Review Article Benefits of Hesperidin for Cutaneous Functions

Mao-Qiang Man (),^{1,2} Bin Yang (),¹ and Peter M. Elias²

¹Dermatology Hospital, Southern Medical University, Guangzhou 510091, China ²Department of Dermatology, University of California San Francisco and Veterans Affairs Medical Center, San Francisco, CA 94121, USA

Correspondence should be addressed to Mao-Qiang Man; mqman@hotmail.com

Received 22 January 2019; Accepted 19 March 2019; Published 2 April 2019

Academic Editor: Anderson Luiz-Ferreira

Copyright © 2019 Mao-Qiang Man et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Hesperidin is a bioflavonoid, with high concentration in citrus fruits. In addition to its well-known benefits for cardiovascular function, type II diabetes, and anti-inflammation, recent studies have demonstrated multiple benefits of hesperidin for cutaneous functions, including wound healing, UV protection, anti-inflammation, antimicrobial, antiskin cancer, and skin lightening. In addition, hesperidin enhances epidermal permeability barrier homeostasis in both normal young and aged skin. The mechanisms by which hesperidin benefits cutaneous functions are attributable to its antioxidant properties, inhibition of MAPK-dependent signaling pathways, and stimulation of epidermal proliferation, differentiation, and lipid production. Because of its low cost, wide availability, and superior safety, hesperidin could prove useful for the management of a variety of cutaneous conditions.

1. Introduction

In humans, no organ has attracted as much attention as the skin does, because of both cosmetic and medical concerns. For cosmetic concerns, average daily costs of facial care for an American woman can be as much as \$8.00 [1]. In 2017, the sale value of skin care products exceeds \$26 billion per year in China alone [2]. Recent studies showed that topical applications of certain skin care products exert a variety of benefits for both chronic and photoaged skin, antimicrobials, and anti-inflammation [3–7]. Because use of skin care products has become increasingly popular, much work has been focused on the identification of ingredients with multiple benefits on the skin in the development of skin care products.

Because skin suffers from as many diseases as any other organ in the body, proper management of cutaneous conditions is of substantial importance. Over a lifetime, everyone will eventually suffer from some cutaneous problems, because the skin interfaces with the environment, making it more vulnerable to external physical, chemical, and microbial stress. In addition to their psychosocial impact and the quality of life for affected patients and their families, certain chronic cutaneous disorders can also contribute to the development of other systemic diseases. For example, both psoriasis and eczematous dermatitis increase circulating levels of proinflammatory cytokines [8-10], which appear to play a pathogenic role in the development of cardiovascular diseases, obesity, type II diabetes, and Alzheimer's disease [11-14]. Because of its vast size, even subclinical inflammation in the skin can dramatically increase serum cytokine levels, which could be linked to some of these age-associated disorders [12, 15]. Due to the complexity of cutaneous functions and the potential risk of developing multiple disorders in the skin, ingredients that exert multiple benefits to the skin are much desirable. In search for these ingredients, hesperidin would appear to be a potential candidate. Studies have demonstrated that both topical and systemic administrations of hesperidin can benefit a variety of cutaneous functions in both normal and diseased skin. In this review, we comprehensively summarize the benefits of hesperidin for cutaneous functions.

2. Sources and Chemical Properties of Hesperidin

Hesperidin was first isolated from the inner portion of orange peels in 1828. Hesperidin together with other similar bioflavonoids was formerly called "vitamin P" (reviewed in [16]). Hesperidin is abundant in citrus fruits, including lemon, orange, lime, and grapefruit. The content of hesperidin in citrus fruits varies greatly with species, part of the fruit itself, geographic sites of cultivation, and processing procedures (Table 1) [17-22]. For example, hesperidin content in fresh Satsuma pulp is 73 mg per kilogram and 157 mg per kilogram in fresh peel [20]. Generally, hesperidin content is higher in citrus peel than in the other parts of the citrus fruits. But lemon seeds contain more hesperidin than peel by methanol extraction [23]. Hand-squeezed Florida orange juice contains 335-351mg hesperidin per litter while Israel Ortanique citrus juice contains 273-287 mg per litter [24]. Juice from pigmented citrus contains more hesperidin than that from nonpigmented citrus [25]. It is likely that immature citrus may contain more hesperidin than ripen citrus does [26]. Pasteurization with heat did not decrease hesperidin content in citrus juice at least stored at 4°C for up to 12 days. Instead, hesperidin content increases following pasteurization of citrus juice at 90°C for 20 seconds [27]. Hesperidin content ranges from 555 to 761 mg per litter in single-strength juice and from 470 to 614 mg per litter in concentrated juice, suggesting that processing procedure affects hesperidin content in citrus juice [28]. In addition to citrus fruits, peppermint (Mentha x piperita L.) also contains hesperidin, whose content increases following UVB irradiation [29]. Methanol extract of Porphyra dentata, a red

edible seaweed, contains 5% hesperidin [30]. Hesperidin (3,5,7 trihydroxyflavanone 7-rhamnoglucoside, C₂₈H₃₄O₁₅) is also named hesperetin 7-rutinoside or 7-O-glycoside hesperitin, with a molecular weight of 610.57. The melting and boiling points of hesperidin are 250-255°C and 576.16°C, respectively. It is stable for at least for 2 years if stored at -20°C. Although hesperidin alone barely dissolves in aqueous solution, it dissolves well in both propylene glycol and poly(ethylene glycol)-400 [71]. Reaction of hesperidin with chitooligosaccharide yields hesperidinchitooligosaccharide complex, which renders it water soluble and further exhibits superior antioxidant activity to hesperidin alone [72]. Moreover, mix of hesperidin and other flavonoids such as theaflavin-3 3'-digallate can increase the solubility of hesperidin in 10% dimethyl sulfoxide [73]. Additionally, alpha glucosyl hesperidin is also water soluble and therefore commonly used in both topical and systemic preparations. Pertinently, the area under the curve for serum hesperidin in rats was over 3-fold higher following orally administrations of glucosyl hesperidin than hesperidin itself [74].

3. Safety

Hesperidin is generally safe for both topical and systemic administrations. Topical applications of 2% hesperidin for 9 days caused no adverse cutaneous reactions in mice [32]. Similarly, intragastrically given Daflon-500 mg, containing 10% hesperidin, at daily dose of 100 mg did not show signs or symptoms of side effects in rats [75]. Likewise, oral administrations of diets, containing either methyl hesperidin or hesperidin, did not show signs or symptoms of side effects in mice and rats, either [76, 77]. Moreover, no adverse events were observed in mice followed daily intraperitoneal injections of phosphorylated hesperidin at dose of 20mg/kg body weight for over 4 weeks [78]. Although one study showed that orally given Daflon-500 mg twice-daily for 60 days caused minor, temporal side effects such as headache and faintness [79], other studies showed that oral Daflon-500 mg is safe in humans [80, 81].

4. Benefits of Hesperidin for Cutaneous Functions

Nowadays, the benefits of bioflavonoids, including hesperidin, on human health have been well appreciated. A large number of studies have demonstrated that systemic administrations of hesperidin exhibit benefits for a variety of diseases, including cardiovascular diseases, diabetes, Alzheimer's disease, and cancer [82–88]. Likewise, the benefits of hesperidin on various cutaneous conditions have also been well illustrated (Table 2).

4.1. Epidermal Permeability Barrier Function. Epidermal permeability barrier, residing in the stratum corneum, prevents movement of agents and water through the stratum corneum. Importantly, recent studies demonstrate that epidermal permeability barrier plays crucial role in the pathogenesis of both cutaneous and possibly systemic disorders [89-91]. Thus, skin care product makers have been striving to develop products that can potently improve epidermal permeability barrier function. Because of the high incidence of adverse cutaneous reactions to skin care products, identification of safe and effective ingredients is becoming emergent [92, 93]. Our group has demonstrated that twice-daily applications of 2% hesperidin to young mouse skin for 6 days accelerated permeability barrier recovery in a model of acute barrier disruption although basal transepidermal water loss rates, stratum corneum hydration, and skin surface pH remained unchanged [31]. Aged skin displays delayed permeability barrier recovery and elevated skin surface pH [94, 95], which both possibly contribute to the development of certain agingassociated disorders. Regimens that can improve epidermal permeability barrier function, particularly at gene levels, are limited. At least on study showed that topical applications of 2% hesperidin twice-daily for 9 days markedly accelerated permeability barrier recovery, along with significant reduction in skin surface pH in aged mice [32]. In addition to normal skin, topical hesperidin can also prevent abnormalities of epidermal permeability barrier induced by topical glucocorticoid in mice. For example, repeated topical applications of glucocorticoids delayed barrier recovery. But if hesperidin was topically given following each glucocorticoid application to mice, abnormalities in both permeability barrier recovery and skin surface pH were normalized [33]. No adverse reactions were observed following topical applications of hesperidin. Taken together, these studies demonstrate that

Sheries	Geographic Site of Cultivation	Extraction Solvent	Peel (ma/a dried)	Ref
C reticulate "Ervithrosa"	Shimen County Hunan China	Methanol	74 736 + 0 845	[17]
C. reticulate "Unshiu"	Chahe, Hubei, China	Methanol	60.540 ± 0.763	[17]
C. reticulate "Unshiu"	Yangshuo County, Guangxi, China	Methanol	70.232 ± 0.487	[17]
C. reticulate "Subcompressa"	Yongquan, Zhejiang, China	Methanol	100.525 ± 1.398	[17]
C. reticulate "Subcompressa"	Huangyan, Zhejiang, China	Methanol	62.678 ± 0.697	[17]
C. reticulate "Chachi"	Gujin, Jiangmen, Guangdong, China	Methanol	62.919 ± 0.543	[17]
C. reticulate "Chachi"	Huicheng, Jiangmen, Guangdong, China	Methanol	59.012 ± 0.787	[17]
C. reticulate "Chachi"	Luokeng, Jiangmen, Guangdong, China	Methanol	74.973 ± 0.845	[17]
C. reticulate "Chachi"	Daze, Jiangmen, Guangdong, China	Methanol	54.075 ± 0.578	[17]
C. reticulate "Chachi"	Yamen, Jiangmen, Guangdong, China	Methanol	88.087 ± 1.062	[17]
C. reticulate "Chachi"	Shuangshui, Jiangmen, Guangdong, China	Methanol	51.921 ± 0.768	[17]
C. reticulate "Chachi"	Siqian Town, Jiangmen, Guangdong, China	Methanol	50.137 ± 0.301	[17]
C. reticulate "Chachi"	Gujing, Jiangmen, Guangdong, China	50% Method	37	[18]
C. reticulate "Chachi"	Meijiang, Jiangmen, Guangdong, China	50% Method	42	[18]
C. reticulate "Chachi"	Wengyuan,Shaoguan, Guangdong, China	50% Method	79	[18]
C. reticulate "Chachi"	Xiaogan, Jiangmen, Guangdong, China	50% Method	30	[18]
C. reticulate "Chachi"	Shuangshui, Jiangmen, Guangdong, China	50% Method	53	[18]
C. reticulate "Chachi"	Wengyuan, Shaoguan, Guangdong, China	50% Method	66	[18]
C. reticulate "Chachi"	Shuangshui, Jiangmen, Guangdong, China	50% Method	46	[18]
C. Sinensis (linn.) Osbeck	North of Iran		5.2-6.2	[19]
C. Sinensis	North of Iran	Doteclation athous first fallowing his mathonsal	5.2-6.1	[19]
C. reticulate Blanco	North of Iran	reubiatum euter mist, tomowed by internation	4.5-6.1	[19]
C. unshiu Marc	North of Iran		1.3-5.8	[19]
C. unshiu Marc	Neretva Valley, Croatia	1 2M HCI in 800% mathanallantar	0.42	[20]
C. reticulate Blanco	Neretva Valley, Croatia	1.21V1 FTC1 111 00/70 111501141101/ WARE1	0.47	[20]
lemon juice	Adana, Turkey		*113.6±11.0/litter	[21]
Pulp removing	Adana, Turkey		$*81.5 \pm 12.7$ mg/litter	[21]
Pasteurization	Adana, Turkey		$*117.4 \pm 32.5$ mg/litter	[21]
lemon	Spain		*536 ± 234 mg/ml	[22]
Grapefruit	Spain		*33 ± 3 mg/ml	[22]
* Content in juice.				

