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Lung cancer: Animal model of lung cancer, molecular carcinogenesis of lung cancer, and antitumor effect of *Ocimum sanctum* against lung cancer

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ABSTRACT

Lung cancer is the leading cause of fatalities related to cancer globally. There are numerous ways to treat lung cancer, including surgery, chemotherapy, and radiation. Since these treatments have not yet shown satisfactory results, more research into the underlying mechanisms and different approaches to therapy and prevention are needed. Animal models are essential to the study of lung cancer because they offer priceless information about the etiology, course, and possible treatments for the illness. The therapeutic application of phytochemicals and medicinal plants to treat cancer-related compounds has gained attention subsequently. In addition to discussing the molecular carcinogenic and antitumor effects of the herbal treatment *Ocimum sanctum* (OS) in connection to lung cancer, this review will address the current awareness regarding lung cancer in animal models. The multitude of animal models used in lung cancer research—such as genetically modified mice, carcinogen-induced models, and xenograft induction—provides a solid foundation for understanding the illness. By easing the examination of the environmental and genetic factors involved and enhancing the analysis of possibilities for treatment, these models eventually assist in the further development of lung cancer therapy. Additionally, using the herb plant OS is essential for both treating and preventing lung cancer. Standardizing dosages and enforcing laws on the use of herbal medications require more in-depth investigation.

Keywords: Animal model, Carcinogenesis mechanism, Lung cancer, *Ocimum sanctum*.

Introduction

Cancer has been a major global public health concern in recent times. According to the National Cancer Institute, cancer is the abnormal growth of newly formed cells that surpass normal limits and then spread to other organs and the other side of the body (Brown *et al.*, 2023). Oncogene activation and/or tumor suppressor gene inactivation often cause uncontrolled cell proliferation and the inhibition of the mechanism responsible for programmed cell death (Siegel *et al.*, 2015). Lung cancer, frequently referred to as lung carcinoma, is a kind of malignant tumor that arises in the lung tissue as a result of an uncontrolled proliferation of cells (Mustafa *et al.*, 2016). Approximately 9.6 million individuals worldwide succumbed to cancer in 2018. Curiously, genetic variables account for about 5%–10% of these cases, but lifestyle and environment have a significantly greater impact, making up 90%–95% (Anand *et al.*, 2008).

Research findings from 2012 indicated that Asia has the highest incidence of lung cancer. The World Health Organization reports that Asia has more cases than every other region, with a rise of more than 51.4%. Breast cancer, stomach cancer, liver cancer, and colon cancer are the most common cancer kinds in Asia. Lung cancer is the sixth among the top causes of death for women in Indonesia and has the highest mortality rate among men. Indonesia has the third highest rate of lung cancer, accounting for 8.6% of new cases and causing the highest number of deaths, which represents 12% of all cancer-related deaths (Arfan and Andarini, 2022). The elevated prevalence can likely be attributed to a multitude of unfavorable lifestyle factors, including smoking, inadequate dietary habits, air pollution, and insufficient public consciousness on early cancer detection. Smoking is responsible for almost 90% of lung cancer cases in Indonesia, making it the primary cause of cancer-related deaths in the country (Mustafa *et al.*, 2016; Kristina *et al.*, 2019).

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Unlike in humans, lung cancer is uncommon in pet animals (dogs and cats) because of restrictions on occurrence rates and prognosis. Animals suffering from lung cancer encounter difficulties in receiving therapy because of the presence of nonspecific clinical signs such as coughing, vomiting, difficulty breathing, fatigue, and vomiting. This leads to a lack of awareness of how lung cancer begins and develops in dogs and cats (Fowler *et al.*, 2020). Presently, surgical procedures, biopsy, radiation, and chemotherapy are the cornerstones of cancer prevention and therapy for both people and animals (Miller and Zachary, 2017). The amount that the disease has spread is a crucial factor that impacts how effectively lung cancer treatments work. Surgical surgery is a successful treatment for malignancies that have not spread, but advanced-stage tumors present considerable therapeutic hurdles. In addition, cancer treatments are expensive and have side effects.

Animal models are essential for the scientific investigation of lung cancer because they provide significant details on the biology, course, and possible therapies of the disease. By using these models, scientists gain insight into the complex interactions that occur between cancer cells and the tumor microenvironment, which is important to understand the dynamics of tumors and how well therapies work. The development of orthotopic lung cancer models, in which tumors are inserted directly into lung tissue, is one instance of this. In contrast with ectopic models, these kinds of models have been demonstrated to yield more realistic results that closely resemble lung cancer in humans. The development of animal models for lung cancer is important because it allows for an improved understanding of the complex process of carcinogenesis, precise evaluation of tumor growth and metastasis, and the efficacy of treatment medications in a setting that closely mimics the real disease environment.

Fortunately, indigenous knowledge also gives us significant insights into the efficient use of natural resources. Nowadays, these materials are widely used as complementary therapies and prophylactics. Under medical supervision, they can also be used in addition to traditional therapies. As a popular herbal remedy in South and Southeast Asia, basil has a number of benefits, such as anti-inflammatory abilities, the ability to stimulate nerve cell proliferation, neuroprotection, and the ability to induce apoptosis in breast cancer (Kusindarta *et al.*, 2016; Kusindarta *et al.*, 2018; Kusindarta *et al.*, 2018a; Raditya *et al.*, 2020; Wihadmadyatami *et al.*, 2023). The processes of molecular carcinogenesis, the development of animal models for the study of lung cancer, and the possible use of *Ocimum sanctum* (OS) as an herbal Ayurvedic medicine for lung cancer prevention and therapy will all be addressed in this review.

Methodology

The National Center for Biotechnology Information, PubMed, Science Direct, Google Scholar, Springer Link, and SAGE journal databases were all employed in a methodical and thorough evaluation of the online scientific literature. The research encompassed all citations from 2000 to 2022. The following keywords were used in the online search: OS, antioxidant properties, cytotoxicity, antitumor, anticancer, antitumor, antimicrobial, antibacterial, anti-inflammation, conventional ethnomedicine, and phytochemical substances, and pharmacological features.

Lung cancer

Approximately 13% of all cancer diagnoses are related to lung cancer, making it as one of the leading causes of mortality globally (Barta *et al.*, 2019). Epigenetic alterations and hereditary DNA damage are the causes of lung cancer. Normal cell functions, such as DNA repair, programmed cell death (apoptosis), and cell growth, are impacted by these alterations. Lung cancer can be worsened or prevented by a number of risk factors, such as genetics, asbestos, air pollution, radon gas, and smoking (Larsen and Minna, 2011). The leading cause of cancer is still smoking (Yuan *et al.*, 2009), and secondhand smoke exposure is a significant risk factor for lung cancer in both humans and animals (Brown *et al.*, 2007; Kim *et al.*, 2012). Lung cancer risk is raised through asbestos exposure in addition to air pollution, particularly from automobile emissions (Zhou *et al.*, 2015). Lung cancer is a condition that is influenced by radon exposure (Fowler *et al.*, 2020). Viral infections can lead to infectious lung cancer in sheep infected with the Jaagsiekte sheep retrovirus (Rai *et al.*, 2000). Research is presently being done to determine whether retroviral infections contribute to lung cancer in humans (Miller *et al.*, 2017). Lung cancer progresses in part due to chronic inflammation (Freeman *et al.*, 2023).

Proto-oncogenes, which encode growth-promoting proteins, and tumor suppressor genes, which encode molecules that negatively regulate cell proliferation, are the two categories of oncogenes that have been found. A protein with dominant behavioral effects can be produced by a mutation in one of the proto-oncogenic paired alleles. On the other hand, gene products that become cancerous cells require the activation of tumor suppressor genes before they may appear. Most patients with lung cancer have altered gene coding for proteins, which regulates or controls cell proliferation. Lung cancer etiology may be significantly influenced by these molecular alterations (Huang *et al.*, 2021). Tumor suppressor gene inactivation or oncogene activation is the first step in the pathophysiology of lung cancer. Tumor invasion, angiogenesis, apoptosis, and cell proliferation are all regulated by the epidermal growth factor receptor (EGFR) (Mustafa *et al.*, 2016).

Lung cancer is classified histologically into two categories: non-small cell lung cancer (NSCLC) and small-cell lung cancer (SCLC). Small cells with minimal cytoplasm and no discernible nucleoli are the hallmarks of SCLC, which typically affects smokers. Because SCLC problems metastasis earlier and double more quickly, they are always diagnosed as systemic diseases. NSCLC comes in five different forms: the two most common histological subtypes of NSCLC are (1) squamous cell carcinoma, which develops in the central bronchi and is associated with smoking; (2) adenocarcinoma, which develops in the bronchioles and alveoli and is most common in women and non-smokers. (3) Large cell carcinoma, which arises from a lack of differentiation in small cells and glandular or squamous cells; (4) carcinoid tumors, which include two subtypes: typical and atypical (Travis *et al.*, 2013; Siddiqua *et al.*, 2022); and (5) adenosquamous carcinoma, which is defined as having more than 10% mixed glandular and squamous components—it has a worse prognosis than squamous carcinoma and adenocarcinoma.

