

Antitumor Effect of Hyperoside Loaded in Charge Reversed and Mitochondria-Targeted Liposomes

Yufei Feng¹
Guozhao Qin¹
Shuyuan Chang¹
Zhongxu Jing²
Yanyan Zhang¹
Yanhong Wang¹

¹Key Laboratory of Chinese Materia Medica in Ministry of Education, Heilongjiang University of Chinese Medicine, Harbin, Heilongjiang, People's Republic of China; ²Heilongjiang Provincial Administration of Traditional Chinese Medicine, Harbin, Heilongjiang, People's Republic of China

Introduction: Hyperoside (HYP), a flavonol glycoside compound, has been shown to significantly inhibit the proliferation of malignant tumors. Mitochondria serve as both “energy factories” and “suicide weapon stores” of cells. Targeted delivery of cytotoxic drugs to the mitochondria of tumor cells and tumor vascular cells is a promising strategy to improve the efficacy of chemotherapy.

Objective: We report a novel dual-functional liposome system possessing both extracellular charge reversal and mitochondrial targeting properties to enhance drug accumulation in mitochondria and trigger apoptosis of cancer cells.

Methods: L-lysine was used as a linker to connect 2,3-dimethylmaleic anhydride (DMA) and 1,2-distearoyl-sn-glycero-3-phosphoethanolamine (DSPE) to yield a new compound, DSPE-Lys-DMA (DLD). Then, DLD was mixed with other commercially available lipids to form charge reversed and mitochondria-targeted liposomes (DLD-Lip). The size, morphology, zeta potential, serum stability, and protein adsorption of the HYP loaded DLD-Lip (HYP/DLD-Lip) were measured. The release profile, cellular uptake, in vitro and in vivo toxicity, and anticancer activity of HYP/DLD-Lip were investigated.

Results: The results showed that the mean diameter of the liposomes was less than 200 nm. The zeta potential of the liposomes was negative at pH 7.4. However, the zeta potential was positive at weak acidic pH values with the cleavage of the DMA amide. The charge reversion of HYP/DLD-Lip facilitated the cellular internalization and mitochondrial accumulation for enhanced antitumor effect. The strongest tumor growth inhibition (TGI 88.79%) without systemic toxicity was observed in DLD/HYP-Lips-treated CBRH-7919 tumor xenograft BALB/C mice.

Conclusion: The charge reversed and mitochondria-targeted liposomes represented a promising anticancer drug delivery system for enhanced anticancer therapeutic efficacy.

Keywords: mitochondria-targeted, liposome, charge reversal, antitumor, hyperoside

Introduction

Traditional Chinese medicine has a long history of tumor treatment, including paclitaxel and vincristine, which have significant antitumor effects and have been widely used clinically.¹⁻³ In recent years, HYP, a flavonol glycoside compound extracted from *Hypericum perforatum* and *Phyllanthus rhizoma*, have been shown to significantly inhibit the proliferation of malignant tumors.^{4,5} The anti-tumor mechanism of HYP occurs primarily through the regulation of calcium-related mitochondrial pathways.⁶ Mitochondrial calcium ion overload can induce cytochrome C and apoptosis-inducing factors to be released from the mitochondria into the cytoplasm, eventually leading to apoptosis.^{7,8} However, as with most

Correspondence: Yanhong Wang
Tel +86451-87266893
Email 799378826@qq.com

chemotherapy drugs, some disadvantages of HYP, such as low solubility and lack of mitochondrial targeting, limit its further application.^{9,10} Therefore, it is urgently important to design and prepare a safe and effective drug delivery system to improve the anti-tumor effect of HYP.

Designing a tumor microenvironment-stimulated drug delivery system based on the unique microenvironment of tumor tissue, such as stimulating responsive nanoparticles, microcapsules, polymer-drug conjugates, and prodrugs can be expected to improve the effect of anticancer drugs.^{11–13} Liposomes, which are often used as carriers of anti-tumor drugs, are vesicles with a cell membrane structure composed of phospholipids and cholesterol. The carrier, because of its good biocompatibility and degradability, can be enhanced by penetration and retention (EPR) in the tumor. This has been extensively studied because of the accumulation of tumor sites and the ability to load hydrophilic and hydrophobic drugs.^{14,15} However, with circulation in the blood stream, liposomes are easily captured and cleared by the reticuloendothelial system and the mononuclear macrophage system. Polyethylene glycol (PEG) modification can increase the circulation time of liposomes in the blood,^{16,17} but its existence will affect the efficiency of drug entry into the cell. Because the metabolism of tumor tissues mainly depends on anaerobic glycolysis, its metabolites produced by respiration, lactic acid and CO₂, cause the acidification of the tumor microenvironment. Tumor tissue has the characteristics of low pH, which is manifested in the lower pH value (6.8) of tumor cell interstitium compared with the pH value (7.4) of normal tissue; the pH value of lysosomes is 4.5.¹⁸ This allows for the preparation of liposomes with controllable pH-responsive to increase the possibility of anticancer activity. Accordingly, more studies have focused on the functionalization of liposomes with a PEG deshielding mechanism with long circulation characters by a degradable pH-sensitive bond between PEG and lipids, such as hydrazone and hydrazine bonds.^{19–25} In an acidic environment, the rupture of acid-sensitive chemical bonds will remove PEG on the liposome surface and enhances cellular uptake.²⁶ Liposomes also can be decorated with carboxylic acid amide²⁷ or maleic anhydride amide,^{28,29} which can be used to achieve charge reversed effect. In blood, liposomes with negative charge can avoid non-specific protein adsorption to prolong the circulation time. However, when these liposomes exposed in the weakly acidic environment of the tumor tissue, the amide bond will be destroyed to induce positive charge on the

