Original article

IncRNome: a comprehensive knowledgebase of human long noncoding RNAs

Deeksha Bhartiya^{1,†}, Koustav Pal^{2,†}, Sourav Ghosh³, Shruti Kapoor³, Saakshi Jalali¹, Bharat Panwar⁴, Sakshi Jain², Satish Sati³, Shantanu Sengupta³, Chetana Sachidanandan³, Gajendra Pal Singh Raghava⁴, Sridhar Sivasubbu³ and Vinod Scaria^{1,}*

¹GN Ramachandran Knowledge Center for Genome Informatics, CSIR Institute of Genomics and Integrative Biology, Mall Road, Delhi 110007, India, ²CSIR Open Source Drug Discovery Unit, Council of Scientific and Industrial Research, Anusandhan Bhavan, Delhi 110001, India, ³Genomics and Molecular Medicine, CSIR Institute of Genomics and Integrative Biology, Mall Road, Delhi 110007, India and ⁴Bioinformatics Centre, CSIR Institute of Microbial Technology, Sector 39-A, Chandigarh 160036, India

*Corresponding author: Tel: +91 9650466002; Fax: +91 11 27667471; Email: vinods@igib.in

[†]These authors contributed equally to this work.

Submitted 17 October 2012; Revised 8 April 2013; Accepted 11 April 2013

Citation details: Bhartiya, D., Pal, K., Ghosh, S., et al. IncRNome: a comprehensive knowledgebase of human long noncoding RNAs. Database (2013) Vol. 2013: article ID bat034; doi:10.1093/database/bat034.

The advent of high-throughput genome scale technologies has enabled us to unravel a large amount of the previously unknown transcriptionally active regions of the genome. Recent genome-wide studies have provided annotations of a large repertoire of various classes of noncoding transcripts. Long noncoding RNAs (lncRNAs) form a major proportion of these novel annotated noncoding transcripts, and presently known to be involved in a number of functionally distinct biological processes. Over 18 000 transcripts are presently annotated as lncRNA, and encompass previously annotated classes of noncoding transcripts including large intergenic noncoding RNA, antisense RNA and processed pseudogenes. There is a significant gap in the resources providing a stable annotation, cross-referencing and biologically relevant information. IncRNome has been envisioned with the aim of filling this gap by integrating annotations on a wide variety of biologically significant information into a comprehensive knowledgebase. To the best of our knowledge, lncRNome is one of the largest and most comprehensive resources for lncRNAs.

Database URL: http://genome.igib.res.in/IncRNome

Introduction

The availability of technology to annotate transcriptomes at the genome-scale and single-nucleotide resolution has in the recent years provided a new outlook at the transcribed regions within the Human genome (1–3). Contrary to the popular belief, a large number of genomic loci have been presently annotated to be transcriptionally active (4). Many of these regions do not have the potential to encode for functional proteins and thus constitute a class of transcripts, popularly annotated as noncoding RNA (5). The noncoding RNA transcripts have been classified into a number of subclasses, with the most popular classification being based on their size, such as the class of small noncoding RNAs, which include the well-annotated microRNAs (miRNAs) (6), small nucleolar RNAs (snoRNAs), long noncoding RNAs (lncRNAs) and so on.

Long noncoding RNAs (IncRNAs), by definition, are transcripts that are >200 nucleotides in length and do not have the potential to encode for proteins exceeding lengths of \geq 30 amino acids (7, 8). Transcriptome annotation in recent years has significantly expanded the repertoire of IncRNAs,

[©] The Author(s) 2013. Published by Oxford University Press.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/ licenses/by/3.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited. Page 1 of 6

not just in humans, but also in other model systems like mouse (9) and zebrafish (10, 11). Although noncoding transcripts with >200 nucleotide lengths have been clubbed together in a general classification of lncRNAs, the members of this class have significant differences in their biological function, genomic loci and regulation. This class includes previously known classes of ncRNAs including the large intergenic noncoding RNA, transcribed pseudogenes, antisense transcripts and several others, including the annotated classes of functionally distinct transcripts such as Xist, which is involved in X inactivation (12) and Hotair (13), involved in epigenetic regulation.

