

Prevalence and risk factors for efavirenz-based antiretroviral treatment-associated severe vitamin D deficiency

A prospective cohort study

Hanna Nylén, PhD^a, Abiy Habtewold, PhD^b, Eyasu Makonnen, PhD^b, Getnet Yimer, PhD^b, Leif Bertilsson, PhD^c, Jürgen Burhenne, PhD^d, Ulf Diczfalusy, PhD^a, Eleni Akillu, PhD^{c,*}

Abstract

Initiation of efavirenz-based combination antiretroviral therapy (cART) is associated with Vitamin D deficiency, but the risk factors including efavirenz pharmacokinetics for cART-induced severe vitamin D deficiency (SVDD) and the impact of anti-tuberculosis (TB) cotreatment are not explored. We investigated the prevalence of SVDD in HIV and TB-HIV coinfecting patients and associated risk factors for treatment-induced SVDD.

Treatment-naïve Ethiopian HIV patients with (n=102) or without (n=89) TB co-infection were enrolled prospectively and received efavirenz-based cART. In TB-HIV coinfecting patients, rifampicin-based anti-TB treatment was initiated 4 or 8 weeks before starting cART. Plasma 25-hydroxyvitamin D (25 [OH]D), cholesterol and 4-beta hydroxycholesterol concentrations were measured at baseline, 4th, 16th, and 48th week of cART. Plasma efavirenz concentrations were determined at 4th and 16th weeks of cART.

TB-HIV patients had significantly lower plasma 25 (OH)D₃ levels than HIV-only patients at baseline. TB co-infection, low Karnofsky score, high viral load, and high CYP3A activity as measured by plasma 4β-hydroxycholesterol/cholesterol ratios were significant predictors of low 25 (OH)D₃ levels at baseline. In HIV-only patients, initiation of efavirenz-based cART increased the prevalence of SVDD from 27% at baseline to 76%, 79%, and 43% at 4th, 16th, and 48th weeks of cART, respectively. The median 25 (OH)D₃ levels declined from baseline by -40%, -50%, and -14% at 4th, 16th, and 48th weeks of cART, respectively.

In TB-HIV patients, previous anti-TB therapy had no influence on 25 (OH)D₃ levels, but the initiation of efavirenz-based cART increased the prevalence of SVDD from 57% at baseline to 70% and 72% at the 4th and 16th weeks of cART, respectively. Median plasma 25 (OH)D₃ declined from baseline by -17% and -21% at week 4 and 16 of cART, respectively.

Our results indicate low plasma cholesterol, high CYP3A activity, and high plasma efavirenz concentrations as significant predictors of early efavirenz-based cART-induced vitamin D deficiency. Low plasma 25 (OH)D₃ level at baseline is associated with TB co-infection and HIV diseases progression. Initiation of efavirenz-based cART is associated with high incidence of SVDD, whereas rifampicin based anti-TB therapy co-treatment has no significant effect. Supplementary vitamin D during cART initiation may be beneficial for HIV patients regardless of TB coinfection.

Abbreviations: cART = combination antiretroviral therapy, CYP = cytochrome P450, HIV = human immunodeficiency virus, SVDD = severe vitamin D deficiency, TB = tuberculosis, VDR = vitamin D receptor.

Keywords: 25-hydroxyvitamin D, cART, CYP2B6, CYP3A4, cytochrome P450, drug metabolism, HIV, TB, tuberculosis, VDR, vitamin D receptor, vitamin D

Editor: Gary Maartens.

This work was supported by grants from European and Developing Countries Clinical Trials Partnership (Grant number: CG_TA.05.40204_005) and from Swedish Research Council (Grant number: 2015-03295)

The authors report no conflicts of interest.

^a Department of Laboratory Medicine, Division of Clinical Chemistry, Karolinska Institutet at Karolinska University Hospital, Stockholm, Sweden, ^b Department of Pharmacology, School of Medicine, College of Health Sciences, Addis Ababa University, Addis Ababa, Ethiopia, ^c Department of Laboratory Medicine, Division of Clinical Pharmacology, Karolinska Institutet, Stockholm, Sweden, ^d Department of Clinical Pharmacology and Pharmacoepidemiology, University of Heidelberg, Heidelberg, Germany.

* Correspondence: Professor Eleni Akillu, Division of Clinical Pharmacology, Department of Laboratory of Medicine, Karolinska Institutet, Karolinska University Hospital Huddinge C-168, SE-141 86 Stockholm, Sweden (e-mail: Eleni.Akillu@ki.se).

Copyright © 2016 the Author(s). Published by Wolters Kluwer Health, Inc. All rights reserved.

This is an open access article distributed under the Creative Commons Attribution License 4.0 (CCBY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Medicine (2016) 95:34(e4631)

Received: 9 May 2016 / Received in final form: 21 July 2016 / Accepted: 27 July 2016

<http://dx.doi.org/10.1097/MD.0000000000004631>

1. Introduction

Vitamin D deficiency is associated with many chronic illnesses, including autoimmune, cardiovascular, and infectious diseases like tuberculosis (TB), HIV disease progression, and mortality.^[1–3] Plasma 25 (OH)D, the primary circulating form of vitamin D, serves as an indicator of vitamin D status.^[4] Vitamin D₃ is formed naturally from 7-dehydrocholesterol, a cholesterol precursor, in the skin upon exposure to sunlight. Thus factors influencing 7-dehydrocholesterol levels may result in altered vitamin D and cholesterol levels. Efavirenz-based combination antiretroviral therapy (cART) is associated with changes in both vitamin D^[5–9] and cholesterol concentrations,^[10] but it is unclear whether there is a correlation between the variation in vitamin D and cholesterol levels during cART and whether this effect is related to drug concentrations.

TB is the most common opportunistic infection and the leading cause of death in people living with HIV. Concomitant HIV and TB treatments are challenging because significant drug–drug interactions, overlapping toxicities, and immune reconstitution inflammatory syndrome.^[11–13] Efavirenz and rifampicin, the cornerstones of first-line cART and anti-TB treatments, respectively, are potent inducers CYP2B6 and CYP3A enzymes and transporter proteins.^[10,14,15] Efavirenz is mainly metabolized by genetically polymorphic CYP2B6 and to some extent by CYP3A enzymes. Vitamin D is involved in the regulation of CYP3A and CYP2B6 enzyme expression.^[16,17] However, CYP3A catalyzes the 4-hydroxylation of 25 (OH)D₃, and hence CYP3A induction may contribute to drug-induced vitamin D deficiency.^[18,19] Long-term CYP2B6 and CYP3A induction by efavirenz is influenced by pharmacogenetic factors relevant for efavirenz disposition.^[10,15,20–22] Accordingly, host genetic factors affecting the plasma concentration of CYP3A inducers may also potentially affect the vitamin D levels in patients on long-term treatment with efavirenz or rifampicin. Indeed, a CYP2B6 genotype-based efavirenz dose adjustment was recommended recently to optimize treatment outcomes.^[23,24] Implication of inter-individual variations in efavirenz plasma concentration for efavirenz-induced vitamin D deficiency is yet to be investigated.

Previous studies reported that efavirenz-based cART lowers the 25 (OH)D level.^[5–9] Poor absorption, lower exposure to the sunshine, and darker skin pigmentation are known risk factors for low 25 (OH)D levels.^[25] Data regarding predictors of 25 (OH)D levels during cART are scarce in HIV patients living in tropical areas, where there is an all-year round sunshine. The risk factors for cART-induced severe vitamin D deficiency and the implication of anti-TB co-treatment remain to be investigated, especially in sub-Saharan Africa, where both HIV and TB are major public health problems. In the present study, we hypothesized that high CYP3A induction by rifampicin co-treatment and inter-individual variability in efavirenz disposition may play a role for between-patient variability in vitamin D status during cART. Therefore, the objectives of this study were to identify the prevalence and associated risk factors for low 25 (OH)D levels and severe vitamin D deficiency before starting treatment and during efavirenz-based cART alone or together with rifampicin-based anti-TB co-therapy in HIV or TB-HIV co-infected patients, respectively.

