

# Risk for contralateral breast cancer among carriers of the *CHEK2*\*1100delC mutation in the WECARE Study

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The protein encoded by the *CHEK2* gene is involved in cellular repair of DNA damage. The truncating mutation, *CHEK2*\*1100delC, seems to increase the risk for breast cancer. We investigated whether the *CHEK2*\*1100delC mutation carrier status increases the risk for asynchronous contralateral breast cancer (CBC) and whether it interacts with radiation therapy (RT) or chemotherapy in regard to CBC risk. The germline mutation frequency was assessed in 708 women with CBC and 1395 women with unilateral breast cancer (UBC) in the Women's Environment, Cancer and Radiation Epidemiology (WECARE) Study whose first primary breast cancer was diagnosed before age 55 years and during 1985–1999. Seven women with CBC (1.0%) and 10 women with UBC (0.7%) were *CHEK2*\*1100delC variant carriers (rate ratio (RR) = 1.8, 95% confidence interval (CI) = 0.6–5.4 for CBC vs UBC). Carriers who received RT for their first breast cancer, compared with non-carriers not treated with RT, had an RR of developing CBC of 2.6 (95% CI = 0.8–8.7). We found no significant associations between the *CHEK2*\*1100delC mutation and CBC overall or among those treated with RT. However, the sampling variability was such that modest increases in risk could not be excluded. Nonetheless, because this is a rare mutation, it is unlikely to explain a major fraction of CBC in the population.

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The *CHEK2* gene codes for a protein that is involved in cell-cycle control and DNA repair. A protein-truncating mutation in the gene, 1100delC, was associated with a two-fold increase in breast cancer risk in the CHEK2 Breast Cancer Case–Control Consortium pooling project, which comprised 10 860 breast cancer cases and 9065 controls from 10 studies in five countries (The CHEK2 Breast Cancer Case–Control Consortium, 2004). The prevalence of the *CHEK2*\*1100delC variant among women with and without breast cancer varies among countries (Meijers-Heijboer *et al*, 2002; Vahteristo *et al*, 2002; Osorio *et al*, 2004; Huzarski *et al*, 2005; Kilpivaara *et al*, 2005; Bernstein *et al*, 2006; Weischer *et al*, 2007).

A higher prevalence of the mutation has been found among patients with bilateral breast cancer (Broeks *et al*, 2004; Kilpivaara *et al*, 2005) and among patients with a family history of breast cancer (Meijers-Heijboer *et al*, 2002; Vahteristo *et al*, 2002; Friedrichsen *et al*, 2004). Carriers of the *CHEK2*\*1100delC mutation are hypothesised to have an increased sensitivity to exposures such as radiation therapy (RT) and chemotherapy that cause DNA double-strand breaks, as the *CHEK2* regulates cell-cycle checkpoint response pathways that respond to such damage (Nevanlinna and Bartek, 2006). We investigated the risk for contralateral breast cancer (CBC) associated with the *CHEK2*\*1100delC germline mutation among approximately 2100 women with breast cancer who were cases and controls in the multicentre Women's Environment, Cancer and Radiation Epidemiology (WECARE) Study (Bernstein *et al*, 2004). In addition, we explored the joint effect of being a carrier of mutated *CHEK2* and having received RT or chemotherapy for breast cancer.

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<sup>14</sup>See Appendix.

