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Association of sexually-transmitted infection and African–American race with *Streptococcus agalactiae* colonization in pregnancy

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Abstract

Background: Group B *Streptococcus* (GBS) remains a significant cause of neonatal infection, but the maternal risk factors for GBS colonization remain poorly defined. We hypothesized that there may be an association between antibiotic exposure during pregnancy and GBS colonization and/or the presence of inducible clindamycin resistance (iCLI-R) in GBS isolates from GBS-colonized pregnant women.

Methods: A retrospective cohort study was performed at Louisiana State University Health Sciences Center – Shreve-port including demographic and clinical data from 1513 pregnant women who were screened for GBS between July 1, 2009 and December 31, 2010.

Results: Among 526 (34.8%) women who screened positive for GBS, 124 (23.6%) carried GBS strains with iCLI-R (GBS-iCLI-R). While antibiotic exposure, race, sexually-transmitted infection (STI) in pregnancy, GBS colonization in prior pregnancy and BMI were identified as risk factors for GBS colonization in univariate analyses, the only independent risk factors for GBS colonization were African–American race (AOR = 2.142; 95% CI = 2.092–3.861) and STI during pregnancy (AOR = 1.309; 95% CI = 1.035–1.653). Independent risk factors for GBS-iCLI-R among women colonized with GBS were non-African–American race (AOR = 2.13; 95% CI = 1.20–3.78) and younger age (AOR = 0.94; 95% CI = 0.91–0.98). Among GBS-colonized women with an STI in the current pregnancy, the only independent risk factor for iCLI-R was *Chlamydia trachomatis* infection (AOR = 4.31; 95% CI = 1.78–10.41).

Conclusions: This study identified novel associations for GBS colonization and colonization with GBS-iCLI-R. Prospective studies will improve our understanding of the epidemiology of GBS colonization during pregnancy and the role of antibiotic exposure in alterations of the maternal microbiome.

Keywords: Group B *Streptococcus*, Pregnancy, Inducible clindamycin resistance, Azithromycin, Sexually transmitted infections

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Introduction

Streptococcus agalactiae (Group B Streptococcus (GBS)) is a dynamic colonizer of the gastrointestinal and genitourinary tracts, frequently causing urinary tract infections, chorioamnionitis, postpartum endometritis, and bacteremia in pregnant women [1, 2]. GBS is also the most common cause of sepsis and meningitis in infants



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younger than three months of age [3]. With the nearelimination of *Streptococcus pneumoniae* and *Haemophi*lus influenzae meningitis due to vaccination, GBS is now the most frequent cause of meningitis in childhood [4, 5]. Approximately 50% of infants born to GBS-colonized mothers acquire GBS in utero or during parturition, with 1–2% of colonized infants developing GBS disease [3]. The implementation of universal screening for GBS in pregnancy and use of intrapartum antibiotic prophylaxis (IAP) reduced the incidence of early-onset GBS disease in the United States from 1.8 cases per 1000 live births to 0.26 cases per 1000 live births from the early 1990's to 2010 [6]. However, the incidence of late-onset GBS has remained unaffected, with ~0.26 cases per 1000 infants diagnosed annually [6]. As such, understanding the risk factors for maternal GBS colonization continues to be an important facet of the development of new strategies to improve maternal and neonatal outcomes.

We previously reported high rates of GBS colonization and high rates of inducible clindamycin resistance (iCLI-R) among GBS isolates from women who received obstetric care at LSUHSC-Shreveport [7]. Because of the high rates of sexually-transmitted infections (STIs) in our population and the common resistance mechanisms for iCLI-R that have been described for GBS and *Staphylococcus aureus*, we hypothesized that exposure to antibiotics during pregnancy may be a risk factor for colonization with GBS and/or colonization with a GBS strain that displays an iCLI-R phenotype. To address this question we analyzed the clinical and demographic data from the medical records of the group of women on which our earlier report was based.

Methods

The LSUHSC-S Institutional Review Board for Human Subjects Research approved this study protocol prior to data collection.

