

The prevalence of potentially zoonotic intestinal parasites in dogs and cats in Moscow, Russia

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Summary

This study was aimed to determine the prevalence of *Toxocara canis/cati*, *Strongyloides stercoralis*, *Giardia* spp., and *Cryptosporidium* spp., which occur and are potentially zoonotic to humans in domestic dogs and cats in Moscow (Russia). The fecal flotation method and larvae detection by microscopy of a direct feces smear were performed to detect *Toxocara*, *Giardia* spp., and *Cryptosporidium* spp. The total parasitic prevalence in dogs was as follows: *Giardia* spp.: 10.2 % (226/2208), *Cryptosporidium* spp.: 2.7 % (60/2208), *T. canis*: 2 % (45/2208), *S. stercoralis* larvae: 1.1 % (25/2208). The younger animals under were infected more than those over 12 months of age ($p < 0.001$). The prevalence rates were along these lines: *Giardia* spp. (18.2 %), *Cryptosporidium* spp. (5.7 %), *T. canis* (3 %), *S. stercoralis* larvae (2.3 %). The overall prevalence in cats was as follows: *Giardia* spp. - 5.2 % (71/1350), *Cryptosporidium* spp. - 4.8 % (65/1350), *T. cati* - 4.1 % (56/1350). Similarly to dogs, the infection rates were higher in cats under 12 months of age *Giardia* spp. (8.2 %), *Cryptosporidium* spp. (8.6 %), *T. cati* (7.5 %). Analysis of combined infections in dogs revealed the following combinations: *Giardia* spp. and *Cryptosporidium* spp. (35.5 %) larvae of *S. stercoralis* sp. and *Giardia* spp. (32.3 %), *T. canis* and *Giardia* spp. (22.6 %), *T. canis* and *Cryptosporidium* spp. (6.6 %), *T. canis* and *S. stercoralis* and (3.2 %), respectively. In cats, only two coinfections by *Giardia* spp. and *Cryptosporidium* spp. (58.3 %), and *T. cati* with *Giardia* spp. (41.7 %) were noticed. Further research is needed to study the spread of parasitic diseases in pet animals. The data will improve countermeasures to prevent these diseases' spread among animals and humans.
Keywords: cat; dog; *Toxocara canis/cati*; *Strongyloides stercoralis*; *Giardia* spp.; *Cryptosporidium* spp.; infection prevalence; Russia

Introduction

Pets serve as companionship animals and provide emotional support for people worldwide (Luis Enrique *et al.*, 2018). Thus, becoming full members of a family, dogs and cats often relieve loneliness and bring joy to their owners (Ursache *et al.*, 2021; Gillespie & Bradbury, 2017; Luis Enrique *et al.*, 2018), and many pet

owners worldwide exist (Gillespie & Bradbury, 2017; Blanciardi *et al.*, 2004). For instance, in 2019, there were approximately 106.4 million cats as pets in Europe, and there were 4.2 million domestic dogs in Australia in 2017 (Ursache *et al.*, 2021; Gillespie & Bradbury, 2017). There is often a close relationship between humans and pets, especially in urban environments where the same space is shared (Ilic *et al.*, 2017).

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One important challenge is to prevent the spread of parasitic infections (Gillespie & Bradbury, 2017; Palmer *et al.*, 2008; Baneth *et al.*, 2016), where protozoans and helminths may cause gastrointestinal distress, resulting in diarrhea, vomiting, and loss of appetite (Bouzid *et al.*, 2015; Burgess *et al.*, 2017; Ursache *et al.*, 2021). Sometimes very intense infections lead to a dramatic deterioration of animal health, even with fatal outcomes (Ballweber *et al.*, 2010; Luis Enrique *et al.*, 2018). However, the intestinal parasites in dogs and cats proceed more often without clinical manifestation (Bilgic *et al.*, 2020; Moreira *et al.*, 2018; Li *et al.*, 2019; Mircean *et al.*, 2012; Stafford *et al.*, 2020); what represents an epidemiological risk because asymptomatic animals can be a source of infection to humans when potentially zoonotic parasites are involved (Bilgic *et al.*, 2020; Ilic *et al.*, 2016; Nguyen *et al.*, 2022).

Worldwide published research papers show that intestinal parasites in dogs and cats are widespread (Thompson, 2008; Mircean *et al.*, 2012; Liu *et al.*, 2014; Unterkofler *et al.*, 2022; Sweet *et al.*, 2020; Nguyen *et al.*, 2022; Abere *et al.*, 2013; Silva *et al.*, 2020). For example, Sweet *et al.* (2020) reported that the overall prevalence of cat intestinal parasites in the continental United States could range from 0.03 % to 33 %. Nguyen *et al.* (2022) reported that in Vietnam, infestation in dogs reaches 77.7 %.