2. Materials and methods

2.1. Study design and settings

The study design was prospective, comparative, observational, open-label, parallel assignment, 2-arm pharmacogenetic and

pharmacokinetic cohort study to identify the prevalence and risk factors for severe vitamin D deficiency at baseline and during efavirenz-based cART with or without rifampicin-based anti-TB therapy. The study was conducted between June 2008 and June 2011 at HIV and TB clinics in Addis Ababa (latitude 9- 1° N), Ethiopia.

All patients gave written, informed consent to participate in this study. The study protocol received ethics approvals from the Institutional Review Board (IRB) of School of Medicine, Addis Ababa University, National Ethics Review Committee of Ethiopia. The study also received approval from IRB of Karolinska Institutet (Stockholm, Sweden) and was conducted as per International Conference for Harmonization-Good Clinical Practice (ICH-GCP) guidelines.

2.2. Study participants

This study was conducted as one of the substudies designed under the umbrella of the broad clinical research project entitled “The HIV-TB Pharmagene Study” in Ethiopia. Details of the main study design, patient enrolment process, and inclusion criteria with follow-up and drug treatments were reported previously.^[26] Briefly, for the main study, newly diagnosed HIV-infected (n=285) and TB-HIV co-infected (n=208) patients were recruited prospectively and enrolled in parallel and followed up to 48 weeks. The eligibility criteria were ≥18 years of age, not pregnant, and CD4 count ≤200 cells/mm³. None of the study participants received isoniazid prophylaxis or other TB treatment for 2 years before enrollment. Treatment adherence was assessed by self-report.

For the present study, a total of 191 patients (102 TB-HIV co-infected patients and 89 HIV only infected patients), with complete set of plasma samples collected at baseline, and at the 4th and 16th weeks of cART, were used to monitor the change in vitamin D during cART. The sample size was calculated for each treatment group considering a moderate effect size, E=0.3, to detect the change in vitamin D levels before and after efavirenz exposure (paired *t* test) with 80% of study power and α=5%, the desired sample size was calculated as 87 in each group. Plasma sample collected at 48 weeks of cART from 42 HIV-only patients was also available and used.

2.3. Treatment

All HIV patients received cART (600 mg efavirenz-based cART containing either zidovudine/lamivudine/efavirenz or stavudine/lamivudine/efavirenz). Plasma samples for the determination of plasma cholesterol, 4β-hydroxycholesterol, and 25 (OH)D were taken before the initiation of treatment (week 0) and at weeks 4, 16, and 48 of treatment.

All TB-HIV patients (n=102) initiated rifampicin-based anti-TB treatment 4 (n=69) or 8 weeks (n=33) before the initiation of efavirenz-based cART. The short course anti-TB treatment consisted of an initiation phase with rifampicin/isoniazid/pyrazinamide /ethambutol for 2 months followed by a continuation phase with rifampicin/isoniazid for 4 months. Samples for the determination of plasma cholesterol, 4β-hydroxycholesterol and 25 (OH)D concentrations were collected before starting anti-TB treatment (corresponding to week -4 or -8 weeks of cART), at the initiation of cART (week 0) and weeks 4 and 16 of cART. The study population, follow-up period, and study sampling time points are presented in Figure 1.

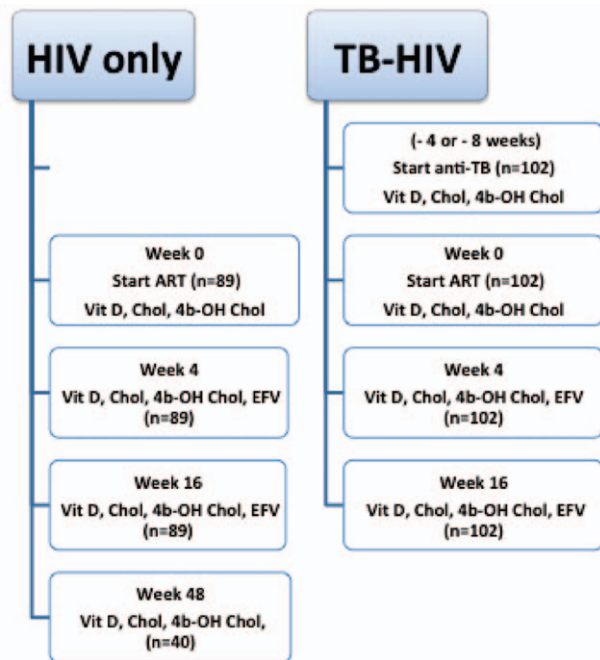


Figure 1. Presentation of the study design, study population, and follow-up period. Study time point for determination of plasma vitamin D (Vit D), cholesterol (Chol), 4 β -hydroxycholesterol (4b-OH chol), and efavirenz (EFV) levels during follow-up period is indicated. HIV-only=HIV patients treated with efavirenz-based cART alone, TB= tuberculosis, TB-HIV=co-infected patients treated with concomitant efavirenz-based cART plus rifampicin-based anti-TB therapy.

Outcome variables were plasma 25 (OH)D level and vitamin D status; vitamin D deficiency (VDD) defined as circulating levels of 25 (OH)D₃ <20 ng/mL (50 nmol/L), and levels between 21 and 29 ng/mL (51–72.5 nmol/L), considered as insufficient.^[27] Severe vitamin D deficiency (SVDD) was defined as plasma 25 (OH)D₃ level <25 nmol/L.^[25] The predicting variables were patient's clinical and laboratory parameters at baseline and during treatment, plasma cholesterol and efavirenz concentrations, and CYP3A enzyme activity as defined by 4 β -hydroxycholesterol/cholesterol ratio.

2.4. Quantification of plasma 25 (OH)D concentrations

25 (OH)D₃ and 25 (OH)D₂ in plasma were determined by isotope dilution liquid chromatography-tandem mass spectrometry (LC-MS/MS) as described previously.^[28] Briefly, 25 (OH)D was extracted from plasma using a Hybrid SPE precipitation 96-well plate (Supelco, Sigma-Aldrich) and isopropanol in acetonitrile (12% v/v, 1% formic acid). 25 (OH)D₃/D₂ Plasma Control (level 1) and 25 (OH)D₃/D₂ Plasma Calibration Standard were used as quality control and calibrator, respectively (Chromsystems, Munich, Germany). ²H₆-25-hydroxyvitamin D₃ was used as internal standard (Synthetica AS, Oslo, Norway). Analysis was done on Acquity Quattro Premier LC-MS/MS (Waters, Milford, MA) system equipped with a UPLC BEHC18 column (1.7 μ m id, 2.1 \times 50 mm, Waters, Milford, MA). The mobile phase was a gradient of 50% to 95% acetonitrile in water (0.1% formic acid). The lower limit of detection (LOD) of 25 (OH)D₃ was 2.5 nmol/L and the lower limit of quantification (LOQ) 7 nmol/L. The linear range was 2.5 to 625 nmol/L. The relative standard deviation (CV) was 16% (at 33.5 \pm 5.4 [n = 39] [mean \pm sd]). The LOQ of 25 (OH)D₂ was 12 nmol/L.

2.5. Quantification of plasma cholesterol concentrations

Cholesterol was determined in plasma using a commercial enzymatic method (Cholesterol Chod-PAPP, Roche Diagnostics GMBH, Mannheim, Germany) on a Roche/Hitachi Modular Instrument. The between-day variation was 1.3% at 5 mmol/L.

2.6. Quantification of efavirenz plasma concentrations

Plasma samples for determination of efavirenz were collected at weeks 4 and 16 of cART in both treatment groups. Plasma efavirenz concentrations were determined using LC-MS/MS as described previously.^[20,26,29] Briefly, protein precipitation was performed using acetonitrile containing internal standards (¹³C₆-efavirenz and ²H₄-8-hydroxy-efavirenz, respectively). Analysis was performed using a Synergi Fusion RP chromatography column (Phenomenex, Torrance, CA) and mobile phases containing ammonium acetate (5 mmol/L, acidic), methanol, and acetonitrile. The lower limits of quantification in plasma were 10.0 ng/mL. The efavirenz calibration range was 10 to 10000 ng/mL. The method was validated according to the FDA validation guidelines and fulfilled all criteria concerning accuracy, precision, recovery, linearity, and stability.

