

1 **COVID-19 vaccine effectiveness against SARS-CoV-2 infection in the United States prior to the Delta and Omicron-**  
2 **associated surges: a retrospective cohort study of repeat blood donors**

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15 **Running title:** Vaccine effectiveness in repeat blood donors

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17 Clinical Diagnostics and Roche.

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1 **ABSTRACT**

2 To inform public health policy, it is critical to monitor COVID-19 vaccine effectiveness (VE), including against  
3 acquiring infection. We estimated VE using self-reported vaccination in a retrospective cohort of repeat blood donors who  
4 donated during the first half of 2021, demonstrating a viable approach for monitoring of VE via serological surveillance.  
5 Using Poisson regression, we estimated an overall VE of 88.8% (95% CI: 86.2–91.1), adjusted for demographic  
6 covariates and variable baseline risk. Time since first reporting vaccination, age, race-ethnicity, region, and calendar time  
7 were statistically significant predictors of incident infection.

8 Key words: COVID-19, SARS-CoV-2; vaccines; vaccine effectiveness; blood donors

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## 1 INTRODUCTION

2 COVID-19 vaccines have a critical role in preventing symptomatic and serious disease caused by SARS-CoV-2  
3 [1], and in reducing viral transmission [2]. Vaccines approved or authorized for emergency use in the US by the Food and  
4 Drug Administration had high efficacy against severe disease in Phase III trials, ranging from 67% to 95% [1, 3-5].  
5 However, long-term management of the SARS-CoV-2 pandemic remains a substantial challenge, especially as variants of  
6 concern emerge. To inform public health policy, it is critical to monitor effectiveness, as differentiated from vaccine  
7 efficacy, of COVID-19 vaccines at the population-level over longer periods of time [6]. Numerous individual-level and  
8 environmental factors (e.g., mitigation practices, vaccine types, and vaccination coverage) impact vaccine effectiveness  
9 (VE), underscoring the importance of and need for population-based studies of VE against SARS-CoV-2 infection.

10 Evaluating VE is challenging given the multitude of endpoints relevant to SARS-CoV-2 infection. Many COVID-  
11 19 VE studies have evaluated severe endpoints, such as mortality and hospitalizations, often in higher risk populations.  
12 Mild and asymptomatic infections account for the vast majority of SARS-CoV-2 infections, with serosurveys indicating  
13 that many more infections occur than diagnosed cases [7]. Vaccine breakthrough infections are frequently asymptomatic  
14 with one study finding that among >10,000 breakthrough infections, over a quarter were asymptomatic [8]. Asymptomatic  
15 individuals (including with vaccine breakthrough infections) can transmit SARS-CoV-2 to others, and thus preventing  
16 asymptomatic infection is important to decrease widespread community transmission [9]. It is therefore important to  
17 monitor VE against acquiring infection to inform COVID-19 vaccine policy.

18 Constraints on large-scale population-level monitoring of VE include the cost and logistical challenges of  
19 enrolling and following cohorts of vaccinated and unvaccinated individuals. Case control study designs may be subject to  
20 bias if controls are not representative of the population from which cases are drawn. Molecular assays, applied to  
21 respiratory swab samples, are the gold standard for diagnosing SARS-CoV-2 infection. However, molecular assays have a  
22 limited detection window, necessitating frequent follow-up that reduces the feasibility of ongoing large-scale surveillance,  
23 higher cost, and lower throughput. In contrast, serological assays that detect binding antibodies (Abs) against the Spike  
24 (S) and Nucleocapsid (NC) viral proteins are lower cost, higher throughput, and can identify SARS-CoV-2 infections after  
25 resolution.

1 We estimated VE using a retrospective cohort of repeat blood donors who donated during the first half of 2021 at  
2 Vitalant, a major US blood collection organization (BCO). This proof-of-concept study demonstrates a viable approach  
3 for continued near real-time monitoring of VE via serological surveillance among repeat blood donors.