Toxocara sp. that parasitizes dogs and cats poses a threat to pub-

lic health because it can cause human disease in types such as visceral toxocariasis, neuro-toxocariasis, ocular toxocariasis and latent toxocariasis (Ursache *et al.*, 2021). The protozoa *Cryptosporidium* spp. and *Giardia duodenalis* parasitize in the gastrointestinal tract of humans and other vertebrates (Li *et al.*, 2019). Among the approximately 40 *Cryptosporidium* species, *C. hominis*, *C. parvum*, *C. meleagridis*, *C. canis*, and *C. felis* are the most common in humans (Feng *et al.*, 2018; de Oliveira *et al.*, 2021). So, the potential role of domestic animals as a source of human infection with *Giardia* spp. is the most debated topic (Bouzid *et al.*, 2015; Mircean *et al.*, 2012; Mravcova *et al.*, 2019; Thompson & Monis, 2004; Thompson *et al.*, 2008). It has been confirmed that genotypes A and B affect many animal species and humans, while genotypes C and D are found in dogs and F in cats and are considered species-specific (Feng and Xiao, 2011; Uiterwijk *et al.*, 2020; Jothikumar *et al.*, 2021). According to the Companion Animal Parasite Council (CAPC), immunocompromised people should limit their exposure to *Giardia*-infected pets (CAPC guideline, 2019). *S. stercoralis* is an endemic parasite in tropical and subtropical regions, but recently there have been more confirmed reports of such infections in central and northern Europe, where humans and dogs became infected (Basso *et al.* 2019; Bourgoin *et al.* 2018; Cervone *et al.* 2016; Eydal & Skirnisson 2016; Liberato

Table 1. Prevalence of *Cryptosporidium* spp., *Giardia* spp., *S. stercoralis* and *T.canis* in dogs.

Type of Infection		2018	2019	2020	2021	2018 – 2021
Dogs (≤ 12) total		104	144	303	399	950
<i>Giardia</i> spp.	N	24	30	55	64	173
	%	23.08	20.83	18.15	16.04	18.21
<i>Cryptosporidium</i> spp.	N	5	19	13	18	55
	%	4.81	13.19	4.29	4.51	5.79
<i>T. canis</i>	N	4	5	14	6	29
	%	3.85	3.47	4.62	1.50	3.05
<i>S. stercoralis</i>	N	3	3	7	9	22
	%	2.88	2.08	2.31	2.26	2.32
Dogs (> 12) total		215	285	324	434	1258
<i>Giardia</i> spp.	N	14	12	11	16	53
	%	6.51	4.21	3.40	3.69	4.21
<i>Cryptosporidium</i> spp.	N	0	1	2	2	5
	%	0.00	0.35	0.62	0.46	0.40
<i>T. canis</i>	N	3	6	3	4	16
	%	1.40	2.11	0.93	0.92	1.27
<i>S. stercoralis</i>	N	1	1	1	0	3
	%	0.47	0.35	0.31	0.00	0.24
Dogs total		319	429	627	833	2208
<i>Giardia</i> spp.	N	38	42	66	80	226
	%	11.91	9.79	10.53	9.60	10.24
<i>Cryptosporidium</i> spp.	N	5	20	15	20	60
	%	1.57	4.66	2.39	2.40	2.72
<i>T.canis</i>	N	7	11	17	10	45
	%	2.19	2.56	2.71	1.20	2.04
<i>S. stercoralis</i>	N	4	4	8	9	25
	%	1.25	0.93	1.28	1.08	1.13

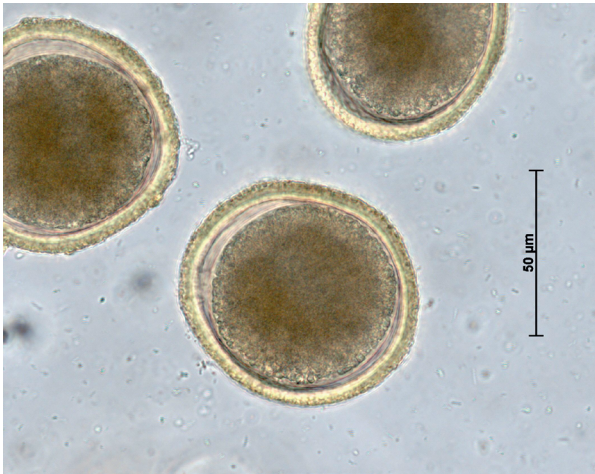


Fig.1. *Toxocara cati* eggs

et al. 2022; Jaleta *et al.* 2017; Raicevic *et al.* 2021; Unterkofler *et al.*, 2022). Recently, the scientific community has been interested in determining the role of helminth canine hosts (i.e., *S. stercoralis*) in the transmission of this infection to humans (Paradies *et al.*, 2017). Data on the epidemiology of canine strongyloidiasis are also limited at this time. Most likely due to the limitations in current diagnostic methods (Paradies *et al.*, 2017).

Humans are infected with zoonotic parasites by consuming contaminated food and water or by direct fecal-oral infection from infected animals (Luis Enrique *et al.*, 2018; Li *et al.*, 2019; Silva *et al.*, 2020). The occurrence of prolonged *Giardia* invasions or disseminated *S. stercoralis* or *Cryptosporidium* spp. infections are likely possible in immunocompromised individuals (Unterkofler *et al.*, 2022; Paradies *et al.*, 2017).

Our work aimed to examine the current situation regarding the prevalence of potentially zoonotic intestinal parasites: *Cryptosporidium* spp. and *Giardia* sp., *S. stercoralis*, and *Toxocara* sp. in domestic dogs and cats in urban environments, and to establish the frequency of coinfections.

Materials and Methods

Two thousand two hundred eight dogs' fecal samples were examined in 2018 – 2021. Sampling included 950 animals aged 1 to 12 months and 1258 over 12 months. Meanwhile, one thousand three hundred fifty cats' fecal samples were examined, where 531 were under the age of 12 months, and 819 were over 12 months of age. The information about the age and likelihood of the animals visiting the outdoor environment was obtained from the animal owners. All examined dogs were kept in apartments and had daily outdoor exposure. Cats were kept solely indoors. In the case of *S. stercoralis* larvae detection, anamnesis regarding the state of the animal and the defecation type was collected from the pet owners. Finally, all data from the owners were processed after receiving their written or verbal consent.

