Type 1 diabetes results from an immune assault on  $\beta$ -cells that progresses over time until the remaining  $\beta$ -cells are unable to keep up with demand and the ensuing hyperglycemia leads to clinical diagnosis. Many textbooks state that this occurs when 80% of the islets are destroyed; but in truth, the amount of residual insulin secretion at the time of clinical diagnosis is variable. The standard model is that those with type 1A diabetes continue to undergo immune-mediated  $\beta$ -cell attack after clinical diagnosis, rapidly leading to the complete absence of  $\beta$ -cells (1).

In the last few decades, clinical research data have increasingly been challenging the notion that  $\beta$ -cells are completely destroyed soon after clinical diagnosis. Once individuals receive exogenous insulin, measurement of  $\beta$ -cell function requires assaying C-peptide, which is secreted in equimolar concentrations with insulin from  $\beta$ -cells. Stimulating the  $\beta$ -cell with a standard liquid "mixed meal" allows for assessment of the  $\beta$ -cell's ability to handle daily activities. Controlling for time of day, administration of exogenous insulin, and fasting glucose level, the mixed-meal tolerance test (MMTT) is a highly reproducible and easily performed test (2). We now know that among type 1 diabetic patients enrolled in clinical trials to preserve  $\beta$ -cell function, it is unusual for control or placebotreated subjects starting with a reasonable amount of C-peptide at diagnosis to completely loose function in the first 2 years (3-13). Outside of these highly controlled clinical trial situations, residual C-peptide soon after diagnosis has been well documented (14-16). The SEARCH for Diabetes in Youth Study of antibody-positive youth with diabetes reported that more than 30% of children within the first year of diagnosis have fasting C-peptide values within the fifth percentile of normal healthy adolescents and that 11% of youth 5 or more years from diagnosis have potentially clinically significant fasting C-peptide levels (17). At the other end of the spectrum, 1) the Joslin Medalist Study demonstrated that 64% of individuals who had lived with type 1 diabetes for more than 50 years had measureable C-peptide (18), 2) our data of unselected subjects at least 30 years from diagnosis found detectable levels in 50% of subjects upon initial testing, and 3) others

also found persistence of C-peptide in some individuals with long-standing disease (19). Recent studies using pathologic specimens also note some patchiness to  $\beta$ -cell loss in those who had type 1 diabetes (1). Further, studies in pregnancy have suggested that an increase in  $\beta$ -cell function may occur (20,21). All these data support the concept that some  $\beta$ -cells may survive for a long time and that their function may wax and wane over time. Such data hold out the hope that attenuation of immune destruction could result in resurgence of endogenous islet function even in those with long-standing disease.

COMMENTARY (SEE WANG, LOVEJOY, AND FAUSTMAN, P.465)

Many articles refer to a peak-stimulated C-peptide level of 0.2 pmol/mL as the clinically relevant value. This is due to a post hoc analysis of Diabetes Control and Complications Trial (DCCT) data in which individuals in the intensively treated group who sustained a C-peptide value of at least 0.2 pmol/mL during an MMTT had less hypoglycemia, retinopathy, and proteinuria (22). Since the DCCT excluded individuals whose C-peptide at entry was greater than 0.5 pmol/mL (23), it is not known whether greater levels of C-peptide would have even greater clinical benefit. Other data pointing to the clinical relevance of some endogenous insulin secretion come from islet transplant studies where, despite an inability to sustain glycemic control without exogenous insulin therapy, even limited function of transplanted islets attenuates major hypoglycemic episodes in this population, which is selected for transplant largely due to having hypoglycemic unawareness (24). The threshold value for such clinical relevance is unknown.

The reliability of such reports depends on robust measures of C-peptide. In recent years, the National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK) has sponsored C-peptide assay standardization workshops to assure cross-laboratory reliability of data (25). The assays have been shown to reliably measure C-peptide in plasma at concentrations to a lower level of ~0.03 pmol/mL. The fact that C-peptide is reliably measured in plasma does not, of course, speak to the clinical relevance of the concentrations found.

