

Original article:

**PLASMA VASCULAR ENDOTHELIAL GROWTH FACTOR B IS
ELEVATED IN NON-ALCOHOLIC FATTY LIVER DISEASE
PATIENTS AND ASSOCIATED WITH BLOOD PRESSURE AND
RENAL DYSFUNCTION**

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ABSTRACT

Vascular endothelial growth factor B (VEGF-B) is a critical metabolic regulator in insulin resistance, and lipid distribution. We intended to ascertain the relationship between circulating VEGF-B and non-alcoholic fatty liver disease (NAFLD) in the general public. We recruited a total of 194 general participants for a routine physical health examination; of these, 84 participants were identified with NAFLD and 110 without NAFLD based on ultrasonographic findings. Homeostasis model assessment of insulin resistance (HOMA-IR), body mass index (BMI), HbA1c, liver function, kidney function, plasma VEGF-B levels and indexes of metabolic syndrome (blood pressure, fasting plasma glucose, fasting lipids) were evaluated. Plasma VEGF-B values were significantly higher in individuals with NAFLD compared to those without NAFLD ($P = 0.022$), and analysis of covariance confirmed this result. VEGF-B showed a positive correlation with γ -glutamyl transpeptidase (γ -GT) and HOMA-IR in univariate analysis ($q = 0.242$; $P = 0.001$; $q = 0.174$; $P = 0.019$, respectively). Multiple linear regression analysis showed that γ -GT and ALT were independently correlated with VEGF-B even after adjusted for gender and age ($q = 0.286$; $P = 0.01$; $q = 0.237$; $P = 0.033$, respectively). Moreover, plasma VEGF-B showed a powerful correlation with blood pressure and renal dysfunction. Plasma VEGF-B might be a new clinical variable related to NAFLD and could be a proper biomarker for the early detection of hypertension and renal dysfunction. However, further studies with large cohorts' size are warranted to validate our findings.

Keywords: Vascular endothelial growth factor B, non-alcoholic fatty liver disease, blood pressure, renal dysfunction, metabolism

INTRODUCTION

Vascular endothelial growth factors (VEGFs) are a family of signal proteins acting as important regulators of angiogenesis during physiological and pathological conditions (Olsson et al., 2006; Zafar et al., 2018). Vascular endothelial growth factor-B (VEGF-B) is a critical member of the VEGF protein family and is abundantly expressed in most tissues and organs (Aase et al., 1999; Holmes and Zachary 2005; Li et al., 2001, 2012). Traditional studies on VEGF-B largely concentrated on their effects on neurotropy, angiogenesis, and neuroprotection, while some ruled out the association between VEGF-B overexpression and tumor growth, invasion, diabetic retinopathy and others (Abedin et al., 2010; Falk et al., 2009; Olofsson et al., 1996; Zhong et al., 2011). Few recent studies identified that VEGF-B is tightly related to metabolism and obesity (Gomez-Ambrosi et al., 2010; Hagberg et al., 2010, 2012). Another study found that the overexpression of VEGF-B in the mouse heart is related to the increase of ceramide and triglycerides, which led to cardiac hypertrophy (Karpanen et al., 2008). Chen et al. reported that adipose VEGF-B repression induced adipose tissues transferred toward white adipose for energy storage, and the glucose metabolism and lipid metabolism are broadly changed (Chen et al., 2020).

There are limited clinical data which reported the pathological roles of VEGF-B in obese subjects and those with metabolic syndrome. Gomez-Ambrosi et al. reported that the serum VEGF-B levels were significant higher in the obese group compared with the lean group (Gomez-Ambrosi et al., 2010). Sun et al. compared the concentration of VEGF-B between individuals with type 2 diabetes (T2DM) and healthy controls and highlighted that there was no significant difference between the two groups (Sun et al., 2014). However, the findings in this study indicated that the use of thiazolidinediones can influence the circulating VEGF-B levels (Sun et al., 2014). Cheng et al. found that individuals with polycystic ovary syndrome (PCOS) had higher circulating VEGF-B levels than

the age-matched healthy controls (Cheng et al., 2016). Another study reported that plasma VEGF-B elevated in newly diagnosed T2DM and impaired glucose regulation (IGR) patients, compared with healthy controls (Wu et al., 2017). These studies drew a substantial association between VEGF-B and insulin resistance (IR), T2DM.

Clinical and preclinical studies have demonstrated IR as one of the core predisposing factors for metabolic diseases like obesity, T2DM, PCOS, and non-alcoholic fatty liver disease (NAFLD) (Eckel et al., 2005; Kitade et al., 2017; Macut et al., 2016). Up to now, there are no existing data on circulating VEGF-B levels in NAFLD individuals. Therefore, in this study, we intended to investigate and compare the plasma VEGF-B levels between cohorts with NAFLD and without NAFLD. Additionally, we also aimed to investigate relationships between plasma VEGF-B level and blood pressure (BP), kidney function, glucose, and lipid metabolism.