Collected fecal samples were submitted to the «Pasteur» laboratory in Moscow for endoparasite testing. Samples were sent to the laboratory diagnosis when infection symptoms were present or during the therapy outcome control or routine monitoring. The fecal flotation method using a zinc sulfate solution with a density of 1.24, as described by Zajac A (2012), for detecting intestinal parasites was performed. For larvae detection, direct feces smear microscopy was performed. The microscopy was performed with a Lomo microscope at 100X and 400X magnifications (Joint-stock company Lomo, Russia).

Ethical Approval and Informed Consent

The study protocol was reviewed and approved by the scientific and methodological commission of VNIIP - a branch of the Federal State Budget Scientific Institution "Federal Scientific Center VIEV" (Protocol No. 1 dated January 19, 2018). The procedures used in this study are in line with the principles of the Declaration of Helsinki and the European Convention for the Protection of vertebrate animals used for experimental and other scientific purposes. Written informed consent was obtained from the owners for the participation of their animals in this study.

Statistics

Data analysis was performed using the statistical package SPSS version 26.0. We assessed the statistical significance of observed differences in the degree of infestation in animals of two age groups using the Chi-square criterion, with a threshold set at 0.05 (p-value).



Fig.2. Larva of *Strongyloides* sp. in dog, stage L1.

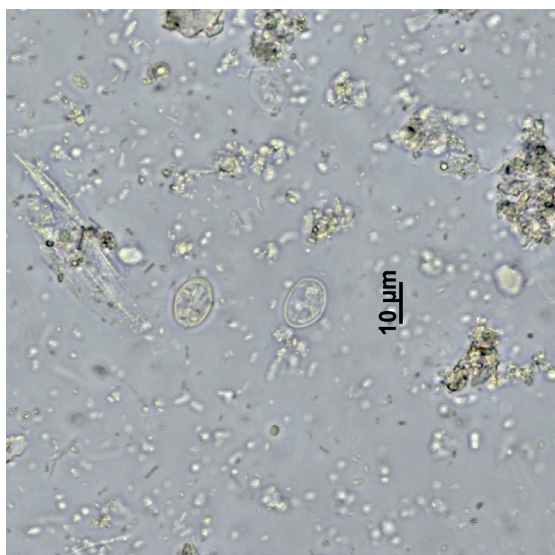


Fig.3 Cysts of *Giardia* spp.

Results

The results showed that *Giardia* spp. cysts (Fig. 3) were detected the most frequently (Table1). The overall prevalence was 10.24 %, with a width of 18.2 % in dogs under 12 months. The infection rates with other parasites were as follows: *Cryptosporidium* spp. - 5.79 % , *T. canis* - 3 % , larvae of *S. stercoralis* (Fig.2) - 2.3 %. The infestation rates in dogs older than 12 months were like this: *Giardia* spp. - 4.21 % , *Cryptosporidium* spp. - 0.4 % , *T. canis* - 1.27 % , and *S. stercoralis* larvae - 0.24 % . Animals under 12

months of age were infected more than those over 12 months of age ($p < 0.001$).

The infection rates in cats were found in this manner (Table 2). *Giardia* spp. - 8.2 % , *Cryptosporidium* spp. - 8.6 % , *T. cati* (Fig.1) - 7.5 % . Cats older than 12 months showed the following prevalences: *Giardia* spp. - 3.3 % , *Cryptosporidium* spp. (Fig.4) - 2.3 % , *T. cati* - 2.3 % . The study revealed that in cats under one year of age, *Cryptosporidium* spp. and *Giardia* spp. were detected at the same level; meanwhile, the *T. cati* eggs were present to a lesser extent. In cats under 12 months of age ($p < 0.001$), the prevalence was higher when compared with older than 12 months cats.

In the analysis of combined infections in dogs, we observed the following (Table 3). Coinfections of *Giardia* spp. and *Cryptosporidium* spp. (35.5 %), larvae of *S. stercoralis* and *Giardia* spp. (32.3 %), *T. canis* and *Giardia* spp. (22.6 %). The combination of *T. canis* and *Cryptosporidium* spp. (6.6 %) or *T. canis* and *S. stercoralis* (3.2 %) in dogs were observed much less frequently. In cats, only two coinfections were found and were caused by *Giardia* spp. and *Cryptosporidium* spp. (58.3 %) or *T. cati* and *Giardia* spp. (41,7 %).

Discussion

Our study was focused on the research of helminths and protozoan prevalence where *T. canis/cati*, *S. stercoralis*, *Giardia* spp., and *Cryptosporidium* spp. in domestic dogs and cats living represent a potential health threat to humans.

Our study has shown that *Giardia* spp. cysts were found most frequently in domestic dogs, especially in young animals up to 12 months old (18.2 %), while the other parasites from the studied

Table 2. Prevalence of *Cryptosporidium* spp., *Giardia* spp. and *T.cati* in cats.