Functionally, the IncRNA class encompasses a wide variety of distinct functions like X-chromosome inactivation, modulation of chromatin structure, regulation of transcriptional and posttranscriptional processes and epigenetic modifications (14). The biological function of IncRNAs is modulated through interaction with other biomolecules in the cell, such as DNA, RNA and proteins (15). Recent evidence has also indicated putative regulatory roles for smaller RNAs processed from IncRNAs, as well as for IncRNAs themselves that harbor regulatory motifs (16, 17). IncRNAs could be regulated in a different way than protein-coding genes (18). Recent evidence also suggests the role of IncRNAs in several diseases including a number of cancers like lung cancers, colorectal and blood neoplasia (7, 19). Candidate IncRNAs like NEAT2 and MALAT1 have been studied in detail with their relations with metastasis in cancers (20-22). Additional candidates like ANRIL have been implicated in diseases like atherosclerosis, (23, 24) while a number of candidate genome-wide association loci map to regions presently annotated as IncRNA genes (25). It has been also suggested that a conceptual understanding of IncRNA as a function of the biological interactions would help to understand disease processes and develop potential drug targets (26).

There are several comprehensive databases for other ncRNAs like miRNAs (27–30), snoRNAs (31); however, there is a paucity of such databases integrating biologically significant annotations for IncRNAs. Although there are IncRNA databases coming up like IncRNAdb (32), NONCODE (33), etc., the extent of IncRNA annotations still remains stringent. IncRNome has been formulated to integrate annotations on a wide variety of biologically significant information into a comprehensive knowledgebase. To the best of our knowledge, IncRNome is one of the largest catalogs for IncRNAs till date, and is available online at the URL: http://genome.igib.res.in/IncRNome.

Database design and architecture

The lncRNome database has been designed keeping in mind both experimental and computational biologists, so as to provide ready access to biologically relevant data as

per the needs of a user. To this end, the structure was designed following consultation with a number of experimental and computational biologists. We created the database to serve as a comprehensive, user-friendly and biologically relevant knowledgebase on human IncRNAs built on MySQL 5.6 and having a PHP-based web interface. In brief, each IncRNA gene has a single page with basic linkouts to other relevant databases, annotation sets and relevant categories of information linked in tabs. Five categories of information are presently available linked with each IncRNA, which includes (i) General Information, (ii) Sequence and Structure, (iii) Interactions and Processing, (iv) Variations and Conservation and (v) Epigenetic Modifications. These categories are connected to the genome browser along with the conservation scores of all IncRNA transcripts (Supplementary File S1).

The category 'General Information' hosts information like the gene name, Ensembl gene ID, gene type, gene status, Ensembl transcript ID, transcript name, transcript type, transcript status, chromosome, strand and genomic loci, all of which have been fetched from Gencode release 12 (http://www.gencodegenes.org) (34). The gene names were used to map the HGNC ID, Refseq ID, Havana gene ID, Havana transcript ID, NCBI ID and chromosomal loci from HUGO Gene Nomenclature Committee website (35). The length was calculated using the genomic loci. The details about IncRNA description, disease associations, interactions, overexpression and references were manually curated through literature. The alternate transcripts were derived using in-house scripts and all IncRNAs were provided stable internal IDs.

The IncRNA sequences were downloaded from UCSC Genome Browser Database (36), and the structures were predicted using RNAfold version 1.8.5. Both the parenthesis structure and the minimum free energy structure predicted using the default parameters have been provided.

