#### Check for updates

#### **OPEN ACCESS**

EDITED BY Harry D. Dawson, Agricultural Research Service, (USDA), United States

REVIEWED BY Fu Li, Chengdu Institute of Biology, (CAS), China Allen D. Smith, Agricultural Research Service, (USDA), United States

\*CORRESPONDENCE Liming Mao Immao@ntu.edu.cn Xiaorong Zhou zhouxiaorong@ntu.edu.cn Shengping Hou sphou&228@163.com

SPECIALTY SECTION This article was submitted to Mucosal Immunity, a section of the journal Frontiers in Immunology

RECEIVED 07 June 2022 ACCEPTED 09 August 2022 PUBLISHED 24 August 2022

#### CITATION

Xu Q, Sun W, Zhang J, Mei Y, Bao J, Hou S, Zhou X and Mao L (2022) Inflammasome-targeting natural compounds in inflammatory bowel disease: Mechanisms and therapeutic potential. *Front. Immunol.* 13:963291. doi: 10.3389/fimmu.2022.963291

#### COPYRIGHT

© 2022 Xu, Sun, Zhang, Mei, Bao, Hou, Zhou and Mao. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Inflammasome-targeting natural compounds in inflammatory bowel disease: Mechanisms and therapeutic potential

Qiuyun Xu<sup>1</sup>, Weichen Sun<sup>1</sup>, Jie Zhang<sup>1</sup>, Youmin Mei<sup>2</sup>, Jingyin Bao<sup>3</sup>, Shengping Hou<sup>4\*</sup>, Xiaorong Zhou<sup>1\*</sup> and Liming Mao<sup>1,3\*</sup>

<sup>1</sup>Department of Immunology, School of Medicine, Nantong University, Nantong, China, <sup>2</sup>Department of Periodontology, Nantong Stomatological Hospital, Nantong, China, <sup>3</sup>Basic Medical Research Center, School of Medicine, Nantong University, Nantong, China, <sup>4</sup>The First Affiliated Hospital of Chongqing Medical University, Chongqing Eye Institute, Chongqing Key Laboratory of Ophthalmology, Chongqing, China

Inflammatory bowel disease (IBD), mainly including Crohn's disease and ulcerative colitis, seriously affects human health and causes substantial social and economic burden. The pathogenesis of IBD is still not fully elucidated, whereas recent studies have demonstrated that its development is associated with the dysfunction of intestinal immune system. Accumulating evidence have proven that inflammasomes such as NLRP3 and NLRP6 play a prominent role in the pathogenesis of IBD. Thus, regulating the activation of inflammasomes have been considered to be a promising strategy in IBD treatment. A number of recent studies have provided evidence that blocking inflammasome related cytokine IL-1 $\beta$  can benefit a group of IBD patients with overactivation of NLRP3 inflammasome. However, therapies for targeting inflammasomes with high efficacy and safety are rare. Traditional medical practice provides numerous medical compounds that may have a role in treatment of various human diseases including IBD. Recent studies demonstrated that numerous medicinal herb derived compounds can efficiently prevent colon inflammation in animal models by targeting inflammasomes. Herein, we summarize the main findings of these studies focusing on the effects of traditional medicine derived compounds on colitis treatment and the underlying mechanisms in regulating the inflammasomes. On this basis, we provide a perspective for future studies regarding strategies to improve the efficacy, specificity and safety of available herbal compounds, and to discover new compounds using the emerging new technologies, which will improve our understanding about the roles and mechanisms of herbal compounds in the regulation of inflammasomes and treatment of IBD.

#### KEYWORDS

traditional medicine, inflammatory bowel disease, NLRP3, inflammasome, IL-1 $\beta$ , natural compounds

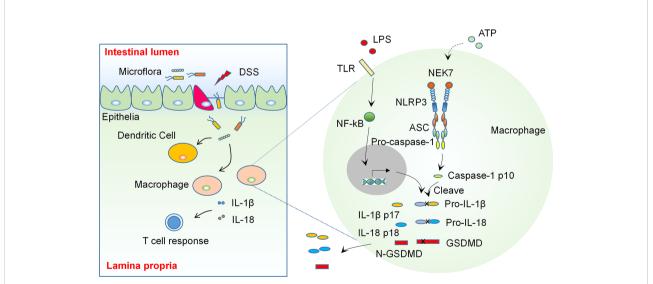
## Introduction

Inflammatory bowel disease (IBD), mainly including Crohn's disease and ulcerative colitis, is a term to describe a group of chronic relapsing inflammatory disorders occurred in the gastrointestinal (GI) tract (1-3). These diseases are associated with high risks to develop colorectal cancer, thus extremely affect people's health and cause serious social and economic burden (4-6). Recent advances have shown that the development of IBD is owing to an aberrant and persistent immune responses to the commensal microflora in the intestine and colon of genetically susceptible individuals (3), whereas the exact pathogenesis of these diseases is still not fully elucidated. The entry of luminal organisms or their products to the lamina propria can be sensed by the tissue resident immune cells such as macrophages and dendritic cells, which process and present antigens to facilitate the activation of adaptive immunity. Meanwhile, the activation of these cells may produce various inflammatory mediators, such as pro-inflammatory cytokines and chemokines, contributing to the amplification of immune responses in the lamina propria of the GI tract.

As prominent mediators of inflammatory response, the inflammasomes are considered to be critical regulators in the development of IBD. Recent studies have demonstrated that the activation of inflammasomes play an essential role in the pathogenesis of IBD (7). The aberrant activation of inflammasomes and the resultant production of proinflammatory cytokines in colonic macrophages may lead to an imbalanced inflammatory response and cause tissue damage in the colon, which may trigger the onset of IBD (see Diagram in Figure 1). Therefore, targeting inflammasomes may be a promising strategy for treatment of IBD. A growing body of studies have shown that targeting IL-1 $\beta$ , a downstream cytokine activated by inflammasomes, using neutralizing antibodies is effective in ameliorating colonic inflammation in both animal models of colitis and IBD patients (8), whereas the high cost of antibody based therapy may limit its application (9). Some emerging natural compounds derived from traditional medicine may provide new options for IBD intervention via modulating inflammasomes and have been shown to be effective in an increasing number of studies (10-13). This review aims at summarizing these studies regarding the role of ingredients or natural compounds from traditional medicinal herbs in colitis and related mechanisms, and providing an outlook for future studies on investigation of IBD targeting therapies.

# Inflammasomes are critical regulators of IBD

Inflammasomes are a group of large molecular weight multiprotein complexes formed by a sensor protein of the



#### FIGURE 1

The NLRP3 Inflammasome is an Essential Regulator in the Pathogenesis of Inflammatory Bowel Disease. Some environmental factors cause epithelial damage, which allows the entry of luminal organisms or their products to the lamina propria, the latter then induces activation of inflammasomes and production of pro-inflammatory cytokines such as  $L-1\beta$  and L-18 in colon macrophages. These cytokines regulate the adaptive immunity and cause an imbalanced inflammatory response and tissue damage in genetically susceptible individuals (Left). The activation of NLRP3 inflammasome needs two signals: A stimulation to TLRs or TNF receptors induces transcription and appropriate post-translational modification of NLRP3 inflammasome components (signal 1). Some extracellular stimulants such as ATP induce inflammasome assembly (signal 2) and cause autocleavage of caspase–1, which then catalyzes the maturation of precursor of  $L-1\beta$ , L-18 and GSDMD, the latter then regulate the adaptive immunity and cause proptosis.

NOD-like receptor family or HIN-200 family, the adaptor protein ASC and the effector caspase-1 in response to stimulation of microbial or damage associated molecular patterns (14, 15). Upon activation, inflammasome formation allows autocatalytic cleavage of caspase-1, which then processes precursors of IL-1 $\beta$  and IL-18 to allow the release of mature cytokines from cells, which may regulate Th17 and Th1 cells, respectively, and amplify the immune responses in the tissue. Meanwhile, the activation of caspase-1 induces a form of programmed cell death, pyroptosis (16–18), which regulates the progression of a variety of inflammatory diseases, including IBD.

To date, a number of PRR sensor proteins, including NLRP3, NLRP1, NLRC4, AIM2 and Pyrin, have been confirmed to form inflammasomes and regulate the activation of caspase-1 by many studies. Moreover, many other PRR sensors, mainly including NLRP2, NLRP6, NLRP7, NLRP9 (19, 20), NLRP12 and IFI16, have also been reported to have inflammasome-forming ability, although these findings need to be further confirmed by other researchers.

NLRP3 is one of the most studied NOD-like receptors that can form an inflammasome. The activation of NLRP3 needs two kinds of stimuli (see Diagram in Figure 1): The first one may derive from extracellular or intracellular receptors, such as TLRs or TNF receptors. This stimulus triggers the transcription of inflammasome associated genes and also induces appropriate post-translational modifications of the translated proteins, which facilitate the activation of inflammasome and subsequent cleavage of cytokine precursors. The second stimulus may come from the formation of mitochondrial reactive oxygen species (ROS), K+ efflux, membrane perturbations, or extracellular ATP, which induces oligomerization of NLRP3 and triggers assembly of the inflammasome (18, 21).

A number of recent reports demonstrate that the overactivation of NLRP3 inflammasome caused by various genetic abnormalities lead to development of colitis (22–24), whereas some other studies provide evidence that deficiency of NLRP3 and inflammasome related genes may induce more severe colitis in mice (25). Thus, the NLRP3 inflammasome may play a role in regulating a delicate immune balance in the colon, loss or overactivation of NLRP3 may both break down the balance and thus lead to the onset of colonic inflammation (14).

Regulation of NLRP3 inflammasome activation or its downstream cytokines may be a promising strategy for treating the IBD. A number of studies have shown that blockade of IL-1 $\beta$  or IL-1R signaling using antibodies can effectively ameliorate colon inflammation in Crohn's disease and mouse model of colitis (7). However, antibody-based therapies are challenged by their high cost due to the high biologic dosage used in treatment and maintenance of diseases, which lead to a high economic burden to the patients (9). Moreover, the side effects of antibody-based therapies in treatment of IBD, such as serious infection and malignancy after administration of anti-TNF antibody, has long been observed and are still significant concerns of current IBD clinicians, which may limit the application of this type of therapies (26, 27). Thus, other alternative therapeutic strategies need to be developed for the intervention of IBD.