TABLE 1: Hesperidin content in citrus fruits.

Animal Models/Human Diseases/Cells	Treatment	Benefits	Mechanisms	Ref.
Epidermal Permeability Bar	rier Function		↑ Dwoliforeetion.	
Young mice	Topical applications of 2% hesperidin twice-daily for 6 days.	†Acute barrier recovery	$\begin{bmatrix} 1 & 1 & 0 \\ 0 & 1 \end{bmatrix}$	[31]
			Lamenar bouy secretion. ↑ Differentiation;	
Aged mice	Topical applications of 2% hesperidin twice-daily for 9 days.	Acute barrier recovery; ↓ Skin surface pH;	↑ Lipid production; ↑ NHEl and sPLA2 expression:	[32]
	~		↑ Lamellar body formation and secretion	
			\uparrow Proliferation;	
Glucocorticoid-treated	One hour after each topical application of 0.05%	Acute barrier recovery;	Filaggrin expression; I. Lipid processing;	[22]
mice	cionetasoi propionale, 2% nesperium was appued. Territoritoritoritoritoritoritoritoritorit	↓ okin suriace pri;	↑ Antioxidation;	[cc]
	ITEALITICATION WERE INTECTUALLY TOL 2 UAYS		↑ Lamellar body formation and secretion;	
			$\uparrow \beta$ -glucocerebrosidase activity.	

TABLE 2: Benefits of hesperidin on cutaneous functions.

ed.
ıtinu
Cor
ä
TABLE

Animal Models/Human Diseases/Cells	Treatment	Benefits	Mechanisms	Ref.
Protecting UV Irradiation Keratinocytes	Keratinocytes first treated with 50 μ M hesperidin for 1 hr, followed by UVB irradiation (30 mJ/cm ²)	 DNA damage Ipid peroxidation Protein carbonylation A prototic index 	↓ Absorb UVB ↓ Reactive oxygen species ↓ Bcl-2 expression	[34]
	Keratinocytes were treated with hesperidin (220 μ g/ml) for 24 hr, followed by UVA irradiation (10 J/cm ²). Cells were incubated with hesperidin for additional 6 or 24 hr	↑ Cell viability ↓ MDA content ↓ TNF-α, IL-1β and IL-6 mRNA Levels	↑ ↑ SOD activity	[35]
	Keratinocytes were treated with hesperidin (50, 100, 200, 400, 600, 1 000mg/L) for 24 hr, followed by UVB irradiation (15 mJ/cm ²).	L CXCR2 expression	Not determined	[36]
Dermal fibroblasts	Fibroblasts irradiated with UVA at dose 10 J/cm ² and treated with hesperetin containing extract at various concentration for 72 hr	\downarrow Matrix metalloproteinase expression $\downarrow \beta$ -galactosidase expression $\uparrow Collocor historehosio$	Not determined	[37]
	Pretreated fibroblasts with 3 and 30 μM hesperetin glucuronide, followed by UVA irradiation at 500 kJm $^{-2}$	Vecrotic cell death	Not determined	[38]
Miss	Mice treated topically with hesperidin (3 mg/ml) daily for 10 days, 30 min after each application of hesperidin, mice were irradiated with 180 mJ/cm ² UVB.	 U Skin erythema & edema U Epidermal proliferation U Lipid peroxidation Unthal Amore 	1 Catalase and superoxide dismutase activity	[39]
	Mice were treated topically 1% hesperidin methyl chalcone before and after one irradiation with 4.14 mJ/cm ² UVB	Lipid peroxidation	 Nuclear factor erythroid 2-related factor 2; Glutathione peroxidase-1, glutathione reductase & heme oxygenase-1 	[40]
	Hesperidin methyl chalcone at the dose of 300 mg/kg was intraperitoneally given 1 hr before and 7 hr after, irradiation with 4.14 mJ/cm ² UVB	 J Skin edema, neutrophil recruitment and matrix metalloproteinase-9 activity; U Cytokine expression; Myeloperoxidase activity; U Lipid peroxidation 	î Glutathione levels and catalase activity.	[41]
	Orally administered 0.1 mL of water containing 100 mg/kg body weight hesperidin daily, while mice were irradiated 3 times at 48 h intervals per week for 12 weeks. Does of UVB were increased 60 mJ/cm ² per exposure at week 1 to 90 mJ/cm ² at week 7.	↓ Transepidermal water loss; ↓ matrix metalloproteinase-9 expression & activity; ↓ Cytokine expression ↓ wrinkle formation	↓ Phosphorylation of mitogen activated protein kinase & extracellular signal-regulated kinases	[42]
	Single UVB irradiation at dose of 180 m J/cm ²	↓ ⊂yclobutane pyrmume dimers; ↑ n53 expression	Not determined	[43]

5

	TAF	BLE 2: Continued.		
Animal Models/Human Diseases/Cells	Treatment	Benefits	Mechanisms	Ref.
Guinea pigs	The dorsal skin was exposed to UVB 3 times a week (every other day) for 2 consecutive weeks. The total energy dose of UVB was 1 J/cm ² per exposure. One week later, 1% hesperetin was topically applied daily to the hyperpigmented areas (2 mg/cm^2) for 4 successive weeks.	↓ Transepidermal water loss;	Not determined	[44]
Pigmentation	Cells incubated with 20 µg/mL of <i>Citrus</i> extracts or 3-50 µM of hesperetin for 48 hr.	↑ Melanin content; ↑ Tyrosinase protein; ↑ Tyrosinase activity.	↑ Melanogenesis-related proteins; ↑ β-Catenin expression; ↑ Phosphorylated glycogen synthase kinase-3β	[45, 46]
B16 mouse melanoma cells	Cells incubated with hesperidin (32.25mg/mL) for 3 days	Minimum inhibition of melanogenesis	Not determined	[47, 48]
	Cells were treated with hesperidin (50, 100, 200, 400, 600, 1000mg/L) for 24 hr, followed by UVB irradiation (15 mJ/cm ²).	↓ Tyrosinase activity; ↓ Melanin content;	Not determined	[36]
	Cells incubated with 5-20 μM hesperidin for 3 days	↑ Melanin content;	Not determined	[49]
	Cells incubated with 50 μM hesperidin for 3 days	↓ Melanin content; ↓ Tyrosinase protein; ↓ Tyrosinase-related protein 1,2	↓ Melanogenesis-related proteins; ↑ p-Erkl/2; ↑ Proteasome activity	[50]
	Cells incubated with <i>Citrus</i> extracts (12.5, 25.0, and 50.0 μ g/mL) for 3 days	 Tyrosinase protein & activity; Tyrosinase-related protein 1,2 	<pre>↓ Microphthalmia-associated transcription factor (MITF) proteins</pre>	[51]
Human epidermal	Cells incubated with 3-50 $\mu{\rm M}$ of hesperetin for 48 hr.	↑ Melanin content; ↑ Tyrosinase activity	Not determined	[46]
melanocytes	Cells incubated with 50 μ M hesperidin for 3 days Cells incubated with 0 4mo/ml <i>Citrus</i> extracts for 3 days	 ↓ Melanin content; ↓ Tyrosinase activity ↓ Tyrosinase activity 	Not determined Not determined	[50]
	and an antima and an and an and an and a and a and a	the summer of the second secon		[1]

TABLE 2: Continued.

6

	Ref.	[49]	[52]	[53]		[44]	[49]	[52]
	Mechanisms	Not determined	Not determined	Not determined		Not determined	Not determined	Not determined
ABLE 2: Continued.	Benefits	↓ Tyrosinase activity Img/ml of <i>Citrus</i> extracts	induced over 40% inhibition of tyrosinase activity 1.75 mg/ml of calamondin	peel extract, containing hesperidin, induced 90% inhibition of tyrosinase activity		↓ Pigmentation	↓ Pigmentation	>8% increase in skin brightening
17	Treatment	aatic assay of mushroom tyrosinase			The dorsal skin was exposed to UVB 3 times a week (every other day) for 2 consecutive weeks. The total	week later, 1% hesperetin was to four per exposure. One week later, 1% hesperetin was topically applied daily to the hyperpigmented areas (2 mg/cm^2) for 4 successive weeks.	The epidermis was treated topically with 0.2% hesperidin for 14 days	Topical applications of cream containing 0.4 mg/ml <i>Citrus</i> extracts for 56 days
	Animal Models/Human Diseases/Cells	Enzym				Guinea pigs	Reconstructed human epidermis	Humans

Animal Models/Human Diseases/Cells	Treatment	Benefits	Mechanisms	Ref.
Cutaneous Wound Healing Dermal fibroblast	Fibroblasts were incubated with mixture containing 0.05 mg/ml hesperidin for 24 or 96 hr.	↑ Wound closure.	↑ Collagen synthesis	[54]
Diabetic rats	After wound, rats were given oral hesperidin (25-100 mg/kg body weight) for 21 days.	↓ Wound closure.	↑ VEGF-c, Ang-1/Tie-2, TGF-β and Smad-2/3 mRNA expression; ↑ SOD and GSH levels; ↑ MDA and NO levels:	[55]
	After wound, rats were given oral hesperidin (10-80 mg/kg body weight) for 20 days.	1 Wound closure.	↑ VEGFR1 and VEGFR2 levels; ↓ TNFα, IL-6; ↑ SOD and GSH levels; ↓ MDA levels;	[56]
Humans with venous ulcers	Fifteen patients were treated orally with diosmin/hesperidin (450/50 mg, twice daily) for 90 days. Another 15 patients treated with pycnogenol (50 mg orally, 3 times daily) served as controls	No differences in wound healing time between two groups	Not determined	[57]
	Fifty-three patients received Daflon 500 mg, and 52 received placebo for 2 months	↓ Wound healing time; ↓ Hospitalization duration	Not determined	[58]
<i>y</i> irradiated mice	Mice were given oral hesperidin (100mg/kg body weight) once 1 hr before γ irradiation. Wound was made prior to irradiation.	↑ Wound contraction; ↓ Wound healing time.	↑ NO; ↑ DNA synthesis; ↑ Collagen; ↑ Hexosamine; ↑ Densities of bold vessels and fibroblasts	[59]
	Mice given 1, 2, 5 or 10 % of hesperidin ointment topically covering the whole excision wounds, twice daily after exposure to 6 Gy $\tilde{\Gamma}^3$ -radiation until complete healing of wounds.	↑ Wound contraction; ↓ Wound healing time.	Anti-oxidative stress	[60]

TABLE 2: Continued.