2.7. Statistical analysis

Comparison of median plasma 25 (OH)D₃ levels between treatment groups was done using the Mann-Whitney test. Plasma 25 (OH)D₃, cholesterol, efavirenz concentrations, and 4 β -hydroxycholesterol/cholesterol ratios were log-transformed (base 10) before applying the *t* test, repeated measure analysis of variance (ANOVA), and regression analysis. Pairwise comparison of data from baseline within and between treatment groups was made using paired and unpaired *t* test, respectively. For each patient, the percent change in plasma 25 (OH)D₃ level from baseline to the 4th, 16th and 48th weeks of cART was calculated using the following formula:

$$\% \text{ change in plasma } 25(\text{OH}) \text{ D}_3 = \left[\frac{25(\text{OH}) \text{ D}_3 \text{ at week } x - 25(\text{OH}) \text{ D}_3 \text{ at baseline}}{25(\text{OH}) \text{ D}_3 \text{ at baseline}} \right] \times 100$$

Repeated measure ANOVA was used to analyze the change in log plasma 25 (OH)D₃ levels over time. Univariate followed by multivariate linear regression analysis was performed to identify predictors of low plasma 25 (OH)D₃ levels at baseline and during treatment. Predictor variables that resulted in a *P* value <0.1 in the univariate regression analysis was entered into a backward stepwise multivariate regression analysis to identify significant predictors in the final model. Likewise logistic regression was done to determine predictors of severe vitamin D deficiency before and during cART. Statistical analyses were performed using Statistica version 12 (StatSoftInc, Tulsa, OK) and SPSS Statistics (IBM Corporation, Somers, NY) software, version 23.0. GraphPad Prism version 5.0 for Windows (Graph Pad, La Jolla, CA) was used for graphical presentations. A *P* value <0.05 was considered significant.

3. Results

The baseline sociodemographic, clinical, and laboratory characteristics of study participants stratified by treatment group are presented in Table 1. The best measure of vitamin D status is the

Table 1
Baseline demographic, clinical, and laboratory characteristics of the study participants.

	HIV-only, n=89	TB-HIV, n=102
Sex		
Male	20 (22.5%)	47 (46.1%)
FEMALE	69 (77.5%)	55 (53.9%)
HIV stage		
1	1 (1.2%)	
2	7 (8.1%)	1 (1.0%)
3	38 (44.2%)	56 (56.6%)
4	40 (46.5%)	42 (42.4%)
Type of ART		
d4T30/3TC/EFV	47 (54.7%)	38 (38.4%)
d4T40/3TC/EFV	5 (5.8%)	1 (1.0%)
TDF/3TC/EFV		26 (26.3%)
ZDV/3TC/EFV	34 (39.5%)	34 (34.3%)
Age, y	35 (30-42)*	32 (28-40)*
BMI	19.5 (18.2-22.5)*	18.7 (17.2-19.9)*
Hb, g/L	12.7 (11.6-13.6)*	11.3 (10.0-12.9)*
WBC count ($\times 10^3$ cells/ μ L)	4.4 (3.7-5.4)*	5.7 (7.3-4.2)*
Neutrophils (%)	55 (44-63)*	67 (59-75)*
Platelets ($\times 10^3$ cells/ μ L)	233 (171-302)*	312 (240-404)*
Aspartate aminotransferase, U/L	32.5 (27-40)*	41 (34-73)*
Alanine transaminase, U/L	28 (21-127)*	30 (21-45)*
Alkaline phosphatase, U/L	106 (79-124)*	114 (89-174)*
Total bilirubin, mg/dL	0.38 (0.3-0.6)*	0.40 (0.3-0.7)*
Direct bilirubin, mg/dL	0.1 (0.1-0.1)*	0.1 (0.1-0.2)*
Albumin, g/dL	4.0 (2.5-4.5)*	3.4 (3.0-4.0)*
Urea, mg/dL	24 (19-30)*	24 (20-31)*
Serum creatinine, mg/dL	0.8 (0.7-1.0)*	0.9 (0.7-1.1)*
CD4 counts, cells/ cm^3	116 (68-160)*	100 (55-135)*
Log plasma HIV viral load	2.59 (2.226-2.86)*	5.09 (4.44-55.1)*

3TC=lamivudine, EFV=efavirenz, TDF=Tenofovir, ZDV= zidovudine, d4T30 (stavudine 30 mg for patients weighing <60 kg), d4T40 (stavudine 40 mg for patients weighing \geq 60 kg).
*Median (interquartile range).

concentration of 25 (OH)D₃ + 25 (OH)D₂ in plasma. Very few patients had 25 (OH)D₂ levels above the lower LOQ; hence, all statistical calculations were done using only plasma 25 (OH)D₃ data.

3.1. Plasma 25 (OH)D₃ levels at baseline and during treatment

Comparisons of the median plasma 25 (OH)D₃ levels at baseline and during cART and the median percent change in plasma 25 (OH)D₃ levels during therapy between the 2 treatment groups are presented in Table 2. The pretreatment median plasma 25 (OH)D₃ concentration was significantly lower in TB-HIV coinfect

patients than in HIV patients without TB. In the HIV-only cohort, initiation of efavirenz-based cART dramatically lowered the median plasma 25 (OH)D₃ levels by -40%, -50%, -14% at week 4, 16, and 48 of cART, respectively (Table 2). Although in TB-HIV co-infected patients, previous treatment with rifampicin-based anti-TB drugs for 4 or 8 weeks resulted in no significant change in plasma 25 (OH)D₃ from baseline. However, initiation of efavirenz-based cART co-treatment led to a gradual decline in plasma 25 (OH)D₃ level and the median percent change of 25 (OH)D₃ from baseline by 4th and 16th of weeks of cART was -17% and -21%, respectively.

Comparisons of change in log plasma 25 (OH)D₃ levels from baseline after 4 weeks of efavirenz-based cART alone versus 4 or 8 weeks of rifampicin-based anti-TB therapy alone are presented in Figure 2. TB-HIV co-infected patients had significantly lower mean log plasma 25 (OH)D₃ levels compared to HIV-only patients at baseline ($P < 0.001$, Fig. 2), which became reversed soon after initiating therapy. The mean log plasma 25 (OH)D₃ levels became significantly lower in HIV-only patients after 4 weeks of efavirenz-based cART compared to 4 to 8 weeks of rifampicin-based anti-TB treatment only in TB-HIV patients ($P = 0.03$). Initiation of cART in TB-HIV patients on anti-TB therapy lowered the mean plasma 25 (OH)D₃ levels, and there was no significant difference in the mean plasma 25 (OH)D₃ levels between the 2 treatment groups at weeks 4 and 16 of cART.

The overall change in the mean log plasma 25 (OH)D₃ levels profile from baseline during efavirenz-based cART alone versus rifampicin-based anti-TB co-treatment is presented in Figure 3. In HIV-only patients, within-treatment group analysis over time (using repeated measure ANOVA) indicated that efavirenz-based cART significantly reduced the mean log plasma levels of 25 (OH)D₃ ($P < 0.001$). Paired *t* tests indicated significantly lower plasma 25 (OH)D₃ levels at week 4 ($P < 0.001$, geometric mean ratio [GMR]= 1.606; 95% confidence interval [CI] of the mean GMR = 1.46-1.76) and at week 16 ($P < 0.001$, GMR = 1.99; 95% CI of GMR = 1.76-2.21) compared with the baseline pretreatment value. As cART continued, the plasma 25 (OH)D₃ levels were restored, and no significant difference from baseline was found after 48 weeks of cART ($P = 0.12$) in HIV-only patients.

In TB-HIV co-infected patients, within-treatment group analysis indicated that efavirenz-based cART significantly reduced mean log plasma levels of 25 (OH)D₃ ($P < 0.0001$). Paired *t* tests indicated significantly lower 25 (OH)D₃ level at week 4 ($P = 0.01$, GMR = 1.17; 95% CI of GMR = 1.04-1.34) and week 16 ($P < 0.001$, GMR = 1.41, 95% CI of GMR = 1.18-1.69) compared to the baseline value. Data on plasma 25 (OH)D₃ at week 48 of cART in TB-HIV co-infected patients were not available.