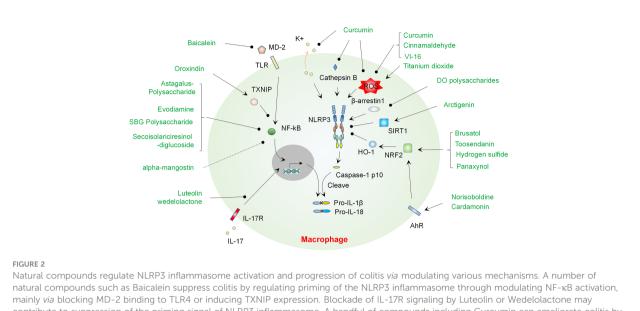
Previous studies have reported a number of small molecule inhibitors of NLRP3 inflammasome, such as MCC950 and CY-09 (28, 29), while their roles in IBD have not been evaluated. Studies in recent literature identified numerous new compounds derived from natural herbs may have a role in suppressing colitis in animal models through inhibiting inflammasomes. Here, we summarized the main findings of these studies and discussed the roles of these compounds in IBD models and the underlying mechanisms in inhibiting inflammasomes. This work may contribute to further mechanistical studies of these inhibitors in suppressing the activation of inflammasomes and may also be of benefit to future translational investigations.

## Traditional medicine derived compounds affect IBD *via* modulating the NLRP3 inflammasome

Traditional medicine has long been used globally to treat a variety of human diseases, including infectious diseases, autoimmune diseases, cancers and IBD (30–34). Although the effects and exact mechanisms of traditional medicine are not yet fully validated and clearly addressed, it provides large numbers of compounds that may play a role in intervention of IBD. A variety of bioactive components derived from medicinal herbs have been found to play an inhibitory role in colitis *via* multiple mechanisms, and a few of them can prevent colitis at least partially by regulating the activation of inflammasomes *via* various mechanisms such as regulating NF- $\kappa$ B and ROS. Herein, we give an overview of these studies, discuss the cellular targets of compounds and their mechanisms, and provide an outlook for future studies (Summarized in Figure 2 and Table 1).

#### NF-κB

NF-κB is a transcription factor that plays a prominent role in diverse physiological processes including inflammatory responses. The activation of NF-κB promotes the production of pro-inflammatory cytokines in innate cells and regulates the activation and differentiation of lymphocytes. In the process of inflammasome activation, NF-κB provides the priming signal and promotes the transcription of inflammasome components, such as NLRP3 and Pro-IL-1β.



mainly via blocking MD-2 binding to TLR4 or inducing TXNIP expression. Blockade of IL-17R signaling by Luteolin or Wedelolactone may contribute to suppression of the priming signal of NLRP3 inflammasome. A handful of compounds including Curcumin can ameliorate colitis by suppressing the NLRP3 inflammasome via regulating ROS production, K+ efflux or cathepsin B release. Polysaccharides from DO may suppress  $\beta$ -arrestin-1 and thus inhibit the activation of NLRP3 inflammasome. A number of compounds such as Norisoboldine can promote activation of AhR and then activate NRF2 or SIRT1 signaling, which contribute the suppression of the NLRP3 inflammasome and amelioration of colitis. DO, *Dendrobium officinale*.

Recent studies found that a number of herbal compounds can affect the priming of inflammasome via regulating the activation of NF-KB. A good example for this type of compounds is oroxindin, a bioactive extract with antiinflammatory activity derived from Astragalus membranaceus (AM), which is a medicinal plant wildly used in Chinese medicine for treatment of many human diseases (53, 54). Liu et al (11) found that oroxindin can suppress the activation of NF-kB signaling via inducing the expression of thioredoxininteracting protein (TXNIP), an inhibitory regulator of NF-kB pathway (55). Considering the previously reported role of TXNIP in interacting and activating NLRP3 (56), this regulatory molecule may regulate inflammasome activation in both priming and activating steps. This may also explain the observation in Liu et al. study showing that the formation of NLRP3 inflammasome was also suppressed by oroxindin. The authors used a colitis model induced by dextran sulfate sodium (DSS), a sulfated polysaccharide that is toxic to colon epithelium and causes epithelial cell injury, the entry of luminal organisms or their products induce inflammatory responses in the lamina propria (57, 58). Their data showed that oroxindin can suppress macrophage infiltration to the colon and attenuated colon pathological changes. On this basis, the role of oroxindin in suppressing NLRP3 inflammasome function may mainly dependent on its inhibition on TXNIP associated NF-KB signaling. It should be noted that another compound extracted from AM, Astagalus Polysaccharide (AP), also showed antiinflammatory effects (59, 60) and anti-colitis role via regulating the NLRP3 inflammasome. Its impact on inflammasome may be mediated by inhibiting the NF- $\kappa$ B pathway, since treatment with AP suppressed the expression of NLRP3, ASC, caspase-1, IL-18, and IL-1 $\beta$  (35). Interestingly, polysaccharide from *Scutellaria Baicalensis Georgi* (SBG) (37) also has anti-NF- $\kappa$ B effects, through which it inhibits NLRP3 inflammasome activation and mitigates DSS induced colitis.

Another compound that may play anti-colitis role through regulating NF- $\kappa$ B and the NLRP3 inflammasome is Evodiamine (EVO), a bioactive component with anti-inflammatory role (61) derived from *Evodia Rutaecarpa* (ER), which is a herbal plant that has been used clinically in treating some human inflammatory diseases, especially for headache and abdominal pain (62). To study the role of EVO in colitis, Shen et al (36) used murine DSS colitis model and found that treatment of EVO ameliorated DSS induced colonic symptoms, inhibited the production of pro-inflammatory cytokines such as TNF- $\alpha$ , IL-1 $\beta$  and IL-6, and restrain the activation of NLRP3 inflammasome. Regarding the mechanisms, they found that EVO can suppress activation of NF- $\kappa$ B and thus suppress the expression of inflammasome components.

#### $\beta$ -arrestin1

 $\beta$ -arrestin1 is a ubiquitously expressed protein that is identified initially as an inhibitor and adaptor of the G protein-coupled receptors with the purpose of regulating many

Bioactive Compound	Source	Target	Mechanism	Disease model	Impact to host	Toxicity Data	References
Astagalus Polysaccharide	Astragalus membranaceus	NF-ĸB	Inhibit NF-ĸB activation and expression of inflammasome components	DSS- colitis	Reduce DAI and histological injury scores	Not available	Tian et al. (35)
Oroxindin	Astragalus membranaceus	TXNIP, NF-κB	Promote expression of TXNIP and suppress NF-κB activation	DSS- colitis	Suppress macrophages infiltration and attenuate pathological changes in colonic tissue	Not available	Liu et al. (11)
Evodiamine	Evodia rutaecarpa	NF-κB	Inhibit NF-κB activation	DSS- colitis	Ameliorate mice body weight loss, DAI, colon length shortening, colonic pathological damage	Not available	Shen et al. (36)
SBG Polysaccharide	Scutellaria Baicalensis Georgi	NF-ĸB	Inhibit activation of NF-kB	DSS- colitis	Decrease DAI, colonic pathological damage, and reduce MPO activity	No observed toxicity (500 mg/ ml, 0.4 ml/10 g p.o., 7 days in mouse)	Cui et al. (37)
DO polysaccharides	Dendrobium officinale (DO)	$\beta$ -arrestin1	Inhibit β-arrestin1 expression	DSS-colitis	Decrease mortality, alleviate colonic pathological damage	No observed toxicity (200 mg/ kg/day p.o. for 7 days in mouse)	Liang et al. (10)
Arctigenin	Fructus Arctii (Great Burdock Achene)	SIRT1	Suppress SIRT1 activation	DSS- colitis	Suppressed colon inflammation	Not available	Pu et al. (38)
Baicalein	Scutellaria baicalensis Georgi	MD-2	Suppress MD-2 binding to TLR4 and block TLR4/ MyD88 Signaling	TNBS colitis	Decrease the activity of MPO and the expression of pro-inflammatory mediators.	Not toxic to RAW264.7 cells (200µM within 48h)	Luo et al. (12)
Cinnamaldehyde	Cinnamon	ROS	Prevent the production of ROS	DSS- colitis	Reduce loss of body weight, DAI, colon shortening and infiltration of inflammatory cells	Not toxic to RAW264.7 cells (100µM within 24h)	Qu et al. (39)
Flavonoid VI-16	Fruit and vegetable	mitochondria ROS	Reduce the mitochondrial ROS	DSS- colitis	Reduce colitis severity	Not available	Zhao et al. (40)
Curcumin	Curcuma longa species	K+ efflux, ROS and cathepsin B	Suppress K+ efflux, intracellular ROS formation and cathepsin B release	DSS- colitis	Suppress the colitis severity	Not available	Gong et al. (41)
Titanium dioxide	Food additive	ROS	Induce ROS production	DSS- colitis	Increase the severity of colitis	Not available	Ruiz et al. (42)
Luteolin and wedelolactone	Wedelia chinensis	IL-17 signaling pathway	Suppress genes in IL-17 signaling pathway	DSS- colitis	Suppress the colitis severity	Not available	Lin et al. (43)
Brusatol	Brucea javanica	NRF2	Activate NRF2	TNBS colitis	Attenuate diarrhea, colonic shortening, macroscopic damage and histological injury	Not toxic to RAW264.7 cells (200nM within 24h)	Zhou et al. (44)
Toosendanin	Melia toosendan Sieb et Zucc	NRF2/HO-1	Upregulate NRF2/ HO-1 expression	DSS- colitis	Ameliaorate DAI, shortened colon length, pathological damage of the colon tissues	Not available	Fan et al. (45)
Hydrogen sulfide	Diet	NRF2	Upregulate NRF2 and Reduce ROS generation	DSS- colitis	Attenuate colitis severity, reduce colon shortening and colonic pathological damages	Not available	Qin et al. (46)
Norisoboldine	Radix Linderae	AhR	Activate AhR, elevated NRF2 and reduce level of ROS	TNBS colitis	Alleviate colitis related symptoms	Not toxic to THP- 1 cells (30µM within 24h)	Lv et al. (13)

#### TABLE 1 Summary of Natural Compounds that Regulates Inflammasomes in Colitis Models.

(Continued)