MATERIALS AND METHODS

Cohorts

A total of eighty-four cohorts (60 males, 24 females), aged 18–70 years, with fatty liver diagnosed by ultrasound, were recruited in the NAFLD group in this study. Whereas, one hundred and two adult individuals (49 males and 53 females), aged 18–70 years without any ultrasound evidence of NAFLD, were recruited in the control group. These cohorts attended the Healthcare Center of Union Hospital of Tongji Medical College, Huazhong University of Science and Technology for a routine physical health examination between May 2017 and August 2017. The cohorts were recruited if they met following inclusion criteria: negative tests for the presence of hepatitis B surface antigen and antibody to hepatitis C virus; no history of current or past excessive alcohol drinking (a threshold of < 20 g/d for women and < 30 g/d for men); no active or previous history of liver cirrhosis and other chronic liver diseases; no active history insulin treatment; and female

participant not to be pregnant at the time of recruitment. The cohorts undertook a complete health examination, including anthropometric measurements (body weight, height), BP measurement, fasting plasma glucose (FPG), HbA1c, blood lipids, fasting insulin, liver function test, and kidney function tests. HOMA-IR was calculated as [fasting insulin ($\mu\text{U/mL}$) \times FPG (mmol/L)/22.5]; and BMI was calculated using the following formula: [weight (kg)/ height² (m)]. Hypertension was defined as systolic blood pressure (SBP) \geq 140 mmHg, or diastolic blood pressure (DBP) \geq 90 mmHg.

The Ethical Committee of Tongji Medical College, Huazhong University of Science and Technology approved the current study, and our study was accordant with the declaration of Helsinki. We got the written informed consent from each participant before recruited them to the study.

VEGF-B assay

The circulating VEGF-B levels were tested by a commercially available human VEGF-B ELISA kit (USCN Science Co, Wuhan, China) in accordance with the manufacturer's protocol. The detection range of the VEGF-B ELISA test was 15.6 to 1000 pg/mL, and intra-assay and inter-assay variations were $< 10\%$ and $< 12\%$, respectively. The detection limit of this ELISA kit is 5.5 pg/mL.

Determination of fatty liver

The fatty liver was determined using ultrasonography by a proficient radiologist with extensive experience in abdominal ultrasound examinations using a high-resolution B-mode topographic ultrasound system with a 3.5 MHz probe (HDI 5000, Philips, Bothell, WA, USA). The fatty liver ultrasonography assessment consisted of at least two of the following findings: diffusely increased echogenicity ('bright') liver with liver echogenicity higher than kidney or spleen, deep attenuation of ultrasound signal, and vascular blurring (Farrell et al., 2007).

Statistical analysis

We used Statistical Package for Social Sciences, version 22.0 (SPSS, Chicago, IL, USA) to analyze the data. Results were shown as mean \pm SD. The differences of continuous variables between the NAFLD group and the control group, the hypertension group and the control group were analyzed using t-tests. We adjusted age, gender and FPG to perform the analysis of covariance (ANCOVA) between the NAFLD group and the control group. We analyzed the relationship between variables by Spearman's correlation coefficient test. Multivariate analysis was performed between sex, age, BMI, liver function, kidney function, HOMA-IR, HbA1c, blood lipids, BP and VEGF-B. All P values presented are two-tailed, and values less than 0.05 are recognized as statistically significant.

RESULTS

The mean age of the cohorts was 48.77 ± 11.01 years. Of all cohorts, 58.2 % were male ($n = 113$), and 41.8 % were female ($n = 81$). The average BMI recorded was 25.44 ± 4.79 kg/m², and 24 of them had diabetes. The cohort group allocation was based on medical history and liver ultrasonography, and the entire cohort was divided into groups, of these 102 patients (52.58 %) without NAFLD in the control group, and 84 patients (47.42 %) those have NAFLD (most of them presented mild liver lipid accumulation) in the NAFLD group. The clinical characteristics of these groups are displayed in Table 1. Briefly, age, estimated glomerular filtration rate (eGFR), alkaline phosphatase (ALP), and total cholesterol (TC) did not show a significant difference. Instead, FPG, HOMA-IR, BP, LDL-C, triglycerides (TG), BMI, alanine transaminase (ALT), aspartate aminotransferase (AST), γ -glutamyl transpeptidase (γ -GT), uric acid (UA) and HbA1c were all significantly higher in the cohorts of the NAFLD group, and levels of HDL-C were significantly lower in the NAFLD group. The VEGF-B measured values were significantly higher ($P = 0.022$) in NAFLD individuals

compared to non-NAFLD subjects (Figure 1a). Analysis of covariance between the two groups also indicated that VEGF-B were higher ($p=0.047$) in subjects with NAFLD after adjusting for FPG, gender and age. The

VEGF-B measured values were significantly higher ($P = 0.003$) in hypertension individuals compared to control subjects (Figure 1b).