The third category comprises IncRNA interactions with proteins and other RNAs, IncRNA processing, predicted open reading frames (ORFs) and various motifs. The database hosts 937 quadruplex and 40 hairpins motifs present in IncRNAs. Both the motifs have been predicted using tools developed 'in-house', Quadfinder (37) and HairpinFetcher, respectively. It also hosts 3716 miRNA binding sites on IncRNA. More than 10000 binding sites for nine other proteins, which have been summarized in the section 'Datasets and Features', have been provided. These datasets have been mapped using PAR-CLIP (38) and CLIP-Seq datasets as described in the later sections. There are 6808 predicted protein-binding sites also provided in the database, which were predicted using Support Vector Machine-based evaluation of interaction propensities. The 1692 small RNA processing sites have also been provided as described in the sections below. The fourth category consists of 345351 genomic variations mapped to IncRNAs. The

database of single nucleotide polymorphisms (dbSNP) SNPs were downloaded from UCSC genome browser and mapped to IncRNAs. Conservation scores of 66 573 sites within IncRNAs have been provided in this category. The fifth category provides 11 790 epigenetic marks in the promoters of IncRNAs. The datasets were downloaded from the NIH Human Epigenome Roadmap project and mapped to IncRNA promoters. The detailed methods are available as Supplementary methods.

The database also features a comprehensive search option, which enables users to search through IncRNome using different keywords, such as, IncRNA names, Ensembl IDs, known targets, SNPs, diseases, etc. In addition, a separate browse option also allows users to browse the database through either using the chromosome numbers or different IncRNA biotypes. The database also features a genome browser, which can be used to browse through the genome for representative features and also provides a visual representation of the associated genomic annotations available within the database mentioned above along with the conservation scores of IncRNAs.

Datasets and features

Long noncoding RNA annotations

IncRNA annotations were derived from Gencode release 12 (http://www.gencodegenes.org) (34), which consists of 11790 IncRNA genes and 18855 transcripts. The IncRNAs transcripts are classified into 10 different biotypes, the statistics of which has been provided in the Figure 1. In addition, the datasets of IncRNAs and their HGNC IDs were derived from the Human Gene Nomenclature Committee website (35), which consisted of 1073 IncRNAs. Additional mappings were derived for 99 human IncRNAs from IncRNAdb and from literature through manual curation and overlapped with each other based on genomic coordinates (Figure 2). A stable internal ID is also provided for easy access and to enable cross-referencing between the different IDs regularly used by different sequence databases. The consensus IDs forms the primary reference key within IncRNome and has also been used to reference alternate transcript isoforms. Wherever appropriate, all IncRNAs have also been linked back to relevant databases such as Ensembl, HGNC and NCBI for quick cross-reference.

The manual annotation of the functionally characterized IncRNAs is provided, which includes information about the disease associations, expression and functional significance. The annotations are collected from literature surveys and manual curations.

Sequence and structure and motifs

The IncRNA sequences were downloaded from UCSC Genome Browser using genomic locations of individual

transcripts (36). RNA structures were computed using RNAfold with default parameters, which is part of the Vienna RNA package version 1.8.5. Our group has previously suggested the presence of G-quadruplex motifs in IncRNAs that could have potential regulatory functions (39). To enable researchers to further take up experiments in this area, predictions of potential G-quadruplex forming motifs in entire IncRNA transcripts predicted using Quadfinder have been included (37), as well as potential hairpin structures in the IncRNA have been identified using HairpinFetcher.

IncRNA processing

A recent study conducted by our lab has pointed to a subset of IncRNAs, which could be potentially processed to small RNAs having downstream regulatory functions by having a dual transcriptional output (40). The same analysis was replicated on the present large datasets of IncRNAs. In brief, smallRNA clusters were derived from DeepBase (41), a comprehensive database of smallRNA annotations derived from smallRNA sequencing experiments available in the public domain and overlaid on the IncRNA annotations to derive information on potential IncRNAs that could be processed to smallRNAs.