Adenocarcinoma usually develops from mucosal glands and represents approximately 40% of all lung cancer cases. The development of adenocarcinoma usually begins in the marginal area of the lung, and in many cases, it is found in scars or areas of chronic inflammation (Li and Lu, 2018), making it more difficult to attribute the cause to the bronchial wall. This cancer grows and spreads along the alveolar wall, using the alveolar wall as a starting point for growth (Travis *et al.*, 2013). Cells from an adenocarcinoma may distribute locally, entering the pericardium, pleura, diaphragm, or bronchi. The spread will proceed to the mediastinum, big blood arteries, trachea, esophagus, spine, or neighboring lobes if problems from other disorders are present. Before spreading to the mediastinal or subcarinal lymph nodes and ultimately the contralateral lung, lymph node metastases start in the parabronchial lymph nodes. Extensions to the opposing lobe, pleural nodules, malignant pleural or pericardial effusions, and distant locations (liver, brain, or bones) are examples of remote metastases (Nana-Sinkam and Shojaee, 2017). EGFR mutations and oncogenic rearrangements of anaplastic lymphoma kinase fusion are present in a subgroup of NSCLCs and make them more sensitive to tyrosine kinase inhibitors (Shojaee and Nana-Sinkam, 2017). Thyroid transcription factor-1, cytokeratin-7, and napsin A are the traditional histochemical indicators of adenocarcinoma. There are three subtypes of pulmonary adenocarcinoma: acinar, papillary, and mixed. Adenocarcinoma can spread to the liver, bones, central nervous system, and adrenal glands, just like small cell carcinoma does. Adenocarcinoma is more likely to be confined than small cell carcinoma, particularly when it presents as peripheral lung nodules (Travis *et al.*, 2013).

Animal models of lung cancer

Animal models play a crucial role in lung cancer mechanism, prevention, and medication due to the intricate nature of oncology research and the ability of animal models to produce significant data with meaningful clinical implications, including disease processes, pharmacological treatments, and early detection approaches (Onaciu *et al.*, 2020). Animal models are experimental animals that closely resemble human diseases (Li *et al.*, 2021). Although *in vitro* studies offer valuable cellular insights, their conclusions could be more extensive in their capacity to capture the intricate pathogenic connections *in vivo* animals (Tratar *et al.*, 2018). *In vivo* research using animal models is valuable for assessing possible treatments' effectiveness, including traditional herbal remedies. Furthermore, the impact of signaling systems on lung cancer can also be demonstrated. One way to uncover biomarkers is by combining data from animal models with human clinical research (Domínguez-Oliva *et al.*, 2023). External or exogenous factors, such as lifestyle and site of residence, communicate with internal factors, especially genetics, affecting the onset and progression of cancer (Rudolph *et al.*, 2016). Therefore, integrating *in vitro* and *in vivo* studies yields more compelling findings (Tratar *et al.*, 2018). A number of animal models have been developed to mimic lung cancer; four common approaches are used to generate lung cancer models in mice: the genetically modified mouse model, the chemically induced model, the cell line-derived xenograft (CDX) model, and the patient-derived xenograft (PDX) model (Fig. 1) (Onaciu *et al.*, 2020). Using these models, scientists may study how lung cancer develops, evaluate cutting-edge therapeutic approaches, and investigate the effects of various treatments. The advantages and disadvantages of every method used to create the lung cancer model are listed in Table 1.

Chemically induced lung cancer animal model

Chemically induced cancer animal models use carcinogens to transform normal cells into tumors. Chemically induced animal models have the advantage of mimicking the occurrence of human cancer from the beginning of the carcinogenic process (Liu and Johnston, 2002.) providing a valuable platform to study the pathogenesis, progression, and potential therapeutic interventions for lung cancer. However, the main drawback of this method is that it takes 30–50 weeks to form tumors after using carcinogens (Minicis *et al.*, 2013). Various chemical substances have been employed in Several studies to induce lung carcinomas in animals through intraperitoneal injection such as N-nitroso diethylamine (NDEA) (Çiçek *et al.*, 2022), 3-methylcholanthrene and diethylnitrosamine (DEN) (Zhu *et al.*, 2017). In animal models of adenocarcinoma-type lung cancer, N-nitrosamines such as 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (Brown *et al.*, 2007) and DEN

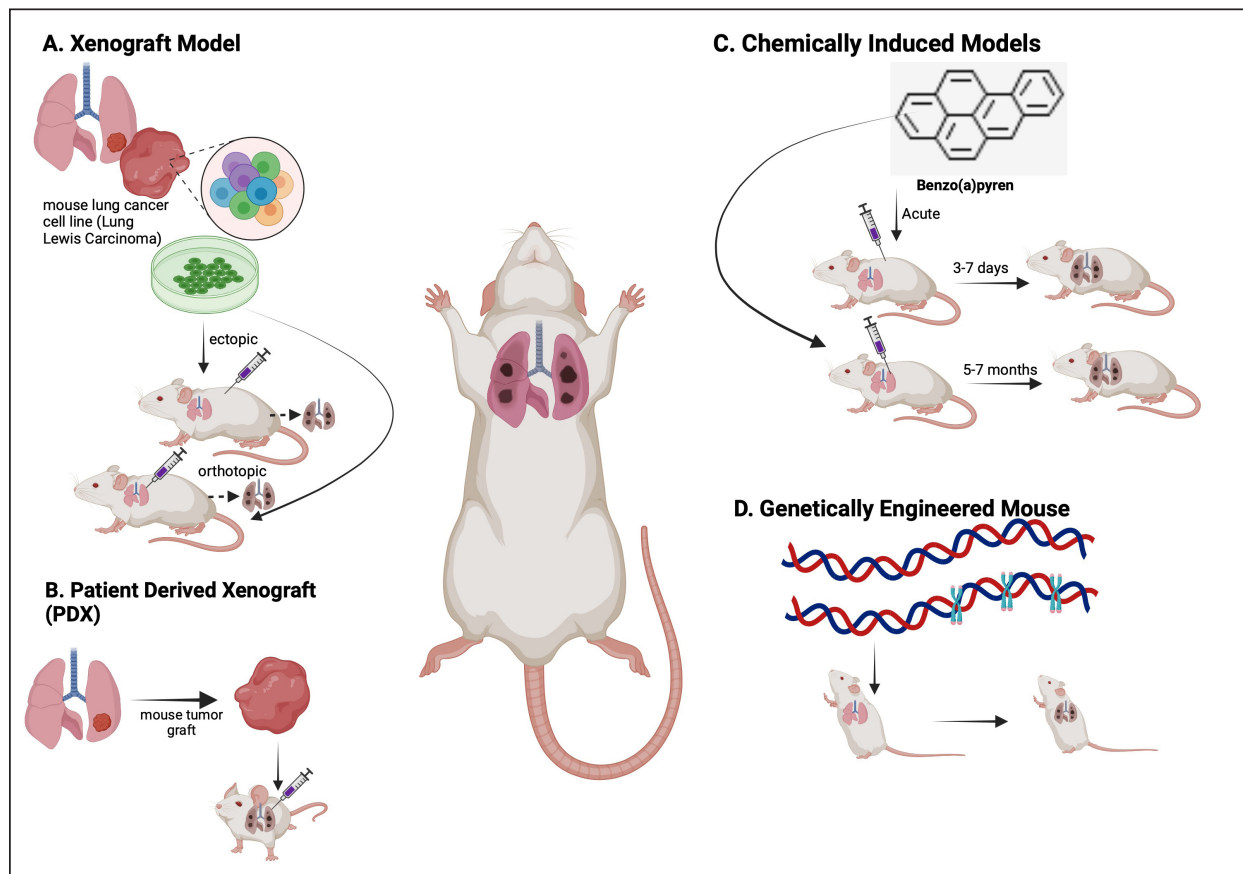


Fig. 1. The four most frequently used techniques to generate mouse lung cancer models include chemically induced models, PDX model, CDX model, and genetically engineered mouse model.

(Mervai *et al.*, 2018) have been shown to selectively induce lung adenocarcinoma in animals so that the pathogenesis of lung cancer can be studied. In addition, chemical induction models using substances such as benzo(a)pyrene (BaP) have been developed to induce lung cancer in mice, providing insight into the effects of these carcinogens on lung tumorigenesis (Wang *et al.*, 2021). Urethane, a chemical carcinogen present in tobacco smoke, has been shown to induce lung tumors in mice by causing activating mutations in Kras (Minowada and Miller, 2009).

Cell lines for lung cancer induction in animal models

A xenotransplantation paradigm in which cancer cell lines are injected into immunodeficient mice is known as the CDX model (Brennecke *et al.*, 2014). For this aim, the NSCLC adenocarcinoma cell line A549 has been widely employed, created an orthotopic lung cancer animal model by implanting a variety of cells into BALB/c nude mice, demonstrating the adaptability and significance of the A549 cell line (Lee *et al.*, 2010). Furthermore, a lung cancer animal model in nude mice has been established using the human large-cell lung cancer cell line H460 (Ye *et al.*, 2016). Furthermore, the C57BL/6 mouse-derived murine lung cancer cell

lines LLC and CMT167 are crucial for researching the tumor microenvironment and determining how sensitive lung cancers are to PD-1/programmed cell death ligand 1 (PD-L1) antibody blocking (Du *et al.*, 2016).