#### TABLE 1 Continued

Bioactive Compound	Source	Target	Mechanism	Disease model	Impact to host	Toxicity Data	References
Cardamonin	Cardamom	AhR	Activate AhR/ NRF2/NQO1 pathway	DSS- colitis, TNBS- colitis	Reduce colitis severity	Not toxic to THP- 1 and BMDM cells (100µM within 24h)	Wang et al. (47)
Palmatine	A number of herbs	PINK1/ Parkin	Enhance the expression of PINK1 and parkin	DSS- colitis	Attenuate body weight loss and colon shortening, reduced DAI and histopathologic score	No observed toxicity (521 mg/ kg/day, p.o.,90 days in mouse)	Mai et al. (48)
alpha-mangostin	A number of herbs	Not clear	Reduce NLRP3, caspase 1, IL-18, and IL-1β expression	LPS- colitis	Reduce severity of intestinal villi detachment, reduce congestion and hemorrhage, reduce epithelial cell nuclei deformation and the mitochondria swelling.	Not available	Yin et al. (49)
Secoisolariciresinol diglucoside	Lignans	NF-kB	Disruption of NF- kB activation and suppression of NLRP1 inflammasome	DSS- colitis	Attenuate the severity of colon inflammation and macrophage infiltration to the colon	Not toxic to RAW264.7 cells (25µM within 24h)	Wang et al. (50)
Apigenin	Plant and fruit	NLRP6	Reshape gut microbiota	DSS- colitis	Protect mice from colon damage	Not available	Radulovic et al. (51)
Panaxynol	Ginseng	NRF2	Activate NRF2 and reduce ROS	DSS- colitis	Reduce colitis severity	No observed toxicity (1mg/kg/ day, p.o., 7 days in mouse)	Chaparala et al. (52)

DAI, Disease activity index; AhR, Aryl hydrocarbon receptor

immunological pathways and cell death (63, 64). It can also bind with a number of signaling molecules in MAPK pathways and thus regulate their activities in macrophages (64). Regarding its role in inflammasome activation,  $\beta$ -arrestin1 is required for full activation of NLRP3 and NLRC4 inflammasomes (65). Regulating  $\beta$ -arrestin1 may affect the development of colitis, in Liang et al.'s study (10), the authors studied the impact of polysaccharides extracts from *Dendrobium officinale* (DO) on colonic inflammation induced by DSS. DO is a widely used herb in traditional Chinese medical practice to treat gastrointestinal disorders. The data showed that polysaccharides from DO (DOPS) can inhibit  $\beta$ -arrestin1 expression and suppress activation of the NLRP3 inflammasome in both *in vivo* and *in vitro* experiments.

#### Sirtuin 1 (SIRT1)

SIRT1 is a transcription factor that regulates various biological pathways. It can function as a histone deacetylase in the presence of nicotinamide adenine dinucleotide (NAD) (66). SIRT1 can suppress the production of pro-inflammatory cytokines by regulating deacetylation of NF- $\kappa$ B p65, thus it also affects the priming of NLRP3 inflammasome components. Moreover, SIRT1 may inhibit NLRP3 inflammasome activation by decreasing expression of CD40/CD40L (67) or suppressing oxidative stress (68). A recent study found that (38) SIRT1 may be inhibited by Arctigenin, one of the major bioactive components of *Fructus Arctii* (Great Burdock Achene), which is a herbal plant widely used in traditional Chinese medicine. It has been reported that Arctigenin has extensive pharmacological effects in a variety of diseases such as diabetes (69). Regarding its impact on colitis, Pu et al (38) found that Arctigenin can mitigate DSS induced colon inflammation *via* suppressing NLRP3 inflammasome. This effect was mediated by SIRT1, since knock-downing of SIRT1 decreased anti-inflammasome activity of Arctigenin.

#### Myeloid differentiation protein-2 (MD-2)

MD-2 is a critical protein that assists the TLR4 receptor in sensing LPS (70). Thus, it may regulate the priming step of NLRP3 inflammasome activation. A study by Luo et al (12) found that Baicalein (5,6,7-trihydroxyflavone), a bioactive ingredient isolated from the root of SBG with potent anti-inflammatory and anti-cancer effects *via* multiple mechanisms (71, 72), may regulate the progression of colitis by targeting MD-2. The researchers employed a mouse model of colitis induced by intrarectally administration of 2,4,6-trinitrobenzene sulfonic acid (TNBS), a haptenic agent that renders colonic proteins immunogenic to the host immune system and drives an

10.3389/fimmu.2022.963291

inflammatory response in the lamina propria. Baicalein can alleviate colitis severity induced by TNBS by inhibiting the activation of NLRP3 inflammasome and production of IL-1 $\beta$ in the colon. Further studies showed that the effect of baicalein may be associated with its role in blocking MD-2 binding with TLR4 and thus suppressing down-stream pathways, such as NFkB and p38 MAPK. Thus, the impact of baicalein on the activation of the NLRP3 may be through regulating the priming signal, whereas if baicalein affects the activating signal of NLRP3 inflammasome need to be studied in future.

# K<sup>+</sup> efflux, ROS production and cathepsin B release

 $K^+$  efflux, ROS production and cathepsin B release are the three major upstream signals that stimulate the activation step of the NLRP3 inflammasome formation. ROS is a term to describe a group of unstable molecules that contains oxygen and easily react with other molecules. The accumulation of ROS in cells may result in damage of DNA, RNA and protein, and ultimately cause cell death. The aberrant production of ROS is associated with pathogenesis of diverse inflammatory diseases and cancers.

Regulating these signals may affect inflammasome activation and colitis. For instance, Gong et al. (41) showed that curcumin, the principal curcuminoid of Curcuma longa species with antioxidative activity (73), can suppress  $K^+$  efflux, intracellular ROS formation and cathepsin B release, and thus inhibit NLRP3 inflammasome activation. Regarding the role of curcumin in development of colitis, it can attenuate DSS induced colitis symptoms. In addition, some clinical trials have demonstrated that curcumin is efficient and safe chemical for management of IBD (74).

Regulating the level of ROS may contribute to the progression of colitis. Qu et al (39) found that Cinnamaldehyde (CA) can inhibit colitis development by suppressing the NLRP3 inflammasome via regulating ROS. CA is a major bioactive ingredient with impressive antibacterial and anti-inflammatory activity derived from cinnamon (75, 76). The researchers showed that CA was effective in preventing DSS induced colitis associated conditions, it prevented activation of the NLRP3 inflammasome and production of pro-inflammatory cytokines in the colon. For the mechanisms, the authors found that CA can decrease the production of ROS and additionally suppress the phosphorylation of AKT, mTOR and COX2 in macrophages. Previous reports have demonstrated that ROS is an important regulator of the NLRP3 inflammasome (77-79), thus the anti-colitis and antiinflammasome effects of CA may exert via down-regulating ROS. Another example on this direction is provided by the study of Zhao et al. (40). They studied the role of VI-16, a synthetic flavonoid compound, in colitis. Flavonoids are a group of bioactive polyphenols abundant in fruits and vegetables (80, 81). Previous studies have reported that a number of flavonoids affect the activation of inflammasome (82), but the mechanism is not fully clarified. Using DSS induced colitis model, Zhao et al. (40) showed that VI-16 administration protected mice from colon inflammation, and this protection was dependent on NLRP3 expression in hematopoietic cells. Regarding the mechanisms, VI-16 was able to reduce the level of mitochondrial ROS and thus block TXNIP-NLRP3 interaction.

An opposite example regarding the role of ROS in colitis is provided by the study of Ruiz et al (42), they studied the possible role of Titanium dioxide (TiO<sub>2</sub>) in colonic inflammation. TiO<sub>2</sub> is a natural oxide of titanium that widely used in food products as additives or in pharmaceutical formulations, although a minor human health risk of TiO<sub>2</sub> nanoparticles has been proposed (83). A number of recent studies have shown that TiO<sub>2</sub> play a role in regulating autophagy, oxidative stress and apoptosis (84, 85). In Ruiz et al.'s study, the researchers found that oral administration of TiO<sub>2</sub> nanoparticles increased the severity of colitis induced by DSS, TiO<sub>2</sub> particles were taken up by intestinal epithelial cells and macrophages, and induced production of ROS, which then triggered the formation of the NLRP3 inflammasome. Hence, intake of food with TiO<sub>2</sub> particles may worsen the disease for patients with colitis.

#### IL-17 pathway

A number of recent studies showed that IL-17 pathway may act as a priming signal to promote expression of NLRP3 and IL-1 $\beta$  (86, 87). Thus, targeting IL-17 signaling may also affect the activation of NLRP3 inflammasome and contribute to progression of colitis. For instance, a study conducted by Lin et al (43) identified two compounds, wedelolactone and luteolin, from the bioactive fraction of *Wedelia chinensis* (WC) extracts, which has been proven to be effective in murine colitis model (88), and found that both compounds can suppress DSS induced colitis, and suppress the expression of NLRP3 and NLRP1 in the colon. Additionally, the inhibitory effects of luteolin on NLRP3 expression is mediated by suppressing genes in IL-17 signaling pathway, while the exact targets of luteolin and related mechanisms need to be identified in future.

# Nuclear factor erythroid 2-related factor 2 (Nrf2)/heme oxygenase-1 (HO-1)

Nrf2 is an important transcription factor that regulates an array of genes in modulating antioxidant and detoxifying systems. A number of previous studies showed that Nrf2 can suppress NLPR3 inflammasome activation by promoting expression of antioxidant genes, such as HO-1 (89), or inhibiting the expression of TXNIP (90). Studies by Zhou et al. (44, 91) found that colonic pathological changes induced by DSS or TNBS can be inhibited by Brusatol, a bioactive ingredient with

anti-inflammatory effect (92) extracted from *Brucea javanica* (BJ), which is a medicinal plant used for treatment of ulcerative colitis (UC) in Chinese medicine (93). For the mechanisms, the authors showed that the compound can activate Nrf2 and suppress the NLRP3 inflammasome activation illustrated by decreased expression of IL-1 $\beta$  and IL-18 in the colon. Thus, Brusatol may exert its anti-inflammasome role by up-regulating Nrf2 mediated antioxidant activity.

Another compound that may suppress colitis and NLRP3 inflammasome through Nrf2/HO-1 pathway is Toosendanin (TSN), a triterpenoid and natural insecticide with antiinflammatory activity derived from *Melia toosendan Sieb et Zucc* (MTSZ), which is traditional herbal medicine used as parasiticide of digestive tract and agricultural insecticide in China (94). For its role in colitis, Fan et al. (45) reported that TSN can attenuate DSS induced colonic inflammation *via* regulating the activation of NLRP3 inflammasome, the compound can also inhibit M1 macrophage polarization and production of pro-inflammatory cytokines and mediators in oxidative stress. These effects of TSN may be associated with its role in upregulating Nrf2/HO-1 expression in the colon and thus preventing M1 macrophage polarization and NLRP3 inflammasome activation.