With this backdrop, in this issue of *Diabetes Care*, Wang et al. (26) report results

from individuals with type 1 diabetes using a highly sensitive C-peptide assay. This assay, performed with ELISA kits from Mercodia AB in Sweden, reportedly can reliably measure C-peptide concentrations to a lower detection limit of 1.5 pmol/L (or 0.0015 pmol/mL). This is  $\sim$ 20–40 times more sensitive than the standard assays. Using fasting serum samples from 182 type 1 diabetic patients recruited over a 10-year period, Wang et al. found that  $\sim$ 79% of subjects within 5 years of diagnosis and 10% between 31 and 40 years from diagnosis have detectable C-peptide in the ranges detectable only by the highly sensitive assay with only two subjects with detectable values who have lived with diabetes more than 40 years. As noted above, this is less than were reported in the Medalist Study, which used standard C-peptide measurements. Thus, while this study tested a large and less highly selected group, these data confirm previous studies that suggest that some  $\beta$ -cell secretion occurs long after diagnosis. Validating this highly sensitive assay in a workshop setting will enable other investigators to confirm these findings in defined populations. An interesting question not directly addressed in this article is the reproducibility of the assay in the same individual over time. There was clear variation in the results in the four subjects repeatedly sampled. While the authors attribute this variation to glycemic status, this is a hypothesis that could be tested by formal assessment under standardized conditions. In our own work, though 50% of subjects had detectable C-peptide in standard assays during arginine stimulation, when the same subjects were retested, this was not consistently confirmed. This variation may be a reflection of the waxing and waning of disease or issues with the assays.

As noted above, even with the conventional C-peptide assays, the clinical relevance of detecting low levels of C-peptide (less than 0.2 pmol/L) in plasma of people with type 1 diabetes is unclear. Wang et al. attempted to address the clinical relevance of the extremely low levels detected in their assay by exploring the relationship of C-peptide and glucose values in both the subjects who had multiple sampling

## Commentary

over time and the cohort of 182 type 1 diabetic patients described above. While these are interesting exploratory analyses, correlations of multiple variables in samples not obtained for the purpose of addressing this question should be interpreted with caution. Formal testing of the hypothesis that very low levels of C-peptide are biologically relevant will require a prospective study design controlling for multiple clinical and demographic variables, standardized testing procedures, and with prespecified outcome measures. Even then, biological relevance does not necessarily equate with clinical relevance.

This article thus serves to highlight the increasing consensus of many studies over the past decades that have found that some  $\beta$ -cells may function long after the clinical diagnosis of type 1 diabetes and that endogenous secretion is clinically important. Unresolved are questions about the clinical relevance of C-peptide less than 0.2 pmol/mL, and whether we can harness small amounts of  $\beta$ -cell function to the clinical benefit of patients.

## CARLA J. GREENBAUM, MD

- From the Diabetes Program, Benaroya Research Institute, Seattle, Washington.
- Corresponding author: Carla J. Greenbaum, cjgreen@ benaroyaresearch.org.

DOI: 10.2337/dc11-2441

© 2012 by the American Diabetes Association. Readers may use this article as long as the work is properly cited, the use is educational and not for profit, and the work is not altered. See http:// creativecommons.org/licenses/by-nc-nd/3.0/ for details.

**Acknowledgments**—No potential conflicts of interest relevant to this article were reported.

The author thanks Srinath Sanda, MD, of the Benaroya Research Institute, for helpful comments in the review of the manuscript.

## 

## References

- Eisenbarth GS. Banting Lecture 2009: An unfinished journey: molecular pathogenesis to prevention of type 1A diabetes. Diabetes 2010;59:759–774
- Greenbaum CJ, Mandrup-Poulsen T, McGee PF, et al.; Type 1 Diabetes TrialNet Research Group; European C-Peptide Trial Study Group. Mixed-meal tolerance test versus glucagon stimulation test for the assessment of beta-cell function in therapeutic trials in type 1 diabetes. Diabetes Care 2008;31:1966–1971