Table 1: Main characteristics of the NAFLD group and controls

Variables	NAFLD	Controls	P-value
Gender (male/female)	60/24	49/53	
Age (years)	49.54±11.22	48.39±10.76	0.476
SBP (mmHG)	129.31±17.61	121.29±17.66	0.003
DBP (mmHG)	80.78±11.32	74.60±11.52	<0.001
Weight (kg)	78.27±16.27	167.57±7.79	<0.001
BMI (kg/m ²)	27.79±5.08	23.09±2.52	<0.001
UA (umol/L)	390.97±97.06	315.96±92.23	<0.001
BUN (mmol/L)	5.103±1.40	5.85±7.45	0.364
Cr (umol/L)	72.49±19.69	67.15±17.90	0.055
CysC (mg/L)	0.93±0.23	0.84±0.18	0,008
eGFR (ml/min/1.73m ²)	104.26±23.85	104.46±22.37	0.954
TB (umol/L)	14.59±5.97	15.61±5.43	0,231
CB (umol/L)	4.05±1.74	4.31±1.75	0.332
ALT (U/L)	30.65±26.31	22.00±6.43	0,004
AST (U/L)	35.33±25.50	19.57±10.40	<0.001
ALP (U/L)	70.90±17.09	69.10±21.78	0.54
γ-GT (U/L)	43.77±45.90	25.03±18.06	0.001
Total Protein (g/L)	73.44±4.07	73.16±4.49	0.662
Albumin (g/L)	46.18±2.38	46.08±2.99	0.809
Globulin (g/L)	27.14±3.52	27.08±3.25	0.913
A/G	1.72±0.25	1.73±0.23	0.891
TG (mmol/L)	1.94±0.96	1.25±0.80	<0.001
TC (mmol/L)	4.83±0.80	4.67±0.77	0.151
HDL-C (mmol/L)	1.19±0.27	1.45±0.36	<0.001
LDL-C (mmol/L)	3.06±0.75	2.82±0.64	0.02
FPG (mmol/L)	5.63±2.04	4.91±1.56	0.011
HbA1C (%)	6.01±1.42	5.55±1.31	0.048
Ins (uIU/mL)	19.50±16.22	7.97±11.58	<0.001
HOMA-IR	4.70±4.17	1.77±2.71	<0.001
VEGF-B (pg/ml)	40.11±16.93	34.73±14.69	0.022

Data presented as mean ± standard deviation

SBP, systolic blood pressure; DBP, diastolic blood pressure; BMI, body mass index; UA, uric acid; BUN, blood urea nitrogen; Cr, Creatinine; CysC, CystatinC; eGFR, estimated glomerular filtration rate; TB, total bilirubin; CB, conjugated bilirubin; ALT, alanine aminotransferase; AST, aspartate aminotransferase; ALP, alkaline phosphatase; γ-GT, γ-glutamyl transpeptidase; TG, total triglyceride; TC, total cholesterol; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; FPG, fasting plasma glucose; HbA1C, glycosylated hemoglobin; Ins, insulin; HOMA-IR, homeostatic model assessment for insulin resistance; VEGF-B, vascular endothelial growth factor B.