	TA	ABLE 2: Continued.		
Animal Models/Human Diseases/Cells	Treatment	Benefits	Mechanisms	Ref.
Inflammation Mouse RAW 264.7 cell line	Incubated with hesperidin (5-250 $\mu { m g/ml})$	Lipopolysaccharide- induced nitric oxide production	Not determined	[30]
	Incubated with hesperidin or hesperetin (40-100 μ M) for 30 min, followed by stimulation with 1 μ g/mL of Lipopolysaccharide	↓ Antioxidative stress; ↓ PGE2; ↓ COX-2 expression; ↓ Nitric oxide production	<pre>↓ NF-κB activation; ↓ JNK1/2 and p38 phosphorylation; ↓ IκBα; ↓ iNOS mRNA; ↓ Antioxidative stress;</pre>	[61]
Keratinocytes	Keratinocytes treated with HES (20 μ g/mL) for 2 hr, followed by incubation with H ₂ O ₂ for 48 hr	↓ IL-8 protein & mRNA; ↓ TNF-α protein & mRNA; ↓ COX-2 expression	UNF-κB activation, phosphorylated IκBα and phosphorylated p38 MAPK	[62]
	Cells were incubated with both heat-killed Propionibacterium acnes and 5-50 μ g/mL of hesperidin for 24 hr.	↓ IL-8 protein & mRNA; ↓ TNF-α protein & mRNA;	Not determined	[63]
Human skin explants	Human skin explants were pre-incubated with hesperidin methyl chalcone (0.2 mg/ml) and then stimulated with SP for 24 hours.	 Proportion of dilated vessels; Total vessel area; IL-8 production. 	Not determined	[64]
Rats	Thirty min prior to carrageenan or dextran injection, hesperidin (50 or 100 mg/kg body weight) was subcutaneous injected	↓ Edema	Not determined	[65]
Mice	Intraperitoneal injection of hesperidin (75 mg/kg), following by subcutaneous injection of carrageenan	↓ Edema	Not determined	[99]
Guinea pigs	Hesperidin (40mg/kg) was orally given 1 hr prior to injection of carrageenan.	↓ Edema	Not determined	[67]

Animal Models/Human Diseases/Cells	Treatment	Benefits	Mechanisms	Ref.
Skin Cancers		DNA fragmentation; Amontation and a second		
A431 human skin	Incubation of cells with hesperetin (10,100,500 $\mu M)$ for 24 hr	and Bax); and Bax);	↑ ERK, JNK, p38, ROS	[68]
		Levels of cyclin 172, DJ, DJ, DJ, DJ, DJ, DJ, DJ, DJ, DJ, DJ		
	Cells treated with hesperidin (10, 25 and 50 μ M) for	↓ Levels of cyclin D, CDK2	↑ ROS	[עס]
	various times	and thymidylate synthase;	↓ ATP content	[20]
		1 Apoptosis		
	Subcutaneous injection of 125 μ l of 1% hesperidin	\downarrow Incidence of tumor and		[02]
Mlice	solution daily I week prior to tumor induction.	number of tumor per	Not determined	[0]
	4	mouse		
Note: SOD: superoxide dismuta cyclooxygenase-2, COX-2; SP: st NH2-terminal kinase; CDK2: c	se; GSH: reduced glutathione: NO: nitric oxide (NO); PGE2: pros thstance P; DMBA: 7,12-dimethylbenz[a] anthracene; TPA: 12-0-tet yclin-dependent kinase 2.	staglandin E2; VEGF: vascular endothelial g tradecanoyl-13-phorbol acetate; ROS: reactive	rowth factor; MPO: myeloperoxidase; MDA: malc : oxygen species; ERK: extracellular signal-regulate	ndialdehyde; COX2: d kinase; JNK: c-Jun

Continued.	
ABLE 2:	
H	

topical hesperidin improves epidermal permeability barrier function in both normal and glucocorticoid-disrupted skin.

4.2. UV-Induced Cutaneous Damage. Because the skin is the outermost layer of the body, it is more vulnerable to ultraviolet (UV) irradiation, leading to the development of photoaging and other cutaneous disorders such as actinic keratosis and skin cancers [96, 97]. Thus, protection against UV irradiation can prevent and/or mitigate UV-induced cutaneous damage. Study showed that treatment of keratinocytes with 50 μ M hesperidin could cause over 70% reduction in apoptotic index induced by UVB irradiation in comparison to vehicle-treated keratinocytes [34]. In addition to UVB, hesperidin can also protect keratinocytes from UVA-induced damage. Li et al. [35] reported that treatment of keratinocytes with hesperidin for 24 hr induced a dose-dependent increase in viability of UVA-irradiated keratinocytes. Pretreatment of keratinocytes with hesperidin at a dose of 220 μ g/ml also significantly reduced UVA-induced oxidative stress and expression levels of proinflammatory cytokines. These data indicate that hesperidin protects keratinocytes from both UVA- and UVB-induced damage.

The wavelength of UVA is 320-400 nm, which can penetrate into the dermis, leading to premature skin aging (photoaging) upon repeated exposure. It appears that hesperidin can also attenuate UVA-induced damage to fibroblasts. Because hesperidin is hydrolyzed to hesperetin by the gut microbiota and absorbed by passive transport in the large intestine [36], some studies use hesperetin instead of hesperidin. Bae et al. [37] reported that treatment of UVA-irradiated human fibroblasts with 0.1% Citrus unshiu peel extract, containing hesperetin (metabolite of hesperidin), decreased expression levels of β -galactosidase, matrix metalloproteinase-1, and the number of senescent cells. Moreover, pretreatment of human fibroblasts with hesperetin glucuronides induced a 25% protection against UV-A-induced necrotic cell death [38].

Hesperidin not only protects cells against UV-induced damage in vitro, but also protects the skin from UVinduced damage in vivo. Pretreatment of mouse skin with topical hesperidin could prevent UVB-induced elevations in cutaneous cytokine expression and lipid peroxidation, while increasing expression levels of antioxidant enzymes such as glutathione peroxidase-1, glutathione reductase, and heme oxygenase-1 in mouse skin, following either single or multiple UVB irradiation [39, 40]. In addition, topical applications of hesperidin also markedly prevented UVB irradiation-induced erythema, edema, and epidermal proliferation [39]. Moreover, intraperitoneal administrations of hesperidin methyl chalcone can prevent UVB irradiationinduced reductions in antioxidant capacity and elevations in both cutaneous cytokine expression and myeloperoxidase activity [41]. Lee et al. [42] reported that daily drinking water containing hesperidin attenuated a number cutaneous abnormalities induced by repeated UVB irradiation, including compromised epidermal permeability barrier, promoted wrinkle formation, increased cytokine expression, and both

expression levels and activity of matrix metalloproteinase-9. Furthermore, pretreatment of mice with topical hesperidin could enhance repair of DNA damage induced by UVB irradiation [43]. Finally, topical hesperetin lowered transepidermal water loss by \approx 50% in guinea pigs subjected to repeated UVB irradiation [44]. Collectively, either topical or oral administrations of hesperidin can protect skin from damage induced by both UVA and UVB irradiation.

4.3. Melanogenesis. For beauty concern, skin whitening is very popular, particularly in Asia. Hesperidin has long been used as a skin whitening agent although the results of its effects on melanogenesis are controversy. Study showed that treatment of murine B16-F10 melanoma cells with 20 μ g/mL citrus extract (containing $362.3 \pm 16.7 \ \mu g/mL$ hesperetin) induced onefold increase in melanin content, while hesperidin alone also increased melanin content by over 20% [45]. Likewise, 50μ M hesperetin increased melanin content by over 80% in murine B16-F10 melanoma cells [46]. In contrast, other studies demonstrated that hesperidin did not affect melanin production in B16F10 murine melanoma cells [47–49]. However, most of other studies showed that both citrus extract and hesperidin inhibited melanogenesis in both murine B16-F10 melanoma cells and human melanocytes [36, 44, 50–53]. For example, treatments with 50μ M hesperidin for 48-72 hr induced 60% reduction in melanin content in murine B16-F10 melanoma cells and ${\approx}30\%$ reduction in human melanocytes [50]. Topical applications of 0.2% hesperidin to reconstructed human epidermis for 14 days reduced pigment by $\approx 25\%$ [49]. In addition, topical applications of hesperetin, a metabolite of hesperidin, lightened skin in UVB-induced hyperpigmentation [44]. Thus, topical applications of hesperidin and its metabolite can reduce epidermal pigmentation in both normal and UVB challenged skin.

4.4. Cutaneous Wound Healing. Cutaneous wounds are very common while management of cutaneous wounds is still a challenge, particularly in certain conditions such as diabetic and venous wounds. A number of studies have demonstrated that hesperidin accelerated wound healing both in *vitro* and in vivo. Wessels et al. [54] reported that addition of a culture medium containing 0.05 % hesperidin for 24 hr accelerated wound closure by 39% in comparison to vehicle control in in vitro scratch models. In diabetic rats, wound almost completely healed (97%) following orally given hesperidin (100 mg/kg body weight) for 21 days while the wound did not close at all in the vehicle-treated controls [55]. In diabetic rats, the benefits of oral hesperidin on wound healing and other biomarkers, including serum glucose and glycated hemoglobin, were comparable to insulin treatments [56]. Besides diabetic wound, treatment of wound in venous insufficient subjects is also troublesome. Although wound healing times were similar in patients treated with oral diosmin/hesperidin (450/50 mg) and with pycnogenol [57], more patients were completely healed in diosmin/hesperidintreated group than in placebo controls (32% versus 13%) after 2-month treatment [58]. Moreover, either topical or oral administrations of hesperidin shortened wound healing time ≈ 3 days in γ ray-irradiated mice [59, 60]. These data demonstrate that either topically or orally given hesperidin can accelerate cutaneous healing under various conditions.

4.5. Inflammation. Benefits of flavonoids on both systemic and local inflammation have been demonstrated [98, 99]. Because nitric oxide is an inflammatory mediator, it is often used as a biomarker to evaluate inflammatory response. In vitro study demonstrated that treatment of mouse RAW 264.7 cells with lipopolysaccharide (LPS) for 24 hr induced over 9-fold increase in nitrite levels. But addition of hesperidin (250 μ g/ml) to culture medium lowered nitrite levels by ≈75% in LPS-treated cells [30]. Yang et al. [61] performed a similar study using hesperetin and its metabolites. The results showed that stimulation of RAW 264.7 cells with LPS markedly increased in both nitric oxide and inducible nitric oxide synthase mRNA levels, which both significantly decreased by coincubation of RAW 264.7 cells with LPS and 10 μ M hesperetin metabolite. Interestingly, hesperetin only at lower dose $(1\mu M)$ lowered nitric oxide and inducible nitric oxide synthase mRNA levels, but hesperetin at dose of 10 μ M had no effect.