PDXs for lung cancer

PDX models have become an invaluable tool in lung cancer research, providing a platform to study tumor biology, test therapeutic regimens, and identify predictive biomarkers. These models involve implanting patient-derived tumor tissue into immunocompromised mice to create xenografts that mimic the characteristics of native tumors, successfully created a broad panel of patient-derived tumor xenografts for NSCLC, despite the considerable time and distance between clinical and animal facilities, providing a stable and reliable preclinical model for human lung cancer research (Ilić *et al.*, 2014). This PDX model recapitulates the histopathology and molecular diversity of NSCLC, offering a platform to study disease heterogeneity and test personalized treatment approaches. Furthermore, PDX models have been instrumental in predicting effective therapies for lung cancer, demonstrating the utility of PDX models

Table 1. Advantages and disadvantages of several methods to lung cancer animal model methods.

Methods	Advantage	Disadvantages
Chemicals induce lung cancer	<ul style="list-style-type: none"> • High reproducibility in generating animal lung cancer models and ensuring consistency of results (Liu and Johnston., 2002). • Cost-effective and accessible for research purposes (Uner <i>et al.</i>, 2022). • Has a high mutation rate in animals, making it possible to study genetic changes and tumor progression (Liu and Johnston, 2002). • Can help identify predictive biomarkers and evaluate the efficacy of potential therapies, aiding personalized treatment approaches (Ilić <i>et al.</i>, 2014). • Facilitates the development of lung tumors with different characteristics, enabling the study of different tumor types and responses to treatment (Ku <i>et al.</i>, 2007). 	<ul style="list-style-type: none"> • These models often show low metastatic potential (Liu and Johnston, 2002). • Shallow dose–response curves, some chemical-induced animal models, such as the A/J mouse model, exhibit shallow dose response curves, which may affect the interpretation of treatment effects (Luettich <i>et al.</i>, 2014). • Depending on exposure to specific carcinogens, which may not fully replicate the complex environmental factors that contribute to lung cancer in humans (Khan <i>et al.</i>, 2022).
CDX model	<ul style="list-style-type: none"> • Relevance to human cancer as cell lines from humans such as A549 and H460, closely resemble the characteristics of human lung cancer, providing a relevant model to study this disease (Chong <i>et al.</i>, 2021). • It is consistent and reproducible to induce lung cancer in animal models, ensuring uniformity in research results (Gautam <i>et al.</i>, 2003). • Cell lines are readily available and easy to culture, enabling the creation of efficient and cost-effective lung cancer models in animals (Vertrees <i>et al.</i>, 2000). • Studies using cell lines can predict treatment response and identify potential therapeutic targets, thus aiding drug development (Wislez <i>et al.</i>, 2005). • It is flexible as it can be used in a variety of studies, including investigating the effects of apoptosis, drug response, and metastatic potential, thus increasing the versatility of lung cancer research (Weng <i>et al.</i>, 2009). 	<ul style="list-style-type: none"> • It lacks the heterogeneity seen in primary tumors, potentially limiting the representation of diverse lung cancer subtypes in animal models (Janker <i>et al.</i>, 2018). • Prolonged culture of cell lines may lead to genetic alterations and changes in cellular behavior, affecting model fidelity (Zhao <i>et al.</i>, 2018). • Some cell lines may show limited metastatic potential, hindering the study of advanced lung cancer and metastasis (Li <i>et al.</i>, 2011). • Cell lines can adapt to <i>in vitro</i> conditions, potentially altering their response to <i>in vivo</i> environments and treatments (Han <i>et al.</i>, 2008). Does not fully capture the immune response and complexity of the tumor microenvironment observed in lung cancer patients, thus limiting the translational relevance of findings (Nolan <i>et al.</i>, 2020).
PDX model	<ul style="list-style-type: none"> • Retains the genetic and histological characteristics of the original patient tumor, thus providing a clinically relevant model for studying lung cancer (Garcia-Oliveira <i>et al.</i>, 2020). • More predictive of patient response to treatment compared to traditional models, allowing for personalized treatment approaches in lung cancer research (Garcia-Oliveira <i>et al.</i>, 2020) • Captures the heterogeneity of human tumors, enabling the study of diverse lung cancer subtypes and responses to therapy (Fu <i>et al.</i>, 2015). • Preserves an intact stromal component, reflecting the tumor microenvironment present in the patient, which may influence treatment response (Garcia-Oliveira <i>et al.</i>, 2020). • Reflects genetic features and tumor progression observed in lung cancer patients, increasing the translational relevance of research findings (Garcia-Oliveira <i>et al.</i>, 2020). 	<ul style="list-style-type: none"> • Requires a long time as it involves transplanting patient tumor tissue into immunodeficient mice, which may delay research progress (Shoji <i>et al.</i>, 2023). • Requires higher costs due to the lengthy process, limiting the scalability of studies that use patient-derived cancers for lung cancer induction in animal models (Shoji <i>et al.</i>, 2023). • Requires ethical approval and compliance with animal welfare regulations, which adds complexity to the experimental procedure (Shoji <i>et al.</i>, 2023) • Have limited availability, especially for rare lung cancer subtypes, potentially limiting the scope of research (Shoji <i>et al.</i>, 2023). • PDX models are usually established in immunodeficient mice, which may not fully capture the immune responses and tumor–host interactions seen in patients with lung cancer (Shoji <i>et al.</i>, 2023).

(Continued)

Methods	Advantage	Disadvantages
A genetically engineered mouse model	Has clinical relevance as GEMMs closely mimic human lung cancer, allowing for the study of disease progression and treatment response (Barck <i>et al.</i> , 2015).	Has fewer somatic mutations than human lung cancer, resulting in limited immune cell infiltration and decreased immune response (Kim, 2024).
	Can predict response to therapy, aiding the development of personalized treatment approaches for lung cancer (Barck <i>et al.</i> , 2015).	Requires ethical approval and compliance with animal welfare regulations, adding complexity to experimental procedures (Houdebine <i>et al.</i> , 2014).
	Captures the heterogeneity of human tumors, enabling the study of diverse lung cancer subtypes (Barck <i>et al.</i> , 2015).	Creating and maintaining GEMMs can be costly and time-consuming, potentially limiting the scalability of research (Ormandy <i>et al.</i> , 2011).
	Reflects genetic features and tumor progression observed in lung cancer patients, thereby increasing the translational relevance of research findings (Barck <i>et al.</i> , 2015)	Obtaining genetically engineered mouse models for certain lung cancer subtypes may be difficult, especially for rare variants (Ormandy <i>et al.</i> , 2011).
	Guides the development of mouse models based on specific genetic events in human cancers, which is important for target validation (Walrath <i>et al.</i> , 2010).	Does not fully capture the complexity of the tumor microenvironment observed in patients with lung cancer, thus affecting the translational relevance of findings (Ormandy <i>et al.</i> , 2011).

in predicting effective heparanase-based therapies for lung cancer, highlighting the relevance of these models in guiding treatment strategies (Katz *et al.*, 2018). Additionally, it emphasizes the clinical significance of PD-L1 in patients with lung cancer using PDX models, demonstrating the ability of these models to reproduce the biological characteristics of human cancers (Ma *et al.*, 2016).

Genetically engineered mouse model for lung cancer

Research on lung cancer has derived greatly from the use of genetically engineered mice models (GEMMs), which offer an invaluable platform for examining the development, spread, and effects of treatment. Animal models of genetically engineered lung cancer are produced by modifying (adding/removing) genes to produce desired functional alterations. Lung cancer that is both universal and unique to the pau will be the outcome of the change (Zhao *et al.*, 2000). The KrasG12D/+ (Kirsten rat sarcoma viral oncogene homolog) mouse model of lung cancer is a well-known GEMM for the disease. It is a p53 Frt/Frt model in which conditional mutations in the p53 tumor suppressor gene and the K-ras oncogene cause adenocarcinoma to develop in the mouse lung (Barck *et al.*, 2015). Utilization of GEMMs with conditional mutations in the K-ras oncogene and p53 tumor suppressor gene to induce lung tumors can exert circadian rhythm disruption effects on lung tumorigenesis (Papagiannakopoulos *et al.*, 2016) and highlight histopathological changes using micro-CT imaging and automated analysis (Barck *et al.*, 2015). Another method eliminates the SWI/SNF ATPase subunits BRM and BRG1 in promoting lung cancer development in GEMM. This study highlights the histopathological changes associated with lung cancer progression in GEMMs. Mutations in growth factors such as SP-C-IgEGF, SP-C-cMyc, and SpC-IgEGF

will result in adenocarcinoma GEMM changes and provide insight into tumor evolution and progression (McFadden *et al.*, 2016). Other GEMM studies can be seen in Table 2.

Molecular carcinogenesis of lung cancer

The development of lung cancer involves a complex process known as molecular carcinogenesis, which includes genetic mutations, epigenetic changes, and interactions with the tumor microenvironment. Gaining a comprehensive understanding of these mechanisms is essential for the development of highly effective preventative and treatment strategies.

