement T, Dunn M, Lunney JK (2013) Opportunities for bead-based multiplex assays in veterinary diagnostic laboratories. *J Vet Diagn Invest* 25:671–691
- Lawson S, Lunney J, Zuckermann F, Osorio F, Nelson E, Welbon C, Clement T, Fang Y, Wong S, Kulas K, Christopher-Hennings J (2010) Development of an 8-plex Luminex assay to detect swine cytokines for vaccine development: assessment of immunity after porcine reproductive and respiratory syndrome virus (PRRSV) vaccination. *Vaccine* 28:5356–5364
- U.S. Veterinary Immune Reagent Network. [www.vetimm.org]
- Rowland RR (2010) The interaction between PRRSV and the late gestation pig fetus. *Virus Res* 154:114–122
- Ladinig A, Wilkinson J, Ashley C, Detmer SE, Lunney JK, Plastow G, Harding JC (2014) Variation in fetal outcome, viral load and ORF5 sequence mutations in a large scale study of phenotypic responses to late gestation exposure to type 2 porcine reproductive and respiratory syndrome virus. *PLoS One* 9:e96104
- Gao Y, Flori L, Lecardonnel J, Esquerre D, Hu ZL, Teillaud A, Lemonnier G, Lefevre F, Oswald IP, Rogel-Gaillard C (2010) Transcriptome analysis of porcine PBMCs after in vitro stimulation by LPS or PMA/ionomycin using an expression array targeting the pig immune response. *BMC Genomics* 11:292
- Diaz I, Darwich L, Pappaterra G, Pujols J, Mateu E (2005) Immune responses of pigs after experimental infection with a European strain of porcine reproductive and respiratory syndrome virus. *J Gen Virol* 86:1943–1951
- Gomez-Laguna J, Salguero FJ, De Marco MF, Pallares FJ, Bernabe A, Carrasco L (2009) Changes in lymphocyte subsets and cytokines during European porcine reproductive and respiratory syndrome: increased expression of IL-12 and IL-10 and proliferation of CD4(-)CD8(high). *Viral Immunol* 22:261–271
- Guo B, Lager KM, Henningson JN, Miller LC, Schlink SN, Kappes MA, Kehrli ME, Jr, Brockmeier SL, Nicholson TL, Yang HC, Faaberg KS (2013) Experimental infection of United States swine with a Chinese highly pathogenic strain of porcine reproductive and respiratory syndrome virus. *Virology* 435:372–384
- Meier WA, Galeota J, Osorio FA, Husmann RJ, Schnitzlein WM, Zuckermann FA (2003) Gradual development of the interferon-gamma response of swine to porcine reproductive and respiratory syndrome virus infection or vaccination. *Virology* 309:18–31
- Miguel JC, Chen J, Van Alstine WG, Johnson RW (2010) Expression of inflammatory cytokines and Toll-like receptors in the brain and respiratory tract of pigs infected with porcine reproductive and respiratory syndrome virus. *Vet Immunol Immunopathol* 135:314–319
- Petry DB, Lunney J, Boyd P, Kuhar D, Blankenship E, Johnson RK (2007) Differential immunity in pigs with high and low responses to porcine reproductive and respiratory syndrome virus infection. *J Anim Sci* 85:2075–2092
- Wang G, Song T, Yu Y, Liu Y, Shi W, Wang S, Rong F, Dong J, Liu H, Cai X, Zhou EM (2011) Immune responses in piglets infected with highly pathogenic porcine reproductive and respiratory syndrome virus. *Vet Immunol Immunopathol* 142:170–178
- Wesley RD, Lager KM, Kehrli ME, Jr (2006) Infection with porcine reproductive and respiratory syndrome virus stimulates an early gamma interferon response in the serum of pigs. *Can J Vet Res* 70:176–182
- Lowe JE, Husmann R, Firkins LD, Zuckermann FA, Goldberg TL (2005) Correlation of cell-mediated immunity against porcine reproductive and respiratory syndrome virus with protection against reproductive failure in sows during outbreaks of porcine reproductive and respiratory syndrome in commercial herds. *J Am Vet Med Assoc* 226:1707–1711
- Albina E, Carrat C, Charley B (1998) Interferon-alpha response to swine arterivirus (PoAV), the porcine reproductive and respiratory syndrome virus. *J Interferon Cytokine Res* 18:485–490
- Baumann A, Mateu E, Murtaugh MP, Summerfield A (2013) Impact of genotype 1 and 2 of porcine reproductive and respiratory syndrome viruses on interferon-alpha responses by plasmacytoid dendritic cells. *Vet Res* 44:33
- Calzada-Nova G, Schnitzlein WM, Husmann RJ, Zuckermann FA (2011) North American porcine reproductive and respiratory syndrome viruses inhibit type I interferon production by plasmacytoid dendritic cells. *J Virol* 85:2703–2713
- Colonna M, Trinchieri G, Liu YJ (2004) Plasmacytoid dendritic cells in immunity. *Nat Immunol* 5:1219–1226
- Dwivedi V, Manickam C, Binjawadagi B, Linhares D, Murtaugh MP, Renukaradhya GJ (2012) Evaluation of immune responses to porcine reproductive and respiratory syndrome virus in pigs during early stage of infection under farm conditions. *Viol J* 9:45
- Delputte PL, Van Breedam W, Barbe F, Van Reeth K, Nauwynck HJ (2007) IFN-alpha treatment enhances porcine Arterivirus infection of monocytes via upregulation of the porcine Arterivirus receptor sialoadhesin. *J Interferon Cytokine Res* 27:757–766
- Souza CJH, Choi I, Araujo KPC, Abrams SM, Kerrigan M, Rowland RR, Lunney JK (2013) Comparative serum immune responses of pigs after a challenge with porcine reproductive and respiratory syndrome virus (PRRSV). In: Proceedings of the 10th IVS International Veterinary Immunology Symposium. Milan, Italy, p 47