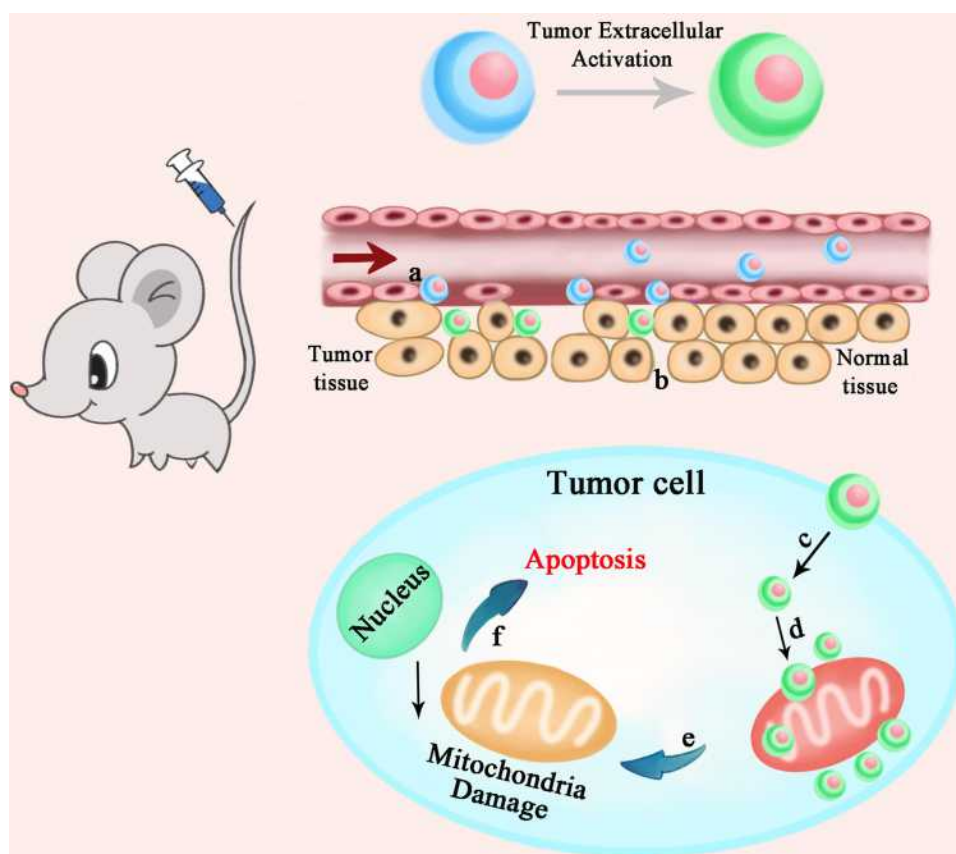
surface of the liposomes, which significantly increases cellular internalization of liposomes.³⁰

Based on the characteristics of the tumor tissue micro-environment and effect of HYP on mitochondria, this study constructed a new type of liposomal drug delivery system. A pH-responsive DSPE-Lys-DMA (DLD) lipid was designed. L-lysine was used as a linker to connect 2, 3-dimethylmaleic anhydride (DMA) and 1,2-distearoyl-sn-glycero-3-phosphoethanolamine (DSPE) to yield a new compound, DLD. The pH-responsive DLD was mixed with other commercially available lipids to form charge reversed, mitochondria-targeted liposomes (DLD-Lip). At pH 7.4, liposomes are negatively charged due to the carboxyl group in 2, 3-dimethylmaleic anhydride. When DLD-Lip accumulated in tumor tissues, the weak acidity in the microenvironment destroyed the dimethylmaleamide bond and the positively charged amino groups were exposed and the surface charge of DLD-Lips was reversed from a negative to positive charge, generating an electrostatic attraction with the tumor cell membrane, which increased the uptake of tumor cells. Then, the electrostatic interaction between highly positively charged liposomes and the mitochondrial membrane (130 ~ 150 mV, internal negative charge) caused it to accumulate in the mitochondria to achieve mitochondrial targeting, and finally triggering HYP to play its therapeutic effect (Scheme 1).

Materials and Methods

Materials

Hyperoside reference standard (batch number: 111521–201708) was purchased from the China Institute for the Control of Pharmaceutical and Biological Products. 1,2-Distearoyl-sn-glycero-3-phosphoethanolamine-N-[methoxy-(polyethylene glycol)-2000] (DSPE-mPEG₂₀₀₀) was purchased from Shanghai Fansheng Biological Technology Co., Ltd. Fetal Bovine Serum was purchased from Zhejiang Tianhang Biological Technology Co., Ltd. Rat liver cancer cell line CBRH-7919 cells were purchased from Shanghai Lianmai Biological Engineering Co., Ltd. Mitochondrial stripping, Caspase 3 activity assay kit, Caspase 9 activity assay kit, and Annexin V-FITC apoptosis detection kit were purchased from Biyuntian Biotechnology Co., Ltd. Female BALB/c nude mice were purchased from the Beijing Weitong Lihua Experimental Animal Technology Co., Ltd. (Beijing, China, license number: SCXK (Beijing) 2016–0006, initial weight 18–22 g), and placed in the



Scheme 1 Illustration of dual-functional liposome (DLD-Lip) with pH response to the tumor microenvironment and mitochondrial-targeting anticancer drug delivery. a) pH-response of DLD in mildly acidic tumor microenvironment for surface charge conversion. b) Positive charge of liposome improved cellular uptake. c) Internalization of liposomes into tumor cells. d) Binding to mitochondria. e) HYP play its therapeutic effect in mitochondria. f) Mitochondria apoptosis.

animal experiment center. The relevant operations of all experimental animals were approved and carried out in accordance with the relevant requirements of the Animal Experimental Management Committee of Heilongjiang University of Traditional Chinese Medicine.

Synthesis of DSPE-Lys-DMA (DLD)

Synthesis of DSPE-Lys

DSPE (1.50 g, 2.00 mmol) was dissolved in anhydrous trichloromethane (30 mL) under nitrogen atmosphere. DIPEA (2.00 mL, 12.00 mmol) was added to the solution under stirring at 0°C. Boc-L-Lys(Boc)-OH (0.83 g, 2.40 mmol), HOBT (0.32 g, 2.40 mmol), and HBTU (0.91 g, 2.40 mmol) dissolved in anhydrous DMF (5 mL) was added to the reaction flask containing DSPE. The solution was stirred under nitrogen in an ice bath for 30 min and at room temperature for another 24 h. The solvents were removed using rotary evaporation and the mixture was purified via column chromatography (silica, CH₂Cl₂: MeOH = 10:1). The product DSPE-(Boc)-L-Lys-Boc was dried in a vacuum.