The skin serves as the first line of defense against to external stimuli. Keratinocytes can produce and release proinflammatory cytokines upon stimulation [90]. Hesperidin can lower cytokine production in keratinocyte cultures. For instance, prior to challenge with H2O2 (100 µM), treatment of keratinocytes with hesperidin (20 μ g/ml) for 2 hr could inhibit IL-8 and TNF α production by 96% and 78%, respectively [62]. Expression levels of cyclooxygenase-2 (COX-2) protein and mRNA also significantly decreased in keratinocytes cotreated with hesperidin versus treated with alone H₂O₂. Evidence indicates that hesperidin can inhibit bacterial pathogen-induced cytokine production, too. Incubation of keratinocytes with both Propionibacterium acnes and 5-5 μ g/mL of hesperidin for 24 hr inhibited IL-8 and TNF α production by 49% and 71%, respectively, which were comparable to the levels inhibited by dexamethasone treatment [63]. Moreover, pretreatment of hesperidin methyl chalcone (0.2 mg/ml) also dramatically decreased the proportion of dilated vessels (48% inhibition), total vessel area (72% inhibition), and IL-8 production (79% inhibition) in human skin explants following stimulation with substance P [64]. Thirty minutes prior to subcutaneous injection of carrageenan (1%), subcutaneous injection of hesperidin at doses of 50 and 100 mg/kg reduced the paw edema by 47 and 63%, respectively, within 5 hr [65]. Hesperidin at dose of 100 mg/kg also decreased dextraninduced edema by 33%. The efficacy of hesperidin on edema was comparable to that produced by oral indomethacin (10 mg/kg). However, hesperidin did not prevent histamineinduced paw edema. Pelzer et al. [66] reported that intraperitoneal injection of hesperidin could also inhibit carrageenaninduced paw edema by 36 to 40% within 7 hr. One hour prior to edema induction with carrageenan on the paw,

oral administration of hesperidin (40 mg/kg/ body weight) could decrease the edema by 50%, 51%, 63% and 77 %, respectively, while indomethacin (10mg/kg) decreased the edema by 65%, 71%, 72% and 74%, respectively, after 1, 2, 3, and 4 hours [67]. These results indicate that hesperidin can prevent and treat cutaneous inflammation induced by various agents.

4.6. Cutaneous Cancers. In addition to the preventive and therapeutic benefits for other cancers [100, 101], studies showed that hesperidin and its metabolite also benefit cutaneous cancers. Smina et al. [68] showed that treatment of A431 cells with hesperetin at as low as 10 μ M induced DNA fragmentation along with significant increase in Bax, an apoptotic protein, expression while reducing expression levels of cyclin B1, D1, D3, and E1 proteins by over 1-fold. A similar study also demonstrated that incubation of A431 cells with 10 μ M hesperidin induced over 10-fold increase in apoptosis and DNA damage [69]. In vivo study demonstrated that hesperidin can prevent the development of skin tumor. For example, daily subcutaneous injection of 125 μ l of 1% hesperidin 1 week prior to induction of skin tumor by topical 12-O-Tetradecanoylphorbol-13-acetate (TPA) resulted in reductions in tumor incidence by 50% and the number of papillomas per mouse by 48% after 20 weeks of TPA applications [70]. Thus, hesperidin could be an alternative regimen for preventing and treating cutaneous cancers.

4.7. Other Cutaneous Functions. Evidence also indicates benefits of hesperidin on other cutaneous functions. Orally given hesperidin 30 min prior to irradiation with γ ray upregulated expression levels of mRNA for vascular endothelial growth factor by over 25 folds [102]. Hesperidin exhibited antimicrobial activity, including the common pathogens in the cutaneous infections such as *Staphylococcus aureus*, *Candida albicans*, *Candida tropicalis*, and *Streptococcus pyogenes*, with the minimum inhibitory concentration of 8.25% for both *Candida albicans* and *Staphylococcus aureus* [103–106]. A clinical trial on humans showed that orally given hesperidin (500 mg/daily) for 28 days markedly reduced facial roughness and 33% reduction in beta-galactosidase, a biomarker of aging, by 6 months [107].

5. Mechanisms

Although a line of evidence shows that hesperidin benefits a number of cutaneous functions, the underlying mechanisms by which hesperidin acts are unclear yet. It appears that hesperidin and its metabolite act via a variety of mechanisms depending on which function regulated by hesperidin.

5.1. Improvements in Epidermal Permeability Barrier Function. Formation of epidermal permeability barrier is highly regulated by multiple keratinocyte functions, including proliferation, differentiation, lipid production, acidification, and antimicrobial peptide expression [108]. In young mice, topical hesperidin mainly upregulated expression levels of filaggrin and stimulated keratinocyte proliferation [31] while in aged mice, topical hesperidin upregulated expression levels of a whole panel of mRNA associated with epidermal permeability barrier, including sodium/hydrogen exchanger (NHE1), secretory phospholipase A2 (sPLA2), differentiation-related proteins (filaggrin, involucrin, and loricrin), lipid synthetic enzymes (fatty acid synthase; 3hydroxy-3-methyl-glutaryl-coenzyme A reductase), and lipid transport protein (ATP-binding cassette subfamily A member 12) [32]. However, in glucocorticoid-treated mouse epidermis, topical hesperidin dramatically increased filaggrin protein and glutathione reductase mRNA expression, β glucocerebrosidase activity, and epidermal proliferation [33]. Thus, the underlying mechanisms by which topical hesperidin improves epidermal permeability barrier function could be attributable to upregulation of these functions, depending on the skin conditions.

5.2. Protection against UV Irradiation. UV irradiation causes skin damage mainly in three aspects, i.e., oxidative stress, DNA fragmentation, and inflammation. Nuclear factor erythroid 2-related factor 2 (Nrf2) is a mast regulator of antioxidant system in the cells [109, 110]. Nrf2 deficiency accelerated UV irradiation-induced photoaging and inflammation [111, 112] while activation Nrf2 can protect UV irradiation-induced apoptosis and inflammation [113, 114]. In addition to upregulation of Nrf2 expression in the senescent rat heart [115], methylhesperidin, methylated derivative of hesperidin, enhanced translocation of Nrf2 from cytoplasm to nuclear, resulting in upregulation of antioxidantrelated gene expression and reduction in reactive oxygen species, consequently leading to protection of epidermal keratinocytes against UVB-induced damage in keratinocyte cultures [116]. Hesperidin-induced reductions in DNA damage and cytokine expression appear to be due to decreased oxidative stress in UV-irradiated keratinocytes [34, 35]. Moreover, hesperidin inhibited UVB irradiation-induced increase in expression levels of phosphorylation of mitogenactivated protein kinase (MAPK) and extracellular signalregulated kinases (ERK) in mice [42]. Hence, UV protection of hesperidin can be primarily due to upregulation of antioxidant and downregulation of MAPK/ERK signaling pathway.

5.3. Melanogenesis. Skin pigmentation is determined by both melanogenesis and melanosome transport. Proteins involved in melanogenesis include tyrosinase, tyrosinaserelated proteins (TRP) and microphthalmia-associated transcription factor (MITF). Upregulation of expression levels of tyrosinase, TRPs, and MITF can increase melanin production [117, 118]. A number of studies demonstrated that hesperidin decreased expression levels and activity of tyrosinase, TRPs, and MITF in both B16 mouse melanoma cells and human melanocytes [36, 50-52]. Moreover, hesperidin could activate α adrenergic receptor, leading to induction of aggregation of melanophores in B. melanostictus, suggesting that hesperidin-induced skin lightening is mediated by adrenergic receptor [119]. Another mechanism whereby hesperidin lightens skin could be attributable to inhibition of melanosome transport in melanocytes, instead of inhibition

of melanogenesis [49]. Therefore, hesperidin lightens skin via inhibition of both melanogenesis and melanosome transport.

5.4. Acceleration of Cutaneous Wound Healing. Wound healing is involved in cell proliferation and migration and vascular formation. Activation of tumor growth factor beta (TGF- β) signaling and vascular endothelial growth factor (VEGF) expression are crucial for wound healing and restoration of epidermal permeability barrier function [120-123]. Study showed that oral administration of hesperidin (50 mg/kg body weight) increased TGF- β and VEGF-c mRNA expression by over 2-fold in a diabetic model of Sprague Dawley rats [55]. Additionally, expression levels of mRNA for VEGF receptors also increased following oral administrations of hesperidin at a dose of 50 mg/kg body weight [56]. Oxidative stress can impede wound healing in both diabetic and normal conditions while antioxidants can improve wound healing [124-126]. In diabetic rats, orally given hesperidin significantly increased cutaneous SOD and GSH content while reducing MDA content, along with acceleration of cutaneous wound healing, indicating the antioxidant property of hesperidin contributes to its acceleration of cutaneous healing [55, 56].

Inflammatory response is required for wound healing in early phase. Topical applications of cytokines such as recombinant human granulocyte-macrophage colony-stimulating factor accelerated cutaneous wound healing [127–129]. However, excessive inflammation can delay wound healing and potentially cause scar formation [130]. Accordingly, antiinflammation could accelerate cutaneous wound healing [131, 132]. Hesperidin decreased cytokine expression, including TNF α , IL-6, and IL-8, in both rat skin and human keratinocyte cultures [56, 62, 63]. Taken together, hesperidininduced acceleration of cutaneous wound healing can be attributable to upregulating expression of VEGF, antioxidant enzymes, and anti-inflammation.

5.5. Attenuation of Inflammation. Development of inflammation is a complex process involving interactions of a number of molecules in various signaling pathways, including p38 mitogen-activated protein kinase (MAPK) pathway [133]. Inhibition of p38 MAPK signaling pathway can markedly lower expression of IL-1 β and IL-6, IL-8, IL-18, and TNF α , in both macrophage culture and mice [134, 135]. Study showed that, prior to H_2O_2 stimulation, treatment of keratinocytes with hesperidin for 2 hr induced over 50% reduction in NF- κ B and phosphorylated p38 MAPK in comparison with those without pretreatment with hesperidin [62]. Likewise, treatment of mouse RAW 264.7 cells with hesperetin metabolite almost completely reversed lipopolysaccharide-induced increase in NF-kB expression in addition to reductions in phosphorylated p38 MAPK and c-Jun N-terminal kinase 1/2 [61]. Thus, hesperidin-induced inhibition of p38 MAPK signaling pathway could contribute its attenuation of inflammation.

5.6. Treatment of Cutaneous Cancers. Although studies have demonstrated that hesperidin and its metabolite exerted

anticancer property both in vitro an in vivo [68-70], the underlying mechanisms are inconclusive. Zhao et al. [69] showed that treatment of A431 human skin carcinoma cells with hesperidin $(25 \,\mu\text{M})$ for 72 hr induced over 1-fold increase in reactive oxygen species, 40% reduction in intercellular ATP content and 80% reduction in SOD content. Using other cell lines, it has been demonstrated hesperidin can induce endoplasmic reticulum stress and activate caspase-9, caspase-8, and caspase-3 activities [136–138]. Moreover, TGF β -Smad signaling pathway, particularly Smad3, plays a key role in the development of certain cancers [139, 140]. Previous study revealed that oral administrations of hesperidin (100 mg/kg body weight) for 18 weeks inhibited TGF β -Smad3 signaling and prevented development of hepatic cancer [141]. Furthermore, several studies have demonstrated that reduction in the formation of micronucleus could contribute to the anticancer properties of hesperidin, at least, in models of some chemically induced cancers. Hesperidin-induced protection in γ ray-induced DNA damage could also be attributable to the reduction in the formation of micronucleus. Additionally, hesperidin can increase apoptosis of cancer cells via upregulation of peroxisome proliferator-activated receptor γ expression.