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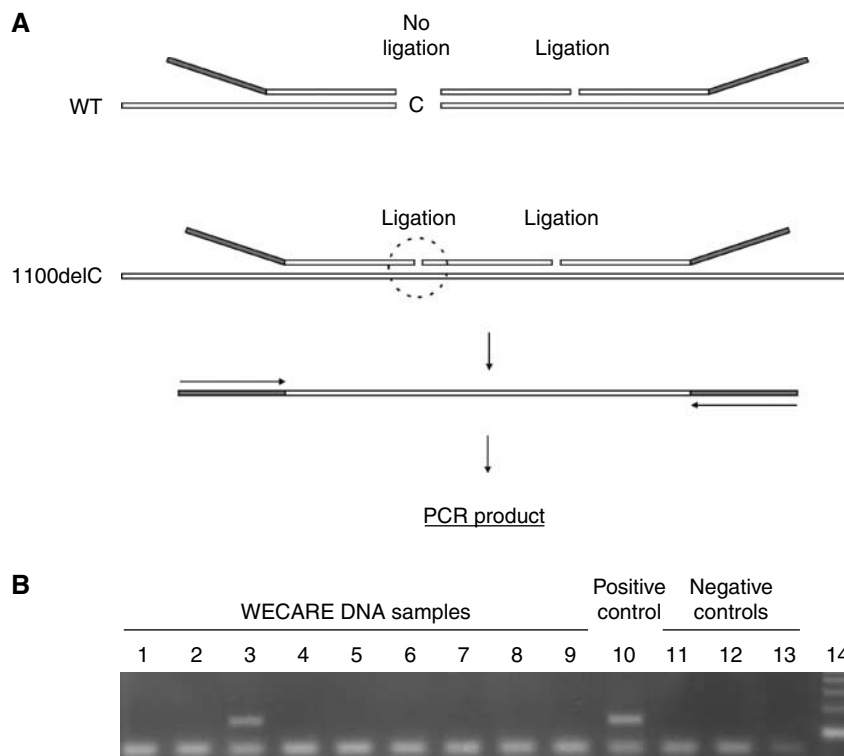
## MATERIALS AND METHODS

### Study population

Cases in this study were women with asynchronous CBC and controls were women with unilateral breast cancer (UBC) obtained from the WECARE Study (Bernstein *et al*, 2004). The participants were identified through five population-based cancer registries: four in the United States (Surveillance Epidemiology and End Results (SEER) registries in Iowa, Seattle, Los Angeles County and Orange County/San Diego) and one in Denmark (Danish Breast Cancer Cooperative Group (DBCG) registry supplemented by data from the Danish Cancer Registry). Case women were eligible for the study if they were under age 55 years at diagnosis of their first invasive breast cancer and were diagnosed with a second primary breast cancer in the contralateral breast (invasive or *in situ*) at least 1 year later, while residing in the same reporting area at the time of diagnosis of both cancers with no history of a prior cancer diagnosis or an intervening cancer diagnosis between their first and second primary breast cancers. All first primary breast cancers were invasive without distant metastases and were diagnosed between 1 January 1985 and 31 December 1999. The second primary breast cancer had to have been diagnosed no later than 31 December 2001. All women were alive at the time of contact. Control women had to fulfil the same eligibility criteria as cases except that they had no CBC during the interval between the matched case woman's first and second breast cancers ('at-risk' period) and had no prophylactic mastectomy during the 'at-risk' period. The reference date for case women was the date of diagnosis of their second primary breast cancer, whereas the corresponding reference date for control women was defined as their date of diagnosis of primary breast cancer plus the 'at-risk'

period for their matched case. Two UBC controls were individually matched to each CBC case on year of birth (5-year strata), year of diagnosis (4-year strata), registry region and race, and counter-matched on RT (as designated by the cancer registry), that is, each triplet comprised two women who had received RT and one woman who had not. Counter-matching was used to improve the statistical efficiency of the study design (Bernstein *et al*, 2004).