The received DSPE-(Boc)-L-Lys-Boc was dissolved in anhydrous dichloromethane (DCM)/TFA (1:1, 10 mL). The solution was stirred under nitrogen for 4 h. The solvents were removed via rotary evaporation, and the sample was then dried in vacuum for 1 h. The product was precipitated in anhydrous diethyl ether. The white precipitate was three times washed with anhydrous diethyl ether and collected using centrifugation. DSPE-Lys, with a yield of 80.23%, was obtained after vacuum drying. The ¹H NMR spectrum of DSPE Lys was shown in [Supporting Information](#).

Synthesis of DSPE-Lys-DMA

Under the protection of nitrogen, DSPE-Lys (1.00 g, 1.14 mmol) was dissolved in 20 mL of anhydrous dichloromethane. In an ice bath, Triethanolamine (TEA, 0.58 g, 5.71 mmol) and DMA (0.57 g, 4.56 mmol) dissolved in dichloromethane were injected into the reaction system, and the reaction system was stirred at room temperature for 24 hours. After the reaction, the organic solvent was removed and the crude product was recrystallized in

acetone solution at 4°C. Finally, the obtained precipitate DSPE-Lys-DMA was washed three times with ether and then dried in a vacuum. The yield was 30%. H-Nuclear Magnetic Resonance (¹H-NMR) and Electrospray Ionization-Mass Spectrometry (ESI-MS) were used to confirm the structure of DSPE-Lys. The ¹H NMR spectrum of DSPE-Lys-DMA was shown in [Supporting Information](#).

Preparation and Characterization of Liposomes

HYP-loaded liposomes were prepared using the thin-film hydration method. Soy lecithin (SPC), cholesterol, DLD, DSPE-mPEG₂₀₀₀ and HYP (mass ratio, 8:2:2.5:1.5:1) were dissolved in a round bottom flask using chloroform/methanol as a mixed solvent (2:1, v/v). The organic solvent was removed using a rotary evaporator to form a homogeneous film on the bottom of the bottle and it was dried under vacuum overnight. The overnight-dried film was hydrated in phosphate buffered saline (PBS, pH 7.4) or ultrapure water to a phospholipid concentration of 1.6 mg/mL. The liposomes were sonicated in a water bath at 37°C for 10 minutes, then further probed at 40 W for 100 s to prepare DLD/HYP-Lip. DLD/HYP-Lip (0.5 mL) was added to 2.0 mL of ultrapure water, placed in an ultrafiltration centrifuge tube, and centrifuged at 4000 rpm for 20 min. Then, the lower layer of the solution was removed and the concentration of HYP was determined by high-performance liquid chromatography (HPLC). The encapsulation efficiency and drug loading were calculated according to the following formulas:

$$EE\% = W/W_0 \times 100\%$$

$$DL\% = W_{drug}/W_{total} \times 100\%$$

W: content of HYP in the liposome after centrifugation

W₀: content of HYP in the liposome

W_{drug}: the weight of HYP

W_{total}: the weight of HYP, cholesterol, and phospholipids

The average particle size of the liposomes was measured using a Dynamic Light Scattering (DLS) Analyzer. For the sample, 100 μL liposomes (phospholipid concentration 5 mg/mL) were diluted in 1 mL buffer solution.

DLD/HYP-Lip (200 μL) was diluted to a certain multiple, added to a special copper mesh, and stained with phosphotungstic acid. The sample was then air dried, followed by observation of the shape of the particles under a transmission electron microscope.

The pH-Induced Potential Changes

To investigate the potential change of DLD/-Lip in different pH buffer solutions, 1 mL of SPC-Lip and DLD/-Lip were added to 15 mL of PBS solution (pH 7.4, pH 6.8) and acetic acid buffer solution (incubate at pH 5.5 and pH 4.5) at 37°C, at 0, 2, 4, 6, 8, and 12 h. A 1.5 mL sample was taken for potential measurement.

Differential Scanning Calorimetry Analysis

To study the interaction between drugs and other excipients in the preparation of liposomes, the crystal characteristics of HYP and other excipients were analyzed by differential scanning calorimetry (DSC). An appropriate amount of the sample was weighed and placed in an aluminum crucible, and the crucible cap was sealed. A blank crucible was used as a reference. The temperature ranged from 40°C to 300°C, the heating rate was 20°C/min, and the shielding gas (high purity nitrogen) flow rate was 60 mL/min. SPC, cholesterol, DSPE-PEG, DLD, HYP, DLD/HYP-Lip, and DLD/HYP-Lip physical mixed samples were analyzed, and the differential scanning calorimeter characteristic curves of each sample were recorded.

Serum Stability Study

The stability of the liposomes in serum was assessed by measuring the change in particle size of SPC-Lip and DLD-Lip in fetal bovine serum. Liposomes with a phospholipid concentration of 5 mg/mL were mixed with an equal volume of fetal bovine serum and incubated at 37°C. At 1, 4, 8, and 24 h, 100 μL of the sample was taken for particle size determination. The liposome particle size at 0 h is the particle size measured in a PBS solution at pH 7.4.

Liposome Adsorption Test

Bovine serum albumin (BSA) was used as a model protein to detect the protein adsorption capacity of liposomes. The liposomes were placed in a pH 7.4 or 6.8 phosphate buffer solution (phospholipid concentration of 0.15 mg/mL, protein concentration of 0.25 mg/mL) containing BSA and incubated at 3°C for 2 h or 4 h, respectively. Three hundred microliters of the solution was centrifuged at 13,000 × g for 15 min. The aggregates of adsorbed protein settled at the bottom, and the supernatant contains unadsorbed protein. BSA was used as the standard solution and the

protein content in the supernatant was determined using the bicinchoninic acid (BCA) protein kit. Then, 25 μL of the sample was added to a 96-well plate and 200 μL of the BCA reaction solution was added. The microplate reader measures the absorbance at 570 nm.