Study design

We identified pregnant women who were screened at 35–37 weeks gestation by vagino-rectal swab for GBS, at prenatal visits between 1 July 2009 and 31 December 2010 [7]. Antimicrobial susceptibility testing, including tests for the presence of inducible resistance to clindamycin, was performed on all GBS isolates from vagino-rectal swabs received by the UH-S Microbiology laboratory personnel according to Clinical Laboratory Standards Institute (CLSI) guidelines. We analyzed a total of 1522 medical records for pertinent clinical and demographic data. Subjects were excluded from the analyses if their records were not obtainable after more than four months of requests (n=8) or if they were employees or spouses of employees involved in the study (n=1) resulting in

data on 1513 pregnant women available for analysis in the study population. The main outcomes of interest were: (1) GBS positivity and (2) iCLI-R among GBS positive women. We studied the relationship between each of our outcomes and antibiotic use in pregnancy (exclusive of intrapartum antimicrobial prophylaxis for GBS colonization), considering both specific antibiotics and antibiotic class. We defined "sexually-transmitted infections (STI) during the current pregnancy" as *C. trachomatis* (CT), *Neisseria gonorrheae* (NG) or *Trichomonas vaginalis* (TV), as confirmed by routine clinical testing at LSUHSC-S.

Statistical analyses

The chi-square test was used to identify categorical variables (e.g. antibiotic exposure by class and individually, STI during pregnancy (CT, NG and TV individually and as a composite variable) significantly associated with GBS colonization and those significantly associated with iCLI-R among GBS positive women. Each antibiotic and antibiotic class was considered separately for association with GBS colonization. The two-sample t-test was used to determine if continuous variables (e.g., age, bodymass index (BMI), gravidity) are significantly associated with GBS colonization for all subjects and with iCLI-R among GBS positive women, as well as among GBS positive women with STI. Multiple logistic regression analysis was used to determine independent risk factors for GBS colonization as well as risk factors for iCLI-R among GBS positive women and among GBS positive women with STI. Our strategy for the multiple logistic regression analysis was to use as independent predictors those found to be significantly associated with the outcome from univariate analysis. We likewise used it for our post hoc analysis to determine risk factors for GBS among women with STI.

Missing data were excluded from the statistical analyses, as the percentage of missing data for the different observed variables accounted for only 0.1–6.2% of the records available for analysis, except for "GBS in prior pregnancy," where 46.3% were appropriately "not applicable" for primigravid mothers. Data on the number of missing information for each variable are reported in the tables.

Results

Demographic and clinical characteristics of the study population

The characteristics of subjects (n=1513) whose records were included in these analyses are detailed in Table 1. A majority of subjects were African–American (AA) (74.7%) and had a term delivery (91.4%). Nearly 35% had GBS colonization, 37% had a history of STI, and 55.1%

Table 1 Characteristics of Pregnant Women in Study (N = 1513)

Characteristic	Number (%) ^a Or mean ± SD, median, range ^a	Number Unknown/NA (%)	
GBS colonization	526 (34.8)	2 (0.1) ^b	
STI during this pregnancy	536 (37.0)	64 (4.2)	
C. trachomatis	332 (22.9)	64 (4.2)	
N. gonorrhea	117 (8.1)	64 (4.2)	
T. vaginalis	296 (20.4)	64 (4.2)	
Antibiotic exposure ^c	786 (55.1)	87 (5.8)	
HIV positive	14 (0.1)	54 (3.6)	
Term delivery	1343 (91.4)	44 (2.9)	
GBS in prior pregnancy	189 (23.2)	700 (46.3)—NA	
Race		22 (1.5)	
African–American	1114 (74.7)		
White	201 (13.5)		
Hispanic	141 (9.5)		
Other	35 (2.3)		
Age (years)	24.4±5.8, 23.0, 13-49	18 (1.2)	
Weight (lbs)	191.6 ± 49.8, 182.0, 96-408	69 (4.6)	
Height (in)	64.0 ± 2.9, 64.0, 49-74	94 (6.2)	
BMI	32.8 ± 7.9, 31.3, 18.8-70.1	94 (6.2)	
Gravidity	$2.7 \pm 1.8, 2.0, 1-20$	2 (0.1)	

^a Calculated from non-missing values

received antibiotic treatment during pregnancy. The median age at the time of delivery was 23.0 years and that of the median BMI was 31.3 kg/m^2 . We tested the demographic and clinical factors listed in Table 1 for association with GBS colonization in the current pregnancy.

