

# Draft Genome Sequences of Histamine- and Non-Histamine-Producing *Photobacterium* Strains

Kristin Bjornsdottir-Butler,<sup>a</sup> Maria Sanchez Leon,<sup>b</sup> Paul V. Dunlap,<sup>c</sup> Ronald A. Benner, Jr.<sup>a</sup>

FDA, Division of Seafood Science and Technology, Gulf Coast Seafood Laboratory, Dauphin Island, Alabama, USA<sup>a</sup>; FDA, Division of Public Health Informatics and Analytics, College Park, Maryland, USA<sup>b</sup>; University of Michigan, Ann Arbor, Michigan, USA<sup>c</sup>

**Histamine-producing bacteria (HPBs) have recently been identified from the marine environment. The identification and characterization of HPBs is important to developing effective mitigation strategies for scombrototoxin fish poisoning. We report here the draft genomes of seven histamine-producing and two non-histamine-producing marine *Photobacterium* strains.**

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Address correspondence to Kristin Bjornsdottir-Butler, [Kristin.Butler@fda.hhs.gov](mailto:Kristin.Butler@fda.hhs.gov).

In the United States, rapid chilling of fish and fish products to  $\leq 4.4^{\circ}\text{C}$  is recommended to control histamine (scombrototoxin) formation (1). *Photobacterium* spp. are ubiquitous in the marine environment and have been isolated from the edible portion of scombrototoxin-containing fish, for example, tuna and mahi-mahi (2, 3). Some strains are able to grow and produce toxic concentrations of histamine at low temperatures (3–5). Indeed, several outbreaks of scombrototoxin poisoning have been linked to *Photobacterium* spp. (5). In addition, variability in histamine production between strains of the same species has been noted (3). Therefore, psychrotrophic histamine-producing *Photobacterium* spp. are of particular concern in the occurrence of scombrototoxin-associated illnesses.

The *Photobacterium* strains sequenced in this study were histamine-producing isolates of *Photobacterium aquimaris* (BS-1 [6] and BS-2 [6]), *Photobacterium phosphoreum* (AK-3 [7], FS-1.1 [7], and FS-3.1 [7]), and *Photobacterium kishitanii* (DSM-2167, *calba*.1.1 [7]) and non-histamine-producing isolates of *Photobacterium aquimaris* (DSMZ-23343), and *Photobacterium damsela* (BT-6 [8]), isolated from various marine sources. The strains were sequenced to confirm their species identifications and the presence or absence of the histidine decarboxylase gene (*hdc*, involved in formation of histamine).

The genomes were sequenced using an Ion PGM sequencer and the Ion OneTouch 2 system with 400-bp reads (Life Technologies, Frederick, MD, USA). Briefly, for DNA purification, single colonies were incubated in 5 ml of Luria 70% seawater (LSW-70) (9) at 20°C with shaking at 200 rpm for 24 h. DNA was extracted with the DNeasy blood and tissue kit according to the manufacturer's instructions (Qiagen, Valencia, CA, USA). DNA concentrations were determined using a Qubit 2.0 fluorometer with the Qubit dsDNA HS assay kit according to the manufacturer's instructions (Life Technologies). DNA was enzymatically fragmented using the Ion Xpress Plus fragment library kit (Life Technologies), and size was determined with E-Gel SizeSelect 2% agarose gels in an E-Gel iBase unit (Life Technologies). The template for the Ion Torrent PGM instrument was prepared with the

Ion PGM Template OT2 400 kit and sequenced with the Ion PGM sequencing kit on an Ion 318 V2 chip according to the manufacturer's instructions (Life Technologies). For each isolate, the genomic sequence single-pass reads were *de novo* assembled using SPAdes software (10) and annotated using the NCBI Prokaryotic Genome Annotation Pipeline ([http://www.ncbi.nlm.nih.gov/genome/annotation\\_prok](http://www.ncbi.nlm.nih.gov/genome/annotation_prok)) (11). Through annotation, 3,971 to 4,212, 4,149 to 4,405, 4,396 to 4,637, and 3,986 genes were identified for the *P. aquimaris*, *P. phosphoreum*, *P. kishitanii*, and *P. damsela* isolates, respectively. The identification of the isolates and the presence of the *hdc* gene was confirmed in the histamine-producing isolates of *P. aquimaris* (BS-1 and BS-2), *P. kishitanii* (DSM-2167 and *calba*.1.1) but not in the histamine-producing isolates of *P. phosphoreum* (AK-3, FS-1.1, FS-3.1).