Angiogenesis and vasculogenesis

Angiogenesis is a common, complex physiological process that is managed by certain molecules that the body produces. Endogenous local or systemic biomolecular signals synchronize the actions of smooth muscle and endothelial cells to repair damaged blood vessels. Vascularization aids in the provision of nutrients in addition to serving as a pathway for neoplasm cell extravasation, or the movement of tumor cells to other organs through circulation (Rajabi and Mousa, 2017). The process of neoplasm cells generating blood vessels that invade and spread within the tumor environment is known as angiogenesis. Neoplasm cells require blood vessel development to survive since it provides them with nutrition and oxygen. Not only does angiogenesis encourage the growth and metastasis of neoplasm cells, but it is also a sign of cancer. Vasculogenesis, sprouting angiogenesis, intussusception, and vasculogenic mimicry are a few angiogenesis mechanisms (Liu *et al.*, 2023).

The genesis and progression of neovascularization are aided by endothelial progenitor cells or EPCs. Neoplasm cells provide proangiogenic substances, including vascular endothelial growth factor (VEGF) and cytokines, which subsequently attract dendritic

Table 2. Gene mutation related genetically engineered mouse model for lung cancer.

Mutation	Description
Oncogene and growth factor	
EGFR mutations	EGFR mutation exon 19 deletions and the L858R point mutation lead to constitutive activation of the EGFR signaling pathway, promoting cell proliferation and survival (McFadden <i>et al.</i> , 2016; Suda and Mitsudomi, 2014). Oncogenic potential of M277E mutation in the EGFR sensitive to EGFR-targeted therapies such as erlotinib (Yu <i>et al.</i> , 2017).
Kirsten rat sarcoma viral oncogene (KRAS) mutations	KRAS mutation G12C and G12D variants are prevalent in lung adenocarcinomas and are associated with poor prognosis. GEMMs harboring KRAS mutations have been instrumental in understanding the biology of lung cancer and evaluating potential therapeutic strategies (Suda and Mitsudomi, 2014).
Serine/threonine-protein kinase B-Raf (BRAF) mutations	BRAF V600E mutation leads to aberrant activation of the MAPK signaling pathway and has been used to explore targeted therapies, including BRAF and MEK inhibitors (Ota <i>et al.</i> , 2021; Villaruz <i>et al.</i> , 2014)
Fibroblast growth factor receptor (FGFR) mutations	FGFR2 and FGFR3 have been implicated in lung squamous cell carcinoma, reported that these mutations are oncogenic and sensitive to FGFR kinase inhibitors, making them important targets for therapeutic intervention (Liao <i>et al.</i> , 2013).
The insulin-like growth factor 1 receptor (IGF-1R) mutation	IGF-1R promotes cell growth and survival and its overexpression has been observed in various lung cancers (Huang <i>et al.</i> , 2016).
HGF and c-Met receptor	The hepatocyte growth factor (HGF) and its receptor c-Met are involved in lung cancer progression. demonstrated that targeting both the c-Met and EGFR pathways resulted in additive inhibition of lung tumorigenesis in transgenic mice (Stabile <i>et al.</i> , 2010).
Tumor suppressor gene	
GandATA4	Lung cancer is deficient in the tumor suppressor GATA4, sensitive to TGFBR1 inhibition. They confirmed the tumor suppressor function of GATA4 <i>in vivo</i> using a transgenic lung cancer mouse model, specifically in Isl-Kras G12D mice, where they targeted GATA4 using lentiviral delivery (Gao <i>et al.</i> , 2019).
BRCA1-associated protein-1 (BAP1)	BAP1 gene deletions are frequently observed in NSCLC and SCLC cell lines. According to Ventii <i>et al.</i> (2008), this work highlights the significance of BAP1 in the biology of lung cancer and its potential as a target for therapeutic intervention (Ventii <i>et al.</i> , 2008).
RBM5	The RBM5 gene, which is found at the 3p21.3 locus and is commonly deleted in lung cancer, functions as a tumor suppressor by changing the expression of genes related to metastasis. RBM5 causes apoptosis and suppresses the development of lung cancer cells (Shao <i>et al.</i> , 2012).
FUS1	In lung cancer models, coexpression of the tumor suppressor genes FUS1 and p53 generates a synergistic effect on tumor suppression. Their research demonstrates that FUS1 is an important target for treatment approaches since it appears to be involved in preventing the formation of lung tumors (Deng <i>et al.</i> , 2007).
Deleted in liver cancer 1 (DLC-1) gene	The DLC-1 gene functions as a suppressor of tumors in NSCLCs in humans. These results provide credence to the theory that tumor suppressor genes might become inactive due to both genetic and epigenetic modifications (Yuan <i>et al.</i> , 2003).
UBE1L	A study was conducted on the potential tumor suppressor gene UBE1L in the setting of K-rasLA2 mice. Although the study indicated that UBE1L loss did not accelerate the development of lung cancer, it does highlight the complexity of tumor suppressor gene interactions in lung cancer models (Yin <i>et al.</i> , 2009).
ING1 and ING2	These genes may be associated with the progression of lung cancer, and more research on their roles could occur employing animal models (Okano <i>et al.</i> , 2006).
15-Hydroxyprostaglandin dehydrogenase (15-PGDH)	15-PGDH in lung cancer as a tumor suppressor. Their research revealed that overexpressing 15-PGDH decreased the growth of tumors in xenograft models, implying that it may be a promising target for treatment (Huang <i>et al.</i> , 2008).
Cyclin D2	Lung cancer is caused by abnormal methylation of the cyclin D2 promoter. The inactivation of tumor suppressor activities may be facilitated by this methylation, underscoring the significance of epigenetic modifications in the development of lung cancer (Virmani <i>et al.</i> , 2003).

cells coming from bone marrow and facilitate their proliferation and development. Endothelial Progenitor Cells (EPC) mobilization merges into a single endothelium layer subsequent to matrix metalloprotease 9 (MMP 9) activation (Zuazo-Gaztelu and Casanovas, 2018). VEGF drives the development of EPCs into mature endothelial cells. Through a paracrine mechanism, EPCs also promote angiogenesis by expressing proangiogenic factors at the site of neovascularization of the tumor stroma or ischemic tissue (Marçola and Rodrigues, 2015; Urbich and Dimmeler, 2004).

Sprouting angiogenesis is the first stage of vasculogenesis. Tumor cells, the microenvironment, and the extracellular matrix (ECM) are all involved in this pathway. Destabilization of endothelial-pericyte connections (maintaining the integrity and maintenance of blood vessels) is the first step in the sprouting angiogenesis stage. The mesenchymal-endothelial transition that endothelial cells go through improves their ability to migrate, invade, and proliferate. Mesenchymal-endothelial transition degrades the ECM and basement membrane with activated proteases (such as MMP), opening migration, and proliferation pathways. The lumen of blood vessels is formed by the polarization of endothelial cells (Urbich and Dimmeler, 2004; Lugano *et al.*, 2020). This event provides a proangiogenic signal—the tip cell becomes invasive and motile. Endothelial cells respond to VEGF signals by expanding filopodia to form new blood vessels along the vascular bed. Endothelial cells in the sprouting angiogenesis stalk (stalk cells) have the potential to proliferate. The proliferative potential of stalk cells forms tubules, allowing expansion to form new blood vessels (Norton and Popel, 2016).

The next step is intussusception microvascular growth. The intussusception microvascular growth mechanism begins with the division of the blood vessels into two new blood vessels, and a transvascular connective tissue column is formed within the lumen of the blood vessel, called a tissue pillar. Transendothelial cell bridges are formed subsequent to the induction of endothelial cells from the opposing wall. The endothelium layer forms as a result of the arrangement of the interendothelial connections. Interstitial pillars are created to reinforce the bridge, and mural cells are enlisted to line the newly created interstitial walls (Burri *et al.*, 2004; Ali *et al.*, 2019). There is vascularogenic mimicry. Tumor cell differentiation is the basis for this process. A matrix network like that of vasculogenesis is created by this process. Red blood cells and plasma make up this structure, which aids in blood circulation. Regardless of angiogenesis, endothelial cells that engage in vasculogenic mimicry can serve as a supplementary circulatory system for tumor cells. Vasculogenic mimicry is caused by low oxygen levels, the release of ECM components, and the activation of transmembrane Metalloproteinase (MP) (Luo *et al.*, 2020).

MMP, integrins, and VEGF are a few biomolecules involved in tumor cell angiogenesis. In addition to being the primary source of pathological angiogenesis, VEGF also causes inflammation, rheumatoid arthritis, diabetic retinopathy, and cancer (Mukhopadhyay *et al.*, 2004). The Zn structure is essential to a class of endopeptidases known as MMPs. MMP plays a role in the disintegration of ECM macromolecules (Wang and Khalil, 2018).