Risk factors for GBS colonization in pregnancy

African–Americans, those with STI, antibiotic exposure, and who were GBS positive in a prior pregnancy had significantly higher GBS colonization rates in the current pregnancy than those with no STI, no antibiotic exposure, non-AA race and no GBS in a prior pregnancy (Table 2a). Among continuous variables, only BMI had significant association with GBS colonization. GBS positive women had higher BMI than GBS negative women (33.7 \pm 8.2 vs. 32.3 \pm 7.8, p < 0.01). Age and gravidity were similar between GBS-positive and GBS-negative women. Although BMI was significantly associated with GBS in univariate analysis, we did not include it as a predictor variable in the multiple logistic regression model because of its significant association with AA race and because of the imprecision of BMI calculations that are not based on pre-pregnancy weights. AA women had higher BMI than non-AA women, but pre-pregnancy weights within one year prior to initiation of prenatal care were only available for a minority of subjects (data not shown). Independent risk factors for GBS colonization were STI during pregnancy and AA race as determined by multivariate analysis (Table 2b). The odds for GBS colonization among AA women was 2.142 times the odds of GBS colonization for non-AA women, while the odds of GBS colonization for women with STI were 1.309 times the odds for women without STI during pregnancy. Using the Pearson test for goodness-of-fit, the combination of AA race and STI yielded the highest *p*-value, suggesting this combination of factors provides the most accurate model of risk factors for GBS colonization. The characteristics of the 526 women with GBS colonization are summarized in Table 3.

Risk factors for iCLI-R among women with GBS strains

Only race and age were significantly associated with iCLR among the GBS colonized women in univariate analyses (Table 4a). Non-AA women had a higher rate of colonization with GBS-iCLR than AA women (33.3% vs. 22.2%, p=0.04). Women with GBS-iCLR were younger than those without GBS-iCLR (23.0 \pm 5.1 vs. 24.4 \pm 5.8, p=0.01). Race and age were also the independent risk factors for iCLR among GBS positive women as determined by multivariate analysis (Table 4b). Adjusted for

b Among the 1513 pregnant women, 2 have missing values for GBS and these 2 women have STI. The STI of these 2 women cannot be correlated to their unknown GBS. For purposes correlating STI and GBS, they are among the 66 with missing values (Table 2)

^c Includes all antibiotic treatments for any infections diagnosed during the pregnancy, excluding IAP

Table 2 Factors associated with GBS colonization (N = 1513). (a) Categorical factors and continuous variables associated with GBS by univariate analysis—number (%) or mean \pm SD, range. (B) Independent risk factors for GBS by multivariate analysis—adjusted odds ratios

Characteristic	GBS positive	GBS negative	<i>p</i> value	# Missing/NA
(a)	,	Š	•	J
Categorical variables				
STI during pregnancy	219/494 (44.3)	315/953 (33.1)	< 0.01**	66
Antibiotic exposure	313/488 (64.1)	471/936 (50.3)	< 0.01**	89
AA race	519/519 (86.7)	662/970 (68.2)	< 0.01**	24
GBS in prior pregnancy	80/268 (29.9)	109/544 (20.0)	< 0.01**	701
Continuous variables				
Age	24.1 ± 5.6 , $14-44$ (n = 520)	$24.6 \pm 5.9, 13-49$ (n = 973)	0.13	10
BMI	$33.7 \pm 8.2, 19.4-70.1$ (n = 491)	$32.3 \pm 7.8, 19-69$ (n = 928)	< 0.01**	94
Gravidity	$2.7 \pm 1.8, 1-12$ (n = 519)	$2.7 \pm 1.9, 1-20$ (n = 978)	0.97	16
Characteristic	Adjusted OR	95% CI for AOR	<i>p</i> value	# Used
(b)				
African–American race	2.142	2.092–3.861	< 0.01**	
STI during pregnancy	1.309	1.035-1.653	0.02*	
Goodness of fit (Pearson test)			0.45	1432

CI confidence interval

Table 3 Characteristics of GBS colonized women (N = 526)

Characteristic	Number (%) ^a Or mean ± SD, median, range ^a	Number Unknown/NA (%)	
iCLI-R	124 (23.6)	_	
STI during pregnancy	220 (44.5)	32 (6.1)	
Antibiotic exposure	313 (64.1)	38 (7.2)	
HIV positive	5 (1.0)	24 (4.6)	
Term delivery	461 (90.0)	14 (2.7)	
GBS in prior pregnancy	80 (29.8)	258 (49.0)—NA	
AA race	450 (86.7)	7 (1.3)	
IAP	384 (80.2)	47 (8.9)	
Age (years)	24.1 ± 5.6 , 23.0 , $14-44$	6 (1.1)	
BMI	$33.7 \pm 8.2, 32.4, 19.4-70.1$	35 (6.7)	
Gravidity	2.7 ± 1.8 , 2.0 , $1-12$	7 (1.3)	

^a Calculated on non-missing values

age, the odds for colonization with GBS-iCLR for non-AA women were 2.13 times the odds for AA women. For every year increase in age, the adjusted odds for GBS-iCLR decreased by 0.06.