- 3. Gottlieb PA, Quinlan S, Krause-Steinrauf H, et al.; Type 1 Diabetes TrialNet MMF/ DZB Study Group. Failure to preserve betacell function with mycophenolate mofetil and daclizumab combined therapy in patients with new-onset type 1 diabetes. Diabetes Care 2010;33:826–832
- Keymeulen B, Vandemeulebroucke E, Ziegler AG, et al. Insulin needs after CD3antibody therapy in new-onset type 1 diabetes. N Engl J Med 2005;352:2598–2608
- Ludvigsson J, Faresjö M, Hjorth M, et al. GAD treatment and insulin secretion in recent-onset type 1 diabetes. N Engl J Med 2008;359:1909–1920
- Orban T, Bundy B, Becker DJ, et al.; Type 1 Diabetes TrialNet Abatacept Study Group. Co-stimulation modulation with abatacept in patients with recent-onset type 1 diabetes: a randomised, double-blind, placebocontrolled trial. Lancet 2011;378:412– 419
- Pescovitz MD, Greenbaum CJ, Krause-Steinrauf H, et al.; Type 1 Diabetes Trial-Net Anti-CD20 Study Group. Rituximab, B-lymphocyte depletion, and preservation of beta-cell function. N Engl J Med 2009; 361:2143–2152
- Pozzilli P, Pitocco D, Visalli N, et al.; IMDIAB Group. No effect of oral insulin on residual beta-cell function in recentonset type I diabetes (the IMDIAB VII). Diabetologia 2000;43:1000–1004
- 9. Wherrett DK, Bundy B, Becker DJ, et al.; Type 1 Diabetes TrialNet GAD Study Group. Antigen-based therapy with glutamic acid decarboxylase (GAD) vaccine in patients with recent-onset type 1 diabetes: a randomised double-blind trial. Lancet 2011;378:319–327
- Montanya E, Fernandez-Castañer M, Soler J. Improved metabolic control preserved beta-cell function two years after diagnosis of insulin-dependent diabetes mellitus. Diabetes Metab 1997;23:314–319
- 11. Buzzetti R, Cernea S, Petrone A, et al.; DiaPep Trialists Group. C-peptide response and HLA genotypes in subjects with recent-onset type 1 diabetes after immunotherapy with DiaPep277: an exploratory study. Diabetes 2011;60:3067–3072
- Sherry N, Hagopian W, Ludvigsson J, et al.; Protégé Trial Investigators. Teplizumab for treatment of type 1 diabetes (Protégé study): 1-year results from a randomised, placebo-controlled trial. Lancet 2011;378: 487–497
- 13. Martin S, Herder C, Schloot NC, et al.; DIATOR Study Group. Residual beta cell function in newly diagnosed type 1 diabetes after treatment with atorvastatin: the Randomized DIATOR Trial. PLoS ONE 2011;6:e17554
- 14. Mortensen HB, Swift PG, Holl RW, et al.; Hvidoere Study Group on Childhood Diabetes. Multinational study in children and adolescents with newly diagnosed type 1 diabetes: association of age, ketoacidosis, HLA

status, and autoantibodies on residual beta-cell function and glycemic control 12 months after diagnosis. Pediatr Diabetes 2010;11:218–226

- Törn C, Landin-Olsson M, Lernmark A, et al. Prognostic factors for the course of beta cell function in autoimmune diabetes. J Clin Endocrinol Metab 2000;85: 4619–4623
- Törn C, Landin-Olsson M, Lernmark A, et al. Combinations of beta cell specific autoantibodies at diagnosis of diabetes in young adults reflects different courses of beta cell damage. Autoimmunity 2001;33: 115–120
- 17. Greenbaum CJ, Anderson AM, Dolan LM, et al.; SEARCH Study Group. Preservation of beta-cell function in autoantibodypositive youth with diabetes. Diabetes Care 2009;32:1839–1844
- Keenan HA, Sun JK, Levine J, et al. Residual insulin production and pancreatic beta-cell turnover after 50 years of diabetes: Joslin Medalist Study. Diabetes 2010; 59:2846–2853
- Nakanishi K, Kobayashi T, Miyashita H, et al. Relationships among islet cell antibodies, residual beta-cell function, and metabolic control in patients with insulin-dependent diabetes mellitus of long duration: use of a sensitive C-peptide radioimmunoassay. Metabolism 1990;39: 925–930
- 20. Butler AE, Cao-Minh L, Galasso R, et al. Adaptive changes in pancreatic beta cell fractional area and beta cell turnover in human pregnancy. Diabetologia 2010; 53:2167–2176
- 21. Nielsen LR, Rehfeld JF, Pedersen-Bjergaard U, Damm P, Mathiesen ER. Pregnancyinduced rise in serum *C*-peptide concentrations in women with type 1 diabetes. Diabetes Care 2009;32:1052–1057
- 22. Steffes MW, Sibley S, Jackson M, Thomas W. Beta-cell function and the development of diabetes-related complications in the Diabetes Control and Complications Trial. Diabetes Care 2003;26:832–836
- 23. The Diabetes Control and Complications Trial Research Group. The effect of intensive treatment of diabetes on the development and progression of long-term complications in insulin-dependent diabetes mellitus. N Engl J Med 1993;329: 977–986
- 24. Shapiro AM, Ricordi C, Hering BJ, et al. International trial of the Edmonton protocol for islet transplantation. N Engl J Med 2006;355:1318–1330
- 25. Little RR, Rohlfing CL, Tennill AL, et al. Standardization of C-peptide measurements. Clin Chem 2008;54:1023–1026
- 26. Wang L, Lovejoy N, Faustman DL. Persistence of prolonged C-peptide production in type 1 diabetes with an ultrasensitive C-peptide assay. Diabetes Care 2012;35: 465–470