Signaling pathways involved in the action of anticancers induced by hesperidin include (a) inhibition of Janus kinase/signal transducer and activator of transcription (JAK/STAT) pathway, activation of which can enhance proliferation, invasion, and metastasis of cancer cells; (b) inhibition of phosphatidylinositol-3-kinase (PI3K)/Akt and the mammalian target of rapamycin (mTOR) pathways, which also play crucial role in proliferation, survivability, invasion and metastasis of cancer cells upon activation; (c) activation of Notch pathway, in which activation of Notch receptors leads to translocation of Notch into nucleus and binding to target genes, resulting in increased apoptosis [142]. Of course, other signaling pathways such as MAPK-ERK, Wingless, and INT-1, NF-κB, and cyclooxygenase-2 pathways have also been proposed to be involved in hesperidin-induced prevention and inhibition of cancers [142]. Therefore, anticancer benefit of hesperidin is likely via multiple mechanisms, including inhibition of TGFβ-Smad3, PI3K/Akt, and JAK/STAT signaling pathways, activation of Notch pathway, reduction in ATP content, and induction of apoptosis.

5.7. Antioxidation. Oxidative stress has been linked to the development of a variety of disorders [143, 144]. As mentioned above, Nrf2 is a key regulator of antioxidant system [109, 110]. In normal condition, Nrf2 is present as Nrf2/Keap1 complex in the cytoplasm and degraded in proteasome. Upon oxidative stress, Nrf2 is separated from Keap1 and enters into nucleus, where Nrf2 can bind to antioxidant response element (ARE) within gene promoter region, leading activation of gene transcription, including reactive oxygen scavenging enzymes such as superoxide dismutase, glutathione peroxidase, glutathione reductase, catalase, and heme oxygenase 1 [109, 145], which all play crucial role in protecting cells from oxidative stress. Previous studies have shown that hesperidin and its metabolite, hesperitin, can increased Nrf2 expression while stimulating degradation of Keap1, resulting

in an increase in nuclear translocation of Nrf2 and production of antioxidant enzymes along with reduction in oxidation [109, 146–148]. Thus, antioxidant property of hesperidin also largely accounts for its benefits in the skin.

In conclusions, either topical or systemic administrations of hesperidin appear to benefit multiple cutaneous functions via divergent mechanisms. Taking citrus juice or other hesperidin-containing products likely could benefit cutaneous functions. However, proper clinical trials are required to validate the benefits of hesperidin for various cutaneous conditions.

Abbreviations

- VEGF: Vascular endothelia growth factor
- SOD: Superoxide dismutase
- GSH: Reduced glutathione
- MDA: Malondialdehyde
- TGF- β : Transform growth factor β
- MAPK: Mitogen-activated protein kinases.

Conflicts of Interest

All authors declare no conflicts of interest.

Authors' Contributions

Mao-Qiang Man originated the concept and literature search and wrote the draft; Bin Yang and Peter M. Elias critically reviewed the manuscript.

Acknowledgments

This work was supported, in part, by the NIH grant AR061106, administered by the Northern California Institute for Research and Education, with resources from the Research Service, Department of Veterans Affairs.

References

 S. Johnson, How much is your face worth? American women average at \$8 per day, 2018,

https://www.huffingtonpost.com/entry/how-much-is-yourface-worth-american-women-average_us_ 58befa65e4b06660f479e594.

- [2] "Market status of skin care products in China," http://china-trade-research.hktdc.com/business-news/article/ 中国消费市场/中国化妆品市场概况/ccm/sc/1/1X000000/ 1X002L09.htm (obtained on December 9th, 2018).
- [3] M. Milani and A. Sparavigna, "Antiaging efficacy of melatoninbased day and night creams: A randomized, split-face, assessorblinded proof-of-concept trial," *Clinical, Cosmetic and Investigational Dermatology*, vol. 11, pp. 51–57, 2018.
- [4] S. J. Choi, S. Lee, K. Kim et al., "Biological effects of rutin on skin aging," *International Journal of Molecular Medicine*, vol. 38, no. 1, pp. 357–363, 2016.
- [5] A. Sethi, T. Kaur, S. K. Malhotra, and M. L. Gambhir, "Moisturizers: The slippery road," *Indian Journal of Dermatology*, vol. 61, no. 3, pp. 279–287, 2016.

- [6] M. Liu, X. Li, X.-Y. Chen, F. Xue, and J. Zheng, "Topical application of a linoleic acid-ceramide containing moisturizer exhibit therapeutic and preventive benefits for psoriasis vulgaris: a randomized controlled trial," *Dermatologic Therapy*, vol. 28, no. 6, pp. 373–382, 2015.
- [7] M. Zasada, R. Debowska, M. Pasikowska, and E. Budzisz, "The assessment of the effect of a cosmetic product brightening the skin of people with discolorations of different etiology," *Journal* of Cosmetic Dermatology, vol. 15, no. 4, pp. 493–502, 2016.
- [8] S. M. Solberg, L. F. Sandvik, M. Eidsheim, R. Jonsson, Y. T. Bryceson, and S. Appel, "Serum cytokine measurements and biological therapy of psoriasis - Prospects for personalized treatment?" *Scandinavian Journal of Immunology*, vol. 88, Article ID e12725, 2018.
- [9] P. M. Brunner, M. Suárez-Fariñas, H. He et al., "The atopic dermatitis blood signature is characterized by increases in inflammatory and cardiovascular risk proteins," *Scientific Reports*, vol. 7, no. 1, Article ID 8707, 2017.
- [10] E. Vakirlis, E. Lazaridou, T. G. Tzellos, S. Gerou, D. Chatzidimitriou, and D. Ioannides, "Investigation of cytokine levels and their association with SCORAD index in adults with acute atopic dermatitis," *Journal of the European Academy of Dermatology and Venereology*, vol. 25, no. 4, pp. 409–416, 2011.
- [11] C. S. Souza, C. C. de Castro, F. R. Carneiro et al., "Metabolic syndrome and psoriatic arthritis among patients with psoriasis vulgaris: Quality of life and prevalence," *The Journal of Dermatology*, vol. 46, no. 1, pp. 3–10, 2019.
- [12] C. Franceschi and J. Campisi, "Chronic inflammation (inflammaging) and its potential contribution to age-associated diseases," *The Journals of Gerontology. Series A, Biological Sciences* and Medical Sciences, vol. 69, supplement 1, pp. s4–s9, 2014.
- [13] A. Mishra and R. D. Brinton, "Inflammation: bridging age, menopause and APOEε4 genotype to alzheimer's disease," *Frontiers in Aging Neuroscience*, vol. 10, p. 312, 2018.
- [14] M. S. Ellulu, I. Patimah, H. Khaza'ai, A. Rahmat, and Y. Abed, "Obesity and inflammation: the linking mechanism and the complications," *Archives of Medical Science*, vol. 4, pp. 851–863, 2017.
- [15] M. Atienza, J. Ziontz, and J. L. Cantero, "Low-grade inflammation in the relationship between sleep disruption, dysfunctional adiposity, and cognitive decline in aging," *Sleep Medicine Reviews*, vol. 42, pp. 171–183, 2018.
- [16] A. Garg, S. Garg, L. J. D. Zaneveld, and A. K. Singla, "Chemistry and pharmacology of the Citrus bioflavonoid hesperidin," *Phytotherapy Research*, vol. 15, no. 8, pp. 655–669, 2001.
- [17] E.-H. Liu, P. Zhao, L. Duan et al., "Simultaneous determination of six bioactive flavonoids in Citri Reticulatae Pericarpium by rapid resolution liquid chromatography coupled with triple quadrupole electrospray tandem mass spectrometry," *Food Chemistry*, vol. 141, no. 4, pp. 3977–3983, 2013.
- [18] X. Ye, F. Song, G. Fan, and F. Wu, "Simultaneous determination of 11 constituents in Citrus reticulate 'Chachi' by high performance liquid chromatography," *Chinese Journal of Chromatography*, vol. 33, no. 4, pp. 423–427, 2015.
- [19] R. Omidbaigi and N. M. Faghih, "Quantitative distribution of hesperidin in Citrus species, during fruit maturation and optimal harvest time," *Natural Product Radiance*, vol. 4, pp. 12– 15, 2004.
- [20] B. Levaj, D.-U. Verica, D. B. Kovačević, and N. Krasnići, "Determination of flavonoids in pulp and peel of mandarin fruits," *Agriculturae Conspectus Scientificus*, vol. 74, no. 3, pp. 221–225, 2009.

- [21] E. Ağçam, A. Akyıldız, and G. A. Evrendilek, "Comparison of phenolic compounds of orange juice processed by pulsed electric fields (PEF) and conventional thermal pasteurisation," *Food Chemistry*, vol. 143, pp. 354–361, 2014.
- [22] B. Abad-García, L. A. Berrueta, S. Garmón-Lobato, A. Urkaregi, B. Gallo, and F. Vicente, "Chemometric characterization of fruit juices from spanish cultivars according to their phenolic compound contents: I. Citrus fruits," *Journal of Agricultural and Food Chemistry*, vol. 60, no. 14, pp. 3635–3644, 2012.
- [23] A. Bocco, M.-E. Cuvelier, H. Richard, and C. Berset, "Antioxidant activity and phenolic composition of citrus peel and seed extracts," *Journal of Agricultural and Food Chemistry*, vol. 46, no. 6, pp. 2123–2129, 1998.
- [24] P. Mouly, E. M. Gaydou, and A. Auffray, "Simultaneous separation of flavanone glycosides and polymethoxylated flavones in citrus juices using liquid chromatography," *Journal of Chromatography A*, vol. 800, no. 2, pp. 171–179, 1998.
- [25] A. R. Proteggente, A. Saija, A. De Pasquale, and C. A. Rice-Evans, "The compositional characterisation and antioxidant activity of fresh juices from sicilian sweet orange (Citrus sinensis L. Osbeck) varieties," *Free Radical Research*, vol. 37, no. 6, pp. 681–687, 2003.
- [26] D. Barreca, E. Bellocco, C. Caristi, U. G. O. Leuzzi, and G. Gattuso, "Flavonoid composition and antioxidant activity of juices from chinotto (Citrus x myrtifolia Raf.) fruits at different ripening stages," *Journal of Agricultural and Food Chemistry*, vol. 58, no. 5, pp. 3031–3036, 2010.
- [27] F. Uçan, E. Ağçam, and A. Akyildiz, "Bioactive compounds and quality parameters of natural cloudy lemon juices," *Journal of Food Science and Technology*, vol. 53, no. 3, pp. 1465–1474, 2016.
- [28] U. Leuzzi, C. Caristi, V. Panzera, and G. Licandro, "Flavonoids in pigmented orange juice and second-pressure extracts," *Journal of Agricultural and Food Chemistry*, vol. 48, no. 11, pp. 5501– 5506, 2000.
- [29] Y. Dolzhenko, C. M. Bertea, A. Occhipinti, S. Bossi, and M. E. Maffei, "UV-B modulates the interplay between terpenoids and flavonoids in peppermint (Mentha×piperita L.)," *Journal of Photochemistry and Photobiology B: Biology*, vol. 100, no. 2, pp. 67–75, 2010.
- [30] K. Kazłowska, T. Hsu, C.-C. Hou, W.-C. Yang, and G.-J. Tsai, "Anti-inflammatory properties of phenolic compounds and crude extract from *Porphyra dentata*," *Journal of Ethnopharmacology*, vol. 128, no. 1, pp. 123–130, 2010.
- [31] M. Hou, M. Man, W. Man et al., "Topical hesperidin improves epidermal permeability barrier function and epidermal differentiation in normal murine skin," *Experimental Dermatology*, vol. 21, no. 5, pp. 337–340, 2012.
- [32] G. Man, T. M. Mauro, Y. Zhai et al., "Topical hesperidin enhances epidermal function in an aged murine model," *Journal of Investigative Dermatology*, vol. 135, no. 4, pp. 1184–1187, 2015.
- [33] G. Man, T. M. Mauro, P. L. Kim et al., "Topical hesperidin prevents glucocorticoid-induced abnormalities in epidermal barrier function in murine skin," *Experimental Dermatology*, vol. 23, no. 9, pp. 645–651, 2014.
- [34] S. R. Hewage, M. J. Piao, K. A. Kang et al., "Hesperidin attenuates ultraviolet b-induced apoptosis by mitigating oxidative stress in human keratinocytes," *Biomolecules & Therapeutics*, vol. 24, no. 3, pp. 312–319, 2016.
- [35] M. Li, X.-F. Lin, J. Lu, B.-R. Zhou, and D. Luo, "Hesperidin ameliorates UV radiation-induced skin damage by abrogation of oxidative stress and inflammatory in HaCaT cells," *Journal of*

Photochemistry and Photobiology B: Biology, vol. 165, pp. 240-245, 2016.