Forty-two percent of the GBS-iCLR cases (52 of 124) were seen in women with STI, so risk factors for iCLR

were analyzed post hoc among the subgroup of GBSpositive women who had STI in the current pregnancy (Table 5). Among the 220 women with STI, 62.3% were infected with C. trachomatis (CT) and 99.5% were treated with antibiotics. Among the 213 women treated with antibiotics, more than half (132 out of 213, 62%) received azithromycin (AZ). Categorical factors significantly associated with GBS-iCLR colonization were CT infection and AZ treatment (Table 6a). GBS-iCLR rates were significantly higher in women with CT and those treated with AZ than those without CT and those treated with antibiotics other than AZ (32.8% and 32.6% vs. 8.4% and 8.6%, respectively; p < 0.01). Among continuous variables, only age was significantly associated with GBS-iCLR colonization (Table 6a). Women with GBS-iCLR were significantly younger than those without GBS-iCLR (21.2 \pm 3.7 years vs. 23.4 \pm 5.2 years, p < 0.01). Thus, factors significantly associated with iCLI-R by univariate analysis were CT infection, AZ treatment and younger age. The only independent risk factor for colonization with GBS-iCLR among GBSpositive women with STI was CT infection (Table 6b). Adjusted for age and AZ treatment (factors associated with GBS-iCLR by univariate analysis), the odds

^{*} Significant association with GBS at 5% level of significance (p value < 0.05)

^{**} Significant association with GBS at 1% level of significance (p value < 0.01)

Table 4 Factors associated with GBS-iCL-R among GBS+ women (N=526). (a) Categorical factors and continuous variables associated with GBS-iCL-R by univariate analysis—number (%) or mean \pm SD, range. (b) Independent risk factors for iCL-R among GBS+ women by multivariate analysis

Characteristic	GBS-iCL-R Pos	GBS-iCL-R Neg	<i>p</i> value	Missing/NA
(a)		-	•	_
Categorical variables				
STI during pregnancy	52/113 (46.0)	168/381 (44.1)	0.72	32
Antibiotic exposure	68/111 (61.3)	245/377 (65.0	0.47	38
AA race	100/123 (81.3)	350/396 (88.4)	0.04*	7
IAP	86/109 (78.9)	298/370 (80.5)	0.71	47
Continuous variables				
Age (years)	$23.0 \pm 5.1, 15-42$ (n = 124)	$24.4 \pm 5.8, 14-44$ (n = 396)	0.014*	4
ВМІ	34 ± 8.2 , $19.4-66.1$ (n = 115)	$33.6 \pm 8.3, 20-70.1$ (n = 376)	0.67	35
Gravidity	$2.6 \pm 1.8, 1-9$ (n = 122)	$2.7 \pm 1.8, 1-12$ (n = 397)	0.33	7
Factor	Adjusted OR	95% CI for AOR	<i>p</i> value	# Used
(b)				
Non-AA race	2.131	1.201–3.782	< 0.01**	
Age	0.943	0.905-0.982	< 0.01**	
Goodness of fit (Pearson)			0.60	515

^{*} Significant Association with iCL-R at 5% level (0.01 < p value < 0.05)

Table 5 Characteristics of GBS Positive Women with STI (N = 220)

Characteristic	Number (%) a Or mean ± SD, median, range a	Number Missing/NA (%)	
iCLI-R	52 (23.6)	_	
Chlamydia trachomatis (CT)	137 (62.3)	-	
Antibiotic exposure	213 (99.5)	6 (2.7)	
AZ treatment	132 (62.0)	7 (3.2)	
AA race	208 (94.6)	-	
Term delivery	198 (92.5)	6 (2.7)	
IAP	171 (82.2)	12 (5.5)	
HIV positive	0 (0)	10 (4.4)	
GBS in prior pregnancy	36 (29.3)	97 (44.0)	
Age (years)	22.9 ± 5.0, 22.0, 15-41	3 (1.4)	
BMI	32.9 ± 8.0, 31.6, 19.4-62.3	12 (5.5)	
Gravidity	2.6 ± 1.8, 2.0, 1–10	1 (0.5)	

^a Calculated from non-missing values

for iCLR among women with CT were 4.31 (95% CI=1.78–10.41) times the odds among those without CT. Although age and AZ treatment had significant

associations with iCLR by univariate analysis, they dropped out as risk factors for iCLR in multivariate analysis because of their significant associations with CT infection (p < 0.01).