Recent studies have provided evidence that high fat or high glucose diet may change the level of intracellular compounds such as Hydrogen sulfide (H<sub>2</sub>S) (95, 96)and alter the colonic homeostasis by regulating ROS production. H<sub>2</sub>S is a gaseous intracellular signal transducer that plays an anti-inflammatory effect in various diseases (97, 98). The role of H<sub>2</sub>S in IBD was evaluated by Qin et al (46) showing that H<sub>2</sub>S attenuated DSS induced colitis, H<sub>2</sub>S treated mice manifested reduced colon shortening and colonic pathological damages. H<sub>2</sub>S treatment inhibited expression of NLRP3 and cleavage of caspase-1 in the colon of mice. In *in vitro* studies, H<sub>2</sub>S can significantly inhibit NLRP3 inflammasome activation in bone marrow macrophages. For the mechanisms, the authors showed that H<sub>2</sub>S can disrupt Nrf2 activation, which may then affect intracellular ROS generation.

#### Aryl hydrocarbon receptor (AhR)

AhR is a universally expressed transcription factor that can bind to a variety of structurally diverse ligands and regulates a range of biological processes such as metabolism of xenobiotic chemicals, immunity and stem cell differentiation (99). It has been shown that AhR can suppress NLRP3 inflammasome activation by inhibiting NLRP3 transcription (100). A study conducted by Lv et al (13) found that a herbal compound, Norisoboldine (NOR) can alleviate colon inflammation induced by TNBS by targeting AhR. NOR is a bioactive alkaloid purified from Radix Linderae (RL), which is the root of *Lindera strychnifolia*, traditional Chinese medicine used in treating various diseases. The researchers showed that NOR alleviated colitis and reduced the levels of NLRP3, cleaved caspase-1 and IL-1 $\beta$  in colons of the mice. In *in vitro* studies, NOR was shown to activate AhR, which then enhanced Nrf2 level and reduced production of ROS. Thus, the anti-colitis role of NOR is mediated by inhibiting the NLRP3 inflammasome activation *via* modulating AhR/Nrf2/ROS pathway.

Cardamonin is a chalcone derived from cardamon with anticancer activity, it also plays a role in anti-infection immunity and the development of inflammatory diseases. A recent study has shown that cardamonin can inhibit the NLRP3 inflammasome activation and thus attenuate experimental colitis induced by both DSS and TNBS (47). In this study, the researchers found that cardamonin can activate the AhR/Nrf2/NQO1 pathway, blocking this pathway can abolish the effect of cardamonin on NLRP3 inflammasome.

#### Mitophagy

Mitophagy is an autophagic pathway that keeps the cell healthy by promoting the mitochondria turnover and preventing the accumulation of damaged mitochondria (101). It regulates the production of ROS and the release of mitochondria DNA. In mammals, mitophagy can be regulated by many factors such as PINK1 and parkin, whose deletion or mutation may cause impaired mitophagy and thereby leading to a variety of inflammatory diseases and cancers (102, 103). Previous studies have demonstrated that mitophagy is a negative regulator of NLRP3 inflammasome activation (104) and an important regulator of gut homeostasis (105). In a study conducted by Mai et al (48) showed that targeting mitophagy by palmatine, an isoquinoline alkaloid with potent antiinflammatory and anti-bacteria effects purified from herbal plants (106), can protect the host from colitis. The authors showed that palmatine attenuated DSS induced colon pathology and reduced the level of MPO, IL-1 $\beta$ , TNF- $\alpha$ , and macrophage infiltration to the colon. In mechanistical studies, they found that palmatine suppressed NLRP3 inflammasome activation, an impact that can be blocked by mitophagy inhibitor Cyclosporin A or PINK1 siRNA. On this basis, the suppression of palmatine on colon inflammation was mediated by inhibiting the NLRP3 inflammasome via modulating PINK1/ Parkin regulated mitophagy.

#### Unknown targets

As described above, studies have identified the targets of many natural compounds in regulating the progression of colitis and the activation of the NLRP3 inflammasome, whereas, for some other compounds, the investigation that identifying their intracellular targets is still in progress. One of these compounds is Alpha-mangostin (alpha-MG), a natural xanthonoid derived from the fruit hull of mangosteen (*Garcinia mangostana* L.) that has anti-cancer activity (107, 108). Being a potent antioxidant, a previous study proved that alpha-MG can reduce LPS induced inflammatory responses in IEC-6 cells (109). Using a high throughput sequencing method, Yin et al (49) found that the genes regulated by alpha-MG were mainly related to inflammation and oxidative stress. Using an LPS induced IBD model in rat, they found that pretreatment of alpha-MG significantly reduced all the pathological changes caused by LPS including detachment of intestinal villi, deformation of intestinal epithelial cell nuclei and swelling of mitochondria, and these effects of alpha-MG was exerted by suppressing the expression of NLRP3, caspase 1, IL-18, and IL-1 $\beta$ .

# Herbal medicine that affects IBD via other inflammasomes

As described above, regulation of the NLRP3 inflammasome using herbal compounds can affect the development of colitis via multiple mechanisms, whereas previous studies have shown that other inflammasome forming NLRs such as NLRP1, NLRP6, NLRC4 and AIM2 are also involved in the pathogenesis of colonic inflammation (110-113). Thus, we searched the literature and found that several compounds affect the development of colitis by targeting NLRP1 and NLRP6. Secoisolariciresinol diglucoside (SDG), a bioactive compound derived from lignans, plays anti-inflammatory and antioxidant roles in various diseases (114, 115). A study performed by Wang et al (50) showed that SDG can attenuated colon inflammation and macrophage infiltration to the colon in a mouse model induced by DSS. Meanwhile, SDG can inhibit the activation of NLRP1 inflammasome and the production of pro-inflammatory cytokines including IL-1 $\beta$ , IL-18 and TNF- $\alpha$  in both colon tissue of DSS treated mice and RAW264.7 macrophages. The effect of SDG on NLRP1 inflammasome was partly dependent on disruption of NF-kB activation.

A previous study showed that apigenin, a flavone that has antioxidant effect and is involved in development of various diseases (116), can suppress DSS induced colitis *via* regulating the activation of NLRP6 inflammasome. In this study, Radulovic et al (51) found that found that mice were protected from DSS induced colitis after cohousing with apigenin treated animals. In contrast, deficiency of NLRP6 disrupted the protective effect of apigenin on the mice. Data of 16S rRNA sequencing showed that apigenin induced a composition change in gut microbiota, which was absent in NLRP6 deficient mice. In addition, the impact of apigenin on colitis was not affected by the absence of caspase-1/11 or ASC. Thus, these studies indicated an inflammasome independent mechanism of NLRP6 whereby apigenin reshaped gut microbiota through NLRP6 and thus protected mice against colitis.

# Herbal compounds that possibly affect colitis *via* regulating inflammasomes

Ginseng has been wildly used in treatment of a variety of human diseases including IBD. Studies have shown that the beneficial activities of ginseng are associated with its metabolites such as compound K (CK), 2-(S)-protopanaxatriol, Rh1, F1, and 20(S)-protopanaxadiol. Seong et al (117) showed that fermented wild ginseng (FWG) alleviated the severity of colitis induced by DSS and reduced the infiltration of macrophages in colonic tissue. FWG also inhibited the expression of IL-1B in LPS treated macrophages. In another study, researchers found that American ginseng extracts and the bioactive component, panaxynol, ameliorated colitis by activating Nrf2 pathway and thus reducing ROS production (52, 118). These evidences suggest that panaxynol may affect colitis by regulating the activation of an inflammasome. Ginsenoside Rf is another bioactive compound extracted from ginseng. A study by Ahn et al (119) reported that ginsenoside Rf can decrease the production of IL-1β, IL-6, TNF-a, NO, and ROS, mediators that are highly activated in IBD, in TNF- $\alpha$  stimulated intestinal epithelial cells and macrophages. Additionally, ginsenoside Rf suppressed TNF-a/LPS-induced NF-kB activity. Hence, ginsenoside Rf has potent intestinal anti-inflammatory effects and may have potential to treat IBD. The effect of ginseng extracts on IL-1ß expression and ROS indicated a possible involvement of inflammasomes in the anti-colitis function.

Indigo naturalis (IN) is herbal medicine extracted from leaves and stems of plants and is a component of crude drugs used in treating many diseases including IBD (120). However, studies have shown that inappropriate application of IN can also aggravate colitis and pulmonary arterial hypertension, thus, using it topically has been suggested by researchers in application of IN in treatment of IBD (120). Another way that may reduce the side effects of IN is to identify the effective components in IN, since the effects of IN may be associated with the bioactive ingredients it contains. For instance, IN derived indigo, indirubin, 6-formylindolo [3,2-b] carbazole (FICZ), and indole-3-carboxaldehyde (IAId) can act as ligands and activate aryl hydrocarbon receptor (AhR), a pathway that can promote mucosal healing by regulating the IL-22 production from type-3 innate lymphocytes cells (120). Considering the inhibitory effect of AhR signaling on NLRP3 inflammasome activation (100), AhR ligands derived from IN may also have an impact on the NLRP3 inflammasome. Thus, the compounds affecting inflammasome regulators may modulate the progression of colitis via altering activation of inflammasomes. It is worthwhile to examine the roles of these compounds in a colitis model, which may contribute to the design of new anticolitis strategies.

### Limitations and possible solutions

Traditional or alternative medicine has been used globally for centuries, the efficacy and safety of a variety of decoctions have been evaluated in large amounts of clinical cases. While the application of traditional medicine, especially the decoctions, is largely dependent on physician's judgement to patients' particular conditions, which to a great extent rely on physician's knowledge and experience, and thus limits the wide application of traditional medicine. A decoction usually contains a number of different herbals or mineral materials for disease treatment, thus it may contain thousands of compounds with various effects. However, the compound composition of a particular herbal plant may largely rely on its growing environment such as soil conditions. Hence it is hard to exactly predict the integrated function of the compounds in a herbal decoction, some adverse events may occur in hospitals using traditional medicine (121). To overcome this problem, integrated analysis of the patients' physiological condition and the functional compounds in an effective herbal decoction is necessary in future studies.

As mentioned previously, most of the herbal derived compounds, such as curcumin and some ginseng products, are water insoluble, unstable and inefficient in transiting across the physiological barriers, hence it is challenging to reach a concentration in the plasma or the affected tissues that is effective to treat a disease. A reported way that can improve the bioavailability of herbal compounds is combined administration with particular adjuvant compounds. For instance, studies decades ago had found that concomitant administration of piperine can largely improve the plasma concentration of curcumin (122). Similarly, a recent study showed that curcumin solubilized in essential turmeric oils showed substantially enhanced anti-inflammatory effects compared with curcumin alone in DSS-induced colitis (123). Further studies showed that some components contain in turmeric oils such as turmerones have anti-inflammatory role (124) and may the therapeutic benefits of curcumin in colitis model.