#### 4 **METHODS**

5 Beginning in June 2020, Vitalant tested all donors for anti-S and anti-NC Abs using the Ortho VITROS anti-  
6 SARS-CoV-2 S Total Ig and Roche Elecsys® NC Anti-SARS-CoV-2 assays. For this analysis, we included donations  
7 from donors who donated at least twice between January 1 to July 6, 2021, with anti-S and anti-NC test results and no  
8 evidence of past infection. Donors self-reported any COVID-19 vaccination at each donation and each interdonation  
9 interval was categorized as vaccinated or unvaccinated time at risk, based on serological status and COVID-19  
10 vaccination report. Specifically, we defined an interdonation interval as vaccinated time at risk if the donor was  
11 vaccinated and had anti-S Abs but not anti-NC Abs at the first timepoint and unvaccinated time at risk if the donor was  
12 not vaccinated at both timepoints and had no anti-S Abs at the first timepoint. Interdonation intervals during which  
13 vaccination took place were excluded because the proportion of vaccinated versus unvaccinated time could not be  
14 determined. A single donor could contribute both unvaccinated and vaccinated time at risk. Exact dates of vaccination and  
15 vaccine manufacturer were not recorded at donation. (Preliminary data from a survey of Vitalant blood donors conducted  
16 subsequently indicates that more than 90% received a Moderna or Pfizer-BioNTech mRNA vaccine.) Survey data further  
17 indicate that approximately 30% of intervals classified as vaccinated exposure time started when donors were partially  
18 vaccinated (i.e., less than 14 days after the final dose), but that most of the vaccinated exposure time accumulated after  
19 being fully vaccinated. Incident SARS-CoV-2 infections were identified using anti-NC seroconversion; vaccine  
20 breakthrough infection defined as seroconversion after self-report of COVID-19 vaccination. The demographic  
21 composition of the donor population is shown in Supplementary Table 1.

22 We used multivariable Poisson regression to evaluate the association between incident SARS-CoV-2 infection  
23 rate and COVID-19 vaccination and reported adjusted incidence rate ratio (IRR). The final model adjusted for the  
24 calendar month during which a donor's follow-up ended (to control for changing baseline risk over time), accumulated  
25 time vaccinated at the start of the final observation interval, sex, age, race-ethnicity, and the Vitalant-defined region in  
26 which donations were collected. Time at risk was treated as an offset in the regression analysis, with uninfected donors  
27 contributing the full interval, and infected donors contributing half of the interval during which infection occurred. A log

1 link function was used, and analysis conducted in R version 4.1.2. VE was defined as 1-IRR. Since a single donor could  
2 contribute both unvaccinated and vaccinated exposure time, and to account for within-donor correlation we computed  
3 95% confidence intervals (CIs) using 10,000 iterations of donor-level bootstrapping (resampling donors with  
4 replacement). As a comparator, we used naïve person-time methods to estimate incidence (number of events/time-at-risk)  
5 in vaccinated and unvaccinated donors and reported unadjusted IRRs and VE.

6 We conducted a simulation study to evaluate VE estimation methods and data inclusion rules with respect to bias  
7 and precision, with a particular focus on the bias that may arise when both incidence and the proportion of the population  
8 vaccinated changes rapidly during the study period, resulting in unvaccinated follow-up time accumulating  
9 disproportionately during a period of higher incidence. We simulated 1,020 datasets similar to the Vitalant repeat donor  
10 cohort, with varying rates of declining incidence, increasing vaccination, and proportions of the population vaccinated by  
11 the end of the period. We evaluated unadjusted IRRs based on conventional person-time methods, Cox proportional  
12 hazards regression on a calendar timescale, and Poisson regression adjusted for calendar time; methods were assessed for  
13 absolute and relative bias and precision (see Supplementary Appendix).

## 14 RESULTS

15 In total, 61,618 donors who donated in 18 geographic regions across the United States contributed 407,449  
16 unvaccinated person-weeks and 326,752 vaccinated person-weeks of time at risk during the study period (January 1 to  
17 July 6, 2021). Most donors (77%) contributed a single interval (median 1, range 1–25), and the median interval length was  
18 56 days (IQR: 28–71). Poisson regression adjusted for the calendar time when a particular donor's follow-up ends (either  
19 with an infection event or still at risk) had low bias, good precision, and confidence interval coverage >95% (see  
20 Supplementary Appendix).

21 We identified 1,653 incident infections in unvaccinated donors, and 100 vaccine breakthrough infections. Kaplan-  
22 Meier survival curves for both groups are shown in Figure 1. Using multivariable Poisson regression, we estimated overall  
23 VE was 88.8% (95% CI: 86.2–91.1), derived from an IRR of 0.11 (0.09–0.14), adjusted for demographic covariates and  
24 variable baseline risk (Table 1). The number of days that the donor had been vaccinated at the start of the final interval  
25 was significant and protective, with a 2% reduction in incidence associated with a 1-day increase in time since vaccination  
26 (IRR 0.98 [0.97,0.99]). Age (1% reduced incidence associated with an additional year of life), Asian or Pacific Islander  
27 race (IRR 0.56 [0.31,0.84]), several geographic regions, and the month during which a donor's follow-up ended, from

1 March onwards, were statistically significant predictors of incident infection, though sex was not (Table 1). The naïve  
2 analysis yielded an unadjusted VE estimate of 92.5% (90.8%–93.8%, Table 1).