Determination of in vitro Release Rate

The in vitro release of DLD/HYP-Lip in the release medium was determined by dialysis. Specifically, 0.1 mL DLD/HYP-Lip and free HYP solution (HYP content 0.5 mg/mL) was added to a pre-treated dialysis bag (molecular weight cut off 6000–8000 Da) and placed in 50 mL of release medium. The sample was then centrifuged at 50 rpm at 37°C. The release medium was 0.1% Tween 80 in PBS solution (pH 7.4, 6.8) and 0.1% Tween 80 acetate buffer solution (pH 5.5, 4.5). At 0.5, 1, 2, 4, 8, 12, and 24 h, 0.5 mL of the release medium was collected and equal volumes of fresh release medium were added to the drug release medium. The content of HYP in the sample was measured, and the cumulative release at each time point was calculated according to the following formula. Then a release curve was drawn.

$$E_r = \frac{V_e \sum_{i=1}^{n-1} C_i + V_0 C_n}{M_{\text{drug}}} \times 100\%$$

Where E_r represents the cumulative release of HYP, V_e represents the displacement volume of the release medium, C_i represents the concentration of the drug released during the i -th replacement sampling, V_0 represents the volume of the release medium, and C_n represents the release medium in the n th sampling. The drug concentration, M_{drug} , represents the HYP content in the liposome.

Cell Lines and Cell Culture

CBRH-7919 cells were derived from rat hepatoma cells and are passaged compared to primary liver cancer cells, making them easier to culture in vitro. CBRH-7919 cells were cultured in 1640 medium containing 10% fetal bovine serum and 1% double antibody.

Cellular in vitro Uptake Efficiency

To observe the in vitro uptake of DLD/HYP-Lip by CBRH-7919 cells, cell uptake of DLD/HYP-Lip was examined by flow cytometry. CBRH-7919 cells were seeded in parallel in a 6-well plate at 1×10^5 per well. The plate was incubated in a 37°C, 5% CO_2 cell incubator for 24 h. After cells were attached, the original culture solution was aspirated and washed twice with

PBS. After washing the PBS was discarded and DLD/HYP-Lip solution (pH 7.4 and pH 6.8) at a concentration of 0.5 $\mu\text{g}/\text{mL}$ was added. Fresh medium without any added drug was used as the control group. Samples were incubated for 4 h. After incubation, the original culture solution was aspirated, washed with cold 4°C PBS three times, 500 μL trypsin digest was added, and then 2 mL complete medium was added to terminate the digestion. The adherent cells were all suspended in complete medium and centrifuged (1000 rpm, 5 min). The supernatant was discarded, followed by resuspending the cells in 1 mL of 4°C PBS. The sample was then centrifuged (1000 rpm, 5 min) twice, and finally the cells were dispersed in 300 μL of 4°C PBS. These cells were then used for flow cytometry to analyze the fluorescence intensity.

In vitro Cytotoxicity Study

The in vitro cytotoxicities of free HYP, SPC/HYP-Lip, pH-sensitive DLD/HYP-Lip, and the blank DLD-Lip against CBRH-7919 cells at pH 7.4 and pH 6.8 were evaluated using 3-(4,5-dimethyl-thiazol-2-yl)-2,5-diphenyl-tetrazolium bromide (MTT assay). CBRH-7919 cells were inoculated in parallel with 5×10^4 cells per well in a 96-well plate. After incubation for 24 h, the cells were exposed to fresh medium with a pH adjusted to either 7.4 or 6.8 containing blank or HYP-loaded liposomes. After 48 hours of incubation, 20 μL of MTT solution (5 mg/mL) was added to each well and the sample was incubated for an additional 4 hours. The absorbance (OD_{490}) was measured at the wavelength of the microplate reader at 490 nm.

Co-Localization into the Mitochondria

Mitochondria localization of SPC/HYP-Lips and DLD/HYP-Lips in CBRH-7919 cells were observed under a laser confocal scanning microscope. CBRH-7919 cells were seeded (1×10^4 cells in each glass-bottomed dish) and cultured for 24 h under 5% CO_2 at 37°C. After 24 h incubation, cells were treated with SPC/HYP-Lips or DLD/HYP-Lips, and cultured for another 12 h under the same conditions (final HYP concentration: 0.5 $\mu\text{g}/\text{mL}$). After incubation, the medium was removed and the cells were washed with cold PBS three times, and then cells were stained with Mitotracker Green FM (75 nM) for 30 min under 5% CO_2 at 37°C. Stained cells were washed with PBS three times to remove free dye and observed by CLSM.

Determination of HYP Content in Mitochondria

CBRH-7919 cells were seeded in parallel in a six-well plate at 1×10^5 cells per well. The plates were incubated in a 37°C, 5% CO₂ cell incubator for 24 h. After the cells were attached, the original culture solution was aspirated, 2 mL was added, and the sample was washed twice with PBS. Then, SPC/HYP-Lip, DLD/HYP-Lip solution at a concentration of 0.5 µg/mL was added; fresh medium without any additive was used as the control group. Samples were incubated in a cell culture incubator at 24 h. After incubation, the original culture solution was aspirated, washed twice with cold 4°C PBS, and then the mitochondria were extracted using a mitochondrial stripping kit. The cells were redispersed in the mitochondrial stripping buffer solution, homogenized in a homogenizer fifteen times, the obtained suspension solution was centrifuged (13,000 rpm, 10 min), and the supernatant was collected and centrifuged again (11,000 rpm, 10 min). For mitochondrial separation, the desired precipitate is mitochondria, and the mitochondria were redispersed in 300 µL of PBS after the supernatant was aspirated. The HYP content in the mitochondria was measured using a flow cytometer.