Table 2
Comparison of median and IQR of plasma 25 (OH)D₃ (nmol/L) levels at baseline and during treatment.

Time on treatment	Treatment group						P
	TB-HIV cohort			HIV-only cohort			
	N	Median (IQR)	Median % change (IQR)	N	Median (IQR)	Median % change (IQR)	
Baseline (pretreatment)	102	23.2 (14.9-33.2)		89	32.5 (24.0-41.86)		<0.0001
Previous RIF-only treatment	76	23.9 (15.2-35.4)	-1% (-17 to 48)				
4 weeks of ART*	102	19.25 (12.2-28.0)	-17% (-37 to 16)	88	19.95 (14.5-25.0)	-40% (-55 to -15)	0.7
16 weeks of ART*	94	16.85 (11.6-27.7)	-21% (-50 to 14)	85	15.20 (10.5-22.3)	-50% (-63 to -31)	0.55
48 Weeks of ART				40	29.20 (19.2-35.4)	-14% (-50 to 23)	

ART=antiretroviral therapy, RIF=rifampicin, IQR=interquartile range.

*TB-HIV patients co-treated with RIF-based anti-TB therapy. Baseline in the TB-HIV cohort was at initiation of treatment with rifampicin (week -4 or -8 before start of ART). Baseline in the HIV-only cohort was at initiation of efavirenz-treatment (week 0).

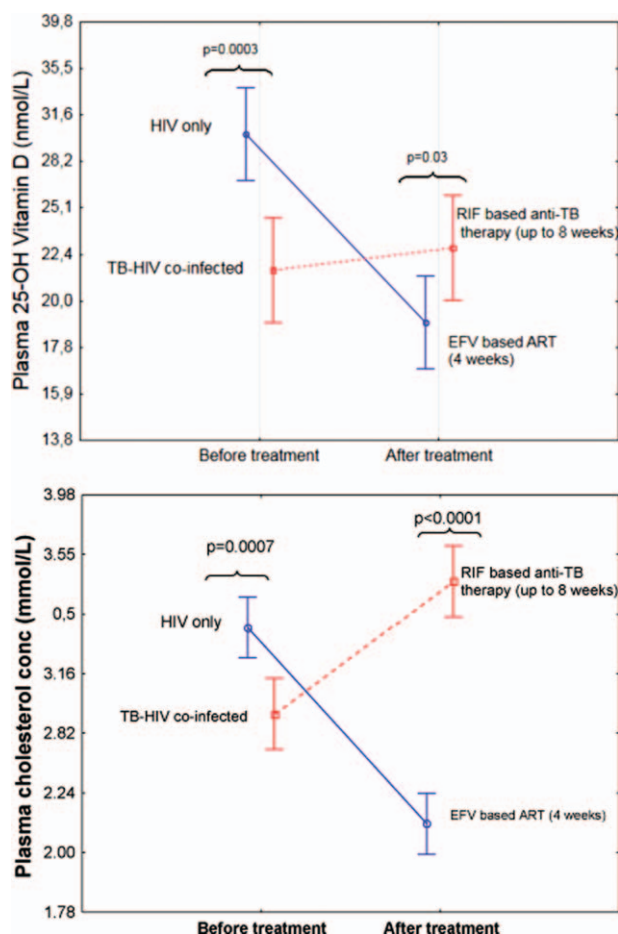


Figure 2. Change in the mean (log) plasma 25 (OH)D₃ (upper panel) and cholesterol levels (lower panel) from baseline by 4 weeks of efavirenz-based cART in HIV-only patients (straight line) versus up to 8 weeks of rifampicin-based anti-tuberculosis (TB) therapy only in TB-HIV co-infected patients (broken line). Between-treatment group differences were analyzed using unpaired *t* test. Vertical bars denote 0.95 confidence intervals of the mean.

3.2. Vitamin D deficiency at baseline and during treatment

All patients had deficient (92%) or insufficient (8%) plasma 25 (OH)D₃ levels at the study enrollment. Despite receiving cART

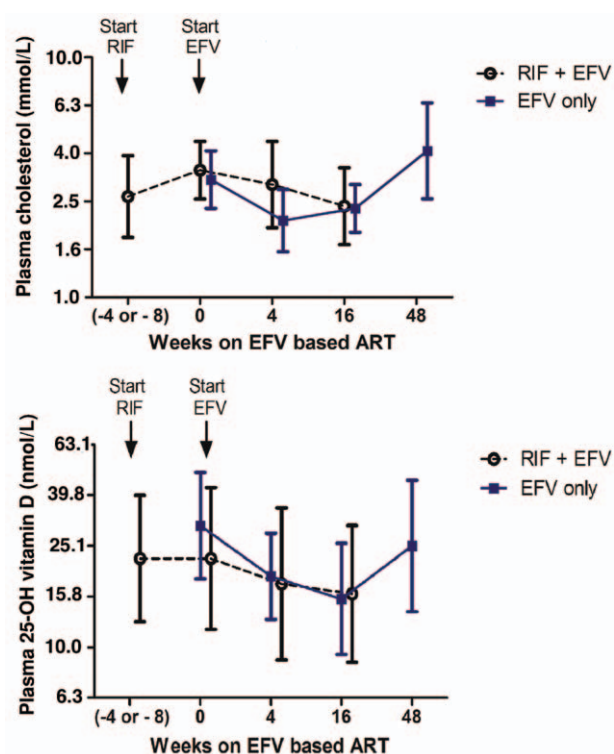


Figure 3. Change in the mean (log) plasma levels of cholesterol (upper panel) and 25 (OH)D₃ (lower panel) from baseline during efavirenz-based cART alone in HIV patients (EFV only, straight line) or with rifampicin-based anti-tuberculosis therapy in TB-HIV co-infected patients (RIF + EFV, broken line). Between-treatment group differences were analyzed using unpaired *t* test. Vertical bars denote 0.95 confidence intervals of the mean.

and anti-TB therapy, none of the patients achieved a sufficient (>72.5 nmol/L) plasma 25 (OH)D₃ level during the study follow-up period.

The prevalence of SVDD before starting treatment (at baseline) and at the 4th, 16th, and 48th weeks of efavirenz-based cART in each treatment group is presented in Table 3. At baseline, 57% (95% CI=48.3%–67.5%) of TB-HIV co-infected patients had SVDD, whereas only 27% (95% CI=15.1%–32.9%) of HIV-

Table 3
Comparison of proportions of patients with severe vitamin D deficiency (defined as plasma 25 (OH)D₃ <25nmol/L) between treatment groups: HIV patients treated with efavirenz-based cART alone (HIV-only cohort) versus TB-HIV co-infected patients co-treated with efavirenz-based cART and rifampicin-based anti-TB therapy (TB-HIV cohort).

Time on treatment		Treatment group				P	Odds ratio (95% CI)
		HIV-only cohort		TB-HIV cohort*			
		n	%	n	%		
Baseline	Nonsevere deficient	65	73.0%	44	43.1%	<0.0001	3.57 (1.939 to 6.574)
	Severe deficient	24	27.0%	58	56.9%		
Anti-TB therapy alone	Nonsevere deficient			35	46.1%	0.33	0.718 (0.376 to 1.371)
	Severe deficient			41	53.9%		
4 weeks of cART	Nonsevere deficient	21	23.9%	31	30.4%	0.31	0.703 (0.353 to 1.400)
	Severe deficient	67	76.1%	71	69.6%		
16 weeks of cART	Nonsevere deficient	18	21.2%	26	27.7%	0.31	0.703 (0.353 to 1.400)
	Severe deficient	67	78.8%	68	72.3%		
48 weeks of cART	Nonsevere deficient	23	57.5%	ND	ND	0.31	0.703 (0.353 to 1.400)
	Severe deficient	17	42.5%	ND	ND		

cART= combination antiretroviral therapy, CI= confidence interval, ND= not determined, TB= tuberculosis.
 * TB-HIV patients were co-treated with rifampicin-based anti-TB therapy.