3 Our simulation study showed substantial bias with a naïve analysis, and that Poisson regression controlling for  
4 calendar time effectively addresses confounding arising from changing baseline hazard over time.

## 5 **DISCUSSION**

6 VE against acquiring any serologically identifiable SARS-CoV-2 infection, was high in blood donors during the  
7 first half of 2021 at 88.8%. Our VE estimate was comparable to previous VE estimates in studies evaluating protection  
8 against SARS-CoV-2 infection based on detection of viral RNA in swab samples, which ranged from 73% to 98.2% [10-  
9 12], and mostly lower than estimates of protection against hospitalization, which ranged from 88.0% to 95.1% [10, 13,  
10 14]. Evidence of increasing protection with longer times since vaccination in these data are consistent with maturing  
11 immune responses during the early period of vaccine implementation, but over longer timescales waning protection would  
12 be expected, as has been observed in other studies [10].

13 A major strength of this study was the broad assessment of a large number of repeat blood donors residing in 18  
14 states during universal screening for SARS-CoV-2 Abs. Our model accounted for spatiotemporal factors and provided a  
15 more generalizable estimate of VE in the US population compared to previous studies in clinical populations. Our  
16 simulation-based exploration of estimation methods identified a robust statistical approach, specifically based on the  
17 criterion of having low bias in situations of time-varying infection and vaccination dynamics. The study constitutes an  
18 important proof of concept for the use of serological surveillance of blood donors to monitor VE over time as variants  
19 have and continue to emerge and vaccine-induced Ab responses wane or are enhanced through booster vaccinations. This  
20 approach further facilitates larger sample sizes and consequently improved precision in VE estimates at lower cost than  
21 clinical cohort studies.

22 Several limitations need to be considered when interpreting these results. The serological assays employed in this  
23 study have high sensitivity and specificity for past infection and vaccination-induced Abs, but lower sensitivity than  
24 molecular assays for acute infection. From the DHQ responses, we did not have detailed information on COVID-19  
25 vaccination timing, number of doses, or vaccine type, and there is the potential for misreporting of vaccination status by  
26 donors.. To limit the impact of potential misclassification of vaccination status, we required serological evidence of an

1 immunological response (presence of anti-S Abs and absence of anti-NC Abs) to corroborate self-reported vaccination,  
2 although donors could still have been classified as vaccinated after receiving only the first of a two-dose series or not long  
3 enough after receiving a single-dose vaccine or the second dose of a two-dose series to have developed robust Ab  
4 responses and be considered fully vaccinated. Sporadic presentation for donation, which for some donors may be very  
5 infrequent, is inherent to repeat blood donor datasets. We developed methods to account for interval censoring of both  
6 vaccination and infection events, but some data were excluded because vaccination status or the ordering of vaccination  
7 and infection events was uncertain during the interval. We used Vitalant collection region as a proxy geographic variable,  
8 however this may not capture with adequate granularity the heterogeneity in local transmission.

9 Blood donors are not representative of the US population, e.g., racial/ethnic minorities are underrepresented,  
10 which may introduce bias if VE differs in underrepresented population groups. Overall, donors are healthier than the  
11 general population, but this is unlikely to bias VE estimates since the healthy donor effect is likely present in both  
12 vaccinated and unvaccinated donors. Vaccinated donors were disproportionately female and older, potential confounders  
13 we controlled for in the analysis. These data were collected before widespread circulation of Delta and Omicron variants  
14 of SARS-CoV-2. Anti-NC seroconversion may be less likely in vaccine breakthrough infections [15], although our  
15 analysis of survey respondents reporting swab-confirmed breakthrough infections who were tested with the NC assay  
16 indicated sensitivity of >90%.

17 Our results showed a high VE against acquiring SARS-CoV-2 infection during the first half of 2021, although  
18 data collection preceded epidemic surges driven by spread of Delta and Omicron variants during the second half of 2021.  
19 We have established repeat donor cohorts for continued monitoring in partnership with another large BCO and the US  
20 Centers for Disease Control and Prevention for long-term serosurveillance, including comparison of VE during periods of  
21 Delta and Omicron predominance. SARS-CoV-2 antibody assays will be complemented with more detailed donor surveys  
22 regarding COVID-19 vaccinations (primary and booster), COVID-19 diagnoses, and symptoms. This next study phase  
23 will also include estimation of VE by vaccine manufacturer and timing of primary and booster doses, as well as  
24 assessment of the severity of vaccine breakthrough infections.