Characteristics of the first and second breast cancers were abstracted from medical records and cancer registry files. Medical records were retrieved to obtain detailed information on the treatment of the first breast cancer (RT, chemotherapy and hormonal therapy). By the use of methods previously described (Bernstein *et al*, 2004), the mean individual radiation dose to the contralateral breast was estimated as 1.29 Gy (range: 0.03–4.68 Gy) (estimation carried out for 85% of those who received RT). Among women treated with chemotherapy, 54% had received CMF, CF or CM (C, cyclophosphamide; M methotrexate; F, fluorouracil), 23% had received CAF, CEF, CMAF, CAM, CMEF or AC (A, adriamycin; E, epirubicin) and 23% had received other or unknown regimens. All the women were interviewed by telephone using a structured questionnaire to obtain information on known breast cancer risk factors. In addition, for the small subset of women for whom medical records did not give information on chemotherapy (7%) and/or hormonal therapy (10%), treatment information was obtained at the interview. Blood samples were drawn by study phlebotomists. Study participants were genotyped for mutations in *BRCA1* and *BRCA2* genes as previously described (Begg *et al*, 2008). All participants provided written informed consent before enrolment into the WECARE Study, and the research protocol was approved by the respective Institutional Review Boards and the ethical committee system in Denmark.



**Figure 1** Detection of *CHEK2*\*1100delC by oligonucleotide ligation. **(A)** Schematic of the method. Total genomic DNA is incubated with three oligonucleotides. On the *CHEK2*\*1100delC sequence, the oligonucleotides hybridise in juxtaposition on the target sequence and can be joined by ligation and subsequently amplified by PCR. On the wild-type *CHEK2* sequence, two of the oligonucleotides are separated by one base (C) and cannot be ligated into an amplifiable probe. **(B)** Analysis of WECARE samples. The controls include DNA samples with (lane 10) and without (lane 11) *CHEK2*\*1100delC, a ligation control without DNA (lane 12), and a negative PCR control (lane 13). Lane 14, 100 bp ladder.

Among 998 CBC patients (cases) and 2112 UBC controls who were eligible and approached to participate in the study, 708 (71%) cases and 1399 (66%) controls were enrolled. Matching was successfully carried out for 694 triplets (matching criteria were relaxed on region or race/ethnicity for 17 triplets) and 11 case-control pairs, whereas three cases had no matched controls. All 708 cases and 1395 of the controls (99.7%) were genotyped for *CHEK2\*1100delC* (four controls excluded).

### Genotyping of *CHEK2\*1100delC*

For detection of *CHEK2\*1100delC*, we developed an assay based on ligation-dependent amplification (Figure 1). The sequences of the oligonucleotides were 5'-[TATGTAAAACGACGGCCAGT]-TGGCAAGTCAACATTATTCCCTTTTGTACTGAATTTTAGATTA-3', 5'-TGATTTTGGGCACTCCAAGATTTTGGG-3' and 5'-AGAGACCTCTCATGAGAACCCTTATGTGGAACC-[ACCCAATTCGCCCTATAATA]-3'; the latter two oligonucleotides were 5'-phosphorylated. Approximately 50 ng of genomic DNA in 5 µl TE buffer was incubated at 98°C for 10 min and then cooled at 25°C. MLPA buffer (1.5 µl; MRC-Holland, Amsterdam, the Netherlands) and 2 fmol of each oligonucleotide were added, and the samples were then heated at 95°C for 1 min and incubated at 37°C for 16 h in a total volume of 8 µl. Ligation of the hybridised oligonucleotides was achieved by the addition of 32 µl of a ligase mix containing 3 µl ligase buffer A (MRC-Holland), 3 µl ligase buffer B (MRC-Holland), 1 U Taq DNA ligase (New England Biolabs, Beverly, MA, USA) and incubation at 54°C for 15 min. Ligation products were amplified with primers 5'-TATGTAAAACGACGGCCAGT-3'

and 5'-TATTATAGGGCGAATTGGGT-3'. Hybridisation, ligation and PCR were performed in 96-well plates, which contained two positive controls (DNA with *CHEK2\*1100delC*), two negative controls (DNA without *CHEK2\*1100delC*) and two ligation controls (without DNA). The PCR products were resolved by agarose gel electrophoresis.

The reliability of the ligation assay was confirmed by re-analysis of 248 DNA samples from WECARE Study participants using denaturing gradient gel electrophoresis (DGGE), as described previously (Bartkova *et al*, 2004). We found complete concordance between the results obtained with these two methods (3 out of 248 samples screened were positive for mutations). The remaining 1855 samples were then analysed with the ligation assay. All positive results were reconfirmed by DGGE analysis or with the ligation assay for samples where long-range amplification was unsuccessful.