Type of Infection		2018	2019	2020	2021	2018 – 2021
Cats (≤ 12) total		80	98	159	194	531
<i>Giardia</i> spp.	N	10	7	15	12	44
	%	12.50	7.14	9.43	6.19	8.29
<i>Cryptosporidium</i> spp.	N	7	10	16	13	46
	%	8.75	10.20	10.06	6.70	8.66
<i>T. cati</i>	N	4	5	17	14	40
	%	5.00	5.10	10.69	7.22	7.53
Cats (> 12) total		134	198	234	253	819
<i>Giardia</i> spp.	N	2	4	10	11	27
	%	1.49	2.02	4.27	4.35	3.30
<i>Cryptosporidium</i> spp.	N	1	4	9	5	19
	%	0.75	2.02	3.85	1.98	2.32
<i>T. cati</i>	N	1	3	8	5	19
	%	0.75	1.52	3.42	1.58	2.32
Cats total		214	296	393	447	1350
<i>Giardia</i> spp	N	12	11	25	23	71
	%	5.61	3.72	6.36	5.15	5.26
<i>Cryptosporidium</i> spp.	N	8	14	25	18	65
	%	3.74	4.73	6.36	4.03	4.81
<i>T. cati</i>	N	5	8	25	18	56
	%	2.34	2.70	6.36	4.03	4.15

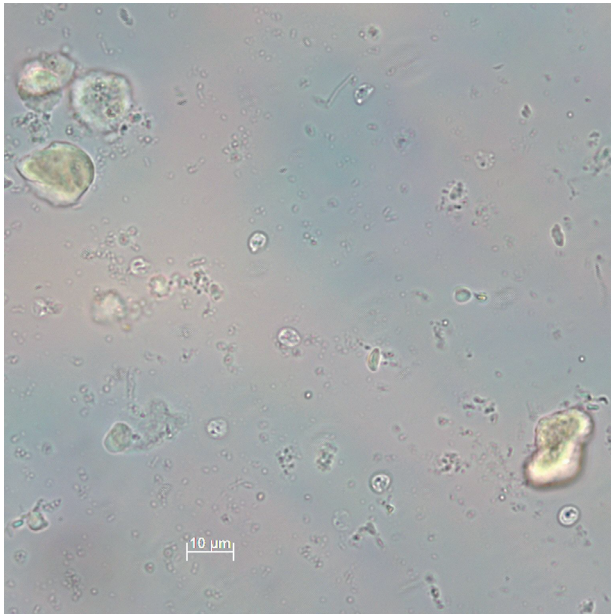


Fig.4. Oocysts of *Cryptosporidium* sp. in cat.

group were detected less often. Many researchers reported similar results (Bouzid *et al.*, 2015; Geurden *et al.*, 2008; Hussein *et al.*, 2017; Liu *et al.*, 2014; Agresti *et al.*, 2022; Piekara-Stepinska *et al.*, 2021). For example, Bouzid *et al.* (2015) reported that the overall prevalence of *Giardia* spp. in dogs and cats is about 15.2 % and 12 %, respectively. Mircea *et al.* (2012) showed that the prevalence of *Giardia* spp. in domestic dogs is up to 4.8 %. According to Li *et al.* (2019), *Giardia* spp. is found in up to 6.9 % of dogs and 9.4 % of cats. We studied domestic animals, so the prevalence of *Giardia* spp. and *Cryptosporidium* was lower than in animals living in the shelters. In shelter dogs, Adeell-Aledon *et al.* (2018) reported *Giardia* spp. infestation up to 40.4 %. Tangtrongsup *et al.* (2020) described the prevalence of *Giardia* spp. up to 25.5 % in dogs and 27.3 % in cats. *Cryptosporidium* in dogs was up to 7.6 % and 12.1 % in cats. Silva (2020) reported a 28 % prevalence of *Giardia* spp. in stray dogs and only 6.2 % in household animals. Despite the high levels of *Giardia* spp. in dogs, we noticed a gradual decrease in infestation intensity during the 2019 – 2021 period. This may be due to the growing awareness of veterinarians and pet owners about the spread of giardiasis. When symptoms of

giardiasis are present, tests are prescribed to exclude them, or anthelmintics against giardiasis are administered for preventive and treatment purposes.

We detected *T. canis* in dogs and cats to a lesser extent than *Giardia* spp. and *Cryptosporidium* spp. (3 % in dogs under 12 months and 7.5 % in cats). Still, the prevalence remained practically at the same level during the observation period. Our data differ significantly from the results of other researchers, where Genchi *et al.* (2021) reported a 25.6 % infection rate of *Toxocara* in cats, while Luis Enrique *et al.* (2018) reported a 25.3 % detection rate of *T. canis* in dogs. This is likely related to the region and category of animals studied. Many researchers have reported a higher prevalence of parasites in animals in countries with hot climates and abundant rainfall. (Li *et al.*, 2019; Liberato *et al.*, 2022; Umar *et al.*, 2017). We believe that preventive deworming prescribed by veterinarians affects the rates of *T. canis* infestation. Thus the infection rate remains low but stable. As Zanzani *et al.* (2014) and Silva *et al.* (2020) noted, introducing preventive deworming can significantly reduce the rate and risk of infection and invasion transmission.