25  
26 **Ethics and informed consent:** Blood donors provided consent for the use of donation data and biospecimens in research  
27 at the time of donation. Consistent with the policies and guidance of the University of California San Francisco

1 Institutional Review Board, Vitalant Research Institute self-certified that use of the deidentified data in this study does not  
2 meet the criteria for human subjects research. Centers for Disease Control and Prevention (CDC) investigators reviewed  
3 and relied on this determination as consistent with applicable federal law and CDC policy (45 C.F.R. part 46, 21 C.F.R.  
4 part 56; 42 U.S.C. § 241[d]; 5 U.S.C. § 552a; 44 U.S.C. § 3501).

5 **Disclaimer:** The findings and conclusions in this report are those of the authors and do not necessarily represent the  
6 official position of the CDC.

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## REFERENCES

1. Tregoning JS, Flight KE, Higham SL, Wang Z, Pierce BF. Progress of the COVID-19 vaccine effort: viruses, vaccines and variants versus efficacy, effectiveness and escape. *Nat Rev Immunol* **2021**; 21:626-36.
2. Singanayagam A, Hakki S, Dunning J, et al. Community transmission and viral load kinetics of the SARS-CoV-2 delta (B.1.617.2) variant in vaccinated and unvaccinated individuals in the UK: a prospective, longitudinal, cohort study. *Lancet Infect Dis* **2022**; 22:183-95.
3. Polack FP, Thomas SJ, Kitchin N, et al. Safety and Efficacy of the BNT162b2 mRNA Covid-19 Vaccine. *N Engl J Med* **2020**; 383:2603-15.
4. Baden LR, El Sahly HM, Essink B, et al. Efficacy and Safety of the mRNA-1273 SARS-CoV-2 Vaccine. *N Engl J Med* **2020**; 384:403-16.
5. Sadoff J, Gray G, Vandebosch A, et al. Safety and Efficacy of Single-Dose Ad26.COV2.S Vaccine against Covid-19. *N Engl J Med* **2021**; 384:2187-201.
6. Hungerford D, Cunliffe NA. Real world effectiveness of covid-19 vaccines. *BMJ* **2021**; 374:n2034.
7. Jones JM, Stone M, Sulaeman H, et al. Estimated US Infection- and Vaccine-Induced SARS-CoV-2 Seroprevalence Based on Blood Donations, July 2020-May 2021. *JAMA* **2021**; 326:1400-9.
8. Team CC-VBCI. COVID-19 Vaccine Breakthrough Infections Reported to CDC - United States, January 1-April 30, 2021. *MMWR* **2021**; 70:792-3.
9. Jefferson T, Spencer EA, Brassey J, et al. Transmission of Severe Acute Respiratory Syndrome Coronavirus-2 (SARS-CoV-2) from pre and asymptomatic infected individuals: a systematic review. *Clin Microbiol Infect* **2022**; 28:178-89.
10. Tartof SY, Slezak JM, Fischer H, et al. Effectiveness of mRNA BNT162b2 COVID-19 vaccine up to 6 months in a large integrated health system in the USA: a retrospective cohort study. *Lancet* **2021**; 398:1407-16.
11. Butt AA, Omer SB, Yan P, Shaikh OS, Mayr FB. SARS-CoV-2 Vaccine Effectiveness in a High-Risk National Population in a Real-World Setting. *Ann Intern Med* **2021**; 174:1404-8.
12. Andrejko KL, Pry J, Myers JF, et al. Prevention of COVID-19 by mRNA-based vaccines within the general population of California. *Clin Infect Dis* **2022**; 74:1382-1389.