The role of VEGF in angiogenesis

The primary modulator of blood vessel development is homodimeric glycoprotein VEGF, sometimes referred to as VEGF-A. By starting the proteinase cascade that results in the creation of plasmin and MMP, VEGF generates feedback for VEGF activity (Liu *et al.*, 2023). Some of the growth factors that may promote the synthesis of VEGF include platelet-derived growth factors (PDGF), fibroblast growth factors (FGF), epidermal growth factors, tumor necrosis factors (TNF), transforming growth factors, interleukin-1 (IL-1), and the hypoxic state of cells (Bao *et al.*, 2009). Increased endothelial cell migration, increased endothelial cell mitosis, increased methane monooxygenase activity, increased $\alpha v \beta 3$ activity, blood vessel lumen layout, and more are all related to VEGF's function in angiogenesis, chemotaxis for granulocytes and macrophages, as well as indirect vasodilation via endothelial-derived relaxing factors or nitric oxide (NO) release (Lamalice *et al.*, 2007). As a crucial modulator of normal physiological angiogenesis, VEGF expression abnormalities are a key marker of the illness process. Pathogenesis, which includes neoplasia and inflammatory diseases, is influenced by VEGF.

Increased VEGF expression has been seen in neoplastic cell biopsies. In a variety of human cancer types, VEGF is known to be the starting point for carcinogenesis and disease development (Ceci *et al.*, 2020). Neoplastic cells are essential for the formation of VEGF; without a sufficient vascular supply, they will not proliferate because they will not have enough oxygen and nutrients. As a key player in tumor angiogenesis, VEGF promotes the formation of new blood vessels from surrounding capillaries, giving tumors access to the nutrients and oxygen they require to proliferate (Lugano *et al.*, 2020). The absence of adequate vascularization in neoplasm cells causes a hypoxic environment, which, in turn, triggers new vascularization. The tumor's state also alters the microenvironment, resulting in elevated growth factor levels and neovascularization triggers tumor growth (Chen *et al.*, 2023). The growing distance of dense cancerous cells from the nearest blood arteries causes the cells to undergo hypoxia. Von Hippel-Lindau does not suppress hypoxia-inducible factor-1 (HIF-1) or scatter in hypoxic circumstances (Kaelin, 2004). Because of this, HIF-1 can dimerize and attach to the VEGF gene promoter, which stimulates the production of VEGF. VEGFR on endothelial cells then

receives this VEGF, potentially causing angiogenesis (Fig. 3).

The role of MMP in angiogenesis

In the process of angiogenesis, endothelial cells migrate, proliferate, and create new matrix components while new capillaries grow out of the endothelial vessels that are already there to facilitate the breakdown of the vascular basement membrane. MMPs are involved in metastasis, neovascularization of tumors, and illnesses. MMPs control transitional epithelium-mesenchyme, a molecular shift that controls the appearance and function of epithelial cells. These configurations include the division of the components of the basement membrane, the breakdown of the ECM, and the intercellular disintegration and adhesion characteristics of the cell matrix (Quintero-Fabián *et al.*, 2019).

MMPs, frequently referred to as matrixes, belong to the Zn²⁺ endopeptidase superfamily of metzincin proteases. Against different ECM substrates, MMPs have distinct proteolytic activity (Kelwick *et al.*, 2015). The substrate's specificity and the overall structural domain's shape are used to categorize MMPs. Six subfamilies make up the categorization of MMPs: membrane-type MMP (MT)-MMPs, collagenases, gelatinases, stromelysins, matrilysins, and other MMPs (Quintero-Fabián *et al.*, 2019). Proteases, which are precursors of all MMPs, must undergo proteolytic cleavage in a physiological setting in order to facilitate the release of propeptide domains or zymogen activation, which in turn produces MMPs (Jabłońska-Trypuć *et al.*, 2016). Endogenous inhibitors and posttranslational proteolytic cleavage control MMP activity (Cabral-Pacheco *et al.*, 2020).

MMPs have a role not only in the abnormalities associated with cancer but also in the control of angiogenesis, vasculogenesis, and lymphangiogenesis. MMPs take role in the degradation of certain substrates and ECM. VEGF, basic fibroblast growth factor (bFGF), cytokines (IL-1 and -6), tumor growth factor- α and - β (TGF), TNF- α , PDGF, angiogenin, and hormones are among the angiogenic agents that can activate MMPs. In order to control proteolytic activity and support invasive and morphogenic processes linked to angiogenesis, endothelial cells generate vesicles containing MMP-2 and MMP-9, which are activated by VEGF and FGF-2 (Fig. 5) (Taraboletti *et al.*, 2007; Jabłońska-Trypuć *et al.*, 2016).

The modulation of ECM remodeling (ECM remodeling) is influenced by MMP-2 and MMP-9. These molecules regulate laminin, glycosaminoglycans, fibronectin, aggrecan, and latent protein signaling. They are activated and deactivated by proteolytic cleavage, which releases biological activity that leads to cellular control (Mott and Werb, 2004). Gelatinases, such as MMP-2 and MMP-9, are categorized according to substrate specificity and are in charge of breaking down gelatin (Luchian *et al.*, 2022). MMP-2, having a molecular weight of 72 kDa, is a member of the gelatinase type

A subgroup. MMP-2 degrades fibronectin, elastin, Type IV, V, VII, and X nonfibrillar collagen, and the basement membrane (Chuliá-Peris *et al.*, 2022).

Cell migration, enhanced collagen affinity, elevated TGF- β bioavailability, neurodegeneration through neuron apoptosis, axon growth, mesenchymal cell differentiation with inflammatory phenotypes, anti-inflammatory effects, proliferation, and IGF-1 cleavage in the development of cancer are all functions of MMP-2 (Gonzalez-Avila *et al.*, 2019). Tumor angiogenesis and MMP-2 expression are related. IL-8, an angiogenic agent, increases MMP-2 expression and activity in tumor cells to promote invasion. Through tumor neovascularization, metastasis, and ECM degradation, MMP-2 is crucial to the development of cancer (Quintero-Fabián *et al.*, 2019).

MMP-9, a 92 kDa type B gelatinase, is what it is called. MMP-9 degrades certain substrates, such as fibrillin, gelatin, and collagen and nonfibrillar collagen (Type IV, V, VII, X, and XIV), found in the basement membrane. MMP-9 biologically influences decreased IL-2 response, enhanced collagen affinity, and pro- and anti-inflammatory action. In addition to tumor cell resistance, TGF- β activation in cancer growth causes hypertrophic chondrocytes to undergo death and incorporates new osteoblast functional units. Additionally, MMP-9 was discovered when it was stimulated by VEGF during and after the metastasis of cancerous cells (Debbarma *et al.*, 2020). Fortunately, during carcinogenesis, MMP-9 releases VEGF, which causes the angiogenic release and encourages the migration of endothelial cells. Increased vascular permeability brought on by MMP-9 expression stimulates the production of neovascularization in malignant tissue and regulates angiogenic factors, both of which contribute to the progression of cancer (Wihadmadyatami *et al.*, 2020).

It has been shown in several investigations that MMP-2 and MMP-9 may diminish collagen type IV, despite being often imposed in human malignancies. Moreover, the effects of MMP-2 and MMP-9 were demonstrated using an *in vivo* experimental paradigm that produced the angiogenic phenotype and keratinocyte tumor invasion. Tumor growth and invasion are accelerated by MMP-2 and MMP-9 activity, which causes cancer angiogenesis by cleaving latent TGF- β in a way that is reliant on CD44. The aforementioned results may explain the pivotal function that MMP-2 and MMP-9 play in tumor angiogenesis by activating proangiogenic factors (Webb *et al.*, 2017; Quintero-Fabián *et al.*, 2019; Wihadmadyatami *et al.*, 2020).

The role of integrin $\alpha 5 \beta 1$ and $\alpha v \beta 3$ in angiogenesis

The transmembrane glycoprotein subunits that make up integrin, known as alpha (α) and beta (β), are not covalently bonded and interact with the ECM. The β subunit binds to the cytoskeleton and affects several signaling pathways, whereas the α subunit controls the specificity of the integrin ligand. By connecting

cytoskeletal actin to ECM molecules, integrins serve as cell surface protein receptors that regulate cell adhesion to the ECM. This process produces mechanical adhesion. Signal transduction pathways govern many fundamental cell functions, including cell division, proliferation, and migration, which set the stage for the interaction between integrin components and ECM. Proper cell attachment and localization are ensured by the substantial participation of integrins in ECM components, which provide signals related to cell survival (Kim *et al.*, 2010; Bachmann *et al.*, 2019; Valdembrì and Serini, 2021). Arginine-glycine aspartic acid (RGD) strands make up short-knot fibronectin peptides, which integrins $\alpha 5 \beta 1$ and $\alpha v \beta 3$ react with (Ludwig *et al.*, 2021).