^{**} Significant Association with iCL-R at 5% level (p value < 0.01)

Table 6 Factors associated with GBS-iCL-R among GBS positive women with STI (N=220). (a) Factors associated with GBS-iCL-R among GBS positive women with STI by univariate analysis—number (%) or mean \pm SD, range. (b) Independent risk factor for iCL-R among GBS+ women with STI by multivariate analysis—adjusted odds ratio

GBS-iCL-R Pos	GBS-iCL-R Neg	<i>p</i> value	Missing/NA
	.,	•	3
45/52 (86.5)	92/168 (54.8)	< 0.01**	-
43/50 (86.0)	89/163 (54.5)	< 0.01**	7
$21.2 \pm 3.7, 15-34$ (n = 52)	$23.4 \pm 5.2, 15-41$ (n = 165)	< 0.01**	3
$31.3 \pm 6.0, 19.4-45.4$ (n = 48)	$33.4 \pm 8.5, 20-62.3$ (n = 160)	0.06	12
$2.5 \pm 2.0, 1-9$ (n = 52)	$2.6 \pm 1.7, 1-10$ (n = 167)	0.85	1
Adjusted OR	95% CI for AOR	<i>p</i> value	# Used
4.31	1.78–10.41	< 0.01	
		0.79	217
	$45/52 (86.5)$ $43/50 (86.0)$ $21.2\pm3.7, 15-34$ $(n = 52)$ $31.3\pm6.0, 19.4-45.4$ $(n = 48)$ $2.5\pm2.0, 1-9$ $(n = 52)$ Adjusted OR	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

Discussion

In this retrospective study, we correlated clinical and microbiologic data from a large population of pregnant women in order to identify risk factors for maternal colonization with GBS and GBS-iCLR. We hypothesized that antibiotic exposure in pregnancy, particularly macrolide exposure, is associated with colonization with GBS strains with inducible resistance to clindamycin (GBSiCLR) due to selective pressure on the maternal flora. Our analyses revealed that GBS colonization in pregnant women was significantly associated with STI in the current pregnancy, a novel finding, as well as AA race, a previously known risk factor. The association of GBSiCLR with non-African-American race and younger age and the association of GBS-iCLR with CT infection and azithromycin exposure among women with an STI in the current pregnancy are unique findings that suggest numerous hypotheses to be tested in future, prospective studies.

Prior studies have identified numerous risk factors for GBS infection in infants, including pre-term delivery, prolonged rupture of membranes, maternal chorioamnionitis, maternal colonization with GBS during labor and delivery, multiparity, maternal GBS colonization in a previous pregnancy, low maternal levels of anti-GBS antibody, certain maternal sexual behaviors, African–American race and GBS bacteriuria during pregnancy, many of which are also risk factors for maternal colonization during a subsequent pregnancy [8–14]. However, relatively few studies have explicitly focused on risk factors for maternal colonization with GBS or correlated

microbiologic and clinical data for large cohorts of patients. The landmark VIP study from 1984 to 1989 did not find an association of GBS colonization with STIs, but the rates of GBS colonization and STIs were significantly lower in that multi-center study and variable between the geographically distributed study sites [15]. Colonization with GBS in a prior pregnancy has been identified as a risk factor for GBS colonization in a subsequent pregnancy by several studies [9–11, 16]. African– American race, age > 21 years and marijuana use within the past four months were identified as risk factors for rectal GBS colonization among non-pregnant women in a longitudinal study [17]. The same study found that recent vaginal intercourse, vaginal yeast or E. coli colonization and abnormal vaginal flora (Nugent score ≥ 4), but not STI, hormonal contraception or specific sexual practices, were associated with vaginal GBS colonization among young adult women [17]. The study by Rocchetti and colleagues identified frequency of sexual intercourse, alterations in vaginal flora and prior miscarriage as risk factors for GBS colonization in pregnancy [18].

Studies comparing the vaginal flora of GBS+ and GBS— women in pregnancy have yielded conflicting results, leaving uncertainty about the importance of alterations of the vaginal microbiome as a contributing or protective factor in GBS colonization [19, 20]. Recent studies failed to detect an association between GBS and lactobacilli in pregnant Polish and Guatemalan women using culture-based and molecular testing, respectively [20, 21]. A large study of the vaginal microbiome in non-pregnant women found strong positive and negative associations between

GBS colonization and several bacterial taxa, but no association between GBS and *Lactobacillus* species was found, suggesting that vaginal microbiome community state type is not a reliable predictor of GBS colonization in non-pregnant women [22]. Longitudinal studies of pregnant women have begun to characterize the diversity and dynamics of the vaginal microbiome and identified unique community state types and, more recently, the relative abundance of various *Lactobacilli* as important correlates of premature delivery [23, 24]. These emerging studies and novel research modalities suggest the potential use of non-culture techniques to assess risk for adverse maternal and infant outcomes.