### Statistical methods

Rate ratios (RRs) and corresponding 95% confidence intervals (CIs) were estimated by univariate conditional logistic regression models accounting for the counter-matched design (details described previously; Bernstein *et al*, 2004). Multivariable models were adjusted for exact age at diagnosis of the first primary breast cancer, age at menarche (<13/≥13 years), menopausal status at reference date (premenopausal/age at menopause <45 years/age at menopause ≥45 years), number of full-term pregnancies (0, 1, 2, 3, ≥4), family history of breast cancer among first-degree relatives (yes/no), lobular histology of the first primary (yes/no), stage of

**Table 1** Frequency of *CHEK2\*1100delC* carrier status among women with asynchronous CBC and UBC in the WECARE Study according to age at first breast cancer, race, study centre, family history of breast cancer and estrogen receptor status of first breast cancer

	Asynchronous CBC (n = 708)		UBC (n = 1395)	
	Carrier	Non-carrier	Carrier	Non-carrier
All (n and %)	7 (1.0)	701 (99.0)	10 (0.7)	1385 (99.3)
Age at first breast cancer (n and %)				
<45 years	2 (0.7)	289 (99.3)	6 (1.1)	562 (98.9)
≥45 years	5 (1.2)	412 (98.8)	4 (0.5)	823 (99.5)
Race (n and %)				
Non-Hispanic white	7 (1.1)	642 (98.9)	10 (0.8)	1276 (99.2)
Hispanic white	0 (—)	24 (100.0)	0 (—)	46 (100.0)
Black	0 (—)	21 (100.0)	0 (—)	39 (100.0)
Asian	0 (—)	13 (100.0)	0 (—)	22 (100.0)
Other	0 (—)	1 (100.0)	0 (—)	2 (100.0)
Study centre (n and %)				
Los Angeles County, CA	1 (0.5)	198 (99.5)	1 (0.3)	390 (99.7)
Seattle	1 (1.0)	98 (99.0)	0 (—)	193 (100.0)
Iowa	3 (2.7)	110 (97.3)	4 (1.8)	224 (98.2)
Orange County/San Diego, CA	0 (—)	118 (100.0)	2 (0.9)	227 (99.1)
Denmark	2 (1.1)	177 (98.9)	3 (0.8)	351 (99.2)
Family history of breast cancer (n and %)				
At least one first-degree relative with breast cancer <sup>a</sup>	3 (1.3)	223 (98.7)	5 (1.7)	284 (98.3)
At least one second-degree relative with breast cancer <sup>a</sup>	3 (1.1)	274 (98.9)	2 (0.5)	409 (99.5)
At least one relative with bilateral breast cancer <sup>a</sup>	1 (6.7)	14 (93.3)	0 (—)	18 (100.0)
Unknown family history, adopted	0 (—)	11 (100.0)	0 (—)	26 (100.0)
Estrogen receptor status, first breast cancer (n and %)				
Positive	4 (1.2)	334 (98.8)	4 (0.5)	739 (99.5)
Negative	3 (1.6)	190 (98.4)	3 (0.9)	335 (99.1)
Other or unknown	0 (—)	177 (100.0)	3 (1.0)	311 (99.0)

CBC = contralateral breast cancer; WECARE = Women's Environment, Cancer and Radiation Epidemiology; UBC = unilateral breast cancer. <sup>a</sup>Women may be in more than one of these categories.

first breast cancer (local/regional), RT (yes/no), chemotherapy (yes/no) and hormonal therapy (yes/no). As results of the univariate and multivariable models were quite similar, we present RRs from the univariate models. In addition, we present RRs from models adjusted for *BRCA1/2* carrier status (deleterious *BRCA1/2* mutations yes/no/unknown). The TPHREG procedure in SAS release 9.1 (SAS Institute Inc., Cary, NC, USA) was used for the statistical analyses. Further information about the SAS programming used for counter-matched WECARE data is provided in the appendix of Bernstein *et al* (2004).