- [36] R. Z. Zhang, W. Y. Zhu, F. Xie, X. S. Ge, and H. L. Jin, "Effect of hesperidin on B16 and HaCaT cell lines irradiated by narrowband-UVB light," *Journal of Clinical Dermatology*, vol. 37, pp. 146–149, 2008.
- [37] J. T. Bae, H. J. Ko, G. B. Kim, H. B. Pyo, and G. S. Lee, "Protective effects of fermented citrus unshiu peel extract against ultraviolet-a-induced photoageing in human dermal fibrobolasts," *Phytotherapy Research*, vol. 26, no. 12, pp. 1851– 1856, 2012.
- [38] A. R. Proteggente, S. Basu-Modak, G. Kuhnle et al., "Hesperetin glucuronide, a photoprotective agent arising from flavonoid metabolism in human skin fibroblasts," *Photochemistry and Photobiology*, vol. 78, no. 3, pp. 256–261, 2003.
- [39] A. Petrova, L. M. Davids, F. Rautenbach, and J. L. Marnewick, "Photoprotection by honeybush extracts, hesperidin and mangiferin against UVB-induced skin damage in SKH-1 mice," *Journal of Photochemistry and Photobiology B: Biology*, vol. 103, no. 2, pp. 126–139, 2011.
- [40] R. M. Martinez, F. A. Pinho-Ribeiro, V. S. Steffen et al., "Topical formulation containing hesperidin methyl chalcone inhibits skin oxidative stress and inflammation induced by ultraviolet B irradiation," *Photochemical & Photobiological Sciences*, vol. 15, no. 4, pp. 554–563, 2016.
- [41] R. M. Martinez, F. A. Pinho-Ribeiro, V. S. Steffen et al., "Hesperidin methyl chalcone inhibits oxidative stress and inflammation in a mouse model of ultraviolet B irradiation-induced skin damage," *Journal of Photochemistry and Photobiology B: Biology*, vol. 148, pp. 145–153, 2015.
- [42] H. J. Lee, A.-R. Im, S.-M. Kim, H.-S. Kang, J. D. Lee, and S. Chae, "The flavonoid hesperidin exerts anti-photoaging effect by downregulating matrix metalloproteinase (MMP)-9 expression via mitogen activated protein kinase (MAPK)-dependent signaling pathways," *BMC Complementary and Alternative Medicine*, vol. 18, no. 1, article no. 39, 2018.
- [43] S. Jin, B. Zhou, and D. Luo, "Hesperidin promotes cyclobutane pyrimidine dimer repair in UVB-exposed mice epidermis," *Irish Journal of Medical Science*, vol. 180, no. 3, pp. 709–714, 2011.
- [44] Y.-H. Tsai, K.-F. Lee, Y.-B. Huang, C.-T. Huang, and P.-C. Wu, "In vitro permeation and in vivo whitening effect of topical hesperetin microemulsion delivery system," *International Journal* of *Pharmaceutics*, vol. 388, no. 1-2, pp. 257–262, 2010.
- [45] Y.-C. Huang, K.-C. Liu, and Y.-L. Chiou, "Melanogenesis of murine melanoma cells induced by hesperetin, a Citrus hydrolysate-derived flavonoid," *Food and Chemical Toxicology*, vol. 50, no. 3-4, pp. 653–659, 2012.
- [46] I. Usach, R. Taléns-Visconti, L. Magraner-Pardo, and J.-E. Peris, "Hesperetin induces melanin production in adult human epidermal melanocytes," *Food and Chemical Toxicology*, vol. 80, pp. 80–84, 2015.
- [47] C. Zhang, Y. Lu, L. Tao, X. Tao, X. Su, and D. Wei, "Tyrosinase inhibitory effects and inhibition mechanisms of nobiletin and hesperidin from citrus peel crude extracts," *Journal of Enzyme Inhibition and Medicinal Chemistry*, vol. 22, no. 1, pp. 91–98, 2007.
- [48] C. Zhang, Y. Lu, L. Tao, X. Tao, X. Su, and D. Wei, "Tyrosinase inhibitory effects and inhibition mechanisms of nobiletin and hesperidin from citrus peel crude extracts," *Journal of Enzyme Inhibition and Medicinal Chemistry*, vol. 22, no. 1, pp. 83–90, 2007.

- [49] B. Kim, J.-Y. Lee, H.-Y. Lee et al., "Hesperidin suppresses melanosome transport by blocking the interaction of rab27amelanophilin," *Biomolecules & Therapeutics*, vol. 21, no. 5, pp. 343–348, 2013.
- [50] H. J. Lee, W. J. Lee, S. E. Chang, and G.-Y. Lee, "Hesperidin, a popular antioxidant inhibits melanogenesis via Erk1/2 mediated MITF degradation," *International Journal of Molecular Sciences*, vol. 16, no. 8, pp. 18384–18395, 2015.
- [51] S. S. Kim, M.-J. Kim, Y. H. Choi et al., "Down-regulation of tyrosinase, TRP-1, TRP-2 and MITF expressions by citrus presscakes in murine B16 F10 melanoma," *Asian Pacific Journal of Tropical Biomedicine*, vol. 3, no. 8, pp. 617–622, 2013.
- [52] S. Kiefer, M. Weibel, J. Smits, M. Juch, J. Tiedke, and N. Herbst, "Citrus flavonoids with skin lightening effects – safety and efficacy studies," *International Journal of Applied Sciences* SOFW, vol. 136, pp. 46–54, 2010.
- [53] S.-N. Lou, M.-W. Yu, and C.-T. Ho, "Tyrosinase inhibitory components of immature calamondin peel," *Food Chemistry*, vol. 135, no. 3, pp. 1091–1096, 2012.
- [54] Q. Wessels, E. Pretorius, C. M. Smith, and H. Nel, "The potential of a niacinamide dominated cosmeceutical formulation on fibroblast activity and wound healing in vitro," *International Wound Journal*, vol. 11, no. 2, pp. 152–158, 2014.
- [55] W. Li, A. D. Kandhare, A. A. Mukherjee, and S. L. Bodhankar, "Hesperidin, a plant flavonoid accelerated the cutaneous wound healing in streptozotocin-induced diabetic rats: Role of TGF-B/SMADS and ANG-1/TIE-2 signaling pathways," *EXCLI Journal*, vol. 17, pp. 399–419, 2018.
- [56] L. Wang, T. He, A. Fu et al., "Hesperidin enhances angiogenesis via modulating expression of growth and inflammatory factor in diabetic foot ulcer in rats," *European Journal of Inflammation*, vol. 16, 2018.
- [57] R. R. Toledo, M. E. R. D. C. Santos, and T. B. Schnaider, "Effect of Pycnogenol on the Healing of Venous Ulcers," *Annals of Vascular Surgery*, vol. 38, pp. 212–219, 2017.
- [58] J. J. Guilhou, O. Dereure, L. Marzin et al., "Efficacy of Daflon 500 mg in venous leg ulcer healing: A double-blind, randomized, controlled versus placebo trial in 107 patients," *Angiology*, vol. 48, no. 1, pp. 77–85, 1997.
- [59] G. C. Jagetia and K. V. N. M. Rao, "Hesperidin treatment abates radiation-induced delay in healing of deep cutaneous excision wound of mice hemi-body exposed to different doses of γradiation," *Clinical Dermatology Dermatitis*, vol. 1, p. 104, 2018.
- [60] G. C. Jagetia and K. V. N. M. Rao, "Topical application of hesperidin, a citrus bioflavanone accelerates healing of full thickness dermal excision wounds in mice exposed to 6 Gy of whole body γ-radiation," *Transcriptomics*, vol. 3, no. 111, 2015.
- [61] H.-L. Yang, S.-C. Chen, K. J. Senthil Kumar et al., "Antioxidant and anti-inflammatory potential of hesperetin metabolites obtained from hesperetin-administered rat serum: An ex vivo approach," *Journal of Agricultural and Food Chemistry*, vol. 60, no. 1, pp. 522–532, 2012.
- [62] P.-D. Moon and H.-M. Kim, "Antiinflammatory effects of traditional Korean medicine, JinPi-tang and its active ingredient, hesperidin in HaCaT cells," *Phytotherapy Research*, vol. 26, no. 5, pp. 657–662, 2012.
- [63] S. Fu, C. Sun, X. Tao, and Y. Ren, "Anti-inammatory effects of active constituents extracted from Chinese medicinal herbs against *Propionibacterium acnes*," *Natural Product Research* (*Formerly Natural Product Letters*), vol. 26, no. 18, pp. 1746–1749, 2012.