- 1 13. Self WH, Tenforde MW, Rhoads JP, et al. Comparative Effectiveness of Moderna, Pfizer-BioNTech, and Janssen  
2 (Johnson & Johnson) Vaccines in Preventing COVID-19 Hospitalizations Among Adults Without Immunocompromising  
3 Conditions - United States, March-August 2021. *MMWR* **2021**; 70:1337-43.
- 4 14. Rosenberg ES, Holtgrave DR, Dorabawila V, et al. New COVID-19 Cases and Hospitalizations Among Adults, by  
5 Vaccination Status - New York, May 3-July 25, 2021. *MMWR* **2021**; 70:1150-5.
- 6 15. Follmann D, Janes HE, Buhule OD, et al. Anti-nucleocapsid antibodies following SARS-CoV-2 infection in the  
7 blinded phase of the mRNA-1273 Covid-19 vaccine efficacy clinical trial. medRxiv **2022**;2022.04.18.22271936.  
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1 TABLES AND FIGURES

2 Table 1: Vaccine effectiveness estimates against anti-nucleocapsid antibody seroconversion\* using multivariable  
 3 regression and person-time approaches among US blood donors, January-July 2021

Model	Incidence in vaccinated donors <i>infections/10<sup>4</sup></i> <i>person-weeks</i> (95% CI)	Incidence in unvaccinated donors <i>infections/10<sup>4</sup></i> <i>person-weeks</i> (95% CI)	Predictors / covariates**	(adjusted) Incidence Rate Ratio (95% CI)	Vaccine Effectiveness % (95% CI)
Poisson regression			<i>Vaccination status</i>		88.8 (86.2–91.1)
			Unvaccinated	<i>ref</i>	
			<b>Vaccinated</b>	<b>0.11 (0.09,0.14)</b>	
			<b>Days vaccinated at start of final interval</b>	<b>0.98 (0.97,0.99)</b>	
			<i>Sex</i>		
			Female	<i>ref</i>	
			Male	0.99 (0.90,1.09)	
			<b>Age (in years at first donation)</b>	<b>0.99 (0.99,0.99)</b>	
			<i>Race-ethnicity</i>		
			White, Non-Hispanic	<i>ref</i>	
			Black, Non-Hispanic	0.67 (0.34,1.06)	
			<b>Asian/Pacific Islander, Non-Hispanic</b>	<b>0.56 (0.31,0.84)</b>	
			Native American/Alaskan, Non-Hispanic	0.70 (0.22,1.27)	
			Hispanic ethnicity	0.93 (0.76,1.11)	
			Missing/Unknown/Refused	1.24 (0.88,1.65)	
			<i>Vitalant Regional Center***</i>		
			Albuquerque, NM (ALB)	<i>ref</i>	
			Billings, MT (BIL)	1.3 (0.92,1.85)	
			Cheyenne, WY (CYS)	1.28 (0.78,1.91)	
			<b>Denver, CO (DEN)</b>	<b>1.39 (1.10,1.80)</b>	
			El Paso, TX (ELP)	1.42 (0.95,2.06)	
			<b>Fargo, ND (FAR)</b>	<b>2.00 (1.56,2.63)</b>	
			<b>Lafayette, LA (LAF)</b>	<b>1.55 (1.13,2.12)</b>	
			Las Vegas, NV (LAS)	1.16 (0.86,1.59)	
			Lubbock, TX (LBB)	1.25 (0.88,1.77)	
			McAllen, TX (MCA)	0.90 (0.38,1.59)	
			Memphis, TN (MEM)	1.38 (0.94,1.98)	
			Phoenix, AZ (PHX)	1.08 (0.85,1.49)	
			Rapid City, SD (RAP)	1.48 (0.98,2.04)	
			Reno, NV (RNO)	0.82 (0.56,1.18)	
			<b>Sacramento, CA (SAC)</b>	<b>0.59 (0.44,0.79)</b>	
			<b>San Francisco, CA (SFO)</b>	<b>0.30 (0.17,0.47)</b>	
			<b>Spokane, WA (SPK)</b>	<b>1.51 (1.14,2.03)</b>	
			<b>Ventura, CA (VTA)</b>	<b>0.43 (0.27,0.62)</b>	
End of follow-up					
January 2021	<i>ref</i>				
February 2021	0.55 (0.32,1.20)				
<b>March 2021</b>	<b>0.31 (0.19,0.65)</b>				
<b>April 2021</b>	<b>0.39 (0.24,0.83)</b>				
<b>May 2021</b>	<b>0.36 (0.22,0.76)</b>				

			June/July 2021	<b>0.26 (0.16,0.55)</b>	
Naïve person-time	3.1 (2.5–3.7)	40.6 (38.6,42.6)	N/A	0.075 (0.062–0.092)	92.5 (90.8–93.8)

1

2 \* Vaccine breakthrough infections defined as anti-nucleocapsid seroconversion after any self-report of previous COVID-19  
3 vaccination. Number of vaccine doses not collected. Analysis included 100 vaccine breakthrough infections.