The vitronectin receptor, or integrin $\alpha v \beta 3$, is made up of two subunits: the 105-kDa $\beta 3$ subunit and the 125-kDa αv subunit. Integrin $\alpha v \beta 3$ interacts to a variety of ECM molecules that have the triple-peptide ArgGly Asp (RGD) motif, such as polarized versions of collagen and laminin (Hsu *et al.*, 2017; Du *et al.*, 2020), fibronectin, fibrinogen, von Willebrand factor, and vitronectin. Integrin $\alpha v \beta 3$ plays a part in angiogenesis, pathological neovascularization, osteoclast-mediated bone resorption, and neoplasia cell metastasis. Among all the integrins, integrin $\alpha v \beta 3$ is crucial to the angiogenesis process. A high level of $\alpha v \beta 3$ integrin expression on cancerous cells promotes angiogenesis and makes it easier for the cells to proliferate, invade, and undergo apoptosis. bFGF and TNF- α are required for $\alpha v \beta 3$ to start $\alpha v \beta 3$ integrin angiogenesis, according to *in vivo* angiogenesis tests (Chandra Kumar, 2003; Chen *et al.*, 2008).

Additionally, involved in angiogenesis is integrin $\alpha 5 \beta 1$. By directly binding to either angiopoietin-171 or VEGFR-1.72, integrin $\alpha 5$ has been found to increase cell migration. Additionally, it has been demonstrated to cross-communicate with the Tie-2 endothelial receptor (Cascone *et al.*, 2005). Angiogenesis depends on B1 integrins; nonetheless, it remains apparent how particular $\alpha 5 \beta 1$ integrins function in this process. Endothelial cells, endothelial support cells, and pericytes from both normal and angiogenic vessels express integrin $\beta 1$. It is believed that pericytes' production of $\beta 1$ integrin contributes to blood vessel stabilization (Carnevale *et al.*, 2006). $\alpha 5 \beta 1$ interacts with a potent antiangiogenic endostatin molecule in addition to its beneficial effect in angiogenesis (Fig. 4) (Sudhakar *et al.*, 2003) (Fig. 4).

During endothelial cells undergoing neoplastic cell angiogenesis, integrin $\alpha 5 \beta 1$ expression is highly elevated, although it is very low in normal endothelial cells. In endothelial cells, the transcription factor HoxD3, which is associated with the homeobox family, regulates the production of $\alpha 5 \beta 1$. bFGF activates HoxD3 (Avraamides *et al.*, 2008). The ECM proteins Del-1, bFGF, and IL-8, but not VEGF, were the factors that caused integrin $\alpha 5 \beta 1$ to express. The $\alpha 5 \beta 1$ integrin-

mediated adhesion promotes endothelial cell motility and neoplasm cell survival by inhibiting protein kinase A activity. Therefore, during angiogenesis, $\alpha 5 \beta 1$ integrins preserve the survival of endothelial cells (Zhu *et al.*, 2017). Elevated levels of $\alpha 5 \beta 1$ were shown to be significantly correlated with lymph node metastases of NSCLC. The survival rate was worse for patients with NSCLC displaying excess $\alpha 5 \beta 1$ integrin than for those with NSCLC displaying normal levels of $\alpha 5 \beta 1$. $\alpha 5 \beta 1$ integrin promotes the development of NSCLC and the spread of metastases (Aksorn and Chanvorachote, 2019; Sökelland and Schumacher, 2019).

OS: prevention and medication against lung cancer

OS is a native plant of the tropics and other warm climates, known by several names such as tulsi (Hindi), holy basil (English), and kemangi (Indonesia) (Joseph and Nair, 2013). Indonesians use this fragrant herb as a vegetable (Cohen, 2014; Wihadmadayati *et al.*, 2019). This plant has a wide range of morphological traits, which are impacted by where it grows. These plants are often fragrant, herbaceous, sun shrubs with branching, and hairy subquadrangular branches that are between 30 and 60 cm long. The plant has simple, green, or purple leaves that can be ovate, elliptic-oblong, obtuse, or acute. The leaves have pubescent edges on both surfaces and can be slightly toothed with all substrate or dentate margins. The leaves are also dotted with tiny glands and have slender, hairy petioles. Volatile oils are produced by the glandular hairs on the stalked and sessile glands on the aerial section. The flowers are purplish-white in hue and hermaphrodite, grouped in elongated racemes in tight whorls. The seeds are globose-subglobose in form and are brownish-reddish yellow (Malav *et al.*, 2015) (Fig. 2).

The classification of OS:

Kingdom	: Plantae
(Unranked)	: Angiospermae
(Unranked)	: Eudicots
(Unranked)	: Asterids
Order	: Lamiales
Family	: Lamiaceae
Genus	: <i>Ocimum</i>
Species	: OS L.

OS has a variety of applications in different parts of Asia, including religious and therapeutic uses. It is used to treat inflammation, stomach pains, headaches, malaria, and the common cold in traditional medicine (Yamani *et al.*, 2016). OS has been shown in prior research to have antibacterial, antiviral, antidiabetic, antioxidant, antidepressant, wound healing, anti-inflammatory, hepatoprotective, cardioprotective, hypolipidemic, immunomodulatory, radioprotective, and neurodegenerative protection functions (Raghavendra *et al.*, 2009; Yamani *et al.*, 2016; Almatroodi *et al.*, 2020).

OS has an abundance of chemical compounds. The ethanolic leaf extract of OS contains toluene, camphene, octane, benzene, citronellal, sabinene, limonene,



Fig. 2. The OS structure. OS is a perennial plant with a medium stature and a woody base. OS grows to a height of 30–60 cm on a standing-up, multibranched stem with hairy, subquadrangular stems. The oval-shaped, green leaves are narrowed at both ends. The petioles are 1.5–3 cm long and very slender. The petals are clustered into bracteate clusters that are 15–20 cm long on little stems.

ledol, dimetilbenzena, etil-2-metilbutirat, eugenol, terpinolene, β -elemene, isocaryophyllene, iso-eugenol, α -amorf, α -guaiene, α -humulen, α -terpineol, borneol, calamine, nerolidol, carvacrol, geraniol, humulene, oksida, elemol, tetradecanal, (EZ)-famesol, cissquisainenehydrate, α -bisbolol, selin-11-en-4- α -ol, α -murolene, 14-hidroksi- α -humulen, carbohydrates, tannins, flavonoids, saponins, glycosides, terpenoids, and alkaloids. Many chemical components are formed when the leaf is extracted with methanol, including fatty acids, carbohydrates, phenol, tannins, flavonoids, saponin, glycosides, terpenoids, alkaloids, and fixed oils (Singh and Chaudhuri, 2018; Kustiati, *et al.*, 2022a). Furthermore, of the others, phenol was said to be the most prevalent. Using gas chromatography-mass spectrometry and liquid chromatography/mass spectrometry (LC/MS) analysis, methanol was extracted and eugenol, α -farnesene, benzene, 1,2-dimethoxy-4-(1-propenyl), cyclohexane, and 1,2,4-triethyl were found (Borah and Biswan, 2018).

The primary source of fixed oils, including oleic acid, stearic acid, hexuronic acid, palmitic acid, linodilininol, and linolenic acid, is the seeds of this plant. Other phytochemical components found in the aqueous and alcoholic extracts of defatted OS seeds include tannins (tannic acid) and flavonoids (quercetin and rutin) (Sharma *et al.*, 2020). Phenolic substances such as apigenin, circimaritin, isothymucin, eugenol, and rosameric acid are produced when the stem is extracted (Bhowmik *et al.*, 2008). After analyzing the

chemical composition of these leaves' volatile oils, 22 components were identified: α -pinene, α -thujene, α -camphene, β -pinene, α -mycrene, d-limonene, eucalyptol, cis- α -terpineol, sabinene, borneol, bornyl acetate, camphor, eugenol, selinene, caryophyllene oxide, β -guaiene, phytol, and oleic acid (Padalia and Verma, 2011; Dohare *et al.*, 2012; Wihadmadyatami *et al.*, 2019; Kustiati 2022a). A mixture including two lignans, four phenolics, and five terpenoids was obtained by extracting the aerial portion of OS with dichloromethane and methane (Table 3) (Flegkas *et al.*, 2019).

OS leaves, seeds, stems, fruits, flowers, and roots are almost all used as extracts or medications in traditional herbal remedies. However, in Indonesia, the leaves are the most commonly used herb in medicine, followed by the seeds, aerial parts, and entire plants. Similar to previous worldwide ethnobotanical studies conducted in Ghana, Lebanon, Morocco, and Turkey, an ethnobotanical reference study found that leaves were the main dominant plant component used in the composition of traditional treatments.

Several studies have been carried out to comprehend OS's impact on cancer. Ethanol extract of OS (EEOS) has been studied *in vivo* and *in vitro* for its anticancer properties. To determine the molecular target of EEOS, a study was carried out on a rat model of MNN-induced stomach carcinogenesis. As genes that promote proliferation and angiogenesis, the study found that eugenol, ursolic acid, and carvacrol extracted from

Table 3. The summary of phytochemical compound of the OS.