- [64] H. Hernandez-Pigeon, L. Garidou, M. Galliano et al., "Effects of dextran sulfate, 4-t-butylcyclohexanol, pongamia oil and hesperidin methyl chalcone on inflammatory and vascular responses implicated in rosacea," *Clinical, Cosmetic and Investigational Dermatology*, vol. 11, pp. 421–429, 2018.
- [65] J. A. Emim, A. B. Oliveira, and A. J. Lapa, "Pharmacological evaluation of the anti-inflammatory activity of a citrus bioflavonoid, hesperidin, and the isoflavonoids, duartin and claussequinone, in rats and mice," *Journal of Pharmacy and Pharmacology*, vol. 46, no. 2, pp. 118–122, 1994.
- [66] L. E. Pelzer, T. Guardia, A. O. Juarez, and E. Guerreiro, "Acute and chronic antiinflammatory effects of plant flavonoids," *Farmaco*, vol. 53, no. 6, pp. 421–424, 1998.
- [67] W. M. Kaidama and R. N. Gacche, "Anti-inflammatory properties of hesperidin in guinea pigs," *International Journal of Pharmacy and Pharmaceutical Research*, vol. 6, pp. 206–217, 2016.
- [68] T. P. Smina, A. Mohan, K. A. Ayyappa, S. Sethuraman, and U. M. Krishnan, "Hesperetin exerts apoptotic effect on A431 skin carcinoma cells by regulating mitogen activated protein kinases and cyclins," *Cellular and Molecular Biology*, vol. 61, no. 6, pp. 92–99, 2015.
- [69] W. Zhao, Y. Chen, and X. Zhang, "Hesperidin-triggered necrosis-like cell death in skin cancer cell line A431 might be prompted by ROS mediated alterations in mitochondrial membrane potential," *International Journal of Clinical and Experimental Medicine*, vol. 11, pp. 1948–1954, 2018.
- [70] B. Berkarda, H. Koyuncu, G. Soybir, and F. Baykut, "Inhibitory effect of Hesperidin on tumour initiation and promotion in mouse skin," *Research in Experimental Medicine*, vol. 198, no. 2, pp. 93–99, 1998.
- [71] M. K. Anwer, R. Al-Shdefat, S. Jamil, P. Alam, M. S. Abdel-Kader, and F. Shakeel, "Solubility of bioactive compound hesperidin in six pure solvents at (298.15 to 333.15) K," *Journal of Chemical & Engineering Data*, vol. 59, no. 6, pp. 2065–2069, 2014.
- [72] R. Cao, Y. Zhao, Z. Zhou, and X. Zhao, "Enhancement of the water solubility and antioxidant activity of hesperidin by chitooligosaccharide," *Journal of the Science of Food and Agriculture*, vol. 98, no. 6, pp. 2422–2427, 2018.
- [73] R. Xu, Y. Cong, M. Zheng, G. Chen, J. Chen, and H. Zhao, "Solubility and modeling of hesperidin in cosolvent mixtures of ethanol, isopropanol, propylene glycol, and n -propanol + water," *Journal of Chemical & Engineering Data*, vol. 63, no. 3, pp. 764–770, 2018.
- [74] M. Yamada, F. Tanabe, N. Arai et al., "Bioavailability of glucosyl hesperidin in rats," *Bioscience, Biotechnology, and Biochemistry*, vol. 70, no. 6, pp. 1386–1394, 2006.
- [75] M. Damon, O. Flandre, F. Michel, L. Perdrix, C. Labrid, and A. Crastes de Paulet, "Effect of chronic treatment with a purified flavonoid fraction on inflammatory granuloma in the rat. Study of prostaglandin E_2 and $F_2\alpha$ and thromboxane B_2 release and histological changes," *Drug Research*, vol. 37, no. 10, pp. 1149–1153, 1987.
- [76] K. Mayumi, T. Seiko, S. Masa-Aki, H. Masao, F. Shoji, and I. Nobuyuki, "Subchronic toxicity study of methyl hesperidin in mice," *Toxicology Letters*, vol. 69, no. 1, pp. 37–44, 1993.
- [77] K. Kawaguchi, T. Mizuno, K. Aida, and K. Uchino, "Hesperidin as an inhibitor of lipases from porcine pancreas and pseudomonas," *Bioscience, Biotechnology, and Biochemistry*, vol. 61, no. 1, pp. 102–104, 1997.

- [78] B. F. Sieve, "A new antifertility factor (a preliminary report)," *Science*, vol. 116, no. 3015, pp. 373–385, 1952.
- [79] S. S. Shoab, J. B. Porter, J. H. Scurr, and P. D. Coleridge-Smith, "Effect of oral micronized purified flavonoid fraction treatment on leukocyte adhesion molecule expression in patients with chronic venous disease: A pilot study," *Journal of Vascular Surgery*, vol. 31, no. 3, pp. 456–461, 2000.
- [80] M. Cospite, "Double-blind, placebo-controlled evaluation of clinical activity and safety of Daflon 500 mg in the treatment of acute hemorrhoids," *Angiology*, vol. 45, no. 6, pp. 566–573, 1994.
- [81] E. Rabe, G. B. Agus, and K. Roztocil, "Analysis of the effects of micronized purified flavonoid fraction versus placebo on symptoms and quality of life in patients suffering from chronic venous disease: From a prospective randomized trial," *International Angiology*, vol. 34, no. 5, pp. 428–436, 2015.
- [82] N. Sugasawa, A. Katagi, H. Kurobe et al., "Inhibition of atherosclerotic plaque development by oral administration of α -glucosyl hesperidin and water-dispersible hesperetin in apolipoprotein E knockout mice," *Journal of the American College of Nutrition*, pp. 1–8, 2018.
- [83] C. Morand, C. Dubray, D. Milenkovic et al., "Hesperidin contributes to the vascular protective effects of orange juice: a randomized crossover study in healthy volunteers," *American Journal of Clinical Nutrition*, vol. 93, no. 1, pp. 73–80, 2011.
- [84] M. Xue, M. O. Weickert, S. Qureshi et al., "Improved glycemic control and vascular function in overweight and obese subjects by glyoxalase 1 inducer formulation," *Diabetes*, vol. 65, no. 8, pp. 2282–2294, 2016.
- [85] F. Homayouni, F. Haidari, M. Hedayati, M. Zakerkish, and K. Ahmadi, "Hesperidin supplementation alleviates oxidative dna damage and lipid peroxidation in type 2 diabetes: a randomized double-blind placebo-controlled clinical trial," *Phytotherapy Research*, vol. 31, no. 10, pp. 1539–1545, 2017.
- [86] A. Justin Thenmozhi, T. R. William Raja, T. Manivasagam, U. Janakiraman, and M. M. Essa, "Hesperidin ameliorates cognitive dysfunction, oxidative stress and apoptosis against aluminium chloride induced rat model of Alzheimer's disease," *Nutritional Neuroscience*, vol. 20, no. 6, pp. 360–368, 2016.
- [87] A. J. Thenmozhi, T. R. W. Raja, U. Janakiraman, and T. Manivasagam, "Neuroprotective effect of hesperidin on aluminium chloride induced alzheimer's disease in wistar rats," *Neurochemical Research*, vol. 40, no. 4, pp. 767–776, 2015.
- [88] A. Ahmadi and A. Shadboorestan, "Oxidative stress and cancer; the role of hesperidin, a citrus natural bioflavonoid, as a cancer chemoprotective agent," *Nutrition and Cancer*, vol. 68, no. 1, pp. 29–39, 2016.
- [89] P. M. Elias, "Primary role of barrier dysfunction in the pathogenesis of atopic dermatitis," *Experimental Dermatology*, vol. 27, no. 8, pp. 847–851, 2018.
- [90] L. Hu, T. M. Mauro, E. Dang et al., "Epidermal dysfunction leads to an age-associated increase in levels of serum inflammatory cytokines," *Journal of Investigative Dermatology*, vol. 137, no. 6, pp. 1277–1285, 2017.
- [91] M. Man, L. Ye, L. Hu, S. Jeong, P. M. Elias, and C. Lv, "Improvements in epidermal function prevent relapse of psoriasis: a selfcontrolled study," *Clinical and Experimental Dermatology*, 2019.
- [92] L. Huang, Y. Zhong, D. Liu et al., "Adverse cutaneous reactions to skin care products on the face vary with age, but not with sex," *Contact Dermatitis*, vol. 79, no. 6, pp. 365–369, 2018.
- [93] M. Kwa, L. J. Welty, and S. Xu, "Adverse events reported to the US food and drug administration for cosmetics and personal

care products," *JAMA Internal Medicine*, vol. 177, no. 8, pp. 1202–1204, 2017.

- [94] P. M. Elias and R. Ghadially, "The aged epidermal permeability barrier: Basis for functional abnormalities," *Clinics in Geriatric Medicine*, vol. 18, no. 1, pp. 103–120, 2002.
- [95] M. Q. Man, S. J. Xin, S. P. Song et al., "Variation of skin surface pH, sebum content and stratum corneum hydration with age and gender in a large chinese population," *Skin Pharmacology and Physiology*, vol. 22, no. 4, pp. 190–199, 2009.
- [96] J. D'Orazio, S. Jarrett, A. Amaro-Ortiz, and T. Scott, "UV radiation and the skin," *International Journal of Molecular Sciences*, vol. 14, no. 6, pp. 12222–12248, 2013.
- [97] A. Amaro-Ortiz, B. Yan, and J. A. D'Orazio, "Ultraviolet radiation, aging and the skin: Prevention of damage by topical cAMP manipulation," *Molecules*, vol. 19, no. 5, pp. 6202–6219, 2014.
- [98] I. Peluso, C. Miglio, G. Morabito, F. Ioannone, and M. Serafini, "Flavonoids and immune function in human: a systematic review," *Critical Reviews in Food Science and Nutrition*, vol. 55, no. 3, pp. 383–395, 2015.
- [99] K. Kawaguchi, H. Maruyama, T. Kometani, and Y. Kumazawa, "Suppression of collagen-induced arthritis by oral administration of the Citrus flavonoid hesperidin," *Planta Medica*, vol. 72, no. 5, pp. 477–479, 2006.
- [100] T. Tanaka, T. Tanaka, M. Tanaka, and T. Kuno, "Cancer chemoprevention by citrus pulp and juices containing high amounts of β-cryptoxanthin and hesperidin," *Journal of Biomedicine and Biotechnology*, vol. 2012, Article ID 516981, 2012.
- [101] H. A. Al-Ashaal and S. T. El-Sheltawy, "Antioxidant capacity of hesperidin from Citrus peel using electron spin resonance and cytotoxic activity against human carcinoma cell lines," *Pharmaceutical Biology*, vol. 49, no. 3, pp. 276–282, 2011.
- [102] G. Haddadi, A. Abbaszadeh, M. Mosleh-Shirazi, M. Okhovat, A. Salajeghe, and Z. Ghorbani, "Evaluation of the effect of hesperidin on vascular endothelial growth factor gene expression in rat skin animal models following cobalt-60 gamma irradiation," *Journal of Cancer Research and Therapeutics*, vol. 14, no. 7, pp. 1098–1104, 2018.
- [103] Ç. K. Karayıldırım, "Characterization and in vitro evolution of antibacterial efficacy of novel hesperidin microemulsion," *Celal Bayar Üniversitesi Fen Bilimleri Dergisi*, vol. 13, pp. 943–947, 2017.
- [104] A. Corciova, C. Ciobanu, A. Poiata et al., "Inclusion complexes of hesperidin with hydroxypropyl-β-cyclodextrin. Physicochemical characterization and biological assessment," *Digest Journal of Nanomaterials and Biostructures*, vol. 9, no. 4, pp. 1623–1637, 2014.
- [105] Z. Cvetnić and S. Vladimir-Knežević, "Antimicrobial activity of grapefruit seed and pulp ethanolic extract," *Acta Pharmaceutica*, vol. 54, no. 3, pp. 243–250, 2004.
- [106] S. A. Nagem, T. K. Mohammud, S. K. Zair, and E. H. Abass, "Recovery of pure Hesperidin from Iraqi Sweet Oranges Peel and study the effect in some bacteria," *Baghdad Science Journal*, vol. 11, pp. 455–460, 2014.
- [107] A. Guéniche, D. Philippe, E. Buyukpamukcu et al., "The natural flavonoid, hesperidin improves skin aging surface parameters following oral intake," *Research Gate*, 2012, https://www.researchgate.net/publication/327832482_ THE_NATURAL_FLAVONOID_HESPERIDIN_IMPROVES_ SKIN_AGING_SURFACE_PARAMETERS_FOLLOWING_ ORAL_INTAKE.