## RESULTS

The mean length of time since first breast cancer diagnosis was 5.0 years (range: 1–15.6 years) for the total study population. Seven (1.0%) carriers of the *CHEK2\*1100delC* germline mutation were found among the 708 women with CBC, whereas 10 (0.7%) carriers were found among the 1395 women with UBC. Characteristics of carriers and non-carriers among cases and controls are shown in Table 1.

The association between being carrier of the *CHEK2\*1100delC* mutation and risk of developing CBC was not statistically significant (RR = 1.7; 95% CI = 0.6–5.1) (Table 2). Compared to women who did not carry the mutation and who did not receive RT for their first breast cancer, carriers who received RT as treatment had an RR of CBC of 2.6 (95% CI = 0.8–8.4). The RR of CBC among mutation carriers treated with chemotherapy was 1.3 (95% CI = 0.3–6.2) compared with non-carriers who did not receive chemotherapy. We had 90% statistical power at two-sided  $P < 0.05$  to exclude a RR of 6.0 for CBC associated with the *CHEK2\*1100delC* mutation overall, a RR of 6.9 for the combination of the mutation and RT and a RR of 13.1 for the combination of the mutation and chemotherapy. The RR for the *CHEK2\*1100delC* mutation and CBC overall was largely unchanged after the adjustment for *BRCA1/2* carrier status (RR = 1.8; 95% CI = 0.6–5.4) as were the RRs for the mutation in combination with RT or chemotherapy (Table 2).

Results of analyses restricted to non-Hispanic whites (649 women with CBC and 1286 women with UBC) were similar to those for all study subjects combined.

## DISCUSSION

In this population-based multicentre study, we found a low prevalence of *CHEK2\*1100delC* mutation carriers among 708 women with CBC (1.0%) and 1395 women with UBC (0.7%). Several studies have shown higher frequencies (ranging from 1.7 to 14.3%) of mutation among women with bilateral breast cancer (Oldenburg *et al*, 2003; Broeks *et al*, 2004; Kilpivaara *et al*, 2005; Chekmariova *et al*, 2006; Kwiatkowska *et al*, 2006); however, none of these were population-based. Two of these studies reported that the frequency of mutation was significantly increased among women with bilateral breast cancer compared to women with UBC (Broeks *et al*, 2004; Kilpivaara *et al*, 2005). Also, a small but prospective study reported a significant increase in risk for CBC among mutation carriers (de Bock *et al*, 2004). Three previous studies showing a high carrier prevalence among bilateral breast cancer cases were from the North European countries, the Netherlands (Oldenburg *et al*, 2003; Broeks *et al*, 2004) and Finland (Kilpivaara *et al*, 2005). Although the carrier prevalence among Danish WECARE Study participants is low relative to these prior reports, the prevalence among Danish participants with UBC is similar to that observed in a previous Danish study (Weischer *et al*, 2007). Differences in frequencies across studies may also be due to differences in study selection criteria, for example, restriction to certain ages at the onset of breast cancer and/or to subjects with family history defined in various ways. Only one of the previous studies on bilateral breast cancer was restricted to non-*BRCA1/2* mutation cases (Oldenburg *et al*, 2003). In contrast to all the previous studies on bilateral breast cancer, which were based on cases from cancer hospitals or genetics clinics, our study is the first that is population-based.