Protein-RNA interactions

Recent high-throughput experimental methods for analysis of interactions through pull down and sequencing techniques have provided critical insights into the landscape of protein-RNA interactions in the human genome (42). One of the major datasets of protein-RNA interactions is derived from PAR-CLIP (38) experiments for Argonaute (Ago) proteins, which are critical components of the RISC machinery involved in miRNA targeting. A comprehensive mapping of potential Ago binding sites in the IncRNA transcriptome is provided by mapping the reads to the human transcriptome. Experimental datasets also exist for other proteins including IGF2BP2, IGF2BP3, IGF2P1, PTB, PUM2, QKI, TNRC6A, TNRC6B and TNRC6C, which have also been mapped to the IncRNA transcripts. Because the number of experimental datasets for protein-RNA interactions is scarce, we also incorporated a computational prediction method involving Support Vector Machine-based prediction of residues in RNA, which could have probable propensity to interact with proteins (Panwar and Raghava 2012, unpublished results).

Genomic variations and conservation

Genome-wide association studies in the recent past have suggested disease associations, which could be modulated by lncRNAs (43). In addition, a number of genomic loci previously shown to be associated with diseases have now been indicated to fall within lncRNA gene loci. To facilitate further in-depth analysis and experimental validation of

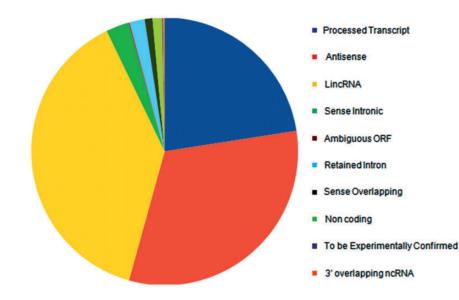


Figure 1. Distribution of Gencode release 12 IncRNAs according to different biotypes.

effect of variations on IncRNA, we have included a comprehensive mapping of genomic variations in IncRNA loci. In brief, the variations corresponding to dbSNP 135 were downloaded (44) and mapped to respective genomic locations of IncRNAs. In addition, disease associated variations were derived from the NIH Catalog of published genomewide association studies and mapped to respective rsIDs. The PhastCons conservation scores were downloaded from UCSC and the genomic loci were mapped to IncRNAs (45).

Epigenetic modifications

A recent report from our group suggests that the promoters of lncRNAs could be potentially regulated by mechanisms that are distinct from protein-coding genes supporting the role of lncRNAs in epigenetic regulation of genes. To capture the epigenetic marks, in terms of DNA methylation and histone marks, we have provided a comprehensive access to epigenetic marks in the promoters of lncRNAs. Briefly the raw datasets were downloaded from the NIH Human Epigenome Roadmap project and mapped and analyzed as described in Sati *et al.* (46). The epigenetic marks are also available for browsing through the genome browser. The datasets and genomic mappings are compiled in Table 1.

Predicted peptides

The open reading frames were predicted for all the lncRNAs using the Sixpack (http://www.ebi.ac.uk/Tools/st/ emboss_sixpack/) tool from EMBOSS. The tool translates the given sequence in six frames and peptides starting with Methionine and with length \geq 10 amino acids.

Conclusions and future perspectives

IncRNome is designed to primarily serve as an evidencebased resource of IncRNAs and their functionality in humans. To this end, we have provided stable reference IDs for IncRNA genes and alternate transcript isoforms of a gene with cross-references to other sequence and annotation databases to ensure interoperability and stable referencing. The knowledge base integrates biologically oriented datasets and resources on IncRNA and manual annotations wherever applicable with the aim of providing a one-stop solution for annotation information on IncRNAs.