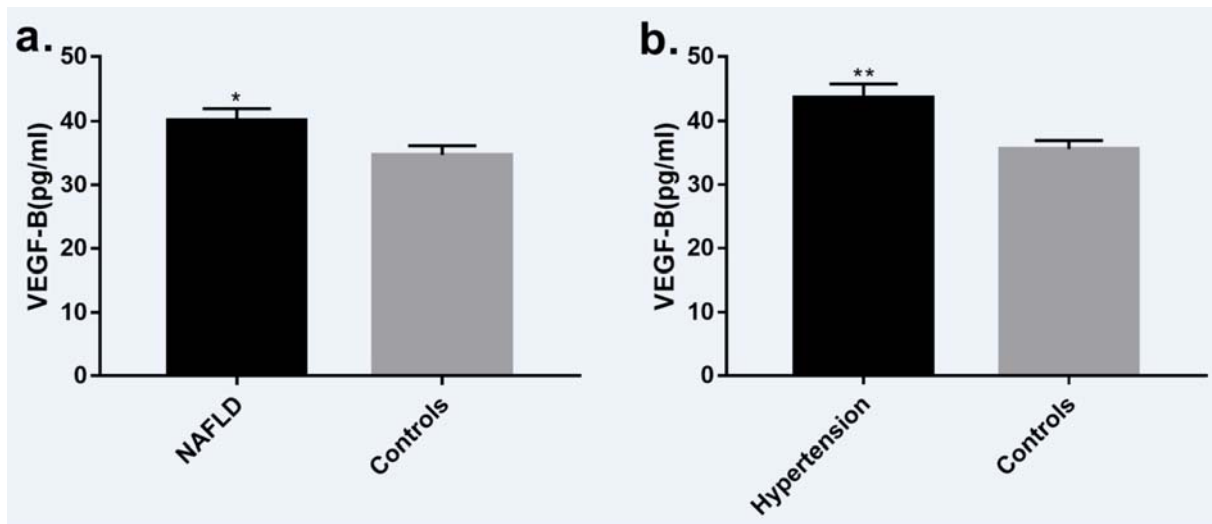


Figure 1: a. Plasma VEGF-B concentration was significantly higher ($P = 0.022$) in subjects with NAFLD ($n=84$) compared with the control group ($n=102$). b. Plasma VEGF-B concentration was significantly higher ($P = 0.003$) in subjects with hypertension ($n=53$) compared with the control group ($n=141$). Data presented as mean \pm SEM. NAFLD: non-alcoholic fatty liver disease

We analyzed the correlation between VEGF-B and the other variables of the two groups by the Spearman correlation coefficient test. Our results revealed a positive correlation between γ -GT ($q = 0.242$; $P = 0.001$), HOMA-IR ($q = 0.174$; $P = 0.019$), SBP, DBP, UA, creatinine (Cr), cystatin C (CysC), TG and VEGF-B. Whereas, we also found a negative correlation between eGFR ($q = -0.185$; $P = 0.01$), HDL-C and VEGF-B. The variables like FPG, HbA1c, BMI, age, ALT, AST, ALP, TC, LDL-C did not show any correlation with VEGF-B level (Supplementary Table 1).

In multivariate regression analysis, we adjusted gender and age to determine the associations between VEGF-B and selected covariates (BMI, liver function, kidney function, HOMA-IR, HbA1c, blood lipids and BP) (Table 2), we found that γ -GT and ALT were independently correlated with VEGF-B ($q = 0.286$; $P = 0.01$; $q = 0.237$; $P = 0.033$, respectively). Besides, we also identified that SBP and DBP were correlated with VEGF-B ($q = 0.345$; $P = 0.002$; $q = 0.284$; $P = 0.01$, respectively), and the eGFR ($q = -0.17$, $P = 0.02$) and other kidney function indexes (UA, Cr, CysC) were also correlated to VEGF-B.

Additionally, our results indicated a significant inverse correlation between VEGF-B and HDL-C ($q = -0.273$; $P = 0.014$).

DISCUSSION

Fatty accumulation within the liver origin from metabolic, and it is one of the main reasons of liver diseases (Pappachan et al., 2014), with a predicted prevalence of approximately 25% in the general population (Satapathy and Sanyal, 2015). NAFLD includes a broad clinical and histological spectrum extending from simple hepatic steatosis to non-alcoholic steatohepatitis, with varying degrees of inflammation and fibrosis, which could potentially lead to cirrhosis. Fatty infiltration in the liver is a critical member of the metabolic disorders, NAFLD individuals is similar to altered body fat distribution individuals, who are at higher risk of cardiovascular disease and diabetes (Kelly et al., 2009; Kotronen and Yki-Jarvinen, 2008; Mariani et al., 2013; Pappachan et al., 2014; Pisto et al., 2014; Sookoian et al., 2011).

Table 2: Age and gender-adjusted multivariate analysis for the association of VEGF-B with selected variables

Variable	β	P-value
SBP	.345	.002
DBP	.284	.010
BMI	.177	.115
UA	.302	.006
BUN	-.035	.755
Cr	.234	.036
CysC	.344	.002
eGFR	-.17	.020
ALT	.237	.033
AST	.179	.109
ALP	.099	.379
γ -GT	.286	.010
TG	.170	.129
TC	.136	.226
HDL-C	-.273	.014
LDL-C	.191	.088
FPG	.089	.431
HbA1C	.042	.706
HOMA-IR	.194	.083

SBP, systolic blood pressure; DBP, diastolic blood pressure; BMI, body mass index; UA, uric acid; BUN, blood urea nitrogen; Cr, Creatinine; CysC, CystatinC; eGFR, estimated glomerular filtration rate; ALT, alanine aminotransferase; AST, aspartate aminotransferase; ALP, alkaline phosphatase, γ -GT, γ -glutamyl transpeptidase; TG, total triglyceride; TC, total cholesterol; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; FPG, fasting plasma glucose; HbA1C, glycosylated hemoglobin; HOMA-IR, homeostatic model assessment for insulin resistance

There are no specific biomarkers of NAFLD. Recently, the essential roles in lipid metabolism, insulin resistance, and adverse influence on diabetes and metabolic disorders have been attributed to VEGF-B (Hagberg et al., 2010, 2012). Mammalian VEGF-B intervenes in maintaining energy balance (Bry et al., 2014; Zafar et al., 2017). By regulating lipid delivery to fat-burning tissues, VEGF-B coordinately increased ectopic lipid deposition, decreased muscle glucose uptake, and caused hyperglycemia (Hagberg et al., 2010, 2012).