Mitochondria-Mediated Cell Apoptosis Pathway

Activity Determination of Caspase 3 and Caspase 9
CBRH-7919 cells were seeded in parallel in a 6-well plate at 1×10^5 per well. The plate was incubated in a 37°C, 5% CO₂ cell incubator for 24 h. After the cells were attached, the original culture solution was aspirated and added. Cells were then washed twice with 2 mL PBS, SPC/HYP-Lip, DLD/HYP-Lip, and HYP solutions at a concentration of 20 µg/mL were added. Fresh medium without any added drug was the control group. Cells were then incubated for 12 h. Following the methods in the Caspase 3 and Caspase 9 activity detection kits, the cell culture solution was aspirated and used. The adherent cells were digested with 500 µL of trypsin and collected into spare cell culture medium. Cells were then centrifuged (1000 rpm, 5 min), the supernatant was discarded, and 2 mL of PBS was added to wash the cells. Then, the cells were centrifuged (1000 rpm, 5 min), the supernatant was discarded, 100 µL of the lysate was added, and samples were transferred into a 600 µL centrifuge tube. In an ice bath, the sample was lysed for 15 min, and centrifuged at 4°C (16,000 rpm, 15

min). The supernatant was transferred to a pre-cooled centrifuge tube in an ice bath. Then, 5 µL sample was taken for determination of protein concentration following the Bradford method. Appropriate samples were added to Caspase 3 and Caspase 9 substrates, and incubated overnight at 37°C in the dark. The activity of Caspase 3 and Caspase 9 was calculated by ELISA at 405 nm.

Apoptosis Test

Apoptosis of CBRH-7919 cells was measured using an Annexin V-FITC apoptosis detection kit and analyzed by flow cytometry. Cells were seeded in parallel in a 6-well plate at 1×10^5 per well and incubated for 24 h. HYP, SPC/HYP-Lip, and DLD/HYP-Lip were added to each well with 20 µg/mL HYP and incubated for 12 h. After incubation, the culture solution was aspirated, washed three times with 4°C PBS; then, 500 µL trypsin digest was added, 2 mL complete medium was added, the digestion was terminated and centrifuged (1000 rpm, 5 min). The supernatant was discarded, the cells were resuspended in PBS and centrifuged (1000 rpm, 5 min). According to the Annexin V-FITC apoptosis detection kit, the cells were resuspended in 195 µL Annexin V-FITC binding solution, 5 µL of Annexin V-FITC, and 10 µL PI staining solution. The samples were incubated at room temperature in the dark. After 20 min, the assay was quantified by flow cytometry.

Animal Tumor Xenograft Models

Animal experiments were performed in accordance with the Experimental Animal Administrative Committee of Heilongjiang University of Chinese Medicine (Ref. No: 2017042501). Approximately 5×10^6 CBRH-7919 cells suspended in PBS (50 mL) were subcutaneously injected into the left backs of BALB/c nude mice under anesthesia. Tumor volume (V) was calculated by measuring the length (L) and width (W), and calculated as $V = L \times W^2 / 2s$. The drug was administered when the tumor was 50–100 mm³.

In vivo Antitumor Efficacy

Mice were randomly assigned into four groups (n=5). Saline, HYP (6.0 mg/kg), SPC/HYP-Lip (6.0 mg/kg, calculated by HYP content), or DLD/HYP-Lip (6.0 mg/kg, calculated by HYP content) was administered via tail vein injection.³¹ Different formulations were injected at 12, 14, 16, 18, and 20 days after CBRH-7919 cell injection. Mice were weighed and tumors were measured every 2 days. Mice were sacrificed on day 25 after tumor inoculation and tumors were removed and then embedded in paraffin

and cut with a microtome (5-mm thick) before staining with hematoxylin and eosin (HE). For quantification of proliferative cells, Ki67 was assessed with Anti-Ki67 [SP6] antibody (Abcam, England). The Ki67 index was calculated as the ratio of proliferative cells to total cells in each field, using five random fields. For quantification of apoptotic cells, tdt-mediated dntp nick-end labeling (TUNEL) assays were used with the in situ cell death detection kit-POD (Roche Group, Switzerland). The apoptotic index was calculated as the ratio of apoptotic cells to total cells in each field, using five random fields.

In vivo Toxicity

Healthy male BALB/c were randomly assigned into four groups ($n = 5$), and saline, HYP (6.0 mg/kg), SPC/HYP-Lip (6.0 mg/kg, calculated by HYP content), or DLD/HYP-Lip (6.0 mg/kg, calculated by HYP content) was administrated via tail vein injection. Different formulations were injected and mice were weighed every 2 days. Then, animal behaviors were assessed.

After 7 days post-dosing, mice were allowed to recover and were weighed every day. After sacrifice, the liver, heart, spleen, lungs, and kidneys were removed and washed with PBS, fixed in 4% formaldehyde, embedded in paraffin, and cut with a microtome (5-mm thick) for HE staining.

HYP Quantification Using HPLC

HYP was measured via HPLC using a Shimadzu HPLC system consisting of a LC-20AT-pump and a SPD-20A Diode array detector (Shimadzu Technologies). A Diamonsil C-18 column (250 mm \times 4.6 mm, 5 μ m; DIKMA Technologies) and a detector wavelength of 360 nm were used. A solvent mixture of acetonitrile - 0.1% phosphoric acid solution (22:78, v/v) was used as an eluent, and the flow rate was 1.0 mL/min.

Statistical Analysis

All data are presented as means \pm SD. A Student's *t*-test or One-way analysis of variance (ANOVA) was used to determine statistical significance. Statistical significance was set at * $p < 0.05$, and ** $p < 0.01$ indicates highly significant differences.

Results and Discussion

Synthesis of DSPE-Lys-DMA (DLD)

The synthetic route of DSPE-Lys-DMA was shown in [Scheme S1 of Supporting Information](#). The carboxylic

group of Boc-L-Lys(Boc)-OH was conjugated to the primary amine group of DSPE and deprotected using trifluoroacetic acid. DMA was connected to the amino groups of the DSPE-Lys using an amidation reaction. The DSPE-Lys-DMA was characterized using matrix assisted laser desorption ionization time-of-flight mass spectroscopy (MALDI-TOF MS) and ^1H NMR in [Supporting Information](#).