Another way that can effectively improve the bioavailability of herbal compounds is by using delivery-aid carriers. Accumulating evidences have shown that nanocarriers, such as lipid nanoparticles and liposomes, can efficiently increase the blood concentration of insoluble herbal compounds. For instance, Yang et al. (125) showed that polymer-based nanoparticles can significantly enhance curcumin delivery to the brain of mice. In another study, researchers found that nanoparticle encapsulation can increase the curcumin oral bioavailability for at least 9-fold compared with concomitant administration of curcumin with piperine (126). Therefore, application of nanoparticle-based carriers may be a promising approach for improving herbal compound bioavailability. Liposomes are vesicles formed by phospholipid bilayers (127), which can solubilize water insoluble compounds and transport these compounds into cells by membrane fusion. Many liposome-based drug delivery systems have been developed and showed enhanced bioavailability for delivery of a great number of water-insoluble herbal compounds in recent studies (128). For instance, a study by Telange et al. (129) showed that the formation of apigenin-phospholipid complex remarkably enhanced water solubility of apigenin for over 36-fold compared to pure apigenin.

Traditional medicine can be considered as a summary of experience from hundreds of ancient physicians in their medical practice, and it has been proven to be effective in many cases. Thus, it at least gives us a hint to discover compounds that are possibly effective to a particular disease. As discussed above, many compounds that prevent inflammation in colitis models described in this review are discovered in this way. According to the findings mentioned here, natural compounds purified from medicinal herbs or dietary materials are promising agents in attenuating colitis severity in animal models. However, their potential side effects on other organs and tissues, their physiological impacts on human cells have not been fully evaluated. Furthermore, the functional ingredients of some decoctions are not identified and the specificity of compounds on inflammasomes or other inflammatory signaling pathways are not clear. Additionally, the exact mechanisms whereby medicinal compounds regulate inflammasome activity are not fully addressed. On this basis, further studies are needed to address these questions focusing on evaluating or reducing side effects, determining drug specificity and working mechanisms, altering compound dosage or combination may be helpful to optimize the efficacy and safety of the compounds. With a satisfied answer of these questions, a clinical trial in IBD patients is a must before recommending the use of these compounds in clinical practice.

As mentioned above, compounds that are effective in studies of animal models for diseases may meet other challenges when applied in translational studies, especially the doses on an mg/kg basis proven to be safe and effective in animal studies sometimes do not display satisfactory outcomes after scaling of the doses based on body weight and applying in human clinical trials. This condition is primarily due to the varied pharmacokinetics in species with different biochemical systems (130, 131). To overcome this problem, U.S. Food and Drug Administration has developed a guidance in estimating the human equivalent dose (HED) for a compound that can act as maximum recommended starting dose (MRSD) in its initial clinical trial based on data derived from animal studies (https://www.fda.gov/ regulatory-information/search-fda-guidance-documents/ estimating-maximum-safe-sta). According to this guidance, the estimation of HED requires studies to determine the No Observed Adverse Effect Level (NOAEL) of a compound in multiple tested animal species. This requires data on systemic toxicology and others, such as bioavailability, metabolite profile

and plasma level of the compound. With the availability of all these data, the conversion of NOAEL to HED needs to employ a scaling system based on body weight (mg/kg) or body surface area  $(mg/m^2)$ , a process that needs to consider multiple factors, such as the method of compound administration and NOAELs in different animal species. However, the in vivo data in majority of studies mentioned in this review did not provide sufficient details to estimate the MRSD of the tested compounds. Only four out of twenty-two studies provided in vivo toxicity data that obtained from IBD model induced in one species of animal (Table 1). Some of other studies did in vitro toxicity tests in a number of cell lines, such as RAW264.7 and THP-1. Thus, the current available data are not enough for the estimation of MRSDs for the compounds mentioned here. More extensive animal studies with multiple species that determine the NOAELs, toxicology, tolerability and pharmacokinetic profiles are necessary for the estimation of its MRSD in future investigation for compounds with potential clinical applications.

#### Conclusion

This review summarized recent studies reporting the role and mechanisms of traditional herbal medicine or diet derived compounds in modulating colitis via regulating inflammasome activity in IBD models. These studies support a point of view that herbal or diet derived compounds may inactivate inflammasomes in the colon and thus attenuate colonic inflammation in animal models, or IBD patients in some cases, by targeting various inflammasome regulators as described above. Whereas the role of inflammasomes in the progression of colon inflammation is complicated since a number of previous studies using animals deficient in inflammasome components such as NLRP3 and caspase-1 proved that inflammasome function is also required in the maintenance of gut homeostasis (7). Thus, the activation of inflammasomes needs an appropriate regulation to keep a balanced immune response in the colon. Modulating inflammasome activity using bioactive compounds purified from traditional medicine or diet might be a promising strategy for treatment of IBD. Establishment of a cellular screening model using new identification technologies might

# References

1. Khor B, Gardet A, Xavier RJ. Genetics and pathogenesis of inflammatory bowel disease. Nat (2011) 474(7351):307-17. doi: 10.1038/nature10209

be helpful to discover new compounds specifically targeting inflammasomes for treatment of IBD. A deep understanding of the pharmacological characters of the compounds in preventing inflammasomes may expediting the development of new strategies with higher efficiency for treatment of IBD and many other inflammatory diseases.

## Author contributions

QX performed the literature search, drew the figures, and wrote the first draft of the manuscript. WS, JZ, YM, and JB helped in manuscript preparation. SH, XZ, and LM revised and edited the final version of the manuscript. All authors contributed to the article and approved the submitted version.

### Funding

This study was supported in part by National Natural Science Foundation of China (32070919, 32170915 and 82172931), Natural Science Foundation of Jiangsu province (BK20201442), Jiangsu Specially-Appointed Professor and Start-up funds for young scientists of Nantong University (03083051).

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

<sup>2.</sup> Graham DB, Xavier RJ. Pathway paradigms revealed from the genetics of inflammatory bowel disease. *Nat* (2020) 578(7796):527-39. doi: 10.1038/s41586-020-2025-2

<sup>3.</sup> Strober W, Fuss I, Mannon P. The fundamental basis of inflammatory bowel disease. J Clin Invest (2007) 117(3):514-21. doi: 10.1172/JCI30587

<sup>4.</sup> Beard JA, Click BH. The burden of cost in inflammatory bowel disease: a medical economic perspective. *Curr Opin Gastroenterol* (2020) 36(4):310–6. doi: 10.1097/MOG.00000000000642

<sup>5.</sup> Burisch J, Jess T, Martinato M, Lakatos PL, EpiCom E. The burden of inflammatory bowel disease in Europe. J Crohns Colitis (2013) 7(4):322-37. doi: 10.1016/j.crohns.2013.01.010

<sup>6.</sup> Collaborators GBDIBD. The global, regional, and national burden of inflammatory bowel disease in 195 countries and territories, 1990-2017: a

systematic analysis for the global burden of disease study 2017. Lancet Gastroenterol Hepatol (2020) 5(1):17-30. doi: 10.1016/S2468-1253(19)30333-4

7. Mao L, Kitani A, Strober W, Fuss IJ. The role of NLRP3 and IL-1beta in the pathogenesis of inflammatory bowel disease. *Front Immunol* (2018) 9:2566. doi: 10.3389/fimmu.2018.02566

8. Mao L, Kitani A, Similuk M, Oler AJ, Albenberg L, Kelsen J, et al. Loss-offunction CARD8 mutation causes NLRP3 inflammasome activation and crohn's disease. J Clin Invest (2018) 128(5):1793–806. doi: 10.1172/JCI98642

9. Brandes A, Groth A, Gottschalk F, Wilke T, Ratsch BA, Orzechowski HD, et al. Real-world biologic treatment and associated cost in patients with inflammatory bowel disease. *Z Gastroenterol* (2019) 57(7):843–51. doi: 10.1055/a-0903-2938

10. Liang J, Chen S, Chen J, Lin J, Xiong Q, Yang Y, et al. Therapeutic roles of polysaccharides from dendrobium officinaleon colitis and its underlying mechanisms. *Carbohydr Polym* (2018) 185:159-68. doi: 10.1016/j.carbpol.2018.01.013

11. Liu Q, Zuo R, Wang K, Nong FF, Fu YJ, Huang SW, et al. Oroxindin inhibits macrophage NLRP3 inflammasome activation in DSS-induced ulcerative colitis in mice via suppressing TXNIP-dependent NF-kappaB pathway. *Acta Pharmacol Sin* (2020) 41(6):771–81. doi: 10.1038/s41401-019-0335-4

12. Luo X, Yu Z, Deng C, Zhang J, Ren G, Sun A, et al. Baicalein ameliorates TNBS-induced colitis by suppressing TLR4/MyD88 signaling cascade and NLRP3 inflammasome activation in mice. *Sci Rep* (2017) 7(1):16374. doi: 10.1038/s41598-017-12562-6

13. Lv Q, Wang K, Qiao SM, Dai Y, Wei ZF. Norisoboldine, a natural aryl hydrocarbon receptor agonist, alleviates TNBS-induced colitis in mice, by inhibiting the activation of NLRP3 inflammasome. *Chin J Nat Med* (2018) 16 (3):161–74. doi: 10.1016/S1875-5364(18)30044-X

14. Kanneganti TD. Inflammatory bowel disease and the NLRP3 inflammasome. N Engl J Med (2017) 377(7):694–6. doi: 10.1056/NEJMcibr1706536

15. Latz E. The inflammasomes: mechanisms of activation and function. Curr Opin Immunol (2010) 22(1):28-33. doi: 10.1016/j.coi.2009.12.004

16. Strowig T, Henao-Mejia J, Elinav E, Flavell R. Inflammasomes in health and disease. Nat (2012) 481(7381):278-86. doi: 10.1038/nature10759

17. He Y, Hara H, Nunez G. Mechanism and regulation of NLRP3 inflammasome activation. *Trends Biochem Sci* (2016) 41(12):1012-21. doi: 10.1016/j.tibs.2016.09.002

18. Swanson KV, Deng M, Ting JP. The NLRP3 inflammasome: molecular activation and regulation to therapeutics. *Nat Rev Immunol* (2019) 19(8):477–89. doi: 10.1038/s41577-019-0165-0

19. Zhu S, Ding S, Wang P, Wei Z, Pan W, Palm NW, et al. Nlrp9b inflammasome restricts rotavirus infection in intestinal epithelial cells. *Nat* (2017) 546(7660):667-70. doi: 10.1038/nature22967