This is the first study to attempt to measure the circulating VEGF-B in NAFLD individuals and to figure out if VEGF-B could be

a predictor of NAFLD. The primary finding of our study is that the mean plasma values of VEGF-B in individuals with NAFLD were significantly higher compared with the control cohorts without NAFLD. Also, Pearson's correlation analysis revealed that circulating VEGF-B levels is positively associated with γ -GT. Besides, gender and age-adjustment, multivariate regression analyses showed γ -GT and ALT were independently correlated with VEGF-B. These results demonstrate that VEGF-B plays a significant role in NAFLD. Our findings are identical with the evidence that inhibition of VEGF-B signaling pathways or decreased expression of VEGF-B relieves excess fatty accumulation in the liver, and normalizes glucose levels, and ameliorates dyslipidemia (Hagberg et al., 2012). Furthermore, an animal study suggested that VEGF-B expression was associated with liver cirrhosis (Ujiie et al., 2020). These studies confirmed that VEGF-B has a vital role in the pathogenesis of NAFLD.

Interestingly, the present results displayed positive relations between circulating VEGF-B concentrations and UA, Cr, and CysC, and negative relation between circulating VEGF-B concentrations and eGFR. VEGF-B expresses in human kidneys and is considered to have critical pathological roles in developing diabetic nephropathy (Falkevall et al., 2017; Lagercrantz et al., 1998). It was reported that patients with macro-albuminuria had higher VEGF-B levels than those with non-albuminuria and microalbuminuria in T2DM (Sun et al., 2014). Moreover, clinical evidence showed that diabetic kidney disease (DKD) patients had elevated VEGF-B levels in the kidney (Falkevall et al., 2017). These consequences demonstrated that VEGF-B reverberated the severeness of diabetic nephropathy and that it probably is an advisable biomarker for the discovery of diabetic nephropathy. Falkevall et al. conducted experiments in DKD mice and showed that renal VEGF-B expression is positive, which is related to the severity of the disease. Inhibition of VEGF-B signal transduction in DKD mice attenuates renal lipotoxicity, resensitizes podocytes to

insulin signal transduction, which hinders the progression of DKD-associated pathologies, and leads to the prevention of renal dysfunction (Falkevall et al., 2017). Our study provided evidence that VEGF-B may also play a role in the pathogenic of kidney disease in normoglycemic subjects. However, the mechanism that VEGF-B acts in renal diseases in normoglycemic subjects is unclear. Further studies are warranted to explain the precise part of VEGF-B in normoglycemic subjects and fatty liver patients.

Furthermore, the mean plasma values of VEGF-B in individuals with hypertension were significantly higher compared with the control cohorts without hypertension, and we found a robust correlation between plasma VEGF-B levels and SBP, DBP. The underlying mechanism that VEGF-B effects on hypertension can partly be explained by a most recent animal study (Zhu et al., 2020). In this study, the author revealed that VEGF-B participates in the progression of obesity-associated hypertension in two mouse models of obesity, and they described that inhibition of VEGF-B may have benefit for the treatment of obesity-associated hypertension (Zhu et al., 2020). As most cohorts in our study had a normal BP range, we suggest that plasma VEGF-B might be a considerable biomarker for the early recognition of hypertension.

Although the exact mechanisms associated with the onset of NAFLD remain uncovered, it has been agreed that IR is a major contributor for the progression of NAFLD (Birkenfeld and Shulman, 2014; Lee et al., 1998; Machado and Cortez-Pinto, 2005). In the present study, Pearson's correlation analysis confirmed that VEGF-B has a significant relationship with HOMA-IR ($r=0.174$, $P=0.019$). Similarly, studies in newly diagnosed T2DM and PCOS patients indicated that circulating VEGF-B concentrations were positively correlated with IR (Cheng et al., 2016; Wu et al., 2017). Besides, Sun et al. discovered that in T2DM patients the circulating VEGF-B levels decreased when treated with thiazolidinediones, which can inhibit peroxi-