While all known GBS isolates retain susceptibility to penicillin in vitro, GBS isolates with resistance to erythromycin and clindamycin have been increasingly reported worldwide, including at our institution [7, 25-27]. Furthermore, GBS isolates with reduced penicillin susceptibility have been identified at several sites around the world, highlighting the need for continued active surveillance for alterations in GBS susceptibility that may again alter the strategies for prevention of GBS disease [28, 29]. The identification of clinical factors associated with changing antimicrobial susceptibility profiles, especially antibiotic exposures, is critical to understanding the role of antimicrobial pharmacodynamics in the promotion of antibiotic resistance in commensal and opportunistic microbes that impact maternal and infant outcomes, especially for drugs like azithromycin that have long half-lives and are used in large doses for treatment of C. trachomatis infection [30, 31]. Furthermore, the consideration of using wide-spread prophylactic dosing of azithromycin for prevention of adverse maternal, neonatal and infant outcomes in underdeveloped countries portends a significant risk of promoting antibiotic resistance in areas of the world that are least-equipped to combat multi-drug resistant organisms [32, 33].

This study has several notable strengths, including the correlation of microbial and clinical data from a large study population, and the identification of novel clinical variables associated with GBS and GBS-iCLR colonization. The predominance of African-Americans in the study is also a strength, given that few studies have focused on this population previously. Weaknesses of this study include the lack of data on GBS serotypes, multilocus sequence types, and molecular characterization of resistance genes prevalent in the GBS isolates, all of which were not possible in the present study, but should be performed in future prospective studies. Additionally, the lack of information regarding maternal sexual, dietary, social and hygiene practices limits the comparability of these results to some prior studies and precludes the evaluation of some potentially confounding factors.

The association of GBS-iCLI-R with C. trachomatis infection and azithromycin exposure supports the hypothesis that azithromycin exerts selective pressure on GBS, potentially through the erm genes that mediate Macrolide-Lincosamide-Streptogramin resistance (MLS_B resistance) in GBS and staphylococci [34, 35]. Erythromycin and clindamycin-resistant Gram-positive organisms often carry erythromycin ribosomal methylation (erm) genes accounting for constitutive (cMLS_B-phenotype) and inducible (iMSL_B-phenotype) resistance in GBS isolates [36]. Genes contributing to erythromycin resistance include ermA(TR), ermB, and mefA, which have been found among macrolide-resistant GBS isolates [25, 37–39]. In a large collection of GBS isolates from numerous sites in Brazil, erythromycin resistance was not associated with particular GBS serotypes, suggesting the independent acquisition of erythromycin resistance genes [37]. Prospective molecular analysis will be necessary to determine the precise genetic mechanism(s) that mediate iCLI-R in our population and the potential changes over time in resistance patterns. These data support a particular focus on antimicrobial stewardship and infection control practices in pregnancy and the need for prospective studies to characterize the impact of antibiotic exposures on the maternal microbiome. Furthermore, they support the need for intensive education of patients and providers regarding prevention of STIs in pregnancy, especially in communities with high rates of STIs.

Conclusion

In summary, this large retrospective study of clinical and microbiological data identified STI potential risk factor for GBS colonization in pregnancy in a population with high rates of STIs. Additionally, the receipt of azithromycin during pregnancy was significantly associated with colonization due to a GBS strain with inducible clindamycin resistance among women with STI in the current pregnancy. These findings have significant implications for the promotion of antimicrobial resistance, especially when public health interventions include widespread use of antibiotics. Prospective studies will be necessary to correlate molecular mechanisms of antibiotic resistance, GBS serotypes and genotypes, clinical factors and changes in the maternal microbiome with maternal and neonatal outcomes and to determine the likelihood that antimicrobial resistance will persist in communities.

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Authors' contributions

GAC was the Principal Investigator for the study, carried out primary statistical analyses and drafted the manuscript; SL, KK, EG, BS, SMA, SK, TC and EL were responsible for data collection, assisted with analyses and performed

background research; GC was the statistical analyst and assisted with preparation of the manuscript; JAB and JAV conceived of the project, supervised data collection, assisted statistical analyses and completed the manuscript for publication. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets generated during and/or analyzed during the current study are not publicly available due to inclusion of Protected Health Information but are available from the corresponding author in de-identified form on reasonable request.