Characterization of Liposomes

The size, polymer dispersity index (PDI), encapsulation efficiency, and drug loading of blank and HYP-loaded liposomes (SPC-Lip, DLD-Lip, SPC/HYP-Lip, and DLD/HYP-Lip) are shown in [Table S1 of Supporting Information](#). The average diameter of the two blank liposomes was similar, approximately 110 nm, while the average diameter of the two HYP drug-loaded liposomes was about 130 nm. Because the drug is encapsulated in the lipid bilayer of liposomes, the particle size of HYP liposomes was larger than that of blank liposomes.^{24,32} The DLD-Lips meter showed higher encapsulation efficiency and drug loading. The encapsulation efficiency of liposomes obtained by reconstitution of long-circulating and pH-sensitive hyperoside liposomes was (94.36 \pm 1.52) %, which was greater than 90%, indicating the optimal formulation of liposomes and a strong encapsulation ability for HYP.³³ The drug loading of the liposomes obtained after reconstitution was 8.5%, indicating that the liposome prepared by the optimal formulation has a strong drug-loading ability for HYP.^{34,35} PDI is (0.164 \pm 0.062), indicating that the particle size of liposomes is very uniform. Transmission Electron Microscope (TEM) was used to perform a morphological analysis of DLD-Lips loaded with DLD/HYP-Lips, the liposome was rounded and spherical, as shown in [Figure 1A](#).

Serum Stability Study

The stability of SPC-Lip and DLD-Lip in serum was examined using an environmental mimic physiological condition containing 50% fetal bovine serum. The results are shown in [Figure 1B](#). In pH 7.4 serum, the average particle size and distribution of DLD-Lips hardly changed after 24 hours (PDI = 0.19 \pm 0.03). These results suggest that DLD-Lips do not aggregate in the serum solution, there are no significant change in the average particle size and distribution of sulfate.³³

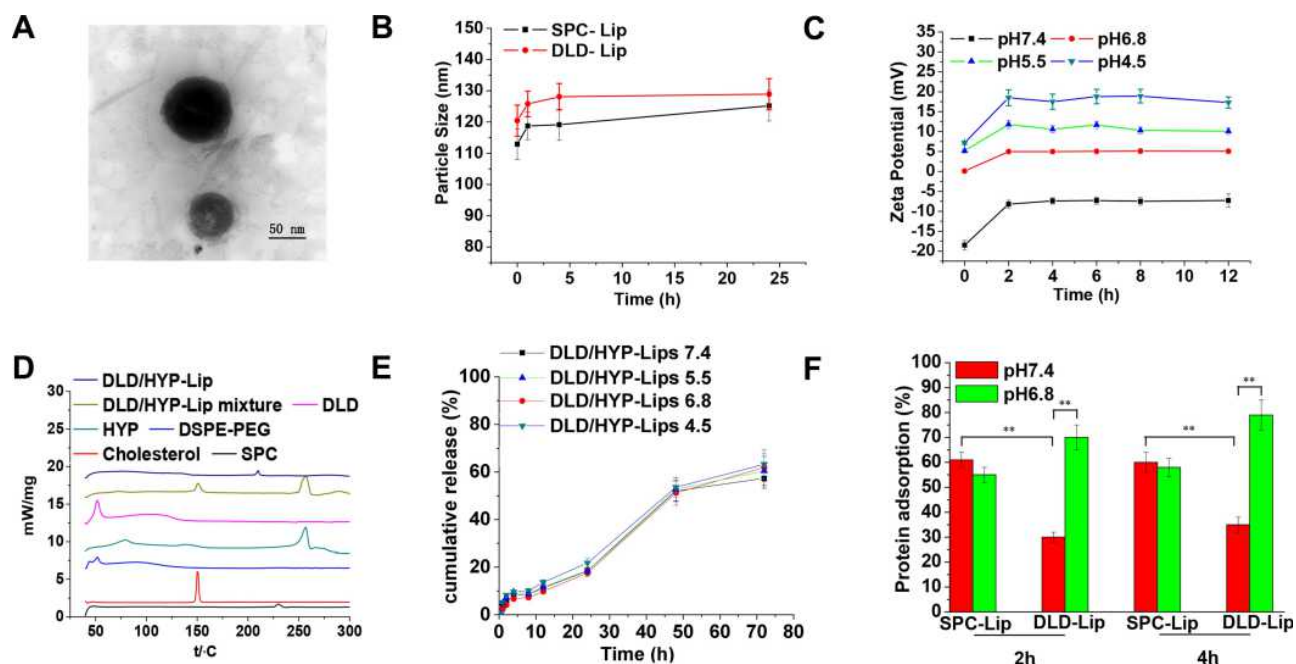


Figure 1 Physicochemical properties of liposomal formulations. TEM image of DLD/HYP-Lips (scale bar = 200 nm) (A). Changes in particle size of SPC-Lip and DLD-Lip in fetal bovine serum (means \pm SD, n=3) (B). Zeta potential variation of DLD/HYP-Lips at different pH values (C). DSC Curve of DLD/HYP-Lip, DLD, HYP, Cholesterol, SPC, DSPE-PEG, and physical mixed sample composed of DLD/HYP-Lip (D). Cumulative release of DLD/HYP-Lip in the release medium (means \pm SD, n=3) (E). Bovine serum albumin (BSA) adsorption on liposomes after 2 and 4 h incubation at 37°C, pH 7.4 or 6.8 (means \pm SD, n = 3) (F). ** $p < 0.01$.