some proliferators-activated receptor- γ activity and result in insulinresistant decrease (Sun et al., 2014). However, multivariate analysis shows that the relation between VEGF-B and HOMA-IR is insignificant. How VEGF-B effects on insulin resistance in NAFLD subjects remain poorly understood, however, animal studies had provided some hints. The endothelial cell-mediated lipid uptake is regulated by VEGF-B and the downstream signaling pathway (Hagberg et al., 2010). And the excess lipid deposition in tissues like skeletal muscle and liver will cause insulin resistance and disrupts the metabolism of nutrients (Perseghin et al., 1999; Samuel et al., 2010). However, the liver absorbs nutrients through the liver sinusoids, in which the permeability is different from endothelial cells, which may be the reason for our multivariate analysis result. No specific study on the relationship between VEGF-B expression levels and insulin resistance in NAFLD has been reported, so we cannot confirm that the underlying mechanism that VEGF-B triggers insulin resistance in NAFLD is identical to the mechanism that VEGF-B triggers in other insulin resistance disorders.

VEGF-B takes part in the glucose homeostasis in various aspects, including pancreatic b-cells function, insulin secretion, and insulin resistance in T2DM (Hagberg et al., 2012; Ning et al., 2020; Wu et al., 2017). However, our data show that there is no correlation between VEGF-B and HbA1c and FPG. In contrast, some authors have reported that VEGF-B levels are positively correlated with HbA1c in newly diagnosis T2DM patients and PCOS patients (Cheng et al., 2016; Wu et al., 2017). Whether these discrepancies depend on the different patient group or are due to the different sample sizes is not for sure and needs further exploration. It is needed to consider that the subjects involved in the present study were included in the healthcare center, and our population had no particular diseases and had no special treatment.

As an important portion of metabolic syndrome, hyperlipidemia is correlated with VEGF-B, too. An animal study found that

vegfb knockout was related to a decrease in plasma triglyceride and LDL and an increase in HDL levels in diabetic mice (Hagberg et al., 2012). Furthermore, Wu et al. suggested that higher VEGF-B levels were correlated to higher blood TG, which indicated that VEGF-B probably affects blood lipids in humans as well (Wu et al., 2017). In our study, blood TG, HDL-C and LDL-C were significantly different between fatty liver individuals and control individuals, and the correlation analyses indicated that plasma VEGF-B levels were significantly associated with blood TG and HDL-C, and multivariate analysis showed that VEGF-B had an inverse correlation with HDL-C, which is identical with the animal research (Hagberg et al., 2012).

We admit that there are some limitations in this study. First, our study was a cross-sectional study, so we couldn't know the causal relationship between VEGF-B and the evaluated covariates. A prospective study with clinical intervention will be a recommended solution to clarify this point. Second, our study had a relatively small sample size, and it is recommended to confirm our results in larger cohorts. Third, although liver ultrasound is accurate to diagnose NAFLD, it is still a semi-quantitative method. It is reported that the best methods for quantitative measure liver fat content is liver biopsy or magnetic resonance spectroscopy (Dasarathy et al., 2009; Noureddin et al., 2013; Shannon et al., 2011). Fourth, we didn't assess the relation between the circulating VEGF-B and the VEGF-B protein or gene expression in liver tissue.

In conclusion, our study is the first study to assess the plasma VEGF-B in NAFLD individuals and reveals that VEGF-B concentrations are independently associated with NAFLD. In addition, our result indicates that plasma VEGF-B might be a considerable biomarker for the early detection of hypertension and kidney dysfunction in the general public.

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Conflict of interest

The authors declare that they have no conflict of interest.

Authors' contributions

All authors contributed to the study conception and design. Material preparation, data collection and analysis were performed by Xiaofeng Ye, Mohammad Ishraq Zafar, Junchao Zeng, Rui Yang and Wen Kong. The first draft of the manuscript was written by Xiaofeng Ye, Wen Kong and Lu-Lu Chen and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

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REFERENCES

- Aase K, Lymboussaki A, Kaipainen A, Olofsson B, Alitalo K, Eriksson U. Localization of VEGF-B in the mouse embryo suggests a paracrine role of the growth factor in the developing vasculature. *Dev Dyn.* 1999; 215(1):12-25.
- Abedin ZR, Ma Z, Reddy EP. Increased angiogenesis in Cdk4(R24C/R24C):Apc(+/-Min) intestinal tumors. *Cell Cycle.* 2010;9:2456-63.
- Birkenfeld AL, Shulman GI. Nonalcoholic fatty liver disease, hepatic insulin resistance, and type 2 diabetes. *Hepatology.* 2014;59:713-23.
- Bry M, Kivela R, Leppanen VM, Alitalo K. Vascular endothelial growth factor-B in physiology and disease. *Physiol Rev.* 2014;94:779-94.