The *CHEK2\*1100delC* allele is hypothesised to interact with environmental factors to promote breast tumorigenesis. Ionising radiation is of particular interest because it induces DNA damage, and the protein of the *CHEK2* gene is a kinase that plays a role in response to such damage by delaying cell-cycle progression to facilitate DNA repair or even induce cell death (Nevanlinna and Bartek, 2006). The germline mutation 1100delC truncates the *CHEK2* protein, thereby abolishing the kinase function (Wu *et al*, 2001). Epidemiological evidence to substantiate increased radiosensitivity among carriers is, however, sparse (Bernstein *et al*, 2006; Broeks *et al*, 2007). Also, although a study conducted in

**Table 2** Risk for asynchronous CBC associated with *CHEK2\*1100delC* mutation carrier status overall and in combination with RT or chemotherapy for the first breast cancer

	No. with asynchronous CBC (n = 705 <sup>a</sup> )	No. with UBC (n = 1395)	Univariate <sup>b</sup>		Adjusted <sup>c</sup>		
			RR	95% CI	RR	95% CI	
<i>Mutation</i>							
No	698	1385	1.00	—	1.00	—	
Yes	7	10	1.7	0.6–5.1	1.8	0.6–5.4	
<i>Mutation × RT</i>							
No × no	359	263	1.00	—	1.00	—	
No × yes	339	1122	1.1	0.9–1.3	1.0	0.9–1.2	
Yes × no	2	2	0.6	0.1–4.2	0.7	0.1–5.4	
Yes × yes	5	8	2.6	0.8–8.4	2.6	0.8–8.7	
<i>Mutation × chemotherapy</i>							
No × no	381	622	1.00	—	1.00	—	
No × yes	317	763	0.6	0.5–0.7	0.6	0.5–0.7	
Yes × no	4	5	1.4	0.3–6.3	1.6	0.4–7.6	
Yes × yes	3	5	1.3	0.3–6.2	1.2	0.3–5.7	

CBC = contralateral breast cancer; CI = confidence interval; RR = rate ratio; RT = radiation therapy; UBC = unilateral breast cancer. <sup>a</sup>Three cases without matched controls excluded. <sup>b</sup>RRs adjusted for counter-matching sampling. <sup>c</sup>RRs adjusted for counter-matching sampling and *BRCA1/2* carrier status.

Belgium suggested that breast cancer patients with a *CHEK2\*1100delC* mutation are, in general, not characterised by a distinct enhanced chromosomal radiosensitivity, this conclusion was based on only four breast cancer cases with the mutation who were compared with healthy controls without the mutation (Baeyens *et al*, 2005). Our result, although not statistically significant, suggests that mutation carriers may have increased radiosensitivity.

The WECARE Study was designed to investigate interactions between genes and exposures that cause DNA double-strand breaks in a study population enriched with hereditary breast cancer cases, owing to the restriction to early-onset breast cancer, and a population heavily exposed to ionising radiation and chemotherapeutic drugs. Detailed information on RT and chemotherapy was obtained by a thorough review of medical records. As DNA samples from WECARE Study participants have also been screened for mutations in *BRCA1* and *BRCA2* genes, we were able to adjust the analyses for *BRCA* carrier status. Patients included in the study survived varying lengths of time from reference date to interview; however, cases and controls within each triplet survived approximately the same length of time. The prevalence of the mutation may have been affected if carriers of the *CHEK2\*1100delC* mutation had a worse survival than non-carriers. Unfortunately, we could not address this question, as we had no information on carrier status among the non-survivors who were not included in the study. Previous studies have shown a worse recurrence-free survival (Schmidt *et al*, 2007) and distant metastases-free survival (de Bock *et al*, 2004) among carriers compared to non-carriers, but overall survival did not differ by carrier status (de Bock *et al*, 2004).

Notably, we found a low carrier frequency for the *CHEK2\*1100delC* mutation in this large population-based sample of early-onset cases of CBC and UBC. The small number of carriers reduced our

ability to study the effects of the mutation on the development of CBC; and accordingly, we could not exclude the possibility that the *CHEK2\*1100delC* mutation is associated with modest increases in risk. In contrast, the rarity of the *CHEK2\*1100delC* mutation indicates that it plays a rather limited role in the aetiology of CBC in the population.