Table 4

Predictors of pre-treatment plasma 25 (OH)D₃ level and severe vitamin D deficiency (25 (OH)D₃ <25 nmol/L) using linear regression and logistic regression analysis respectively.

Variable	Baseline plasma 25-(OH)D ₃				Baseline Severe vitamin D deficiency			
	Univariate		Multivariate		Univariate		Multivariate	
	Beta coefficient (95% CI)	P	Beta coefficient (95% CI)	P	Odds ratio (95% CI)	P	Odds ratio (95% CI)	P
TB co-infection	-0.138 (-0.205 to -0.071)	<0.0001*			3.845 (2.095 to 7.055)	<0.0001*	3.975 (2.084 to 7.582)	0.001
Sex, female	-0.039 (-0.112 to 0.034)	0.30			0.398 (0.420 to 1.411)	0.39		
Karnofsky score	0.007 (0.004 to 0.010)	<0.0001*	0.004 (0.001 to 0.006)	0.02	0.959 (0.934 to 0.985)	0.002*	0.979 (0.951 to 1.001)	0.16
Age, y	0.001 (-0.002 to 0.005)	0.46			0.987 (0.957 to 1.017)	0.37		
Body mass index	0.020 (0.008 to 0.032)	0.001*			0.918 (0.826 to 1.021)	0.11		
Hemoglobin, g/dL	0.020 (0.005 to 0.035)	0.007*			0.914 (0.805 to 1.037)	0.16		
WBC count × 10 ³	-0.005 (-0.016 to 0.006)	0.39			1.031 (0.936 to 1.129)	0.56		
Aspartate aminotransferase, U/L	-0.005 (-0.002 to 0.001)	0.11			1.010 (1.001 to 1.019)	0.02*		
Alanine aminotransferase, U/L	-0.005 (-0.001 to 0.002)	0.26			0.999 (0.989 to 1.010)	0.89		
Alkaline phosphatase, U/L	-0.001 (-0.001 to .000)	0.005*			1.005 (1.000 to 1.010)	0.045*		
Serum Albumin, g/dL	0.139 (0.095 to 0.183)	<0.0001	0.082 (0.033 to 0.131)	0.001	0.465 (0.300 to 0.721)	0.001*	0.594 (0.383 to 0.951)	0.03
Urea, mg/dL	0.002 (-0.001 to 0.006)	0.15			0.977 (0.948 to 1.007)	0.14		
Serum creatinine, mg/dL	0.132 (-0.005 to 0.269)	0.06			0.290 (0.535 to 1.702)	0.29		
CD4 counts, cells/cm ³	0.001 (-0.001 to 0.001)	0.65			0.998 (0.993 to 1.004)	0.55		
Log plasma viral load	-0.056 (-0.080 to -0.031)	<0.0001*	-0.035 (0.060 to -0.11)	0.004	1.557 (1.236 to 1.961)	<0.0001*	1.466 (1.156 to 1.859)	0.002
Log Cholesterol (mmol/L)	0.276 (0.042 to 0.510)	0.02*			0.408 (0.058 to 2.889)	0.37		
Log 4β-OH Chol/Chol ratio	-0.228 (-0.379 to -0.078)	0.003*	-0.156 (-0.302 to -0.10)	0.037	3.769 (1.012 to 14.037)	0.048		

CI = confidence interval, TB = tuberculosis, WBC = white blood cells.

* Predictor variables that resulted in a *P* value <0.1 in the univariate regression analysis was entered into a backward stepwise multivariate regression analysis to identify significant predictors in the final model.

only infected patients had SVDD (*P* < 0.001). In the HIV-only cohort, initiation of efavirenz-based cART significantly increased the proportion of patients with SVDD from 27% at baseline to 76% (95% CI = 67.1%–84.5%) at week 4, and 79% (95% CI = 70.3%–87.3%) at week 16, and 42.5% (95% CI = 32.2%–52.7%) at week 48 cART. Whereas in TB-HIV patients on rifampicin-based anti-TB therapy, initiation of efavirenz-based cART co-treatment increased the proportion of patients with SVDD from 57% at baseline to 70% (95% CI = 69.7%–78.5%) at week 4 and 72% (95% CI = 63.6%–80.9%) at week 16 of cART.

3.3. Plasma cholesterol levels at baseline and during treatment

The pretreatment mean log plasma cholesterol level was significantly higher in HIV patients compared to TB-HIV patients. Initiation of cART in HIV-only patients significantly reduced the plasma level of cholesterol. In contrast, increased plasma levels of cholesterol after 4 to 8 weeks of rifampicin-based anti-TB therapy alone were observed in TB-HIV patients (Fig. 2). The median percent reduction from baseline in cholesterol level by 4 weeks of efavirenz-based cART was -33% and the respective increase by rifampicin-based anti-TB therapy treatment alone was 21%. There was a similar pattern of change in plasma cholesterol and 25 (OH)D₃ level particularly in HIV-only patients treated with efavirenz-based cART (Fig. 3).

3.4. Efavirenz plasma concentrations

The median (IQR) of efavirenz plasma concentrations in HIV-only and TB-HIV patients at week 4 of cART was 1200

(841–1846) ng/mL and 1574 (984–2041) ng/mL, respectively (*P* = 0.45). The median and IQR of efavirenz plasma concentrations in HIV-only and TB-HIV patients at week 16 of cART were 1328 (980–1947) ng/mL and 1236 (826–2013) ng/mL, respectively (*P* = 0.26).

3.5. Predictors of low vitamin D status at baseline and during treatment

Linear, regression analysis was used to identify predictors of low plasma 25 (OH)D₃ at baseline (Table 4). In a univariate analysis, low plasma 25 (OH)D₃ levels at baseline were significantly associated with TB-co-infection, low Karnofsky score, low body mass index, low plasma cholesterol level, low hemoglobin level, low albumin, high viral load, high CYP3A activity (as measured by 4β-hydroxycholesterol to cholesterol ratio), and high serum alkaline phosphatase. In a stepwise multivariate regression analysis, low Karnofsky score, low albumin level, high viral load, and high 4β-hydroxycholesterol to cholesterol ratio remained significant predictors of low 25 (OH)D₃ concentration at baseline.

Univariate logistic regression identified TB co-infection, low Karnofsky score, high aspartate aminotransferase, low albumin levels, high 4β-hydroxycholesterol to cholesterol ratio, and high HIV viral load as significant predictors of SVDD at baseline. Multivariate logistic regression, using a backward stepwise conditional model of all variables with a *P* value <0.1 in the univariate analysis (Table 4), identified TB co-infection, high HIV viral load, and low albumin level as significant predictors of SVDD at baseline.

After 4 weeks of efavirenz-based cART, low Karnofsky score and albumin level at baseline, low plasma cholesterol, high efavirenz plasma concentration, and high 4β-hydroxycholesterol

Table 5

Predictors of plasma 25 (OH)D₃ level and severe vitamin D deficiency (25 (OH)D₃ <25 nmol/L) at 4 weeks of EFV-based cART using simple linear regression and logistic regression respectively.