- Chen Y, Zhao M, Wang C, Wen H, Zhang Y, Lu M, et al. Adipose vascular endothelial growth factor B is a major regulator of energy metabolism. *J Endocrinol.* 2020;244:511-21.
- Cheng F, Zhao L, Wu Y, Huang T, Yang G, Zhang Z, et al. Serum vascular endothelial growth factor B is elevated in women with polycystic ovary syndrome and can be decreased with metformin treatment. *Clin Endocrinol.* 2016;84:386-93.
- Dasarathy S, Dasarathy J, Khiyami A, Joseph R, Lopez R, McCullough AJ. Validity of real time ultrasound in the diagnosis of hepatic steatosis: a prospective study. *J Hepatol.* 2009;51(6):1061-7.
- Eckel RH, Grundy SM, Zimmet PZ. The metabolic syndrome. *Lancet.* 2005;365(9468):1415-28.
- Falk T, Zhang S, Sherman SJ. Vascular endothelial growth factor B (VEGF-B) is up-regulated and exogenous VEGF-B is neuroprotective in a culture model of Parkinson's disease. *Mol Neurodegen.* 2009;4:49.
- Falkevall A, Mehlem A, Palombo I, Heller Sahlgren B, Ebarasi L, He L, et al. Reducing VEGF-B Signaling Ameliorates Renal Lipotoxicity and Protects against Diabetic Kidney Disease. *Cell Metab.* 2017;25:713-26.
- Farrell GC, Chitturi S, Lau GK, Sollano JD. Guidelines for the assessment and management of non-alcoholic fatty liver disease in the Asia-Pacific region: executive summary. *J Gastroenterol Hepatol* 2007;22:775-7.
- Gomez-Ambrosi J, Catalan V, Rodriguez A, Ramirez B, Silva C, Gil MJ, et al. Involvement of serum vascular endothelial growth factor family members in the development of obesity in mice and humans. *J Nutr Biochem.* 2010;21:774-80.
- Hagberg CE, Falkevall A, Wang X, Larsson E, Huusko J, Nilsson I, et al. Vascular endothelial growth factor B controls endothelial fatty acid uptake. *Nature.* 2010;464(7290):917-21.
- Hagberg CE, Mehlem A, Falkevall A, Muhl L, Fam BC, Ortsater H, et al. Targeting VEGF-B as a novel treatment for insulin resistance and type 2 diabetes. *Nature.* 2012;490(7420):426-30.
- Holmes DI, Zachary I. The vascular endothelial growth factor (VEGF) family: angiogenic factors in health and disease. *Genome Biol.* 2005;6(2):209.
- Karpanen T, Bry M, Ollila HM, Seppanen-Laakso T, Liimatta E, Leskinen H, et al. Overexpression of vascular endothelial growth factor-B in mouse heart alters cardiac lipid metabolism and induces myocardial hypertrophy. *Circ Res.* 2008;103:1018-26.
- Kelly TL, Wilson KE, Heymsfield SB. Dual energy X-ray absorptiometry body composition reference values from NHANES. *PloS One.* 2009;4(9):e7038.
- Kitade H, Chen G, Ni Y, Ota T. Nonalcoholic fatty liver disease and insulin resistance: New insights and potential new treatments. *Nutrients.* 2017;9(4):387.
- Kotronen A, Yki-Jarvinen H. Fatty liver: a novel component of the metabolic syndrome. *Arterioscler Thromb Vasc Biol.* 2008;28(1):27-38.
- Lagercrantz J, Farnebo F, Larsson C, Tvrdik T, Weber G, Piehl F. A comparative study of the expression patterns for vegf, vegf-b/vrf and vegf-c in the developing and adult mouse. *Biochim Biophys Acta.* 1998;1398:157-63.
- Lee JH, Rhee PL, Lee JK, Lee KT, Kim JJ, Koh KC, et al. Role of hyperinsulinemia and glucose intolerance in the pathogenesis of nonalcoholic fatty liver in patients with normal body weight. *Korean J Intern Med.* 1998;13:12-4.
- Li X, Aase K, Li H, von Euler G, Eriksson U. Isoform-specific expression of VEGF-B in normal tissues and tumors. *Growth Factors.* 2001;19(1):49-59.
- Li X, Kumar A, Zhang F, Lee C, Tang Z. Complicated life, complicated VEGF-B. *Trends Mol Med.* 2012;18:119-27.
- Machado M, Cortez-Pinto H. Non-alcoholic fatty liver disease and insulin resistance. *Eur J Gastroenterol Hepatol.* 2005;17:823-6.
- Macut D, Tziomalos K, Bozic-Antic I, Bjekic-Macut J, Katsikis I, Papadakis E, et al. Non-alcoholic fatty liver disease is associated with insulin resistance and lipid accumulation product in women with polycystic ovary syndrome. *Hum Reprod.* 2016;31:1347-53.
- Mariani S, Fiore D, Barbaro G, Basciani S, Saponara M, D'Arcangelo E, et al. Association of epicardial fat thickness with the severity of obstructive sleep apnea in obese patients. *Int J Cardiol.* 2013;167:2244-9.
- Ning FC, Jensen N, Mi J, Lindstrom W, Balan M, Muhl L, et al. VEGF-B ablation in pancreatic beta-cells up-regulates insulin expression without affecting glucose homeostasis or islet lipid uptake. *Sci Rep.* 2020;10(1):923.
- Nouredin M, Lam J, Peterson MR, Middleton M, Hamilton G, Le TA, et al. Utility of magnetic resonance imaging versus histology for quantifying changes in liver fat in nonalcoholic fatty liver disease trials. *Hepatology.* 2013;58:1930-40.