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## REFERENCES

- Baeyens A, Claes K, Willems P, De Ruyck K, Thierens H, Vral A (2005) Chromosomal radiosensitivity of breast cancer with a *CHEK2* mutation. *Cancer Genet Cytogenet* **163**: 106–112
- Bartkova J, Guldberg P, Gronbaek K, Koed K, Primdahl H, Moller K, Lukas J, Orntoft TF, Bartek J (2004) Aberrations of the Chk2 tumour suppressor in advanced urinary bladder cancer. *Oncogene* **23**: 8545–8551
- Begg CB, Haile RW, Borg A, Malone KE, Concannon P, Thomas DC, Langholz B, Bernstein L, Olsen JH, Lynch CF, Anton-Culver H, Capanu M, Liang X, Hummer AJ, Sima C, Bernstein JL (2008) Variation of breast cancer risk among *BRCA1/2* carriers: an investigation by the WECARE study group. *JAMA* **299**: 194–201
- Bernstein JL, Langholz B, Haile RW, Bernstein L, Thomas DC, Stovall M, Malone KE, Lynch CF, Olsen JH, Anton-Culver H, Shore RE, Boice Jr JD, Berkowitz GS, Gatti RA, Teitelbaum SL, Smith SA, Rosenstein BS, Borresen-Dale AL, Concannon P, Thompson WD (2004) Study design: evaluating gene-environment interactions in the etiology of breast cancer – the WECARE study. *Breast Cancer Res* **6**: R199–R214
- Bernstein JL, Teraoka SN, John EM, Andrulis IL, Knight JA, Lapinski R, Olson ER, Wolitzer AL, Seminara D, Whittemore AS, Concannon P (2006) The *CHEK2\*1100delC* allelic variant and risk of breast cancer: screening results from the breast cancer family registry. *Cancer Epidemiol Biomarkers Prev* **15**: 348–352
- Broeks A, Braaf LM, Huseinovic A, Nooijen A, Urbanus J, Hogervorst FBL, Schmidt MK, Klijn JGM, Russell NS, van Leeuwen FE, van't Veer LJ (2007) Identification of women with an increased risk of developing radiation-induced breast cancer: a case only study. *Breast Cancer Res* **9**: R26
- Broeks A, de Witte L, Nooijen A, Huseinovic A, Klijn JG, Van Leeuwen FE, Russell NS, van't Veer LJ (2004) Excess risk for contralateral breast cancer in *CHEK2\*1100delC* germline mutation carriers. *Breast Cancer Res Treat* **83**: 91–93
- CHEK2 Breast Cancer Case-Control Consortium (2004) *CHEK2\*1100delC* and susceptibility to breast cancer: a collaborative analysis involving 10,860 breast cancer cases and 9,065 controls from 10 studies. *Am J Hum Genet* **74**: 1175–1182
- Chekmarlova EV, Sokolenko AP, Buslov KG, Iyevleva AG, Ulibina YM, Rozanov ME, Mitiushkina NV, Togo AV, Matsko DE, Voskresenskiy DA, Chagunava OL, Devilee P, Cornelisse C, Semiglazov VF, Imyanitov EN (2006) *CHEK2* 1100delC mutation is frequent among Russian breast cancer patients. *Breast Cancer Res Treat* **100**: 99–102
- de Bock GH, Schutte M, Krol-Warmerdam EM, Seynaeve C, Blom J, Brekelmans CT, Meijers-Heijboer H, van Asperen CJ, Cornelisse CJ, Devilee P, Tollenaar RA, Klijn JG (2004) Tumour characteristics and prognosis of breast cancer patients carrying the germline *CHEK2\*1100delC* variant. *J Med Genet* **41**: 731–735
- Friedrichsen DM, Malone KE, Doody DR, Daling JR, Ostrander EA (2004) Frequency of *CHEK2* mutations in a population based, case-control study of breast cancer in young women. *Breast Cancer Res* **6**: R629–R635
- Huzarski T, Cybulski C, Domagala W, Gronwald J, Byrski T, Szwiec M, Woyke S, Narod SA, Lubinski J (2005) Pathology of breast cancer in women with constitutional *CHEK2* mutations. *Breast Cancer Res Treat* **90**: 187–189
- Kilpivaara O, Bartkova J, Eerola H, Syrjakoski K, Vahteristo P, Lukas J, Blomqvist C, Holli K, Heikkilä P, Sauter G, Kallioniemi OP, Bartek J, Nevanlinna H (2005) Correlation of *CHEK2* protein expression and c.1100delC mutation status with tumor characteristics among unselected breast cancer patients. *Int J Cancer* **113**: 575–580
- Kwiatkowska E, Skasko E, Niwinska A, Wojciechowska-Lacka A, Rachtan J, Molong L, Nowakowska D, Konopka B, Janiec-Jankowska A, Paszko Z, Steffen J (2006) Low frequency of the *CHEK2\*1100delC* mutation among breast cancer probands from three regions of Poland. *Neoplasma* **53**: 305–308
- Meijers-Heijboer H, van den Ouweland A, Klijn J, Wasielewski M, de Snoo A, Oldenburg R, Hollestelle A, Houben M, Crepin E, van Veghel-Plandsoen M, Elstrodt F, van Duijn C, Bartels C, Meijers C, Schutte M, McGuffog L, Thompson D, Easton D, Sodha N, Seal S, Barfoot R,