20. Ngo C, Man SM. NLRP9b: a novel RNA-sensing inflammasome complex. Cell Res (2017) 27(11):1302-3. doi: 10.1038/cr.2017.93

21. Malik A, Kanneganti TD. Inflammasome activation and assembly at a glance. J Cell Sci (2017) 130(23):3955-63. doi: 10.1242/jcs.207365

22. Bauer C, Duewell P, Mayer C, Lehr HA, Fitzgerald KA, Dauer M, et al. Colitis induced in mice with dextran sulfate sodium (DSS) is mediated by the NLRP3 inflammasome. *Gut* (2010) 59(9):1192-9. doi: 10.1136/gut.2009.197822

23. Liu L, Dong Y, Ye M, Jin S, Yang J, Joosse ME, et al. The pathogenic role of NLRP3 inflammasome activation in inflammatory bowel diseases of both mice and humans. *J Crohns Colitis* (2017) 11(6):737–50. doi: 10.1093/ecco-jcc/jjw219

24. Bauer C, Duewell P, Lehr HA, Endres S, Schnurr M. Protective and aggravating effects of Nlrp3 inflammasome activation in IBD models: influence of genetic and environmental factors. *Dig Dis* (2012) 30 Suppl 1:82–90. doi: 10.1159/000341681

25. Zaki MH, Boyd KL, Vogel P, Kastan MB, Lamkanfi M, Kanneganti TD. The NLRP3 inflammasome protects against loss of epithelial integrity and mortality during experimental colitis. *Immun* (2010) 32(3):379–91. doi: 10.1016/j.immuni.2010.03.003

26. Shah B, Mayer L. Current status of monoclonal antibody therapy for the treatment of inflammatory bowel disease. *Expert Rev Clin Immunol* (2010) 6 (4):607–20. doi: 10.1586/eci.10.45

27. Di Paolo A, Luci G. Personalized medicine of monoclonal antibodies in inflammatory bowel disease: Pharmacogenetics, therapeutic drug monitoring, and beyond. *Front Pharmacol* (2020) 11:610806. doi: 10.3389/fphar.2020.610806

28. Corcoran SE, Halai R, Cooper MA. Pharmacological inhibition of the nodlike receptor family pyrin domain containing 3 inflammasome with MCC950. *Pharmacol Rev* (2021) 73(3):968–1000. doi: 10.1124/pharmrev.120.000171

29. Jiang H, He H, Chen Y, Huang W, Cheng J, Ye J, et al. Identification of a selective and direct NLRP3 inhibitor to treat inflammatory disorders. *J Exp Med* (2017) 214(11):3219–38. doi: 10.1084/jem.20171419

30. Yang Y, Islam MS, Wang J, Li Y, Chen X. Traditional Chinese medicine in the treatment of patients infected with 2019-new coronavirus (SARS-CoV-2): A review and perspective. *Int J Biol Sci* (2020) 16(10):1708–17. doi: 10.7150/ ijbs.45538

31. Ma Y, Chen M, Guo Y, Liu J, Chen W, Guan M, et al. Prevention and treatment of infectious diseases by traditional Chinese medicine: a commentary. *APMIS* (2019) 127(5):372–84. doi: 10.1111/apm.12928

32. Dudics S, Langan D, Meka RR, Venkatesha SH, Berman BM, Che CT, et al. Natural products for the treatment of autoimmune arthritis: Their mechanisms of action, targeted delivery, and interplay with the host microbiome. *Int J Mol Sci* (2018) 19(9):2508–28. doi: 10.3390/ijms19092508

33. Kristoffersen AE, Stub T, Broderstad AR, Hansen AH. Use of traditional and complementary medicine among Norwegian cancer patients in the seventh survey of the tromso study. *BMC Complement Altern Med* (2019) 19(1):341. doi: 10.1186/ s12906-019-2762-7

34. Teschke R, Wolff A, Frenzel C, Eickhoff A, Schulze J. Herbal traditional Chinese medicine and its evidence base in gastrointestinal disorders. *World J Gastroenterol* (2015) 21(15):4466–90. doi: 10.3748/wjg.v21.i15.4466

35. Tian Z, Liu Y, Yang B, Zhang J, He H, Ge H, et al. Astagalus polysaccharide attenuates murine colitis through inhibiton of the NLRP3 inflammasome. *Planta Med* (2017) 83(1-02):70–7. doi: 10.1055/s-0042-108589

36. Shen P, Zhang Z, Zhu K, Cao H, Liu J, Lu X, et al. Evodiamine prevents dextran sulfate sodium-induced murine experimental colitis *via* the regulation of NF-kappaB and NLRP3 inflammasome. *BioMed Pharmacother* (2019) 110:786–95. doi: 10.1016/j.biopha.2018.12.033

37. Cui L, Wang W, Luo Y, Ning Q, Xia Z, Chen J, et al. Polysaccharide from scutellaria baicalensis georgi ameliorates colitis *via* suppressing NF-kappaB signaling and NLRP3 inflammasome activation. *Int J Biol Macromol* (2019) 132:393–405. doi: 10.1016/j.ijbiomac.2019.03.230

38. Pu Z, Han C, Zhang W, Xu M, Wu Z, Liu Y, et al. Systematic understanding of the mechanism and effects of arctigenin attenuates inflammation in dextran sulfate sodium-induced acute colitis through suppression of NLRP3 inflammasome by SIRT1. *Am J Transl Res* (2019) 11(7):3992–4009.

39. Qu S, Shen Y, Wang M, Wang X, Yang Y. Suppression of miR-21 and miR-155 of macrophage by cinnamaldehyde ameliorates ulcerative colitis. *Int Immunopharmacol* (2019) 67:22-34. doi: 10.1016/j.intimp.2018.11.045

40. Zhao Y, Guo Q, Zhu Q, Tan R, Bai D, Bu X, et al. Flavonoid VI-16 protects against DSS-induced colitis by inhibiting txnip-dependent NLRP3 inflammasome activation in macrophages *via* reducing oxidative stress. *Mucosal Immunol* (2019) 12(5):1150–63. doi: 10.1038/s41385-019-0177-x

41. Gong Z, Zhao S, Zhou J, Yan J, Wang L, Du X, et al. Curcumin alleviates DSS-induced colitis via inhibiting NLRP3 inflammsome activation and IL-1beta production. *Mol Immunol* (2018) 104:11–9. doi: 10.1016/j.molimm.2018.09.004

42. Ruiz PA, Moron B, Becker HM, Lang S, Atrott K, Spalinger MR, et al. Titanium dioxide nanoparticles exacerbate DSS-induced colitis: role of the NLRP3 inflammasome. *Gut* (2017) 66(7):1216–24. doi: 10.1136/gutjnl-2015-310297

43. Lin TJ, Yin SY, Hsiao PW, Yang NS, Wang IJ. Transcriptomic analysis reveals a controlling mechanism for NLRP3 and IL-17A in dextran sulfate sodium (DSS)-induced colitis. *Sci Rep* (2018) 8(1):14927. doi: 10.1038/s41598-018-33204-5

44. Zhou J, Wang T, Dou Y, Huang Y, Qu C, Gao J, et al. Brusatol ameliorates 2, 4, 6-trinitrobenzenesulfonic acid-induced experimental colitis in rats: Involvement of NF-kappaB pathway and NLRP3 inflammasome. *Int Immunopharmacol* (2018) 64:264–74. doi: 10.1016/j.intimp.2018.09.008

45. Fan H, Chen W, Zhu J, Zhang J, Peng S. Toosendanin alleviates dextran sulfate sodium-induced colitis by inhibiting M1 macrophage polarization and regulating NLRP3 inflammasome and Nrf2/HO-1 signaling. *Int Immunopharmacol* (2019) 76:105909. doi: 10.1016/j.intimp.2019.105909

46. Qin M, Long F, Wu W, Yang D, Huang M, Xiao C, et al. Hydrogen sulfide protects against DSS-induced colitis by inhibiting NLRP3 inflammasome. *Free Radic Biol Med* (2019) 137:99–109. doi: 10.1016/j.freeradbiomed.2019.04.025

47. Wang K, Lv Q, Miao YM, Qiao SM, Dai Y, Wei ZF. Cardamonin, a natural flavone, alleviates inflammatory bowel disease by the inhibition of NLRP3 inflammasome activation *via* an AhR/Nrf2/NQO1 pathway. *Biochem Pharmacol* (2018) 155:494–509. doi: 10.1016/j.bcp.2018.07.039

48. Mai CT, Wu MM, Wang CL, Su ZR, Cheng YY, Zhang XJ. Palmatine attenuated dextran sulfate sodium (DSS)-induced colitis via promoting mitophagy-mediated NLRP3 inflammasome inactivation. *Mol Immunol* (2019) 105:76–85. doi: 10.1016/j.molimm.2018.10.015

49. Yin P, Zou W, Li J, Jin N, Gao Q, Liu F. Using high-throughput sequencing to explore the anti-inflammatory effects of alpha-mangostin. *Sci Rep* (2019) 9 (1):15626. doi: 10.1038/s41598-019-52036-5

50. Wang Z, Chen T, Yang C, Bao T, Yang X, He F, et al. Secoisolariciresinol diglucoside suppresses dextran sulfate sodium salt-induced colitis through inhibiting NLRP1 inflammasome. *Int Immunopharmacol* (2020) 78:105931. doi: 10.1016/j.intimp.2019.105931

51. Radulovic K, Normand S, Rehman A, Delanoye-Crespin A, Chatagnon J, Delacre M, et al. A dietary flavone confers communicable protection against colitis through NLRP6 signaling independently of inflammasome activation. *Mucosal Immunol* (2018) 11(3):811–9. doi: 10.1038/mi.2017.87

52. Chaparala A, Poudyal D, Tashkandi H, Witalison EE, Chumanevich AA, Hofseth JL, et al. Panaxynol, a bioactive component of American ginseng, targets macrophages and suppresses colitis in mice. *Oncotarget* (2020) 11(22):2026–36. doi: 10.18632/oncotarget.27592

53. Auyeung KK, Han QB, Ko JK. Astragalus membranaceus: A review of its protection against inflammation and gastrointestinal cancers. *Am J Chin Med* (2016) 44(1):1–22. doi: 10.1142/S0192415X16500014

54. Zhang L, Shergis JL, Yang L, Zhang AL, Guo X, Zhang L, et al. Astragalus membranaceus (Huang qi) as adjunctive therapy for diabetic kidney disease: An updated systematic review and meta-analysis. *J Ethnopharmacol* (2019) 239:111921. doi: 10.1016/j.jep.2019.111921

55. Kwon HJ, Won YS, Suh HW, Jeon JH, Shao Y, Yoon SR, et al. Vitamin D3 upregulated protein 1 suppresses TNF-alpha-induced NF-kappaB activation in hepatocarcinogenesis. *J Immunol* (2010) 185(7):3980–9. doi: 10.4049/jimmunol.1000990

56. Zhou R, Tardivel A, Thorens B, Choi I, Tschopp J. Thioredoxin-interacting protein links oxidative stress to inflammasome activation. *Nat Immunol* (2010) 11 (2):136–40. doi: 10.1038/ni.1831

57. Chassaing B, Aitken JD, Malleshappa M, Vijay-Kumar M. Dextran sulfate sodium (DSS)-induced colitis in mice. *Curr Protoc Immunol* (2014) 104:15 25 1–15 25 14. doi: 10.1002/0471142735.im1525s104

58. Kiesler P, Fuss IJ, Strober W. Experimental models of inflammatory bowel diseases. *Cell Mol Gastroenterol Hepatol* (2015) 1(2):154–70. doi: 10.1016/j.jcmgh.2015.01.006

59. Ren Q, Zhao S, Ren C, Ma Z. Astragalus polysaccharide alleviates LPSinduced inflammation injury by regulating miR-127 in H9c2 cardiomyoblasts. *Int J Immunopathol Pharmacol* (2018) 32:2058738418759180. doi: 10.1177/ 2058738418759180

60. Zhang Z, Zhang L, Xu H. Effect of astragalus polysaccharide in treatment of diabetes mellitus: a narrative review. *J Tradit Chin Med* (2019) 39(1):133-8.