The interface allows an easy access to various features of IncRNAs comprised within five categories and their sublevels (Supplementary Figure S1). The category 'General' provides all the basic annotations of each IncRNA including genomic loci, the associated diseases and various linkouts. The sequences and the predicted structure of the IncRNA are provided in the category 'Sequence and Structure'. The IncRNA structures are poorly understood and it becomes indispensable to characterize the structures to elucidate the structure-function relationships. Specific IncRNA structures are essential for binding to proteins, RNA and other biomolecules, and to have a better mechanistic insight of IncRNA function, elucidation of its structure becomes important. IncRNome provides information of various hairpin and quadruplex motifs in IncRNAs found to be essential for regulation of a lot of biological processes. Both experimental and prediction datasets on RNA-protein interactions have been provided for IncRNAs revealing various protein and RNA interacting partners of IncRNAs

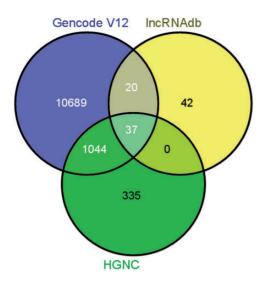


Figure 2. Comparison of annotations between other databases/datasets on long noncoding RNAs.

 Table 1. Total fields in the database along with the genomic loci mapped

Serial No.	Database fields	Total genomic loci mapped
1	Total IncRNAs	18855
2	Hairpins	40
3	Methylation and histone modifications	11 790
4	miRNA binding sites	3716
5	Quadruplexes	937
6	Predicted protein-binding sites on IncRNA	6808
7	Small RNA clusters	
8	Single nucleotide polymorphisms	295 851

(Supplementary Figure S1). Although the exact mechanism how the IncRNA interacts with different partners is still not known, our data provide a startup point to the community to understand the various regulatory interactions of IncRNAs with their respective partners. Genomic variations in IncRNAs have been studied to understand the effect of SNPs on biogenesis and functions of IncRNAs. The diseaseassociated SNPs present in IncRNAs might provide information about genotype to phenotype associations. The distribution of epigenetic marks like DNA methylation and histone modifications across transcription start site (TSS) of IncRNAs might help in evaluating the effect of chromatin modifications on gene expression (Supplementary Figure S1).

Because the field is emerging and many more IncRNAs are being discovered and annotated, thanks to the

availability of a large number of transcriptome sequencing datasets in public domain, IncRNome in the present form has many gaps. The primary gap being the paucity of information on expression of IncRNAs in different tissues. With the availability of genome-wide transcriptome annotation of many tissues in the public domain, we would enrich the database with this information. We intend to collaborate with other international consortiums to enable crosslinking and sharing of resources seamlessly. In future, we envisage the database to be available as a communitycurated and semantically linked interoperable data resource.

Supplementary Data

Supplementary data are available at Database Online.

Acknowledgements

The authors acknowledge Dr V. Sabareesh, Dr Harleen Kaur and Ms Ayesha Pasha for reviewing the manuscript. D.B. and S.S. acknowledge a Senior Research fellowship from CSIR, and S.K. acknowledges support through a Junior Research fellowship from CSIR. The authors acknowledge the funding for this work from the Council of Scientific and Industrial Research (CSIR), India through the project GENCODE-C (BSC00123).

Conflict of interest. None declared.

References

- Ozsolak, F. and Milos, P.M. (2011) RNA sequencing: advances, challenges and opportunities. *Nat. Rev. Genet.*, 12, 87–98.
- Pareek, C.S., Smoczynski, R. and Tretyn, A. (2011) Sequencing technologies and genome sequencing. J. Appl. Genet., 52, 413–435.
- 3. Zhang, J., Chiodini, R., Badr, A. et al. (2011) The impact of next-generation sequencing on genomics. J. Genet. Genomics, 38, 95–109.
- Clark, M.B., Amaral, P.P., Schlesinger, F.J. et al. (2011) The reality of pervasive transcription. PLoS Biol., 9, e1000625.
- 5. Ulveling, D., Francastel, C. and Hube, F. (2011) When one is better than two: RNA with dual functions. *Biochimie.*, **93**, 633–644.
- 6. Osman,A. (2012) MicroRNAs in health and disease–basic science and clinical applications. *Clin. Lab.*, **58**, 393–402.
- Li,X., Wu,Z., Fu,X. et al. (2012) Long noncodingRNAs: insights from biological features and functions to diseases. *Med. Res. Rev.*, 10, 517–553.
- 8. Mercer, T.R., Dinger, M.E. and Mattick, J.S. (2009) Long non-coding RNAs: insights into functions. *Nat. Rev. Genet.*, **10**, 155–159.
- 9. Guttman, M., Amit, I., Garber, M. *et al.* (2009) Chromatin signature reveals over a thousand highly conserved large non-coding RNAs in mammals. *Nature*, **458**, 223–227.
- Pauli, A., Valen, E., Lin, M.F. et al. (2012) Systematic identification of long noncoding RNAs expressed during zebrafish embryogenesis. *Genome Res.*, 22, 577–591.