Group	Compound
Terpenoid	Ursolic acid, oleanolic acid, betulinic acid, stigmasterol, and β -caryophylleneoxide
Lignan	(-)-rabdosiin and shimobashiric acid C
Flavonoid	Luteolin, 7-O- β - _D -glucuronide, and apigenin 7-O- β - _D -glucuronide, quercetin, rutin, vicenin-2, vitexin, isovitexin, vicenin, luteolin, orientin, galuteolin, isorientin, aesculin, caffeic acid, rosmarinic acid, chlorogenic acid, cirsimaritin
Phenolics	(E)-p-coumaroyl 4-O- β - _D -glucoside, 3-(3,4-dihydroxyphenyl) lactic acid, protocatechuic acid, and vanillic acid.
Tannin	Tannic acid

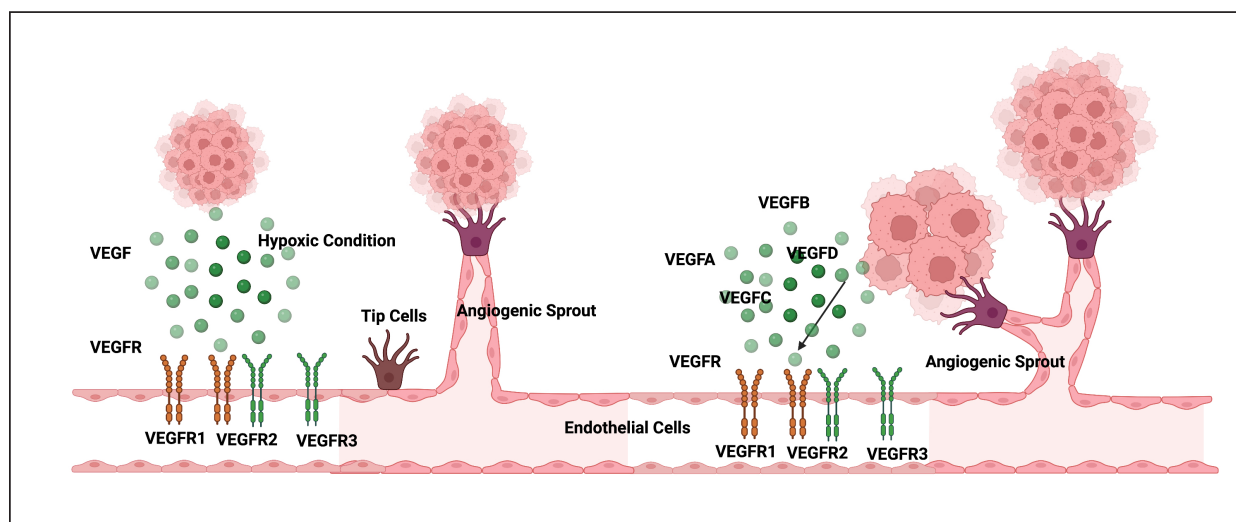


Fig. 3. The development of cancer cells is supported by the angiogenesis that is encouraged by the VEGF. Mutant cells constantly divide, growing bigger and more susceptible to hypoxia. Hypoxic conditions will promote microenvironment cells to secrete VEGF (VEGFA, VEGFB, VEGFC, VEGFD). VEGF will bind to its receptors (VEGFR1, VEGFR2, VEGFR3) and form tip cells on endothelial cells. Tip cells will develop branches (angiogenic sprouts) to form new blood vessel channels. The new blood vessel channels will continue to branch to form vascularization in cancer cells.

the OS leaf were able to lower the expression of PCNA, GST- π , BCL-2, CK, and VEGF in rats with stomach carcinogenesis caused by MNN. According to this discovery, substances also raise the expression of apoptosis-inducing genes such as caspase 3, cytochrome C, and Bax (Manikandan *et al.*, 2007). The study also revealed that the expression of EGFR, Kelch-like ECH-associated protein 1 (KEAP1), and β 2-adrenergic receptor (β 2-AR) was successfully inhibited by quercetin, rutin, and tannic acid that were separated from defatted OS seed extract. The G protein-coupled receptors, which are seven transmembrane receptors, including the β 2-AR, are involved in the development and spread of oral cancer. It was discovered that norepinephrine-stimulated β 2-AR enhanced IL-6 production and cell proliferation in oral squamous cell carcinoma cell lines (Sharma *et al.*, 2020). The lung is one of the tissues with high levels of NRF2 expression

where detoxifying processes take place. It interacts with KEAP1, its own negative regulator, under normal physiological settings. The oxidative stress sensor KEAP1 mediates Nrf2 degradation (Bauer *et al.*, 2013). According to research on an A549 human lung cancer cell, EEOS raises TUNEL 1 positive and the apoptotic sub-G1 in A549 cells, suggesting that enough DNA has been ruined by DNA fragmentation. According to a different study, EEOS's polyphenols and flavonoids cause A549 cells to separate from the ECM, which lowers the cells' viability. Furthermore, the capacity of EEOS to induce apoptosis raises the quantity of intracellular reactive oxygen species (ROS) and the interference activity of superoxide dismutase (SOD) and GP α (Fig. 5) (Wihadmadayati *et al.*, 2019). In A549 cells, angiogenesis is inhibited by downregulating integrin α v β 3, α 5 β 1, MMP-2, and MMP-9 (Wihadmadayati *et al.*, 2020; Kustiati

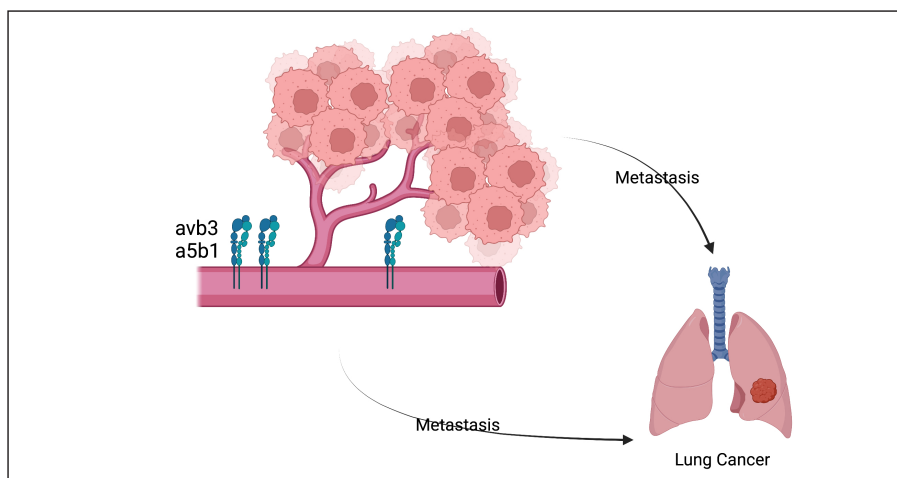


Fig. 4. The interaction of integrins $\alpha v\beta 3$ and $\alpha 5\beta 1$ with ECM regulates metastatic activity. The expression of integrin $\alpha v\beta 3$ and $\alpha 5\beta 1$ will significantly increase during the process of angiogenesis. Integrins will bind to the ECM and enhance cancer cell adhesion. Cancer cell adhesion will help cancer cells move to blood vessels that supply blood to cancer cells so that cancer cells can move to other organs suitable for growth.