Li *et al.* (2019), Sweet *et al.* (2021), Genchi *et al.* (2021), Uiterwijk *et al.* (2019), and other authors reported that the age of animals, especially if they younger (under 12 months) is a significant infection risk factor. It is valid not only for the *Giardia* but also for the other parasite species with more pronounced clinical manifestations (Sweet *et al.*, 2020; 2021; Luis Enrique *et al.*, 2018; Silva *et al.*, 2020; Liberato *et al.*, 2022). Our studies are aligned with those data and show that in dogs and cats, *T. canis/cati*, *S. stercoralis* larvae (in dogs), *Giardia* spp., and *Cryptosporidium* spp. are more frequent in younger animals. In general, parasites detected in older dogs and cats occurred less frequently. The high level of infection in young animals may be due to the immaturity of their immune systems. In addition, in the case of *Toxocara canis*, the transplacental and transmammary routes of transmission promote its occurrence in puppies (Palmer *et al.*, 2008; Gharekhani, 2014). The detection of *S. stercoralis* larvae in puppies is of particular interest. Despite the low prevalence of these parasites in dogs, their presence cannot be left unattended. When *S. stercoralis* larvae are detected, an unformed mucous stool is observed. According to the owners, some puppies were active, but some had prolonged diarrhea associated with loss of appetite and depression. Although

Table 3. Combination mixed infections in dogs and cats.

Type of co-infection	Type of animal			
	Cats		Dogs	
	N	%	N	%
<i>Giardia</i> spp + <i>Cryptosporidium</i> spp.	7	58.3	11	35.5
<i>S. stercoralis</i> + <i>Giardia</i> spp	-	-	10	32.3
<i>T.canis</i> + <i>Cryptosporidium</i> spp.	-	-	2	6.5
<i>T. cati/canis</i> + <i>Giardia</i> spp	5	41.7	7	22.6
<i>T. canis</i> + <i>S. stercoralis</i>				
Total	12	100	31	100

the endemicity of the disease has been reported, we should consider its recent spread into temperate areas. (Liberato *et al.*, 2022; Unterkofler *et al.*, 2022). However, the limited reports of its prevalence may indicate the lack of feasible detection in dogs. It is due to the unsuitable diagnostic methods used for this parasite laboratory analysis (Unterkofler *et al.*, 2022; Paradies *et al.*, 2017; Umur *et al.*, 2017).

Detection of combined infestations plays a significant role when studying the prevalence of intestinal parasites. In our study, combined infections of two different parasites were observed most frequently in dogs than in cats. This observation in dogs can probably be explained by contact with the external environment, whereas the cats included in this study lived only in the apartments (Kostopoulou *et al.*, 2017; Ursache *et al.*, 2021, Genchi *et al.*, 2021; Rojekittikhun *et al.*, 2014).

It is quite common to observe intestinal parasites in animals without clinical manifestation. We should remember that even asymptomatic parasite carriage can lead to various intestinal pathologies. (Bilgic *et al.*, 2020; Luis Enrique *et al.*, 2018; Uiterwijk *et al.*, 2019; Stafford *et al.*, 2020; Liberato *et al.*, 2022; Moreira *et al.*, 2008). Asymptomatic carrier animals excrete protozoan cysts, larvae, and helminth eggs with feces for a long time. Consequently, they become a source of infestation for healthy animals and cause risks for environmental contamination (Bilgic *et al.*, 2020).

Current discoveries in the biology, genetics, and taxonomy of *Giardia* spp., *Cryptosporidium* spp., and *Strongyloides* sp. isolates obtained from different hosts and their molecular similarities reflect the zoonotic potential of these parasites. (Unterkofler *et al.*, 2022; Li *et al.* 2019; Bilgic *et al.*, 2020; Bahramdoost *et al.*, 2021; Agresti *et al.*, 2022).

Our results show the necessity for ongoing surveillance of the prevalence of pet intestinal parasites sharing the same environment as humans. Our research will further study parasites' genetic identity to understand their zoonotic potential. It will lead to the better implementation of preventive measures against the spread of parasites and improvement of the epizootic situation in the urban environment.

Conflict of Interest

The authors declare that they did not have any conflict of interest in conducting this study. Moreover, the authors do not have any potential conflict of interest pertaining to this submission to Helminthologia.

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References

ABERE, T., BOGALE, B., MELAKU, A. (2013): Gastrointestinal helminth

parasites of pet and stray dogs as a potential risk for human health in Bahir Dar town, north-western Ethiopia. *Vet World*, 6(7): 388. DOI: 10.5455/vetworld.2013.388-392

ADELL-ALEDON, M., KÖSTER, P. C., DE LUCIO, A., PUENTE, P., HERNANDEZ-DE-MINGO, M., SÁNCHEZ-THEVENET, P., DEA-AYUELA, M. A., CARMENA, D. (2018): Occurrence and molecular epidemiology of *Giardia duodenalis* infection in dog populations in eastern Spain. *Vet Res*, 14(1): 26. DOI: 10.1186/s12917-018-1353-z

AGRESTI, A., BERRILLI, F., MAESTRINI, M., GUADANO PROCESI, I., LORETTI, E., VONCI, N., PERRUCCI S. (2021): Prevalence, Risk Factors and Genotypes of *Giardia duodenalis* in Sheltered Dogs in Tuscany (Central Italy). *Pathogens*, 11(1): 12. DOI: 10.3390/pathogens11010012

BAHRAMDOOST, Z., MIRJALALI, H., YAVARI, P., HAGHIGHI, A. (2021): Development of HRM real-time PCR for assemblage characterization of *Giardia lamblia*. *Acta Trop*, (224): 106109. DOI: 10.1016/j.actatropica.2021.106109

BALLWEBER, L.R., XIAO, L.H., BOWMAN, D.D., KAHN, G., CAMA, V.A. (2010): Giardiasis in dogs and cats: update on epidemiology and public health significance. *Trends Parasitol*, 26(4): 180 – 189. DOI: 10.1016/j.pt.2010.02.005