**Accession number(s).** The draft genome sequences of the three *P. aquimaris*, two *P. kishitanii*, three *P. phosphoreum*, and one *P. damsela* isolate are available in GenBank under the accession numbers [LZEZ00000000](https://www.ncbi.nlm.nih.gov/nucl/100000000), [LZFA00000000](https://www.ncbi.nlm.nih.gov/nucl/100000000), [LZFB00000000](https://www.ncbi.nlm.nih.gov/nucl/100000000), [LZFC00000000](https://www.ncbi.nlm.nih.gov/nucl/100000000), [LZFD00000000](https://www.ncbi.nlm.nih.gov/nucl/100000000), [LZFE00000000](https://www.ncbi.nlm.nih.gov/nucl/100000000), [LZFF00000000](https://www.ncbi.nlm.nih.gov/nucl/100000000), [LZFG00000000](https://www.ncbi.nlm.nih.gov/nucl/100000000), and [LZFH00000000](https://www.ncbi.nlm.nih.gov/nucl/100000000).

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## REFERENCES

1. FDA. 2011. Scombrototoxin (histamine) formation, p. 113–152. *In* Fish and fisheries products hazards and controls guidance, 4th ed. FDA, Silver Spring, MD. <http://www.fda.gov/downloads/Food/GuidanceRegulation/UCM252400.pdf>.
2. Bjornsdottir-Butler K, Bowers JC, Benner RA. 2015. Prevalence and characterization of high histamine-producing bacteria in Gulf of Mexico fish species. *J Food Prot* 78:1335–1342. <http://dx.doi.org/10.4315/0362-028X.JFP-15-012>.
3. Bjornsdottir-Butler K, McCarthy SA, Dunlap PV, Benner RA. 2016. *Photobacterium angustum* and *Photobacterium kishitanii*, psychrotrophic

- high-level histamine-producing bacteria indigenous to tuna. *Appl Environ Microbiol* 82:2167–2176. <http://dx.doi.org/10.1128/AEM.02833-15>.
4. Dalgaard P, Mejlholm O, Christiansen TJ, Huss HH. 1997. Importance of *Photobacterium phosphoreum* in relation to spoilage of modified atmosphere-packed fish products. *Lett Appl Microbiol* 24:373–378. <http://dx.doi.org/10.1046/j.1472-765X.1997.00152.x>.
  5. Kanki M, Yoda T, Ishibashi M, Tsukamoto T. 2004. *Photobacterium phosphoreum* caused a histamine fish poisoning incident. *Int J Food Microbiol* 92:79–87. <http://dx.doi.org/10.1016/j.ijfoodmicro.2003.08.019>.
  6. Urbanczyk H, Furukawa T, Yamamoto Y, Dunlap PV. 2012. Natural replacement of vertically inherited lux-rib genes of *Photobacterium aquimaris* by horizontally acquired homologues. *Environ Microbiol Rep* 4:412–416. <http://dx.doi.org/10.1111/j.1758-2229.2012.00355.x>.
  7. Ast JC, Dunlap PV. 2005. Phylogenetic resolution and habitat specificity of members of the *Photobacterium phosphoreum* species group. *Environ Microbiol* 7:1641–1654. <http://dx.doi.org/10.1111/j.1462-2920.2005.00859.x>.
  8. Urbanczyk H, Ast JC, Kaeding AJ, Oliver JD, Dunlap PV. 2008. Phylogenetic analysis of the incidence of lux gene horizontal transfer in *Vibrionaceae*. *J Bacteriol* 190:3494–3504. <http://dx.doi.org/10.1128/JB.00101-08>.
  9. Dunlap PV, Jiemjit A, Ast JC, Pearce MM, Marques RR, Lavilla-Pitogo CR. 2004. Genomic polymorphism in symbiotic populations of *Photobacterium leiognathi*. *Environ Microbiol* 6:145–158. <http://dx.doi.org/10.1046/j.1462-2920.2003.00548.x>.
  10. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
  11. Klimke W, Agarwala R, Badretdin A, Chetvernin S, Ciufu S, Fedorov B, Kiryutin B, O'Neill K, Resch W, Resenchuk S, Schafer S, Tolstoy I, Tatusova T. 2009. The National Center for Biotechnology Information's Protein Clusters Database. *Nucleic Acids Res* 37:D216–D223. <http://dx.doi.org/10.1093/nar/gkn734>.