- Olofsson B, Pajusola K, von Euler G, Chilov D, Alitalo K, Eriksson U. Genomic organization of the mouse and human genes for vascular endothelial growth factor B (VEGF-B) and characterization of a second splice isoform. *J Biol Chem*. 1996;271:19310-7.
- Olsson AK, Dimberg A, Kreuger J, Claesson-Welsh L. VEGF receptor signalling - in control of vascular function. *Nat Rev Mol Cell Biol*. 2006;7:359-71.
- Pappachan JM, Antonio FA, Edavalath M, Mukherjee A. Non-alcoholic fatty liver disease: a diabetologist's perspective. *Endocrine*. 2014;45:344-53.
- Perseghin G, Scifo P, De Cobelli F, Pagliato E, Battezzati A, Arcelloni C, et al. Intramyocellular triglyceride content is a determinant of in vivo insulin resistance in humans: a 1H-13C nuclear magnetic resonance spectroscopy assessment in offspring of type 2 diabetic parents. *Diabetes*. 1999;48:1600-6.
- Pisto P, Santaniemi M, Bloigu R, Ukkola O, Kesaniemi YA. Fatty liver predicts the risk for cardiovascular events in middle-aged population: a population-based cohort study. *BMJ Open*. 2014;4(3):e004973.
- Samuel VT, Petersen KF, Shulman GI. Lipid-induced insulin resistance: unravelling the mechanism. *Lancet*. 2010;375(9733):2267-77.
- Satapathy SK, Sanyal AJ. Epidemiology and natural history of nonalcoholic fatty liver disease. *Semin Liver Dis*. 2015;35:221-35.
- Shannon A, Alkhouri N, Carter-Kent C, Monti L, Devito R, Lopez R, et al. Ultrasonographic quantitative estimation of hepatic steatosis in children with NAFLD. *J Pediatr Gastroenterol Nutr*. 2011;53:190-5.
- Sookoian S, Gianotti TF, Rosselli MS, Burgueno AL, Castano GO, Pirola CJ. Liver transcriptional profile of atherosclerosis-related genes in human nonalcoholic fatty liver disease. *Atherosclerosis*. 2011;218:378-85.
- Sun CY, Lee CC, Hsieh MF, Chen CH, Chou KM. Clinical association of circulating VEGF-B levels with hyperlipidemia and target organ damage in type 2 diabetic patients. *J Biol Regul Homeost Agents*. 2014;28:225-36.
- Ujiie N, Nakano T, Yamada M, Sato C, Nakanishi C, Fujishima F, et al. Low-energy extracorporeal shock wave therapy for a model of liver cirrhosis ameliorates liver fibrosis and liver function. *Sci Rep*. 2020;10(1):2405.
- Wu J, Wei H, Qu H, Feng Z, Long J, Ge Q, et al. Plasma vascular endothelial growth factor B levels are increased in patients with newly diagnosed type 2 diabetes mellitus and associated with the first phase of glucose-stimulated insulin secretion function of beta-cell. *J Endocrinol Invest*. 2017;40:1219-26.
- Zafar MI, Zheng J, Kong W, Ye X, Gou L, Regmi A, et al. The role of vascular endothelial growth factor-B in metabolic homeostasis: current evidence. *Biosci Rep*. 2017;37(4):BSR20171089.
- Zafar MI, Mills K, Ye X, Blakely B, Min J, Kong W, et al. Association between the expression of vascular endothelial growth factors and metabolic syndrome or its components: a systematic review and meta-analysis. *Diabetol Metab Syndr*. 2018;10:62.
- Zhong X, Huang H, Shen J, Zaccogna S, Zentilin L, Giacca M, et al. Vascular endothelial growth factor-B gene transfer exacerbates retinal and choroidal neovascularization and vasopermeability without promoting inflammation. *Mol Vis*. 2011;17:492-507.
- Zhu X, Wang Y, Zhu L, Zhu Y, Zhang K, Wang L, et al. SR-A1 prevents obesity-associated blood pressure elevation through suppressing overproduction of VEGF-B in Macrophages. *Cardiovasc Res*. 2020; online ahead of print.