The pH Sensitivity of DLD-Lips

To demonstrate that DLD-Lips changed surface properties in response to tumor extracellular and intracellular environments, zeta potentials of DLD-Lips were tested when they were dispersed in buffer solutions of different pH (pH 7.4, 6.8, 5.5, and 4.5). These pH values simulated physiological pH (pH 7.4), the tumor micro-environment (pH 6.8), endosomes and lysosomes (pH 5.5), and late-lysosomes (pH 4.5). As shown in Figure 1C, when the pH value was 7.4, the zeta potential of DLD-Lips was negative, when pH decreased to 6.8, DLD-Lips surface charges immediately increased to neutral, and plateaued to 5 mV within 1 h, and when the pH values were 5.5 and 4.5, DLD-Lips maintained positively charged surfaces with zeta potentials of \sim 10 mV and 20 mV, respectively. In contrast, traditional liposomes (SPC-Lips) did not undergo charge conversion. This indicated that DLD-Lip was negatively charged in the circulation (pH 7.4) and did not interact as much with negatively charged plasma proteins, leading to high resistance to non-specific protein adsorption and enhanced the stability in the blood.^{36–38} Given that tumor cell membranes are generally negatively charged, when DLD-Lips aggregate at the tumor site (pH 6.8) via the EPR effect, the DLD-Lips charge conversion should enhance cellular uptake.^{25,39} The charge recovery is

attributed to the acid-labile cleavage of the amide formed between amine and DMA. It may be concluded that DLD-liposomes are negatively charged (pH 7.4) while circulating in the blood stream, and the charge will be reversed to positive in the weakly acidic tumor microenvironment and endosomal compartment. Compared with other acid-sensitive chemical bonds, such as hydrazine-modified liposomes cleaved at pH 5.0, the amide formed between amine and DMA has an effect on the weakly acidic tumor micro-environment (pH 6.8), showing an ideal pH response.^{19,40}

Differential Scanning Calorimetry Analysis

Figure 1D shows that the characteristic melting point of SPC was approximately 230°C, that of cholesterol was approximately 150°C, DSPE-PEG was approximately 54°C, and that of DLD was about 50°C. The melting point peak of HYP was approximately 255°C. The melting points of the above compounds are consistent with those reported in the literature.^{41–44} The above five characteristic peaks appear in the DLD/HYP-Lip physical mixed sample. The characteristic melting point peak of DLD/HYP-Lip powder appeared at approximately 210°C, which was different from the curve of the physical mixture. Displacement occurred and it is speculated that the DLD/

HYP-Lip powder was not undergoing pure physical mixing. The findings also showed that the preparation of DLD/HYP-Lip was successful.

Characteristics of HYP Release Behavior

The release characteristics of drug-loaded HYP liposomes in release media with different pH values were determined. The results shown in Figure 1E indicated that the cumulative release of HYP in DLD/HYP-Lips at all pH values was about 20% after 24h, indicating that the weak acidic release medium only promoted the charge reversal without destroying the liposome structure to promote drug release. Release of HYP from DLD/HYP-Lips at all pH values was about 50% after 48h, and release of HYP from DLD/HYP-Lips at all pH values was about 60% after 72h. Therefore, DLD-Lips not only had the function of charge reversal, but also maintained the sustained release of HYP. This sustained release can prevent the sudden release of the carrier during the delivery process in the body and increase the accumulation of the drug in the tumor tissue.^{45,46}

Protein Adsorption Test Results

The properties of the liposome surface have a greater impact on the amount of protein adsorption, and the interaction between them indirectly reflects its circulation time in the blood. If the adsorption of non-specific proteins on the liposome surface is reduced, its circulation time in the blood will be prolonged.^{47,48} Therefore, we used the BCA protein experiment to determine the amount of protein adsorbed on the liposome surface. BSA was used as a model protein to simulate plasma protein. The isoelectric point of BSA is 4.5–5.0,⁴⁹ under the conditions of PH 7.4 and 6.8, the PI value is less than the isoelectric point (PI), and BSA surface is negatively charged, the pHs will not affect the charge on the surface of the BSA, so in cell adhesion experiments and blood compatibility experiments, BSA is usually used as a model protein to detect the protein adsorption capacity of liposomes under the condition of pH 7.4 and 6.8. As shown in Figure 1F, at a physiological pH 7.4, after protein and liposomes were incubated for 2 h, the amount of protein adsorption on the surface of SPC-Lip was (60.97±2.86), DLD-Lip was (30.10±2.14), DLD-Lip was significantly less than that on the SPC-Lip surface. When the incubation time was extended to 4 h, the amount of protein adsorption on the surface of SPC-Lip was (59.84±3.59), DLD-Lip was (34.87±2.91), the protein adsorption capacity of SPC-Lip

and DLD-Lip did not increase significantly compared with 2 h, and the change of the test group was the same as 2 h. However, when the liposome and protein were co-exposed at pH 6.8 after 2 hours of incubation, the amount of protein adsorption on the surface of SPC-Lip was (54.89±2.94), DLD-Lip was (69.98±5.07), the interaction between DLD-Lip and protein was strong, and about 70% of the protein was absorbed by the liposome surface. When the incubation time was extended to 4 h, the amount of protein adsorption on the surface of SPC-Lip was (57.60±3.70), DLD-Lip was (79.04±6.10), the interaction between DLD-Lip and protein was also strong, and about 80% of the protein was absorbed by the liposome surface. The results prove that the change of DLD charge leads to the change of protein adsorption effect. These results are consistent with DLD-Lip's charge reversal phenomenon at pH 6.8.

Cellular Uptake

In order to observe the in vitro uptake of DLD/HYP-Lip by CBRH-7919 cells, the auto fluorescence of HYP was used as a fluorescence marker, and the cell uptake of DLD/HYP-Lip was investigated by flow cytometry. Fluorescent intensity was quantified in the diagram. Varied fluorescent intensities were used to indicate liposome uptake. The results of the cell uptake analysis for DLD/HYP-Lip were shown in Figure 2A and B. For ordinary liposomes, the pH did not change the uptake. In contrast, DLD/HYP-Lips were greatly affected by the pH of the environment. The average fluorescence intensity of SPC/HYP-Lips was similar in CBRH-7919 cells. When the pH value was 7.4, few liposomes were taken up by the CBRH-7919 cells, and the average fluorescence intensity was (9.81±0.71). At pH 6.8, liposome internalization increased seven times, and the average intensity was (70.41±4.96). This may be due to the charge reversal effect of DLD in the slightly acidic environment of the tumor. Under pH 6.8, the negative charge on the surface of DLD was reversed to a positive charge, promoting cell uptake. The results indicate that DLD/HYP-Lip is sensitive to pH, which specifically increase the uptake of HYP in tumor cells in the tumor microenvironment, and improve the effect of the tumor treatment.