61. Tan Q, Zhang J. Evodiamine and its role in chronic diseases. Adv Exp Med Biol (2016) 929:315-28. doi: 10.1007/978-3-319-41342-6\_14

62. Tian KM, Li JJ, Xu SW. Rutaecarpine: A promising cardiovascular protective alkaloid from evodia rutaecarpa (Wu Zhu yu). *Pharmacol Res* (2019) 141:541–50. doi: 10.1016/j.phrs.2018.12.019

63. Jean-Charles PY, Kaur S, Shenoy SK. G Protein-coupled receptor signaling through beta-Arrestin-Dependent mechanisms. *J Cardiovasc Pharmacol* (2017) 70 (3):142–58. doi: 10.1097/FJC.00000000000482

64. Sharma D, Parameswaran N. Multifaceted role of beta-arrestins in inflammation and disease. *Genes Immun* (2015) 16(8):499-513. doi: 10.1038/gene.2015.37

65. Mao K, Chen S, Wang Y, Zeng Y, Ma Y, Hu Y, et al. Beta-arrestin1 is critical for the full activation of NLRP3 and NLRC4 inflammasomes. *J Immunol* (2015) 194 (4):1867–73. doi: 10.4049/jimmunol.1401989

66. Yamamoto H, Schoonjans K, Auwerx J. Sirtuin functions in health and disease. *Mol Endocrinol* (2007) 21(8):1745–55. doi: 10.1210/me.2007-0079

67. Li Y, Yang X, He Y, Wang W, Zhang J, Zhang W, et al. Negative regulation of NLRP3 inflammasome by SIRT1 in vascular endothelial cells. *Immunobiol* (2017) 222(3):552–61. doi: 10.1016/j.imbio.2016.11.002

68. Park S, Shin J, Bae J, Han D, Park SR, Shin J, et al. SIRT1 alleviates LPSinduced IL-1beta production by suppressing NLRP3 inflammasome activation and ROS production in trophoblasts. *Cells* (2020) 9(3):728–45. doi: 10.3390/ cells9030728

69. Zhong Y, Lee K, Deng Y, Ma Y, Chen Y, Li X, et al. Arctigenin attenuates diabetic kidney disease through the activation of PP2A in podocytes. *Nat Commun* (2019) 10(1):4523. doi: 10.1038/s41467-019-12433-w

70. Shimazu R, Akashi S, Ogata H, Nagai Y, Fukudome K, Miyake K, et al. MD-2, a molecule that confers lipopolysaccharide responsiveness on toll-like receptor 4. *J Exp Med* (1999) 189(11):1777–82. doi: 10.1084/jem.189.11.1777

71. Lee W, Ku SK, Bae JS. Anti-inflammatory effects of baicalin, baicalein, and wogonin in vitro and *in vivo. Inflammation* (2015) 38(1):110–25. doi: 10.1007/s10753-014-0013-0

72. Bie B, Sun J, Guo Y, Li J, Jiang W, Yang J, et al. Baicalein: A review of its anticancer effects and mechanisms in hepatocellular carcinoma. *BioMed Pharmacother* (2017) 93:1285–91. doi: 10.1016/j.biopha.2017.07.068

73. Hewlings SJ, Kalman DS. Curcumin: A review of its effects on human health. *Foods* (2017) 6(10):92–102. doi: 10.3390/foods6100092

74. Fallahi F, Borran S, Ashrafizadeh M, Zarrabi A, Pourhanifeh MH, Khaksary Mahabady M, et al. Curcumin and inflammatory bowel diseases: From *in vitro* 

studies to clinical trials. Mol Immunol (2021) 130:20-30. doi: 10.1016/j.molimm.2020.11.016

75. Doyle AA, Stephens JC. A review of cinnamaldehyde and its derivatives as antibacterial agents. *Fitoterapia* (2019) 139:104405. doi: 10.1016/j.fitote.2019.104405

76. Muhammad JS, Zaidi SF, Shaharyar S, Refaat A, Usmanghani K, Saiki I, et al. Anti-inflammatory effect of cinnamaldehyde in helicobacter pylori induced gastric inflammation. *Biol Pharm Bull* (2015) 38(1):109–15. doi: 10.1248/bpb.b14-00609

77. Tschopp J, Schroder K. NLRP3 inflammasome activation: The convergence of multiple signalling pathways on ROS production? *Nat Rev Immunol* (2010) 10 (3):210–5. doi: 10.1038/nri2725

78. Guo X, Chen S, Yu W, Chi Z, Wang Z, Xu T, et al. AKT controls NLRP3 inflammasome activation by inducing DDX3X phosphorylation. *FEBS Lett* (2021) 595(19):2447–62. doi: 10.1002/1873-3468.14175

79. Zhao W, Shi CS, Harrison K, Hwang IY, Nabar NR, Wang M, et al. AKT regulates NLRP3 inflammasome activation by phosphorylating NLRP3 serine 5. *J Immunol* (2020) 205(8):2255–64. doi: 10.4049/jimmunol.2000649

80. Kozlowska A, Szostak-Wegierek D. Flavonoids-food sources and health benefits. *Rocz Panstw Zakl Hig.* (2014) 65(2):79-85.

81. Toh JY, Tan VM, Lim PC, Lim ST, Chong MF. Flavonoids from fruit and vegetables: a focus on cardiovascular risk factors. *Curr Atheroscler Rep* (2013) 15 (12):368. doi: 10.1007/s11883-013-0368-y

82. Lim H, Min DS, Park H, Kim HP. Flavonoids interfere with NLRP3 inflammasome activation. *Toxicol Appl Pharmacol* (2018) 355::93-102. doi: 10.1016/j.taap.2018.06.022

83. Grande F, Tucci P. Titanium dioxide nanoparticles: a risk for human health? *Mini Rev Med Chem* (2016) 16(9):762-9. doi: 10.2174/ 1389557516666160321114341

84. Shen C, Zhang Z, Xie T, Ji J, Xu J, Lin L, et al. Rhein suppresses lung inflammatory injury induced by human respiratory syncytial virus through inhibiting NLRP3 inflammasome activation *via* NF-kappaB pathway in mice. *Front Pharmacol* (2019) 10:1600. doi: 10.3389/fphar.2019.01600

85. Abbasi-Oshaghi E, Mirzaei F, Pourjafar M. NLRP3 inflammasome, oxidative stress, and apoptosis induced in the intestine and liver of rats treated with titanium dioxide nanoparticles: *in vivo* and *in vitro* study. *Int J Nanomed* (2019) 14:1919–36. doi: 10.2147/IJN.S192382

86. Zhang S, Yu N, Zhang R, Zhang S, Wu J. Interleukin-17A induces IL-1beta secretion from RPE cells *Via* the NLRP3 inflammasome. *Invest Ophthalmol Vis Sci* (2016) 57(2):312–9. doi: 10.1167/iovs.15-17578

87. Le Jan S, Muller C, Plee J, Durlach A, Bernard P, Antonicelli F. IL-23/IL-17 axis activates IL-1beta-Associated inflammasome in macrophages and generates an auto-inflammatory response in a subgroup of patients with bullous pemphigoid. *Front Immunol* (2019) 10:1972. doi: 10.3389/fimmu.2019.01972

88. Huang YT, Wen CC, Chen YH, Huang WC, Huang LT, Lin WC, et al. Dietary uptake of wedelia chinensis extract attenuates dextran sulfate sodiuminduced colitis in mice. *PloS One* (2013) 8(5):e64152. doi: 10.1371/ journal.pone.0064152

89. Shi A, Shi H, Wang Y, Liu X, Cheng Y, Li H, et al. Activation of Nrf2 pathway and inhibition of NLRP3 inflammasome activation contribute to the protective effect of chlorogenic acid on acute liver injury. *Int Immunopharmacol* (2018) 54:125–30. doi: 10.1016/j.intimp.2017.11.007

90. Hou Y, Wang Y, He Q, Li L, Xie H, Zhao Y, et al. Nrf2 inhibits NLRP3 inflammasome activation through regulating Trx1/TXNIP complex in cerebral ischemia reperfusion injury. *Behav Brain Res* (2018) 336:32–9. doi: 10.1016/j.bbr.2017.06.027

91. Zhou J, Tan L, Xie J, Lai Z, Huang Y, Qu C, et al. Characterization of brusatol self-microemulsifying drug delivery system and its therapeutic effect against dextran sodium sulfate-induced ulcerative colitis in mice. *Drug Deliv* (2017) 24(1):1667–79. doi: 10.1080/10717544.2017.1384521

92. Tang W, Xie J, Xu S, Lv H, Lin M, Yuan S, et al. Novel nitric oxide-releasing derivatives of brusatol as anti-inflammatory agents: design, synthesis, biological evaluation, and nitric oxide release studies. *J Med Chem* (2014) 57(18):7600–12. doi: 10.1021/jm5007534

93. Huang YF, Zhou JT, Qu C, Dou YX, Huang QH, Lin ZX, et al. Antiinflammatory effects of brucea javanica oil emulsion by suppressing NF-kappaB activation on dextran sulfate sodium-induced ulcerative colitis in mice. *J Ethnopharmacol* (2017) 198:389–98. doi: 10.1016/j.jep.2017.01.042