BANETH, G., THAMSBORG, S.M., OTRANTO, D., GUILLOT, J., BLAGA, R., DEPLAZES, P., SOLANO-GALLEGU, L. (2016): Major Parasitic Zoonoses Associated with Dogs and Cats in Europe. *J Comp Pathol*, 155(1): 54 – 74. DOI: 10.1016/j.jcpa.2015.10.179

BASSO, W., GRANDT, L., MAGNENAT, A., GOTTSTEIN, B., CAMPOS, M. (2019): *Strongyloides stercoralis* infection in imported and local dogs in Switzerland: from clinics to molecular genetics. *Parasitol Res*, 118(1): 255 – 266. DOI: 10.1007/s00436-018-6173-3

BILGIC, B., BAYRAKAL, A., DOKUZEYLUL, B., DODURKA, T. (2020): Zoonotic importance of *Giardia* spp. infections in asymptomatic Dogs. *Van Vet J*, 31 (3): 158 – 160. DOI: 10.36483/vanvetj.813479

BLANCIARDI, P., PAPINI, R., GIULIANI, G., CARDINI, G. (2004): Prevalence of *Giardia* antigen in stool samples from dogs and cats. *Revue Med. Vet.*, 155 (8-9): 417 – 421

BOURGOIN, G., JACQUET-VIALLET, P., ZENNER, L. (2018): Fatal strongyloidiasis in a puppy from France. *Vet Rec Case Rep*, 6(2): e000415. DOI: 10.1136/vetreccr-2016-000415

BOUZID, M., HALAI, K., JEFFREYS, D., HUNTER, P. (2015): The prevalence of *Giardia* infection in dogs and cats, a systematic review and meta-analysis of prevalence studies from stool samples. *Vet Parasitol*, 207(3-4): 181 – 202. DOI: 10.1016/j.vetpar.2014.12.011

BURGESS, L., GILCHRIST, C.A., LYNN, T.C., PETRI JR, W. (2017): Parasitic Protozoa and Interactions with the Host Intestinal Microbiota. *Infect Immun*, 85(8): e00101-17. DOI: 10.1128/IAI.00101-17

COMPANION ANIMAL PARASITE COUNCIL (CAPC) (2019): *Giardia*. Retrieved from <https://capcvet.org/guidelines/giardia/>

CERVONE, M., GIANNELLI, A., OTRANTO, D., PERRUCCI, S. (2016): *Strongyloides stercoralis* hyperinfection in an immunosuppressed dog from France. *Rev. Vet. Clin.*, 51(2): 55 – 59. DOI: 10.1016/j.anicom.2016.05.001