Variable	Plasma 25-hydroxyvitamin D ₃				Severe vitamin D deficiency			
	Univariate		Multivariate		Univariate		Multivariate	
	Beta coefficient (95% CI)	P	Beta coefficient (95% CI)	P	Odds ratio (95% CI)	P	Odds ratio (95% CI)	P
Rifampicin co-treatment	-0.026 (-0.098 to 0.046)	0.47			0.718 (0.376 to 1.371)	0.31		
Sex, female	-0.056 (-0.131 to 0.018)	0.13			1.589 (0.985 to 3.640)	0.06		
Karnofsky score	0.007 (0.004 to 0.010)	<0.0001*	0.007 (0.004 to 0.010)	0.0001	0.956 (0.921 to 0.991)	0.02*		
Albumin	0.068 (0.019 to 0.117)	0.007	0.032 (-0.024 to 0.087)	0.26	0.493 (0.297 to 0.818)	0.006*	0.526 (0.327 to 0.846)	0.007
Type of cART								
TDF-containing cART	-0.093 (-0.318 to 0.133)	0.42			0.825 (0.335 to 2.032)	0.67		
ZDV-containing cART	-0.53 (-0.265 to 0.159)	0.62			0.926 (0.477 to 1.800)	0.82		
D4T-containing cART	-0.071 (-2.80 to 0.139)	0.51			1.225 (0.645 to 2.325)	0.53		
Age, y	0.001 (-0.003 to 0.004)	0.60			0.988 (0.957 to 1.021)	0.47		
Body mass index	0.007 (-0.006 to 0.020)	0.27			0.940 (0.841 to 1.052)	0.28		
Log plasma Chol at week 4 of ART	0.217 (0.008 to 0.426)	0.042*	0.179 (-0.46 to 0.404)	0.11	0.095 (0.013 to 0.702)	0.02*	0.166 (0.022 to 1.258)	0.05
Log 4β-OH/Chol ratio at week 4 of ART	-0.257 (-0.402 to 0.111)	0.001*	-0.201 (-0.359 to -0.043)	0.013	3.881 (0.994 to 14.639)	0.05*		
Log plasma EFV at week 4 of ART	-0.115 (-0.227 to -0.002)	0.04*	-0.095 (-0.232 to -0.009)	0.034	2.244 (0.649 to 7.765)	0.20		

ART = antiretroviral therapy, cART = combination antiretroviral therapy, CI = confidence interval, EFV = efavirenz, TB = tuberculosis, TDF = tenofovir, WBC = white blood cells, ZDV = zidovudine.

* Predictor variables that resulted in a *P* value <0.1 in the univariate regression analysis were entered into a backward stepwise multivariate regression analysis to identify significant predictors in the final model.

to cholesterol ratio at week 4 of cART were significant predictors of a low 25 (OH)D level soon after efavirenz-based cART initiation (Table 5). In a multivariate regression analysis, low Karnofsky score at baseline, high-current efavirenz plasma concentration, and high-current 4β-hydroxycholesterol to cholesterol ratio remained significant predictors of a low 25 (OH)D level at week 4 of cART (Table 5).

In a univariate logistic regression analysis, low albumin at baseline, low Karnofsky score at baseline, low cholesterol, and high 4β-hydroxycholesterol to cholesterol ratio were significant predictors SVDD at week 4 of cART (Table 5). In a multivariate analysis, low albumin at baseline and low-current plasma cholesterol level remained a significant predictor of efavirenz-induced SVDD at week 4 of cART. None of the predicting variables tested were significant predictors of low 25 (OH)D₃ levels at week 16 of cART.

4. Discussion

We performed a prospective observational study to identify predictors of a low 25 (OH)D₃ level before starting therapy and during efavirenz-based cART, anti-TB or a combination thereof in treatment-naïve HIV patients with or without TB co-infection. The main finding of this study includes: a significant association of a low plasma 25 (OH)D₃ level at baseline with TB-co-infection and HIV disease progression; efavirenz-based cART significantly reduces plasma 25 (OH)D₃ levels and this effect is more pronounced when given alone than with rifampicin-based anti-TB therapy; low plasma cholesterol level, high CYP3A activity, and high plasma efavirenz concentration are predictors of early cART-induced low 25 (OH)D₃ level.

Seasonal variation affects vitamin D status, particularly in countries far from the equator, where the lower angle of the sun and more cloud cover during the winter result in less UV-B exposure and hence low production of vitamin D in skin.^[30,31] However, lack of sunshine-derived vitamin D deficiency is not

expected in countries around the equator.^[32,33] Ethiopia is located in east Africa close to the equator (3 degree N to 14.8 degree latitude) and there is abundant sunshine to form vitamin D all year round.^[34] This study was conducted in Addis Ababa, the capital city of Ethiopia, located at latitude 9 degree N. In the present study, we found no significant influence of seasonal variation (rainy versus dry seasons) on plasma 25 (OH)D₃ concentrations. Our finding is consistent with previous findings from countries located near the equator.^[32,33]

Despite the all-year-round sunshine providing abundant UVB radiation, high prevalence of vitamin D deficiency (42%) and insufficiency (49%) in Ethiopian healthy adolescents and children was reported previously.^[35,36] HIV patients and patients with active TB have lower levels of 25 (OH)D₃ than healthy controls and patients with latent TB.^[37,38] Accordingly, all patients who participated in this study had deficient (92%) or insufficient (8%) plasma 25 (OH)D₃ levels at study enrollment, and none of them achieved a sufficient plasma 25 (OH)D₃ level (>72.5 nmol/L) during a 1-year cART follow-up period. The prevalence of vitamin D deficiency in our treatment-naïve HIV-only and TB-HIV patients from Ethiopia is quite high compared to other reports from HIV patients in East Africa. Low prevalence of vitamin D insufficiency at baseline in TB-HIV co-infected (41%) and HIV (35%)-only infected patients from Uganda has been reported.^[39] Likewise, only 9.2% and 43.6% of HIV patients from Tanzania were vitamin D-deficient and -insufficient, respectively.^[40] The finding of a high prevalence of SVDD in Ethiopian HIV patients might be because our study population consisted of very ill patients with low CD4 cell count (<200 cells/mm³) at baseline and a majority presented HIV stage 3 or 4 indicating progression to AIDS (Table 1). Association of nadir CD4 cell count <200 cells/mm³ and HIV/TB disease progression with SVDD was reported previously.^[25,37]

In our regression analysis (Table 4), low plasma 25 (OH)D₃ and SVDD at baseline was significantly associated with high viral load, which is a marker for HIV disease progression.^[41] Our

finding is in line with previous reports from the literature describing association of vitamin D deficiency with HIV disease progression.^[42] Variations in the dietary source containing vitamin D may also contribute to between-population differences in vitamin D status. TB co-infected HIV patients had significantly lower levels of 25 (OH)D₃ than patients without TB co-infection. Our finding further indicates that severe vitamin D deficiency in HIV patients is a risk factor for the development of active TB.^[40,43]

Treatment with rifampicin-based anti-TB treatment (up to 8 weeks) before the start of cART in TB-HIV co-infected patients resulted in no significant change in 25 (OH)D₃ levels from baseline. However, a significant reduction of plasma 25 (OH)D₃ levels was noted soon after initiating cART in both HIV-only and TB-HIV co-infected patients treated. The effect of efavirenz-based cART in lowering 25 (OH)D₃ levels was more pronounced when given alone than when given together with rifampicin-based anti-TB therapy. In HIV only cohort, initiation of efavirenz-based cART increased the proportion of patients with SVDD by 3-fold at week 4 of cART, which persisted until week 16. As therapy continued, vitamin D levels were gradually restored, but 42% of the patients still had SVDD after 48 weeks of cART (Table 3). In contrast, in TB-HIV patients on anti-TB co-treatment initiation of efavirenz-based cART increased the proportion of patients with SVDD from 57% at baseline to 70% and 72% at 4 and 16 weeks of cART, respectively.