94. Shi YL, Li MF. Biological effects of toosendanin, a triterpenoid extracted from Chinese traditional medicine. *Prog Neurobiol* (2007) 82(1):1–10. doi: 10.1016/j.pneurobio.2007.02.002

95. Peh MT, Anwar AB, Ng DS, Atan MS, Kumar SD, Moore PK. Effect of feeding a high fat diet on hydrogen sulfide (H2S) metabolism in the mouse. *Nitric Oxide* (2014) 41::138–45. doi: 10.1016/j.niox.2014.03.002

96. Okamoto M, Yamaoka M, Takei M, Ando T, Taniguchi S, Ishii I, et al. Endogenous hydrogen sulfide protects pancreatic beta-cells from a high-fat dietinduced glucotoxicity and prevents the development of type 2 diabetes. *Biochem Biophys Res Commun* (2013) 442(3-4):227–33. doi: 10.1016/j.bbrc.2013.11.023

97. Murphy B, Bhattacharya R, Mukherjee P. Hydrogen sulfide signaling in mitochondria and disease. *FASEB J* (2019) 33(12):13098–125. doi: 10.1096/fj.201901304R

98. Paul BD, Snyder SH. Gasotransmitter hydrogen sulfide signaling in neuronal health and disease. *Biochem Pharmacol* (2018) 149:101–9. doi: 10.1016/j.bcp.2017.11.019

99. Neavin DR, Liu D, Ray B, Weinshilboum RM. The role of the aryl hydrocarbon receptor (AHR) in immune and inflammatory diseases. *Int J Mol Sci* (2018) 19(12):3851–65. doi: 10.3390/ijms19123851.

100. Huai W, Zhao R, Song H, Zhao J, Zhang L, Zhang L, et al. Aryl hydrocarbon receptor negatively regulates NLRP3 inflammasome activity by inhibiting NLRP3 transcription. *Nat Commun* (2014) 5:4738. doi: 10.1038/ncomms5738

101. Chen G, Kroemer G, Kepp O. Mitophagy: An emerging role in aging and age-associated diseases. *Front Cell Dev Biol* (2020) 8:200. doi: 10.3389/ fcell.2020.00200

102. Guan Y, Wang Y, Li B, Shen K, Li Q, Ni Y, et al. Mitophagy in carcinogenesis, drug resistance and anticancer therapeutics. *Cancer Cell Int* (2021) 21(1):350. doi: 10.1186/s12935-021-02065-w

103. AM A, Ameenudeen S, Kumar A, Hemalatha S, Ahmed N, Ali N, et al. Emerging role of mitophagy in inflammatory diseases: Cellular and molecular episodes. *Curr Pharm Des* (2020) 26(4):485-91. doi: 10.2174/1381612826666200107144810

104. Kim MJ, Yoon JH, Ryu JH. Mitophagy: a balance regulator of NLRP3 inflammasome activation. *BMB Rep* (2016) 49(10):529-35. doi: 10.5483/bmbrep.2016.49.10.115

105. Jackson DN, Theiss AL. Gut bacteria signaling to mitochondria in intestinal inflammation and cancer. *Gut Microbes* (2020) 11(3):285–304. doi: 10.1080/19490976.2019.1592421

106. Long J, Song J, Zhong L, Liao Y, Liu L, Li X. Palmatine: A review of its pharmacology, toxicity and pharmacokinetics. *Biochimie* (2019) 162:176-84. doi: 10.1016/j.biochi.2019.04.008

107. Herrera-Aco DR, Medina-Campos ON, Pedraza-Chaverri J, Sciutto-Conde E, Rosas-Salgado G, Fragoso-Gonzalez G. Alpha-mangostin: Anti-inflammatory and antioxidant effects on established collagen-induced arthritis in DBA/1J mice. *Food Chem Toxicol* (2019) 124:300–15. doi: 10.1016/j.fct.2018.12.018

108. Kim SJ, Hong EH, Lee BR, Park MH, Kim JW, Pyun AR, et al. Alphamangostin reduced ER stress-mediated tumor growth through autophagy activation. *Immune Netw* (2012) 12(6):253–60. doi: 10.4110/in.2012.12.6.253

109. Zou W, Yin P, Shi Y, Jin N, Gao Q, Li J, et al. A novel biological role of alpha-mangostin *via* TAK1-NF-kappaB pathway against inflammatory. *Inflammation* (2019) 42(1):103–12. doi: 10.1007/s10753-018-0876-6

110. Williams TM, Leeth RA, Rothschild DE, Coutermarsh-Ott SL, McDaniel DK, Simmons AE, et al. The NLRP1 inflammasome attenuates colitis and colitisassociated tumorigenesis. *J Immunol* (2015) 194(7):3369–80. doi: 10.4049/ jimmunol.1402098

111. Yin J, Sheng B, Yang K, Sun L, Xiao W, Yang H. The protective roles of NLRP6 in intestinal epithelial cells. *Cell Prolif* (2019) 52(2):e12555. doi: 10.1111/ cpr.12555

112. Ratsimandresy RA, Indramohan M, Dorfleutner A, Stehlik C. The AIM2 inflammasome is a central regulator of intestinal homeostasis through the IL-18/ IL-22/STAT3 pathway. *Cell Mol Immunol* (2017) 14(1):127–42. doi: 10.1038/ cmi.2016.35

113. Carvalho FA, Nalbantoglu I, Aitken JD, Uchiyama R, Su Y, Doho GH, et al. Cytosolic flagellin receptor NLRC4 protects mice against mucosal and systemic challenges. *Mucosal Immunol* (2012) 5(3):288–98. doi: 10.1038/mi.2012.8

114. Rom S, Zuluaga-Ramirez V, Reichenbach NL, Erickson MA, Winfield M, Gajghate S, et al. Secoisolariciresinol diglucoside is a blood-brain barrier protective and anti-inflammatory agent: implications for neuroinflammation. *J Neuroinflammation* (2018) 15(1):25. doi: 10.1186/s12974-018-1065-0

115. Moree SS, Rajesha J. Investigation of *in vitro* and *in vivo* antioxidant potential of secoisolariciresinol diglucoside. *Mol Cell Biochem* (2013) 373(1-2):179–87. doi: 10.1007/s11010-012-1487-4

116. Catarino MD, Alves-Silva JM, Pereira OR, Cardoso SM. Antioxidant capacities of flavones and benefits in oxidative-stress related diseases. *Curr Top Med Chem* (2015) 15(2):105–19. doi: 10.2174/1568026615666141209144506

117. Seong MA, Woo JK, Kang JH, Jang YS, Choi S, Jang YS, et al. Oral administration of fermented wild ginseng ameliorates DSS-induced acute colitis by inhibiting NF-kappaB signaling and protects intestinal epithelial barrier. *BMB Rep* (2015) 48(7):419–25. doi: 10.5483/bmbrep.2015.48.7.039

118. Chaparala A, Tashkandi H, Chumanevich AA, Witalison EE, Windust A, Cui T, et al. Molecules from American ginseng suppress colitis through nuclear factor erythroid-2-Related factor 2. *Nutrients* (2020) 12(6):1850–60. doi: 10.3390/nu12061850

119. Ahn S, Siddiqi MH, Aceituno VC, Simu SY, Yang DC. Suppression of MAPKs/NF-kappaB activation induces intestinal anti-inflammatory action of ginsenoside Rf in HT-29 and RAW264.7 cells. *Immunol Invest* (2016) 45(5):439–49. doi: 10.3109/08820139.2016.1168830

120. Naganuma M. Treatment with indigo naturalis for inflammatory bowel disease and other immune diseases. *Immunol Med* (2019) 42(1):16–21. doi: 10.1080/25785826.2019.1599158

121. Hwang JI, Kim J, Park JW. Adverse events in Korean traditional medicine hospitals: A retrospective medical record review. *J Patient Saf* (2018) 14(3):157–63. doi: 10.1097/PTS.000000000000190

122. Shoba G, Joy D, Joseph T, Majeed M, Rajendran R, Srinivas PS. Influence of piperine on the pharmacokinetics of curcumin in animals and human volunteers. *Planta Med* (1998) 64(4):353–6. doi: 10.1055/s-2006-957450

123. Toden S, Theiss AL, Wang X, Goel A. Essential turmeric oils enhance antiinflammatory efficacy of curcumin in dextran sulfate sodium-induced colitis. *Sci Rep* (2017) 7(1):814. doi: 10.1038/s41598-017-00812-6

124. Liao JC, Tsai JC, Liu CY, Huang HC, Wu LY, Peng WH. Antidepressantlike activity of turmerone in behavioral despair tests in mice. *BMC Complement Altern Med* (2013) 13:299. doi: 10.1186/1472-6882-13-299

125. Yang C, Han M, Li R, Zhou L, Zhang Y, Duan L, et al. Curcumin nanoparticles inhibiting ferroptosis for the enhanced treatment of intracerebral hemorrhage. *Int J Nanomed* (2021) 16:8049–65. doi: 10.2147/IJN.S334965

126. Shaikh J, Ankola DD, Beniwal V, Singh D, Kumar MN. Nanoparticle encapsulation improves oral bioavailability of curcumin by at least 9-fold when compared to curcumin administered with piperine as absorption enhancer. *Eur J Pharm Sci* (2009) 37(3-4):223–30. doi: 10.1016/j.ejps.2009.02.019

127. Has C, Sunthar P. A comprehensive review on recent preparation techniques of liposomes. J Liposome Res (2020) 30(4):336-65. doi: 10.1080/08982104.2019.1668010

128. Lee MK. Liposomes for enhanced bioavailability of water-insoluble drugs: *In vivo* evidence and recent approaches. *Pharmaceutics* (2020) 12(3):264–93. doi: 10.3390/pharmaceutics12030264

129. Telange DR, Patil AT, Pethe AM, Fegade H, Anand S, Dave VS. Formulation and characterization of an apigenin-phospholipid phytosome (APLC) for improved solubility, *in vivo* bioavailability, and antioxidant potential. *Eur J Pharm Sci* (2017) 108:36–49. doi: 10.1016/j.ejps.2016.12.009

130. Nair AB, Jacob S. A simple practice guide for dose conversion between animals and human. *J Basic Clin Pharm* (2016) 7(2):27–31. doi: 10.4103/0976-0105.177703

131. Nair A, Morsy MA, Jacob S. Dose translation between laboratory animals and human in preclinical and clinical phases of drug development. *Drug Dev Res* (2018) 79(8):373–82. doi: 10.1002/ddr.21461