EYDAL, M., SKIRNISSON, K. (2016): *Strongyloides stercoralis* found in

- imported dogs, household dogs and kennel dogs in Iceland. *Icel Agric Sci*, 29(1): 39 – 51. DOI: 10.16886/IAS.2016.04
- FENG, Y., XIAO L. (2011): Zoonotic Potential and Molecular Epidemiology of *Giardia* Species and Giardiasis. *Clin Microbiol Rev*, 24 (1): 110 – 140. DOI: 10.1128/CMR.00033-10
- FENG, Y., RYAN, U., XIAO, L. (2018): Genetic diversity and population structure of *Cryptosporidium*. *Trends Parasitol*, (34):997 – 1011. DOI: 10.1016/j.pt.2018.07.009
- GENCHI, M., VISMARRA, A., ZANET, S., MORELLI, S., GALUPPI, R., CRINGOLI, G., OTRANTO, D. Multicentre Study of Endo-Ectoparasite Infection in Italian Cats. *Parasit Vectors* (preprint under review). DOI: 10.21203/rs.3.rs-558064/v1
- GEURDEN, T., BERKVEN, D., CASAERT, S., VERCRUYSE, J., CLAEREBOUT, E. (2008): A Bayesian evaluation of three diagnostic assays for the detection of *Giardia duodenalis* in symptomatic and asymptomatic dogs. *Vet. Parasitol*, 157(1-2): 14 – 20. DOI: 10.1016/j.vetpar.2008.07.002
- GHAREKHANI, J. (2014): Study on gastrointestinal zoonotic parasites in pet dogs in Western Iran. *Turkiye Parazit Derg*, 38(3): 172 – 176. DOI: 10.5152/tpd.2014.3546
- GILLESPIE, S., BRADBURY, R. S. (2017): A Survey of Intestinal Parasites of Domestic Dogs in Central Queensland. *Trop Med Infect Dis*, 21(4): 60. DOI: 10.3390/tropicalmed2040060
- HUSSEIN, E.M., ISMAIL, O.A., MOKHTAR, A.B., MOHAMED, S.E., SAAD, R.M. (2017): Nested PCR targeting intergenic spacer (IGS) in genotyping of *Giardia duodenalis* isolated from symptomatic infected Egyptian school children. *Parasitol Res*, 116(2): 763 – 771. DOI: 10.1007/s00436-016-5347-0
- ILIC, T., KULISIC, Z., ANTIC, N., RADISAVLJEVIC, K., DIMITRIJEVIC, S. (2017): Prevalence of zoonotic intestinal helminths in pet dogs and cats in the Belgrade area. *J Appl Anim Res*, 45(1): 204 – 208, DOI: 10.1080/09712119.2016.1141779
- JALETA, T., ZHOU, S., BEMM, F., SCHÄR, F., KHIEU, V., MUTH, S., ODERMATT, P., LOK, J., STREIT, A. (2017): Different but overlapping populations of *Strongyloides stercoralis* in dogs and humans-Dogs as a possible source for zoonotic strongyloidiasis. *PLoS Negl Trop Dis*, 11(8): e0005752. DOI: 10.1371/journal.pntd.0005752
- JOTHIKUMAR, N., MURPHY, J., HILL, V. (2021): Detection and identification of *Giardia* species using real-time PCR and sequencing. *J Microbiol Methods*, 189: 106279. DOI: 10.1016/j.mimet.2021.106279
- KOSTOPOULOU, D., CLAEREBOUT, E., ARVANITIS, D., LIGDA, P., VOUTZOURAKIS, N., CASAERT, S., SOTIRAKI, S. (2017): Abundance, zoonotic potential and risk factors of intestinal parasitism amongst dog and cat populations: The scenario of Crete, Greece. *Parasit Vectors*, 10: 43. DOI: 10.1186/s13071-017-1989-8
- LI, J., DAN, X., ZHU, K., LI, N., GUO, Y., ZHENG, Z., FENG, Y., XIAO, L. (2019): Genetic characterization of *Cryptosporidium* spp. and *Giardia duodenalis* in dogs and cats in Guangdong, China. *Parasit Vectors*, 12(1): 571. DOI: 10.1186/s13071-019-3822-z
- LIBERATO, C., IATTA, R., SCARITO, M., GRIFONI, G., DANTE, G., OTRANTO, D. (2022): *Strongyloides stercoralis* in a dog litter: Evidence suggesting a transmammary transmission. *Acta Trop*, 231: 106465. DOI: 10.1016/j.actatropica.2022.106465
- LUIS ENRIQUE, J.P., MORENO, L.R., NÚÑEZ FERNÁNDEZ, F.A., MILLÁN, I.A., RIVERO, L.R., GONZÁLEZ, F.R., PÉREZ RODRÍGUEZ, J.C. (2018): Prevalence of intestinal parasitic infections in dogs from Havana, Cuba: risk of zoonotic infections to humans. *Anim Husb Dairy Vet Sci*, 2(3): 1 – 5. DOI: 10.15761/AHDVS.1000133
- LIU, H., SHEN, Y.J., YUAN, Z.Y., JIANG, Y.Y., XU, Y.Z., PAN, W., HU, Y., CAO, J.P. (2014): Prevalence and genetic characterization of *Cryptosporidium*, *Enterocytozoon*, *Giardia* and *Cyclospora* in diarrheal outpatients in China. *BMC Infect. Dis*, 14: 25. DOI: 10.1186/1471-2334-14-25
- MIRCEA, V., GYÖRKE, A., COZMA, V. (2012): Prevalence and risk factors of *Giardia duodenalis* in dogs from Romania. *Vet Parasitol*, 184(2-4): 325 – 329. DOI: 10.1016/j.vetpar.2011.08.022
- MOREIRA, A., BAPTISTA, C., BRASIL, L., VALENTE, J., BRUHN, F., PEREIRA, D. (2018): Risk factors and infection due to *Cryptosporidium* spp. in dogs and cats in southern Rio Grande do Sul. *Rev Bras Parasitol Vet*, 27(1): 113 – 118. DOI: 10.1590/S1984-296120180012
- MRAVCOVA, K., STRKOLCOVA, G., GOLDOVA, M. (2019): The Prevalence And Assemblages of *Giardia Duodenalis* in Dogs: A Systematic Review in Europe. *Folia Vet*, 63(4): 38 – 45. DOI: 10.2478/fv-2019-0036
- NGUYEN, T., DORNY, P., DINH, T., NGUYEN, V., NGUYEN, H., NGUYEN, T., DAO, H., DERMAUW, V. (2022): Helminth infections in dogs in Phu Tho Province, northern Vietnam. *Curr Res Parasitol Vector Borne Dis*, 2: 100091. DOI: 10.1016/j.crvpbd.2022.100091
- DE OLIVEIRA, A., SUDRE, A., BOMFIM, T., SANTOS, H. (2021) Molecular characterization of *Cryptosporidium* spp. in dogs and cats in the city of Rio de Janeiro, Brazil, reveals potentially zoonotic species and genotype. *PLoS One*, 16(8): e0255087. DOI: 10.1371/journal.pone.0255087
- PALMER, C., THOMPSON, R., TRAUB, R., REES, R., ROBERTSON, I. (2008): National study of the gastrointestinal parasites of dogs and cats in Australia. *J Vet Parasitol*, 151: 181 – 190. DOI: 10.1016/j.vetpar.2007.10.015
- PARADIES P., IARUSSI, F., PARADIES, P., IARUSSI, F., SASANELLI, M., CAPOGNA, A., LIA, R., ZUCCA, D., GRECO, B., CANTACESSI, C., OTRANTO D. (2017): Otranto occurrence of strongyloidiasis in privately owned and sheltered dogs: clinical presentation and treatment outcome. *Parasit Vectors*, 10: 345. DOI: 10.1186/s13071-017-2275-5
- PIEKARA-STĘPINSKA, A., PIEKARSKA, J., GORCZYKOWSKI, M. (2021): *Cryptosporidium* spp. in dogs and cats in Poland. *Ann Agric Environ Med*, 28(2): 345 – 347. DOI: 10.26444/aaem/120467
- RAICEVIC, J., PAVLOVIC, I., COGHILL, T. (2021): Canine intestinal parasites as a potential source of soil contamination in the public areas of Krusevac, Serbia. *J Infect Dev Ctries*, 15(1): 147 – 154. DOI: 10.3855/jidc.12694
- ROJEKITTIKHUN, W., CHAISIRI, K., MAHITTIKORN, A., PUBAMPEN, S., SA-NGUANKIAT, S., KUSOLSUK, T., MAIPANICH, W., UDONSOM, R., MORI, H. (2014): Gastrointestinal parasites of dogs and cats in a refuge in Nakhon Nayok, Thailand. *Southeast Asian J Trop Med Public Health*, 45(1): 31 – 39