- 11. Ulitsky,I., Shkumatava,A., Jan,C.H. *et al.* (2011) Conserved function of lincRNAs in vertebrate embryonic development despite rapid sequence evolution. *Cell*, **147**, 1537–1550.
- Brockdorff,N. (2011) Chromosome silencing mechanisms in X-chromosome inactivation: unknown unknowns. *Development*, 138, 5057–5065.
- 13. Gutschner, T. and Diederichs, S. (2012) The Hallmarks of Cancer: a long non-coding RNA point of view. *RNA. Biol.*, **9**, 703–719.
- 14. Derrien, T., Guigo, R. and Johnson, R. (2011) The long non-coding RNAs: a new (P)layer in the 'Dark Matter'. Front Genet., 2, 107.
- Wilusz,J.E., Sunwoo,H. and Spector,D.L. (2009) Long noncoding RNAs: functional surprises from the RNA world. *Genes Dev.*, 23, 1494–1504.
- Wang,K.C. and Chang,H.Y. (2011) Molecular mechanisms of long noncoding RNAs. *Mol. Cell*, 43, 904–914.
- 17. Yan, B. and Wang, Z. (2012) Long Noncoding RNA: its physiological and pathological roles. *DNA Cell Biol.*, **31** (Suppl. 1), S34–S41.
- Moran, V.A., Perera, R.J. and Khalil, A.M. (2012) Emerging functional and mechanistic paradigms of mammalian long non-coding RNAs. *Nucleic Acids Res.*, 40, 6391–6400.
- 19. Harries, L.W. (2012) Long non-coding RNAs and human disease. *Biochem. Soc. Trans.*, **40**, 902–906.
- 20. Tani,H., Nakamura,Y., Ijiri,K. *et al.* (2010) Stability of MALAT-1, a nuclear long non-coding RNA in mammalian cells, varies in various cancer cells. *Drug Discov. Ther.*, **4**, 235–239.
- Xu,C., Yang,M., Tian,J. et al. (2011) MALAT-1: a long non-coding RNA and its important 3' end functional motif in colorectal cancer metastasis. Int. J. Oncol., 39, 169–175.
- Kim,Y.S., Hwan,J.D., Bae,S. et al. (2010) Identification of differentially expressed genes using an annealing control primer system in stage III serous ovarian carcinoma. BMC Cancer, 10, 576.
- Congrains, A., Kamide, K., Katsuya, T. et al. (2012) CVD-associated non-coding RNA, ANRIL, modulates expression of atherogenic pathways in VSMC. Biochem. Biophys. Res. Commun., 419, 612–616.
- 24. Holdt,L.M. and Teupser,D. (2012) Recent studies of the human chromosome 9p21 locus, which is associated with atherosclerosis in human populations. *Arterioscler. Thromb. Vasc. Biol.*, **32**, 196–206.
- Pasmant, E., Sabbagh, A., Vidaud, M. et al. (2011) ANRIL, a long, noncoding RNA, is an unexpected major hotspot in GWAS. FASEB J., 25, 444–448.
- Bhartiya, D., Kapoor, S., Jalali, S. *et al.* (2012) Conceptual approaches for IncRNA drug discovery and future strategies. *Expert. Opin. Drug Discov.*, 7, 503–513.
- Griffiths-Jones, S., Grocock, R.J., van Dongen, S. et al. (2006) miRBase: microRNA sequences, targets and gene nomenclature. *Nucleic Acids Res.*, 34(Database issue), D140–D144.
- Hsu,S.D., Chu,C.H., Tsou,A.P. et al. (2008) miRNAMap 2.0: genomic maps of microRNAs in metazoan genomes. *Nucleic Acids Res.*, 36(Database issue), D165–D169.