Ethics approval and consent to participate

The conduct of this study was approved by the Institutional Review Board for Human Subjects Research at Louisiana State University Health Sciences Center Shreveport prior to initiation. The IRB study number was 00000152.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests related to the conduct of this study or the content of the manuscript.

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References

- Honig E, Mouton JW, van der Meijden WI. The epidemiology of vaginal colonisation with group B streptococci in a sexually transmitted disease clinic. Eur J Obstet Gynecol Reprod Biol. 2002;105(2):177–80.
- Watt JP, Schuchat A, Erickson K, Honig JE, Gibbs R, Schulkin J. Group B streptococcal disease prevention practices of obstetrician-gynecologists. Obstet Gynecol. 2001;98(1):7–13.
- Edwards MS, Nizet V, Baker CJ. Group B streptococcal infections. Infectious diseases of the fetus and newborn. 7th ed. Philadelphia: W.B. Saunders; 2011. p. 419–69.
- Baker CJ. The spectrum of perinatal group B streptococcal disease. Vaccine. 2013;31(Suppl 4):D3-6.
- 5. Thigpen MC, Whitney CG, Messonnier NE, et al. Bacterial meningitis in the United States, 1998–2007. N Engl J Med. 2011;364(21):2016–25.
- Schrag SJ, Verani JR. Intrapartum antibiotic prophylaxis for the prevention of perinatal group B streptococcal disease: experience in the United States and implications for a potential group B streptococcal vaccine. Vaccine. 2013;31(Suppl 4):D20–6.
- Capraro GA, Rambin ED, Vanchiere JA, Bocchini JA Jr, Matthews-Greer JM. High rates of inducible clindamycin resistance among prenatal group B streptococcal isolates in one northwest Louisiana academic medical center. J Clin Microbiol. 2013;51(7):2469–513.
- Petersen KB, Johansen HK, Rosthoj S, Krebs L, Pinborg A, Hedegaard M. Increasing prevalence of group B streptococcal infection among pregnant women. Dan Med J. 2014;61(9):A4908.
- Colicchia LC, Lauderdale DS, Du H, Adams M, Hirsch E. Recurrence of group B streptococcus colonization in successive pregnancies. J Perinatol. 2015;35(3):173–6.
- Page-Ramsey SM, Johnstone SK, Kim D, Ramsey PS. Prevalence of group B Streptococcus colonization in subsequent pregnancies of group B