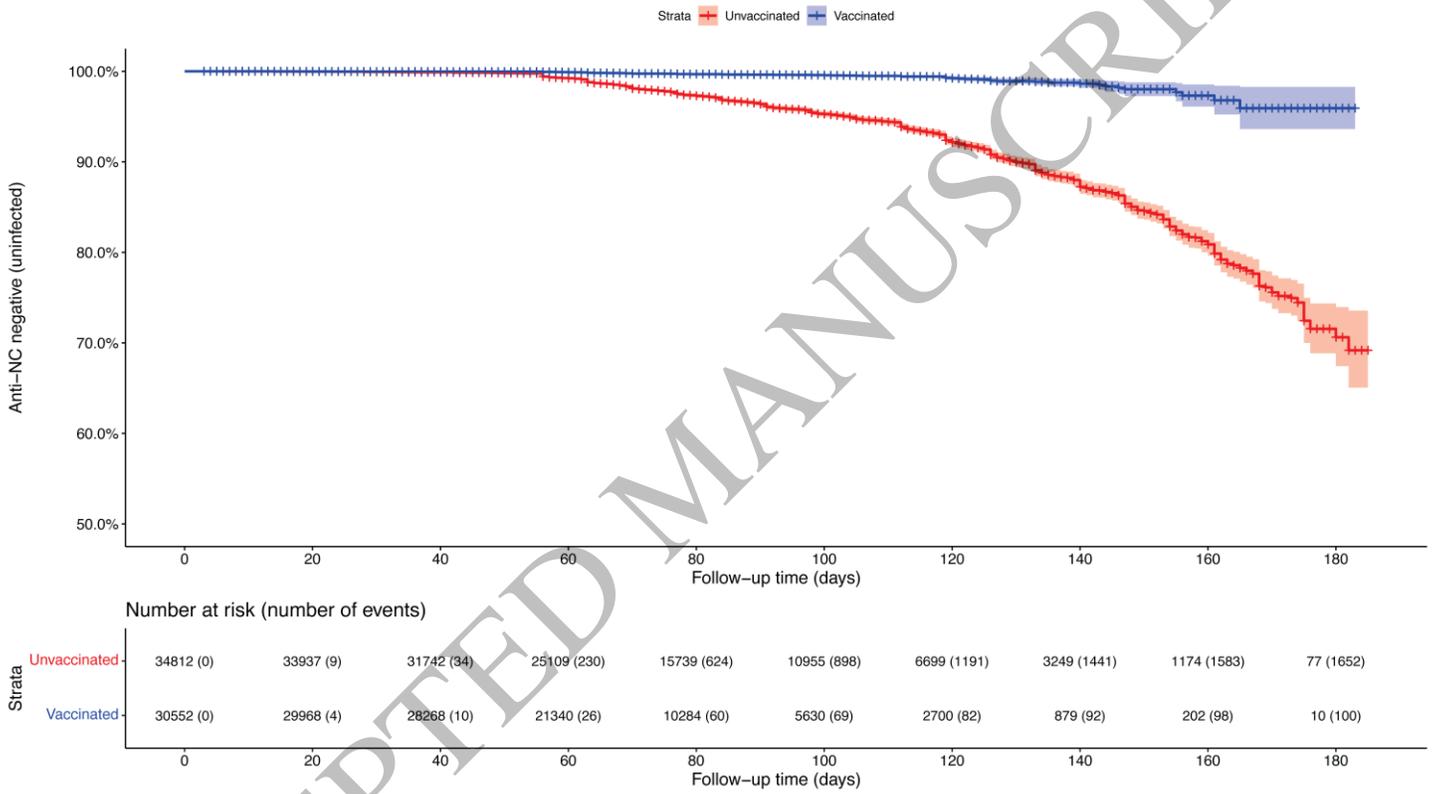
4 \*\* Statistically significant predictors indicated in bold font.

5 \*\*\* Geographic collection areas for Vitalant regional centers are shown in Figure 1 of manuscript Vassallo, RR, Bravo MD, Dumont LJ,  
6 Hazegh K, Kamel H. Seroprevalence of Antibodies to SARS-CoV-2 in US Blood Donors. medRxiv 2020.09.17.20195131; doi:  
7 10.1101/2020.09.17.20195131.

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1 **Figure 1: Kaplan-Meier survival curves by vaccination status comparing time to anti-nucleocapsid antibody**  
 2 **seroconversion among US blood donors, January-July 2021**



6 **Figure 1**  
 7 **190x107 mm (1.0 x DPI)**  
 8