- Kin, T., Yamada, K., Terai, G. et al. (2007) fRNAdb: a platform for mining/annotating functional RNA candidates from non-coding RNA sequences. Nucleic Acids Res., 35(Database issue), D145–D148.
- Wang,X. (2008) miRDB: a microRNA target prediction and functional annotation database with a wiki interface. *RNA*, 14, 1012–1017.
- 31. Ellis, J.C., Brown, D.D. and Brown, J.W. (2010) The small nucleolar ribonucleoprotein (snoRNP) database. *RNA*, **16**, 664–666.
- Amaral,P.P., Clark,M.B., Gascoigne,D.K. et al. (2011) IncRNAdb: a reference database for long noncoding RNAs. Nucleic Acids Res., 39(Database issue), D146–D151.
- Bu,D., Yu,K., Sun,S. et al. (2012) NONCODE v3.0: integrative annotation of long noncoding RNAs. Nucleic Acids Res., 40(Database issue), D210–D215.
- Harrow, J., Denoeud, F., Frankish, A. et al. (2006) GENCODE: producing a reference annotation for ENCODE. Genome Biol., 7 (Suppl. 1), 54.1–54.9.
- Povey, S., Lovering, R., Bruford, E. et al. (2001) The HUGO gene nomenclature committee (HGNC). Hum. Genet., 109, 678–680.
- Kent,W.J., Sugnet,C.W., Furey,T.S. et al. (2002) The human genome browser at UCSC. Genome Res., 12, 996–1006.
- Scaria, V., Hariharan, M., Arora, A. et al. (2006) Quadfinder: server for identification and analysis of quadruplex-forming motifs in nucleotide sequences. Nucleic Acids Res., 34, W683–W685.
- Jayaraj,G.G., Pandey,S., Scaria,V. et al. (2012) Potential G-quadruplexes in the human long non-coding transcriptome. RNA. Biol., 9, 81–86.
- Ascano, M., Hafner, M., Cekan, P. et al. (2012) Identification of RNAprotein interaction networks using PAR-CLIP. Wiley Interdiscip. Rev. RNA, 3, 159–177.
- 40. Jalali,S., Jayaraj,G.G. and Scaria,V. (2012) Integrative transcriptome analysis suggest processing of a subset of long non-coding RNAs to small RNAs. *Biol. Direct*, **7**, 25.
- Yang, J.H. and Qu, L.H. (2012) DeepBase: annotation and discovery of microRNAs and other noncoding RNAs from deep-sequencing data. *Methods Mol. Biol.*, 822, 233–248.
- Konig, J., Zarnack, K., Luscombe, N.M. et al. (2012) Protein-RNA interactions: new genomic technologies and perspectives. Nat. Rev. Genet., 13, 77–83.
- Jin,G., Sun,J., Isaacs,S.D. et al. (2011) Human polymorphisms at long non-coding RNAs (IncRNAs) and association with prostate cancer risk. Carcinogenesis, 32, 1655–1659.
- Sherry,S.T., Ward,M.H., Kholodov,M. et al. (2001) dbSNP: the NCBI database of genetic variation. Nucleic Acids Res., 29, 308–311.
- Siepel, A., Bejerano, G., Pedersen, J.S. et al. (2005) Evolutionarily conserved elements in vertebrate, insect, worm, and yeast genomes. *Genome Res.*, 15, 1034–1050.
- Sati,S., Ghosh,S., Jain,V. et al. (2012) Genome-wide analysis reveals distinct patterns of epigenetic features in long non-coding RNA loci. Nucleic Acids Res., 40, 10018–10031.