Previous *in vitro* studies suggested that a low 25 (OH)D₃ level may be related to an increased catabolism of 25 (OH)D through CYP3A4 enzyme induction.^[18] In line with this, we found a significant correlation between CYP3A enzyme activity, as measured by 4 β -hydroxycholesterol to cholesterol ratio, and low 25 (OH)D concentration both at baseline and week 4 of cART. But as therapy continued, no significant association between CYP3A activity and vitamin D status was observed. We previously reported a significant association between plasma efavirenz concentration and 4 β -hydroxycholesterol/cholesterol ratio in HIV patients.^[10] In the present study, we found a significant correlation between plasma efavirenz concentration and 25 (OH)D₃ concentration. Vitamin D is involved in the regulation of CYP3A enzyme expression.^[18,19] CYP3A4 enzyme activity and expression are induced by 1,25-(OH)D₃ via the vitamin D receptor.^[44] In addition, CYP3A catalyzes the 4-hydroxylation of 25 (OH)D₃ and hence CYP3A induction may contribute to drug-induced vitamin D deficiency.^[18,19]

Rifampicin is a more potent inducer of CYP3A than efavirenz.^[14] Thus, it is plausible to expect a more pronounced effect of rifampicin in lowering the 25 (OH)D₃ level compared to efavirenz-based cART alone. However, we found no significant effect of rifampicin-based anti-TB therapy (given before the initiation of cART), on the vitamin D status (Table 3). Based on our finding, the contribution of CYP3A enzyme induction by rifampicin per se may not be the major underlying mechanism, and perhaps other mechanism such as cholesterol-increasing effect of rifampicin may play a role to counterbalance the effect of efavirenz in modulating vitamin D status as discussed below. We found no significant correlations between CYP3A activity and vitamin D status at 16 and 48 weeks of cART. Probably, the importance of CYP3A catalyzed vitamin D metabolism may diminish, with the increased wellbeing of the patient during long-term cART. Indeed, improved CD4 recovery and vitamin D repletion in HIV patients on cART were reported recently. Consistent with our finding, 25 (OH)D levels have recently been shown to decline up to 24 weeks following initiation of efavirenz-based cART, but to stabilize thereafter.^[45]

Previously, we reported that initiation of efavirenz-based cART in HIV patients significantly lowers the plasma cholesterol level transiently.^[10] 7-dehydrocholesterol is a precursor to both cholesterol and vitamin D. Thus, efavirenz-based cART lowers the vitamin D level possibly by modulating the precursor concentration. To evaluate this, we monitored the change in plasma 25 (OH)D₃ alongside the corresponding plasma cholesterol level in each patient before starting therapy and at different time points during treatment. Notably, TB-HIV co-infected patients had significantly lower pretreatment plasma concentration of both cholesterol and vitamin D compared to HIV-only patients. A similar pattern of change in plasma cholesterol level and vitamin D status over time was observed during cART, particularly when given alone (Fig. 3). Interestingly, initiation of efavirenz treatment significantly lowered both the plasma cholesterol and 25 (OH)D₃ level dramatically in both treatment groups, but this effect was more pronounced when efavirenz-based cART was given alone than with rifampicin-based anti-TB co-treatment. Indeed, there was a significant positive correlation between plasma cholesterol level and vitamin D status during cART, particularly at week 4. Though not significant, a similar trend was observed at 16 and 48 weeks of cART. This may indicate that reduction in 25 (OH)D₃ concentration by early initiation of efavirenz might be secondary to change in cholesterol concentration. In line with this, a recent *in vitro* study reported that treatment of cells with cholesterol resulted in a 3-fold increase in vitamin D relative to cholesterol synthesis, demonstrating that cholesterol feeds back via 7-dehydrocholesterol reductase (DHCR7) increasing vitamin D production.^[46] Indeed our results indicate that rifampicin-based anti-TB therapy alone significantly increased the plasma cholesterol level (Fig. 2). Thus, rifampicin-based anti-TB treatment might counterbalance the vitamin D-lowering effect of efavirenz to some extent by increasing the plasma cholesterol level and hence the vitamin D level in TB-HIV patients co-treated with rifampicin.

Increased plasma levels of 25 (OH)D₃ after 1 year of cART has been reported previously in Japanese male HIV patients.^[47] Similarly, our study shows that at the 48th week of cART in HIV-only patients, both the plasma levels of 25 (OH)D₃ and cholesterol levels restored gradually, which may be because of overall improved health status. In diverse HIV-infected populations, Vitamin D insufficiency and deficiency are associated with HIV disease progression, and virological failure after antiretroviral therapy initiation.^[45] Recent randomized clinical trials indicated that vitamin D supplementation improved CD4 recovery and vitamin D repletion suggesting potential benefit on immunologic recovery during cART.^[48,49]

4.1. Study Limitations

This study has some limitations. First, the study participants consisted of patients who had CD4 cell count <200 at baseline, following the WHO and Ethiopian national HIV treatment guideline valid during the study period. Thus, the result may not be directly extrapolatable to patients with high CD4 cell count at baseline. Second, since vitamin D status is influenced by genetic variations, darker skin, exposure to the sunshine, and dietary sources, the study result from a single study location and Ethiopian population may not be applicable for genetically diverse non-black populations living in the nontropical zone.

5. Conclusions

The finding of this study suggests TB incidence and HIV disease progression are associated with low levels of 25 (OH)D₃ at baseline. Early initiation of efavirenz-based cART is associated with low plasma 25 (OH)D₃ levels and high incidence of SVDD, which sustains up to 16th week of cART. This effect is most pronounced when given alone than with rifampicin-based anti-TB co-treatment. As cART continues, 25 (OH)D₃ levels restore gradually with time. Rifampicin-based anti-TB treatment initiated before cART has no significant effect on the 25 (OH)D₃ levels. High plasma efavirenz concentration, high CYP3A activity, and low plasma cholesterol are predictors of early cART-induced low plasma 25 (OH)D₃ level. Considering the very low levels of 25 (OH)D₃ in both patient groups, supplementary vitamin D may be beneficial not only for TB-HIV patients, but also for HIV-only patients at the initiation of cART and anti-TB treatment.

Acknowledgments

The authors are grateful to the study participants and Hospital staffs for their contribution during data collection.