- [108] K. R. Feingold and P. M. Elias, "Role of lipids in the formation and maintenance of the cutaneous permeability barrier," *Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids*, vol. 1841, no. 3, pp. 280–294, 2014.
- [109] T. Nguyen, P. Nioi, and C. B. Pickett, "The Nrf2-antioxidant response element signaling pathway and its activation by oxidative stress," *The Journal of Biological Chemistry*, vol. 284, no. 20, pp. 13291–13295, 2009.
- [110] H. Ikehata and M. Yamamoto, "Roles of the KEAPI-NRF2 system in mammalian skin exposed to UV radiation," *Toxicology* and Applied Pharmacology, vol. 360, pp. 69–77, 2018.
- [111] A. Hirota, Y. Kawachi, M. Yamamoto, T. Koga, K. Hamada, and F. Otsuka, "Acceleration of UVB-induced photoageing in nrf2 gene-deficient mice," *Experimental Dermatology*, vol. 20, no. 8, pp. 664–668, 2011.
- [112] C. L. L. Saw, A. Y. Yang, M.-T. Huang et al., "Nrf2 null enhances UVB-induced skin inflammation and extracellular matrix damages," *Cell & Bioscience*, vol. 4, no. 1, article 39, 2014.
- [113] A. Hirota, Y. Kawachi, K. Itoh et al., "Ultraviolet A irradiation induces NF-E2-related factor 2 activation in dermal fibroblasts: protective role in UVA-induced apoptosis," *Journal of Investigative Dermatology*, vol. 124, no. 4, pp. 825–832, 2005.
- [114] C. L. Saw, M.-T. Huang, Y. Liu, T. O. Khor, A. H. Conney, and A.-N. Kong, "Impact of Nrf2 on UVB-induced skin inflammation/photoprotection and photoprotective effect of sulforaphane," *Molecular Carcinogenesis*, vol. 50, no. 6, pp. 479– 486, 2011.
- [115] J. Elavarasan, P. Velusamy, T. Ganesan, S. K. Ramakrishnan, D. Rajasekaran, and K. Periandavan, "Hesperidin-mediated expression of Nrf2 and upregulation of antioxidant status in senescent rat heart," *Journal of Pharmacy and Pharmacology*, vol. 64, no. 10, pp. 1472–1482, 2012.
- [116] T. Kuwano, M. Watanabe, D. Kagawa, and T. Murase, "Hydrolyzed methylhesperidin induces antioxidant enzyme expression via the nrf2-are pathway in normal human epidermal keratinocytes," *Journal of Agricultural and Food Chemistry*, vol. 63, no. 36, pp. 7937–7944, 2015.
- [117] I. F. D. S. Videira, D. F. Lima Moura, and S. B. L. M. Vasconcelos Magina, "Mechanisms regulating melanogenesis," *Anais Brasileiros de Dermatologia*, vol. 88, no. 1, pp. 76–83, 2013.
- [118] H. Y. Park, M. Kosmadaki, M. Yaar, and B. A. Gilchrest, "Cellular mechanisms regulating human melanogenesis," *Cellular and Molecular Life Sciences*, vol. 66, no. 9, pp. 1493–1506, 2009.
- [119] J. M. Galgut and S. A. Ali, "Hesperidin induced melanophore aggregatory responses in tadpole of bufo melanostictus via aadrenoceptors," *Pharmacologia*, vol. 3, no. 10, pp. 519–524, 2012.
- [120] K. W. Finnson, S. McLean, G. M. Di Guglielmo, and A. Philip, "Dynamics of transforming growth factor beta signaling in wound healing and scarring," *Advances in Wound Care*, vol. 2, no. 5, pp. 195–214, 2013.
- [121] K. E. Johnson and T. A. Wilgus, "Vascular endothelial growth factor and angiogenesis in the regulation of cutaneous wound repair," *Advances in Wound Care*, vol. 3, no. 10, pp. 647–661, 2014.
- [122] T. A. Wilgus, A. M. Matthies, K. A. Radek et al., "Novel function for vascular endothelial growth factor receptor-1 on epidermal keratinocytes," *The American Journal of Pathology*, vol. 167, no. 5, pp. 1257–1266, 2005.
- [123] P. M. Elias, J. Arbiser, B. E. Brown et al., "Epidermal vascular endothelial growth factor production is required for permeability barrier homeostasis, dermal angiogenesis, and the

development of epidermal hyperplasia: Implications for the pathogenesis of psoriasis," *The American Journal of Pathology*, vol. 173, no. 3, pp. 689–699, 2008.

- [124] D. Kido, K. Mizutani, K. Takeda et al., "Impact of diabetes on gingival wound healing via oxidative stress," *PLoS ONE*, vol. 12, Article ID e0189601, 2017.
- [125] M. Schäfer and S. Werner, "Oxidative stress in normal and impaired wound repair," *Pharmacological Research*, vol. 58, no. 2, pp. 165–171, 2008.
- [126] R. M. El-Ferjani, M. Ahmad, S. M. Dhiyaaldeen et al., "In vivo assessment of antioxidant and wound healing improvement of a new schiff base derived Co (ii) complex in rats," *Scientific Reports*, vol. 6, Article ID 38748, 2016.
- [127] L. Bianchi, A. Ginebri, J. H. Hagman, F. Francesconi, I. Carboni, and S. Chimenti, "Local treatment of chronic cutaneous leg ulcers with recombinant human granulocyte-macrophage colony-stimulating factor," *Journal of the European Academy of Dermatology and Venereology*, vol. 16, no. 6, pp. 595–598, 2002.
- [128] K. Remes and T. Rönnemaa, "Healing of chronic leg ulcers in diabetic necrobiosis lipoidica with local granulocytemacrophage colony-stimulating factor treatment," *Journal of Diabetes and its Complications*, vol. 13, no. 2, pp. 115–118, 1999.
- [129] E. Jaschke, A. Zabernigg, and C. Gattringer, "Recombinant human granulocyte-macrophage colony-stimulating factor applied locally in low doses enhances healing and prevents recurrence of chronic venous ulcers," *International Journal of Dermatology*, vol. 38, no. 5, pp. 380–386, 1999.
- [130] R. G. Rosique, M. J. Rosique, and J. A. Farina Junior, "Curbing inflammation in skin wound healing: a review," *International Journal of Inflammation*, vol. 2015, Article ID 316235, 9 pages, 2015.
- [131] M. Streit, Z. Beleznay, and L. R. Braathen, "Topical application of the tumour necrosis factor-α antibody infliximab improves healing of chronic wounds," *International Wound Journal*, vol. 3, no. 3, pp. 171–191, 2006.
- [132] V.-L. Nguyen, C.-T. Truong, B. C. Q. Nguyen et al., "Antiinflammatory and wound healing activities of calophyllolide isolated from Calophyllum inophyllum Linn," *PLoS ONE*, vol. 12, no. 10, Article ID e0185674, 2017.
- [133] T. Zarubin and J. Han, "Activation and signaling of the p38 MAP kinase pathway," *Cell Research*, vol. 15, no. 1, pp. 11–18, 2005.
- [134] J. Westra, B. Doornbos-van der Meer, P. de Boer, M. A. van Leeuwen, M. H. van Rijswijk, and P. C. Limburg, "Strong inhibition of TNF-alpha production and inhibition of IL-8 and COX-2 mRNA expression in monocyte-derived macrophages by RWJ 67657, a p38 mitogen-activated protein kinase (MAPK) inhibitor," Arthritis Research & Therapy, vol. 6, no. 4, pp. R384– R392, 2004.
- [135] W. Song, L. Wei, Y. Du, Y. Wang, and S. Jiang, "Protective effect of ginsenoside metabolite compound K against diabetic nephropathy by inhibiting NLRP3 inflammasome activation and NF-κB/p38 signaling pathway in high-fat diet/streptozotocin-induced diabetic mice," *International Immunopharmacology*, vol. 63, pp. 227–238, 2018.
- [136] Y. Wang, H. Yu, J. Zhang, J. Gao, X. Ge, and G. Lou, "Hesperidin inhibits HeLa cell proliferation through apoptosis mediated by endoplasmic reticulum stress pathways and cell cycle arrest," *BMC Cancer*, vol. 15, no. 1, article no. 682, 2015.
- [137] J. Zhao, Y. Li, J. Gao, and Y. De, "Hesperidin inhibits ovarian cancer cell viability through endoplasmic reticulum stress signaling pathways," *Oncology Letters*, vol. 14, no. 5, pp. 5569– 5574, 2017.

- [138] R. Banjerdpongchai, B. Wudtiwai, P. Khaw-on, W. Rachakhom, N. Duangnil, and P. Kongtawelert, "Hesperidin from Citrus seed induces human hepatocellular carcinoma HepG2 cell apoptosis via both mitochondrial and death receptor pathways," *Tumor Biology*, vol. 37, no. 1, pp. 227–237, 2016.
- [139] V. Syed, "TGF-β signaling in cancer," Journal of Cellular Biochemistry, vol. 117, no. 6, pp. 1279–1287, 2016.
- [140] M. Petersen, E. Pardali, G. Van Der Horst et al., "Smad2 and Smad3 have opposing roles in breast cancer bone metastasis by differentially affecting tumor angiogenesis," *Oncogene*, vol. 29, no. 9, pp. 1351–1361, 2010.
- [141] A. M. Mahmoud, H. M. Mohammed, S. M. Khadrawy, and S. R. Galaly, "Hesperidin protects against chemically induced hepatocarcinogenesis via modulation of Nrf2/ARE/HO-1, PPARγ and TGF-β1/Smad3 signaling, and amelioration of oxidative stress and inflammation," *Chemico-Biological Interactions*, vol. 277, pp. 146–158, 2017.
- [142] I. Liguori, G. Russo, F. Curcio et al., "Oxidative stress, aging, and diseases," *Clinical Interventions in Aging*, vol. 13, pp. 757– 772, 2018.
- [143] D. Giustarini, I. Dalle-Donne, D. Tsikas, and R. Rossi, "Oxidative stress and human diseases: origin, link, measurement, mechanisms, and biomarkers," *Critical Reviews in Clinical Laboratory Sciences*, vol. 46, no. 5-6, pp. 241–281, 2009.
- [144] F. Ahmadinejad, S. G. Møller, M. Hashemzadeh-Chaleshtori, G. Bidkhori, and M.-S. Jami, "Molecular mechanisms behind free radical scavengers function against oxidative stress," *Antioxidants*, vol. 6, no. 3, p. E51, 2017.
- [145] C. Zhu, Y. Dong, H. Liu, H. Ren, and Z. Cui, "Hesperetin protects against H2O2-triggered oxidative damage via upregulation of the Keap1-Nrf2/HO-1 signal pathway in ARPE-19 cells," *Biomedicine & Pharmacotherapy*, vol. 88, pp. 124–133, 2017.
- [146] H. Ren, J. Hao, T. Liu et al., "Hesperetin suppresses inflammatory responses in lipopolysaccharide-induced RAW 264.7 cells via the inhibition of NF-κB and activation of Nrf2/HO-1 pathways," *Inflammation*, vol. 39, no. 3, pp. 964–973, 2016.
- [147] B. Hemanth Kumar, B. Dinesh Kumar, and P. V. Diwan, "Hesperidin, a citrus flavonoid, protects against l-methionineinduced hyperhomocysteinemia by abrogation of oxidative stress, endothelial dysfunction and neurotoxicity in wistar rats," *Pharmaceutical Biology*, vol. 55, no. 1, pp. 146–155, 2017.
- [148] M. S. Aly, S. R. Galaly, N. Moustafa, H. M. Mohammed, S. M. Khadrawy, and A. M. Mahmoud, "Hesperidin protects against diethylnitrosamine/carbon tetrachloride-induced renal repercussions via up-regulation of Nrf2/HO-1 signaling and attenuation of oxidative stress," *Journal of Applied Pharmaceutical Science*, vol. 7, no. 11, pp. 7–14, 2017.