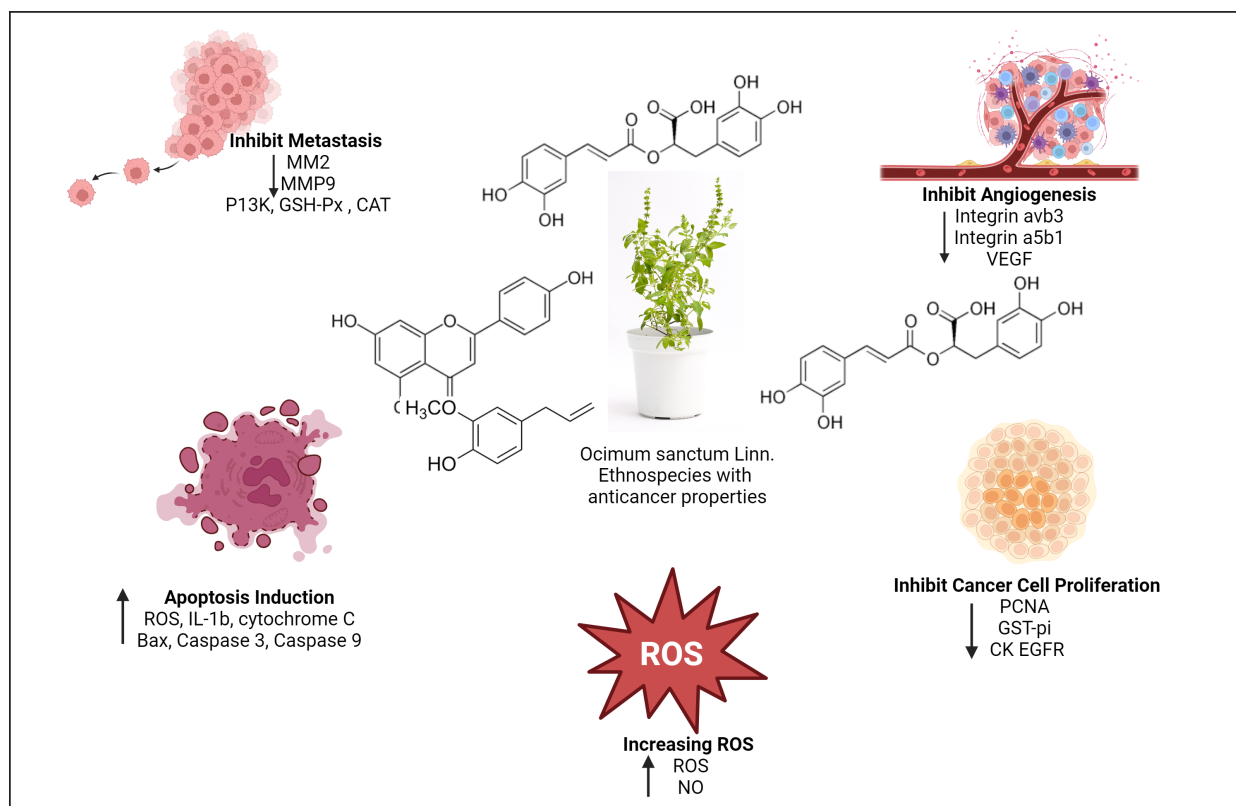


Fig. 5. OSs active ingredients inhibit cancer cells from proliferating and cause them to undergo apoptosis. Several active substances, including quercetin, eugenol, and rosmarinic acid, have been identified in OS Linn. Through the induction of cell death by an increase in ROS and NO, these bioactive substances will limit and shrink the growth of cancer cells. Bax, cytochrome C, caspase 3, caspase 9, and IL-1 β all rise throughout the apoptotic process. Proliferating cell nuclear antigen (PCNA), glutathione S-transferase (GST-pi), and EGFR will all drop in response to a reduction in cancer cell proliferation. OS also inhibits metastasis, which is characterized by decreased expression of ECM (MMP-2, MMP-9), PI3K, glutathione peroxidase (GSH-Px), and catalase (CAT). It does this by reducing integrin $\alpha v\beta 3$, integrin $\alpha 5\beta 1$, and VEGF.

et al., 2022). The anti-metastasis action of EEOS was demonstrated to be mediated by inhibition of phosphatidylinositol 3-kinase (P13K) in research conducted on NCI-H460 NSCLC cells (Kwak *et al.*, 2014). According to Utispan *et al.* (2019), rosmarinic acid in the extract of EEOS may reduce the activity of MP after preventing cell invasion (Utispan *et al.*, 2020) (Fig. 5).

The (-)-rabdosiin of EEOS has been shown to stimulate antiproliferation in human cancer cell lines in another investigation (Flegkas *et al.*, 2019). Applying EEOS to human breast cancer cells has the potential to increase the expression of P53 and Bid (BH3 interacting domain death agonist) as well as the Bax/Bcl2 protein ratio (Manaharan *et al.*, 2013). Singhal *et al.* (2017) reported that vicenin-2, a flavonoid derived from tulsi (holy basil), inhibits the proliferation of prostate cancer (CaP) and increases the antitumor properties of docetaxel *in vitro* and *in vivo* when compared to other active substances of OS Linn, such as erientin and luteolin. Vicenin-2 raises p53 levels, decreases hTERT, PCNA, and pRB, and suppresses pEGFR, pAkt, and the mTOR signaling mediator Pp70S6K (Singhal *et al.*,

2017). Inducing apoptosis, preventing cell invasion, limiting proliferation, restricting metastasis, and inhibiting angiogenesis are among the functions of OS in lung cancer (Fig. 5).

Multiple investigations have indicated that OS's ability to fight cancer is significantly influenced by intrinsic mitochondrial apoptosis. Cytochrome C is released into the cytosol when mitochondrial malfunction brought on by the cleavage of Bid by the intrinsic apoptosis pathway triggers apoptosis. Apoptotic protease activating factor-1, or Apaf1, is an adaptor protein that is activated when cytochrome C in the cytosol attaches to procaspase. This process creates the apoptosome, which in turn attracts procaspase 9. Apoptosis is brought on by the activation of caspase-9 and caspase-3, which causes cell death and DNA fragmentation, among other apoptotic hallmarks (Manaharan *et al.*, 2013, Manaharan *et al.*, 2014; Wihadmadyatami *et al.*, 2019, 2020). Table 4 provides a summary of the OS's function.

Future perspective

The results in this review indicate that mouse models predominantly used in cancer research, particularly in

Table 4. The role of OS ethanolic extract to prevent cancer metastasis.

Role of OS	Gene	Reference
Apoptosis induction	BCI2	(Manaharan <i>et al.</i> , 2014; Manikandan <i>et al.</i> , 2007)
	Bax	(Manaharan <i>et al.</i> , 2014; Manikandan <i>et al.</i> , 2007)
	P53	(Manaharan <i>et al.</i> , 2014)
	Bid	(Shoji <i>et al.</i> , 2023)
	ROS	(Wihadmadyatami <i>et al.</i> , 2019)
	SOD	(Wihadmadyatami <i>et al.</i> , 2019)
	GPα	(Wihadmadyatami <i>et al.</i> , 2019)
	Cytochrome C	(Manikandan <i>et al.</i> , 2007)
	Caspase 3	(Manikandan <i>et al.</i> , 2007)
Oxidative stress	ROS	(Wihadmadyatami <i>et al.</i> , 2019)
Inhibit invasion cell	MMP	(Utispan <i>et al.</i> , 2020)
Inhibit proliferation	PCNA	(Manikandan <i>et al.</i> , 2007)
	GST-pi	(Manikandan <i>et al.</i> , 2007)
	CK	(Manikandan <i>et al.</i> , 2007)
	EGFR	(Sharma <i>et al.</i> , 2020)
Antimetastasic	P13K	(Kwak <i>et al.</i> , 2014)
	MMP-9	(Kim <i>et al.</i> , 2010)
	GSH-Px	(Kim <i>et al.</i> , 2010)
	CAT	(Kim <i>et al.</i> , 2010)
Inhibit angiogenesis	VEGF	(Kustiati <i>et al.</i> , 2022; Manikandan <i>et al.</i> , 2007)
	Integrin αvβ3	(Kustiati <i>et al.</i> , 2022; Wihadmadyatami <i>et al.</i> , 2020)
	MMP-2	(Wihadmadyatami <i>et al.</i> , 2020)
	MMP-3	(Kim <i>et al.</i> , 2010)
	Integrin α5β1	(Kustiati <i>et al.</i> , 2022)

the study of lung cancer. The use of mice as a lung cancer model is because mice have the advantage of genetic tractability and the ability to modify different biological pathways. Nevertheless, these models possess notable constraints that can impact the applicability of results to human patients. A significant drawback of mice models is their inability to completely mimic the genetic and phenotypic diversity exhibited in human lung cancer. Moreover, the immune system of mice, especially in models with reduced immunity, fails to replicate the immunological response observed in humans accurately. The constraint above can substantially influence the examination of interactions between tumors and the immune system and the effectiveness of immunotherapies (Weiss *et al.*, 2015). In order to address this constraint, there is a continuous effort to establish a wider range of animal species as models for studying lung cancer. This novel method is essential for enhancing the disease's comprehension mechanism and therapy solutions. Moreover, applying genetic engineering, environmental factors, and lifestyle factors in these models will increase their significance and effectiveness in preclinical research, ultimately leading to improved outcomes for individuals with lung cancer.

Understanding the interactions between tumor cells, stroma, and microenvironment, which are included in molecular carcinogenesis, is critical for developing effective therapies. Research into the role of immune cells, fibroblasts, and ECM components in lung cancer can provide insights into how these interactions influence tumor growth and metastasis (Pullamsetti *et al.*, 2017). Targeting these interactions may enhance existing therapies' effectiveness and lead to new treatment strategies.

Moreover, as a new approach to treatment strategies, the use of herbal medicine, it is possible to employ phytoconstituents of OS in lung cancer, deeper research involving the use of various animal models and new technologies such as organoids and organs on a chip, is needed related to the functional mechanism, standardization of dosages, and regulations on the use of herbal medication.

Conclusion

Recently, mice have been used extensively as experimental models for lung cancer. Chemically induced mice, lung cancer cell line induction, PDXs, and genetically modified mice are some of the manufacturing techniques used. It is crucial to apply the strategy of using molecular carcinogenesis interactions, such as tumor microenvironment, genetic mutation, and epigenetics, to improve the efficacy of therapy and encourage the development of novel treatment modalities, particularly those involving herbal medicine. One of the most widely used herbal remedies in Southeast and South Asia is OS. Lung cancer therapy and prevention heavily depend on OS. OS possesses

antioxidant, anti-inflammatory, immunomodulatory, antiproliferative, proapoptotic, antiangiogenic, and antimetastatic qualities, which account for its reported chemopreventive and anticancer therapeutic benefits. Furthermore, OS has the ability to control enzymatic activity, including GSH, GST, GSH-Px, and several antioxidant enzymes such as SOD and CAT, as well as signal transduction pathways.

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Conflict of interest

The authors declare that they have no conflict of interest

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Authors' contributions

Conceptualized and Supervision: H.W., D.L.K., D and .A.A.N., Methodology: H.W., Writing-original draft: U.K., H.W., D.L.K., and D.A.A.N.; Writing-review and editing: H.W. and U.K.; Visualized: U.K. and H.W. All authors have read and approved the final manuscript.

Ethical approval

Since this review does not use materials obtained directly from humans or animals, there is no need for ethical clearance.

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