- SILVA, V., SILVA, J., GONCALVES, M., BRANDAO, C., BRITO, V. N. (2020): Epidemiological survey on intestinal helminths of stray dogs in Guimarães, Portugal. *J Parasit Dis*, 44(4): 869 – 876. DOI: 10.1007/s12639-020-01252-2
- STAFFORD, K., KOLLASCH, T., DUNCAN, K., HERR, S., GODDU, T., HEINZ-LOOMER, C., RUMSCHLAG, A., RYAN, W., SWEET, S., LITTLE, S. (2020): Detection of gastrointestinal parasitism at recreational canine sites in the USA: the DOGPARCS study. *Parasit Vectors*, 13(1): 275. DOI: 10.1186/s13071-020-04147-6.
- SWEET, S., SZLOSEK, D., MCCRANN, D., COYNE, M., KINCAID, D., HEGARTY, E. (2020): Retrospective analysis of feline intestinal parasites: trends in testing positivity by age, USA geographical region and reason for veterinary visit. *Parasit Vectors*, 13(1): 473. DOI: 10.1186/s13071-020-04319-4
- SWEET, S., HEGARTY, E., MCCRANN, D., COYNE, M., KINCAID, D., SZLOSEK, D. (2021): A 3-year retrospective analysis of canine intestinal parasites: fecal testing positivity by age, U.S. geographical region and reason for veterinary visit. *Parasit Vectors*, 14: 173. DOI: 10.1186/s13071-021-04678-6
- TANGTRONGSUP, S., SCORZA, A., REIF, J., BALLWEBER, L., LAPPIN, M., SALMAN, M. (2020): Seasonal distributions and other risk factors for *Giardia duodenalis* and *Cryptosporidium* spp. infections in dogs and cats in Chiang Mai, Thailand. *Prev Vet Med*, 174: 104820. DOI: 10.1016/j.prevetmed.2019.104820
- THOMPSON, R.C., MONIS, P.T. (2004): Variation in *Giardia*: implications for taxonomy and epidemiology. *Adv Parasitol*, (58): 137 – 141. DOI: 10.1016/S0065-308X(04)58002
- THOMPSON, R.C., PALMER, C.S., O'HANDLEY, R. (2008): The public health and clinical significance of *Giardia* and *Cryptosporidium* in domestic animals. *Vet J*, 177(1): 18 – 25. DOI: 10.1016/j.tvjl.2007.09.022
- UITERWIJK, M., NIJSSE, R., KOOYMAN, F., WAGENAAR, J., MUGHINI-GRAS, L., PLOEGER, H. (2019): Host factors associated with *Giardia duodenalis* infection in dogs across multiple diagnostic tests. *Parasit Vectors*, 12(1): 556. DOI: 10.1186/s13071-019-3810-3
- UITERWIJK, M., MUGHINI-GRAS, L., NIJSSE, R., WAGENAAR, J., PLOEGER, H., KOOYMAN, F. (2020): *Giardia duodenalis* multi-locus genotypes in dogs with different levels of synanthropism and clinical signs. *Parasit Vectors*, 13(1): 605. DOI: 10.1186/s13071-020-04496-2
- UMUR, S., MERAL, Y., BOLUKBAS, S., GURLER, A., ACICI, M. (2017): First clinical *Strongyloides stercoralis* case in a dog in Turkey. *Turk J Vet Anim Sci*, 41: 312 – 315. DOI: 10.3906/vet-1606-2
- URSACHE, A., GYORKE, A., MIRCEAN, V., DUMITRACHE, M., CODEA, A., COZMA, V. (2021): *Toxocara cati* and Other Parasitic Enteropathogens: More Commonly Found in Owned Cats with Gastrointestinal Signs Than in Clinically Healthy Ones. *Pathogens*, 10(2): 198. DOI: 10.3390/pathogens10020198
- UNTERKÖFLER, M. S., EIPELDAUER, I., MERZ, S., PANTCHEV, N., HERMANN, J., BRUNTHALER, R., BASSO, W., HINNEY, B. (2022): *Strongyloides stercoralis* infection in dogs in Austria: two case reports. *Parasit Vectors*, 15(1): 168. DOI: 10.1186/s13071-022-05270-2
- ZAJAC, A.M., CONBOY, G.A. (2012): *Veterinary Clinical Parasitology, 8th Edition*. Wiley-Blackwell Chichester, 368
- ZANZANI, S.A., GAZZONIS, A.L., SCARPA, P., BERRILLI, F., MANFREDI, M.T. (2014): Intestinal parasites of owned dogs and cats from metropolitan and micropolitan areas: prevalence, zoonotic risks, and pet owner awareness in northern Italy. *Biomed Res Int*, 2014: 696508. DOI: 10.1155/2014/696508