References

- Turnbull ER, Drobniewski F. Vitamin D supplementation: a comprehensive review on supplementation for tuberculosis prophylaxis. *Expert Rev Respir Med* 2015;9:269–75.
- Sudfeld CR, Wang M, Aboud S, et al. Vitamin D and HIV progression among Tanzanian adults initiating antiretroviral therapy. *PLoS One* 2012;7:e40036.
- Klassen KM, Fairley CK, Chen M, et al. Vitamin D deficiency may be associated with a more rapid decline in CD4 cell count to <350 cells/microL in untreated HIV-infected adults. *Curr HIV Res* 2015;13:517–23.
- Calvo MS, Whiting SJ, Barton CN. Vitamin D intake: a global perspective of current status. *J Nutr* 2005;135:310–6.
- Adeyemi OM, Agniel D, French AL, et al. Vitamin D deficiency in HIV-infected and HIV-uninfected women in the United States. *J Acquir Immune Defic Syndr* 2011;57:197–204.
- Brown TT, McComsey GA. Association between initiation of antiretroviral therapy with efavirenz and decreases in 25-hydroxyvitamin D. *Antivir Ther* 2010;15:425–9.
- Conrado T, Miranda-Filho Dde B, Ximenes RA, et al. Vitamin D deficiency in HIV-infected women on antiretroviral therapy living in the tropics. *J Int Assoc Physicians AIDS Care (Chic)* 2011;10:239–45.
- Fux CA, Baumann S, Furrer H, et al. Is lower serum 25-hydroxy vitamin D associated with efavirenz or the non-nucleoside reverse transcriptase inhibitor class? *AIDS* 2011;25:876–8.
- Shahar E, Segal E, Rozen GS, et al. Vitamin D status in young HIV infected women of various ethnic origins: incidence of vitamin D deficiency and possible impact on bone density. *Clin Nutr* 2013;32:83–7.
- Habtewold A, Amogne W, Makonnen E, et al. Pharmacogenetic and pharmacokinetic aspects of CYP3A induction by efavirenz in HIV patients. *Pharmacogenomics J* 2013;13:484–9.
- Lawn SD, Meintjes G, McMiller H, et al. Management of HIV-associated tuberculosis in resource-limited settings: a state-of-the-art review. *BMC Med* 2013;11:253.
- Yimer G, Ueda N, Habtewold A, et al. Pharmacogenetic & Pharmacokinetic Biomarker for Efavirenz Based ARV and Rifampicin Based Anti-TB Drug Induced Liver Injury in TB-HIV Infected Patients. *PLoS One* 2011;6:e27810.
- Mukonzo JK, Okwera A, Nakasujja N, et al. Influence of efavirenz pharmacokinetics and pharmacogenetics on neuropsychological disorders in Ugandan HIV-positive patients with or without tuberculosis: a prospective cohort study. *BMC Infect Dis* 2013;13:261.
- Hariparsad N, Nallani SC, Sane RS, et al. Induction of CYP3A4 by efavirenz in primary human hepatocytes: comparison with rifampin and phenobarbital. *J Clin Pharmacol* 2004;44:1273–81.
- Ngaimisi E, Mugusi S, Minzi O, et al. Effect of Rifampicin and CYP2B6 Genotype on Long-Term Efavirenz Autoinduction and Plasma Exposure in HIV Patients With or Without Tuberculosis. *Clin Pharmacol Ther* 2011;90:406–13.
- Schmiedlin-Ren P, Thummel KE, Fisher JM, et al. Expression of enzymatically active CYP3A4 by Caco-2 cells grown on extracellular matrix-coated permeable supports in the presence of 1alpha,25-dihydroxyvitamin D₃. *Mol Pharmacol* 1997;51:741–54.
- Drocourt L, Ourlin JC, Pascussi JM, et al. Expression of CYP3A4, CYP2B6, and CYP2C9 is regulated by the vitamin D receptor pathway in primary human hepatocytes. *J Biol Chem* 2002;277:25125–32.
- Wang Z, Lin YS, Zheng XE, et al. An inducible cytochrome P450 3A4-dependent vitamin D catabolic pathway. *Mol Pharmacol* 2012;81:498–509.
- Wang Z, Schuetz EG, Xu Y, et al. Interplay between vitamin D and the drug metabolizing enzyme CYP3A4. *J Steroid Biochem Mol Biol* 2013;136:54–8.
- Habtewold A, Amogne W, Makonnen E, et al. Long-term effect of efavirenz autoinduction on plasma/peripheral blood mononuclear cell drug exposure and CD4 count is influenced by UGT2B7 and CYP2B6 genotypes among HIV patients. *J Antimicrob Chemother* 2011;66:2350–61.
- Ngaimisi E, Mugusi S, Minzi O, et al. Long-term efavirenz autoinduction and its effect on plasma exposure in HIV patients. *Clin Pharmacol Ther* 2010;88:676–84.
- Ngaimisi E, Minzi O, Mugusi S, et al. Pharmacokinetic and pharmacogenomic modelling of the CYP3A activity marker 4beta-hydroxycholesterol during efavirenz treatment and efavirenz/rifampicin co-treatment. *J Antimicrob Chemother* 2013;13:484–9.
- Mukonzo JK, Owen JS, Ogwal-Okeng J, et al. Pharmacogenetic-based efavirenz dose modification: suggestions for an African population and the different CYP2B6 genotypes. *PLoS One* 2014;9:e86919.
- Mukonzo JK, Nanzigu S, Waako P, et al. CYP2B6 genotype, but not rifampicin-based anti-TB cotreatments, explains variability in long-term efavirenz plasma exposure. *Pharmacogenomics* 2014;15:1423–35.
- Welz T, Childs K, Ibrahim F, et al. Efavirenz is associated with severe vitamin D deficiency and increased alkaline phosphatase. *AIDS* 2010;24:1923–8.
- Habtewold A, Makonnen E, Amogne W, et al. Is there a need to increase the dose of efavirenz during concomitant rifampicin-based antituberculosis therapy in sub-Saharan Africa? The HIV-TB pharmagene study. *Pharmacogenomics* 2015;16:1047–64.
- Holick MF, Binkley NC, Bischoff-Ferrari HA, et al. Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab* 2011;96:1911–30.
- Nylen H, Bjorkhem-Bergman L, Ekstrom L, et al. Plasma levels of 25-hydroxyvitamin D₃ and in vivo markers of cytochrome P450 3A activity in Swedes and Koreans: effects of a genetic polymorphism and oral contraceptives. *Basic Clin Pharmacol Toxicol* 2014;115:366–71.
- Burhenne J, Matthee AK, Pasakova I, et al. No evidence for induction of ABC transporters in peripheral blood mononuclear cells in humans after 14 days of efavirenz treatment. *Antimicrob Agents Chemother* 2010;54:4185–91.
- Klingberg E, Olerod G, Konar J, et al. Seasonal variations in serum 25-hydroxy vitamin D levels in a Swedish cohort. *Endocrine* 2015;49:800–8.
- Bolland MJ, Chiu WW, Davidson JS, et al. The effects of seasonal variation of 25-hydroxyvitamin D on diagnosis of vitamin D insufficiency. *N Z Med J* 2008;121:63–74.
- Holick MF. McCollum Award Lecture, 1994: vitamin D—new horizons for the 21st century. *Am J Clin Nutr* 1994;60:619–30.
- Wejse C, Olesen R, Rabna P, et al. Serum 25-hydroxyvitamin D in a West African population of tuberculosis patients and unmatched healthy controls. *Am J Clin Nutr* 2007;86:1376–83.
- Gebreegziabher T, Stoecker BJ. Vitamin D insufficiency in a sunshine-sufficient area: southern Ethiopia. *Food Nutr Bull* 2013;34:429–33.
- Wakayo T, Belachew T, Vatanparast H, et al. Vitamin D deficiency and its predictors in a country with thirteen months of sunshine: the case of school children in central Ethiopia. *PLoS One* 2015;10:e0120963.
- Herrador Z, Sordo L, Gadsis E, et al. Micronutrient deficiencies and related factors in school-aged children in Ethiopia: a cross-sectional study in Libo Kemkem and Fogera districts, Amhara Regional State. *PLoS One* 2014;9:e112858.
- Theodorou M, Serste T, Van Gossom M, et al. Factors associated with vitamin D deficiency in a population of 2044 HIV-infected patients. *Clin Nutr* 2014;33:274–9.
- Venturini E, Facchini L, Martinez-Alier N, et al. Vitamin D and tuberculosis: a multicenter study in children. *BMC Infect Dis* 2014;14:652.
- Conesa-Botella A, Goovaerts O, Massinga-Loembe M, et al. Low prevalence of vitamin D deficiency in Ugandan HIV-infected patients with and without tuberculosis. *Int J Tuberc Lung Dis* 2012;16:1517–21.

- [40] Sudfeld CR, Giovannucci EL, Isanaka S, et al. Vitamin D status and incidence of pulmonary tuberculosis, opportunistic infections, and wasting among HIV-infected Tanzanian adults initiating antiretroviral therapy. *J Infect Dis* 2013;207:378–85.
- [41] Ghani AC, de Wolf F, Ferguson NM, et al. Surrogate markers for disease progression in treated HIV infection. *J Acquir Immune Defic Syndr* 2001;28:226–31.
- [42] Viard JP, Souberbielle JC, Kirk O, et al. Vitamin D and clinical disease progression in HIV infection: results from the EuroSIDA study. *AIDS* 2011;25:1305–15.
- [43] Arnedo-Pena A, Juan-Cerdan JV, Romeu-Garcia A, et al. Vitamin D status and incidence of tuberculosis among contacts of pulmonary tuberculosis patients. *Int J Tuberc Lung Dis* 2015;19:65–9.
- [44] Wang K, Chen S, Xie W, et al. Retinoids induce cytochrome P450 3A4 through RXR/VDR-mediated pathway. *Biochem Pharmacol* 2008;75:2204–13.
- [45] Havers F, Smeaton L, Gupte N, et al. 25-Hydroxyvitamin D insufficiency and deficiency is associated with HIV disease progression and virological failure post-antiretroviral therapy initiation in diverse multinational settings. *J Infect Dis* 2014;210:244–53.
- [46] Prabhu AV, Luu W, Sharpe LJ, et al. Cholesterol-Mediated Degradation of 7-Dehydrocholesterol Reductase Switches the Balance from Cholesterol to Vitamin D Synthesis. *J Biol Chem* 2016.
- [47] Koga I, Seo K, Yoshino Y, et al. Increase of 25-hydroxyvitamin D levels after initiation of combination antiretroviral therapy. *J Infect Chemother* 2015;21:737–41.
- [48] Coelho L, Cardoso SW, Luz PM, et al. Vitamin D3 supplementation in HIV infection: effectiveness and associations with antiretroviral therapy. *Nutr J* 2015;14:81.
- [49] Steenhoff AP, Schall JI, Samuel J, et al. Vitamin D (3) supplementation in Batswana children and adults with HIV: a pilot double blind randomized controlled trial. *PLoS One* 2015;10:e0117123.