- Streptococcus-colonized versus noncolonized women. Am J Perinatol. 2013;30(5):383–8.
- Tam T, Bilinski E, Lombard E. Recolonization of group B Streptococcus (GBS) in women with prior GBS genital colonization in pregnancy. J Matern Fetal Neonatal Med. 2012;25(10):1987–9.
- Taylor JK, Hall RW, Dupre AR. The incidence of group B streptococcus in the vaginal tracts of pregnant women in central Alabama. Clin Lab Sci. 2002;15(1):16–7.
- Berardi A, Rossi C, Guidotti I, et al. Factors associated with intrapartum transmission of group B Streptococcus. Pediatr Infect Dis J. 2014;33(12):1211–5.
- Schuchat A, Oxtoby M, Cochi S, et al. Population-based risk factors for neonatal group B streptococcal disease: results of a cohort study in metropolitan Atlanta. J Infect Dis. 1990;162(3):672–7.
- Regan JA, Klebanoff MA, Nugent RP. The epidemiology of group B streptococcal colonization in pregnancy. Vaginal Infections and Prematurity Study Group. Obstet Gynecol. 1991;77(4):604–10.
- Turrentine MA, Colicchia LC, Hirsch E, et al. Efficiency of screening for the recurrence of antenatal group b streptococcus colonization in a subsequent pregnancy: a systematic review and meta-analysis with independent patient data. Am J Perinatol. 2016;33(5):510–7.
- Meyn LA, Krohn MA, Hillier SL. Rectal colonization by group B Streptococcus as a predictor of vaginal colonization. Am J Obstet Gynecol. 2009;201(1):76.e1-7.
- Rocchetti TT, Marconi C, Rall VL, Borges VT, Corrente JE, da Silva MG. Group B streptococci colonization in pregnant women: risk factors and evaluation of the vaginal flora. Arch Gynecol Obstet. 2011;283(4):717–21.
- Kubota T, Nojima M, Itoh S. Vaginal bacterial flora of pregnant women colonized with group B streptococcus. J Infect Chemother. 2002;8(4):326–30.
- Brzychczy-Wloch M, Pabian W, Majewska E, et al. Dynamics of colonization with group B streptococci in relation to normal flora in women during subsequent trimesters of pregnancy. New Microbiol. 2014;37(3):307–19.
- 21. Rick AM, Aguilar A, Cortes R, et al. Group B Streptococci colonization in pregnant guatemalan women: prevalence, risk factors, and vaginal microbiome. Open Forum Infect Dis. 2017;4(1):ofx020.
- 22. Rosen GH, Randis TM, Desai PV, et al. Group B streptococcus and the vaginal microbiota. J Infect Dis. 2017;216(6):744–51.
- 23. Callahan BJ, DiGiulio DB, Goltsman DSA, et al. Replication and refinement of a vaginal microbial signature of preterm birth in two racially distinct cohorts of US women. Proc Natl Acad Sci USA. 2017;114(37):9966–71.
- DiGiulio DB, Callahan BJ, McMurdie PJ, et al. Temporal and spatial variation of the human microbiota during pregnancy. Proc Natl Acad Sci USA. 2015;112(35):11060–5.
- 25. Desjardins M, Delgaty KL, Ramotar K, Seetaram C, Toye B. Prevalence and mechanisms of erythromycin resistance in group A and group B streptococcus: implications for reporting susceptibility results. J Clin Microbiol. 2004;42(12):5620–3.
- Hayes K, Cotter L, Barry L, O'Halloran F. Emergence of the L phenotype in group B Streptococci in the South of Ireland. Epidemiol Infect. 2017;145(16):3535–42.
- Morales WJ, Dickey SS, Bornick P, Lim DV. Change in antibiotic resistance of group B streptococcus: impact on intrapartum management. Am J Obstet Gynecol. 1999;181(2):310–4.
- 28. Seki T, Kimura K, Reid ME, et al. High isolation rate of MDR group B streptococci with reduced penicillin susceptibility in Japan. J Antimicrob Chemother. 2015;70(10):2725–8.
- 29. Banno H, Kimura K, Tanaka Y, et al. Analysis of multidrug resistant group B streptococci with reduced penicillin susceptibility forming small, less hemolytic colonies. PLoS ONE. 2017;12(8):e0183453.
- Asaka T, Manaka A, Sugiyama H. Recent developments in macrolide antimicrobial research. Curr Top Med Chem. 2003;3(9):961–89.
- Workowski KA, Bolan GA. Sexually transmitted diseases treatment guidelines, 2015. MMWR recommendations and reports: morbidity and mortality weekly report recommendations and reports 2015; 64(Rr-03): 1–137.
- Roca A, Oluwalana C, Bojang A, et al. Oral azithromycin given during labour decreases bacterial carriage in the mothers and their offspring: a double-blind randomized trial. Clin Microbiol Infect. 2016;22(6):565.e1-9.
- Keenan JD, Bailey RL, West SK, et al. Azithromycin to reduce childhood mortality in Sub-Saharan Africa. N Engl J Med. 2018;378(17):1583–92.

- Bergal A, Loucif L, Benouareth DE, Bentorki AA, Abat C, Rolain JM. Molecular epidemiology and distribution of serotypes, genotypes, and antibiotic resistance genes of Streptococcus agalactiae clinical isolates from Guelma, Algeria and Marseille, France. Eur J Clin Microbiol Infect Dis. 2015;34(12):2339–48.
- Brzychczy-Wloch M, Borszewska-Kornacka M, Gulczynska E, et al. Prevalence of antibiotic resistance in multi-drug resistant coagulase-negative staphylococci isolated from invasive infection in very low birth weight neonates in two Polish NICUs. Ann Clin Microbiol Antimicrob. 2013;12:41.
- Leclercq R. Mechanisms of resistance to macrolides and lincosamides: nature of the resistance elements and their clinical implications. Clin Infect Dis. 2002;34(4):482–92.
- 37. Dutra VG, Alves VM, Olendzki AN, et al. Streptococcus agalactiae in Brazil: serotype distribution, virulence determinants and antimicrobial susceptibility. BMC Infect Dis. 2014;14:323.
- 38. de Azavedo JC, McGavin M, Duncan C, Low DE, McGeer A. Prevalence and mechanisms of macrolide resistance in invasive and noninvasive group B streptococcus isolates from Ontario, Canada. Antimicrob Agents Chemother. 2001;45(12):3504–8.
- Diekema DJ, Andrews JI, Huynh H, et al. Molecular epidemiology of macrolide resistance in neonatal bloodstream isolates of group B streptococci. J Clin Microbiol. 2003;41(6):2659–61.

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