- Mangion J, Chang-Claude J, Eccles D, Eeles R, Evans DG, Houlston R, Murday V, Narod S, Peretz T, Peto J, Phelan C, Zhang HX, Szabo C, Devilee P, Goldgar D, Futreal PA, Nathanson KL, Weber B, Rahman N, Stratton MR (2002) Low-penetrance susceptibility to breast cancer due to CHEK2\*1100delC in noncarriers of BRCA1 or BRCA2 mutations. *Nat Genet* 31: 55–59
- Nevanlinna H, Bartek J (2006) The CHEK2 gene and inherited breast cancer susceptibility. *Oncogene* 25: 5912–5919
- Oldenburg RA, Kroeze-Jansema K, Kraan J, Morreau H, Klijn JG, Hoogerbrugge N, Ligtenberg MJ, van Asperen CJ, Vasen HF, Meijers C, Meijers-Heijboer H, de Bock TH, Cornelisse CJ, Devilee P (2003) The CHEK2\*1100delC variant acts as a breast cancer risk modifier in non-BRCA1/BRCA2 multiple-case families. *Cancer Res* 63: 8153–8157
- Osorio A, Rodriguez-Lopez R, Diez O, de la Hoya M, Martinez JJ, Vega A, Esteban-Cardenas E, Alonso C, Caldes T, Benitez J (2004) The breast cancer low-penetrance allele 1100delC in the CHEK2 gene is not present in Spanish familial breast cancer population. *Int J Cancer* 108: 54–56
- Schmidt MK, Tollenaar RA, de Kemp SR, Broeks A, Cornelisse CJ, Smit VT, Peterse JL, van Leeuwen FE, Van't Veer LJ (2007) Breast cancer survival and tumor characteristics in premenopausal women carrying the CHEK2\*1100delC germline mutation. *J Clin Oncol* 25: 64–69
- Vahteristo P, Bartkova J, Eerola H, Syrjakoski K, Ojala S, Kilpivaara O, Tamminen A, Kononen J, Aittomaki K, Heikkila P, Holli K, Blomqvist C, Bartek J, Kallioniemi OP, Nevanlinna H (2002) A CHEK2 genetic variant contributing to a substantial fraction of familial breast cancer. *Am J Hum Genet* 71: 432–438
- Weischer M, Bojesen SE, Tybjaerg-Hansen A, Axelsson CK, Nordestgaard BG (2007) Increased risk of breast cancer associated with CHEK2\*1100delC. *J Clin Oncol* 25: 57–63
- Wu X, Webster SR, Chen J (2001) Characterization of tumor-associated Chk2 mutations. *J Biol Chem* 276: 2971–2974

## Appendix

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