Cellular uptake pathways were identified, results showed that all of the cellular uptake of DLD/HYP-Lip decreased remarkably in the presence of amiloride ($p < 0.05$), an inhibitor of macropinocytosis, and similar phenomena was

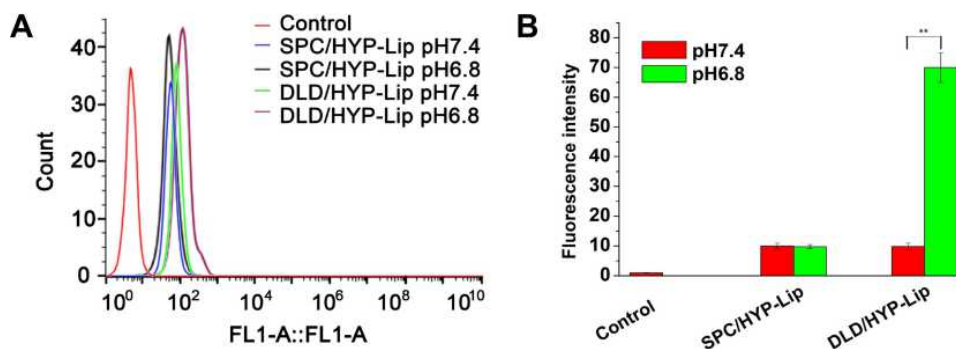


Figure 2 Cellular uptake of DLD/HYP-Lip in CBRH-7919 cells measured quantitatively using FCM (A and B). Data are presented as means \pm SD (n = 3). **p < 0.01.

observed in the presence of chlorpromazine, an inhibitor of clathrin-mediated endocytosis. In contrast, the presence of filipin, an inhibitor of caveolin-mediated endocytosis, had no significant effect on the cellular uptake of DLD/HYP-Lip. It was speculated that DLD-Lips would enter the cells initially in macropinosomes and clathrin coated vesicles by macropinocytosis and clathrin-dependent endocytosis, respectively, and both further in endosomes/lysosomes. Owing to DLD conjugate containing lysines, DLD-Lips might perform the proton sponge effect similar to polyethylenimine (PEI) which leads to the swelling and disruption of the endosomes/lysosomes for cytoplasmic liberation of the intact liposomes.^{50,51} It is noteworthy that internalization through macropinocytosis is more efficient than that through clathrin-mediated endocytosis, since more porous membrane structure of macropinosomes can enhance leakage of inclusions into cytoplasm compared with clathrin-coated nanocarriers, in terms of avoiding endosomal/lysosomal degradation.^{52,53} The experiment content and results (Figure S1) of “Identification of cellular uptake pathways” are shown in Supporting Information.

In order to prove the potential of the prepared mitochondrial targeting liposome DLD/HYP-Lip in tumor treatment, the endosomal escape of the mitochondrial part was studied by confocal fluorescence microscopy. As shown in result (Figure S3) of Supporting Information the incubation time increases, more particles were released into the cytoplasm (red dots in the merged image), indicating that liposomes escape from the lysosome. The results show that DLD/HYP-Lip can successfully target mitochondria, and DLD play a vital role in the targeting process.

In vitro Cytotoxicity Study

Through the MTT test, the cytotoxicity of blank DLD-Lips to CBRH-7919 cells with different lipid concentrations was evaluated at pH 7.4 and 6.8, and the viability of untreated

cells at a specific pH is 100%. Figure 3A shows a blank DLD-Lips, even at high lipid concentrations under two pH conditions, were non-toxic; therefore, DLD-Lips may be a safe drug delivery system. The MTT method was also used to evaluate the cytotoxicity of HYP, SPC/HYP-Lips, and DLD/HYP-Lips to liver cancer CBRH-7919 cells at pH 7.4 and 6.8. At pH 6.8, Figure 3B–D show that DLD/HYP-Lips was also more cytotoxic to CBRH-7919 cells than HYP and SPC/HYP-Lips. DLD/HYP-Lips promoted the death of CBRH-7919 cells. Therefore, these data are consistent with the data from cell uptake studies. In addition, the pH-induced surface charge reversal of liposomes contributed to the internalization of liposomes and the accumulation of anticancer drugs in cells. DLD/HYP-Lips IC₅₀ of CBRH-7919 cells were eight times lower than HYP at the same pH. HYP, SPC/HYP-Lips, and DLD/HYP-Lips IC₅₀ of CBRH-7919 cells were 4.89 μ g/mL, 13.82 μ g/mL, 18.08 μ g/mL at pH 7.4; HYP, SPC/HYP-Lips, and DLD/HYP-Lips IC₅₀ of CBRH-7919 cells were 4.59 μ g/mL, 12.91 μ g/mL, 0.26 μ g/mL at pH 6.8. Therefore, DLD/HYP-Lips had the greatest inhibitory effect on the growth of CBRH-7919 cells, especially at pH 6.8. This enhanced inhibitory effect can be attributed to the higher intracellular uptake by targeting mitochondria to trigger apoptosis.

Mitochondrial Targeting by Confocal Laser Scanning Microscopy (CLSM)

To determine whether the enhanced anticancer effect of DLD Lips stemmed from the mitochondria-mediated apoptosis pathway, mitochondrial targeting was first examined with a CLSM and flow cytometry (FCM). Figure 4A shows CLSM images of mitochondria localization of DLD/HYP-Lips in CBRH-7919 cells. Yellow dots indicate co-localization of green fluorescence from Mitotracker Green FM and red fluorescence from DLD/HYP-Lips. Bright yellow fluorescence indicates that DLD/HYP-Lips

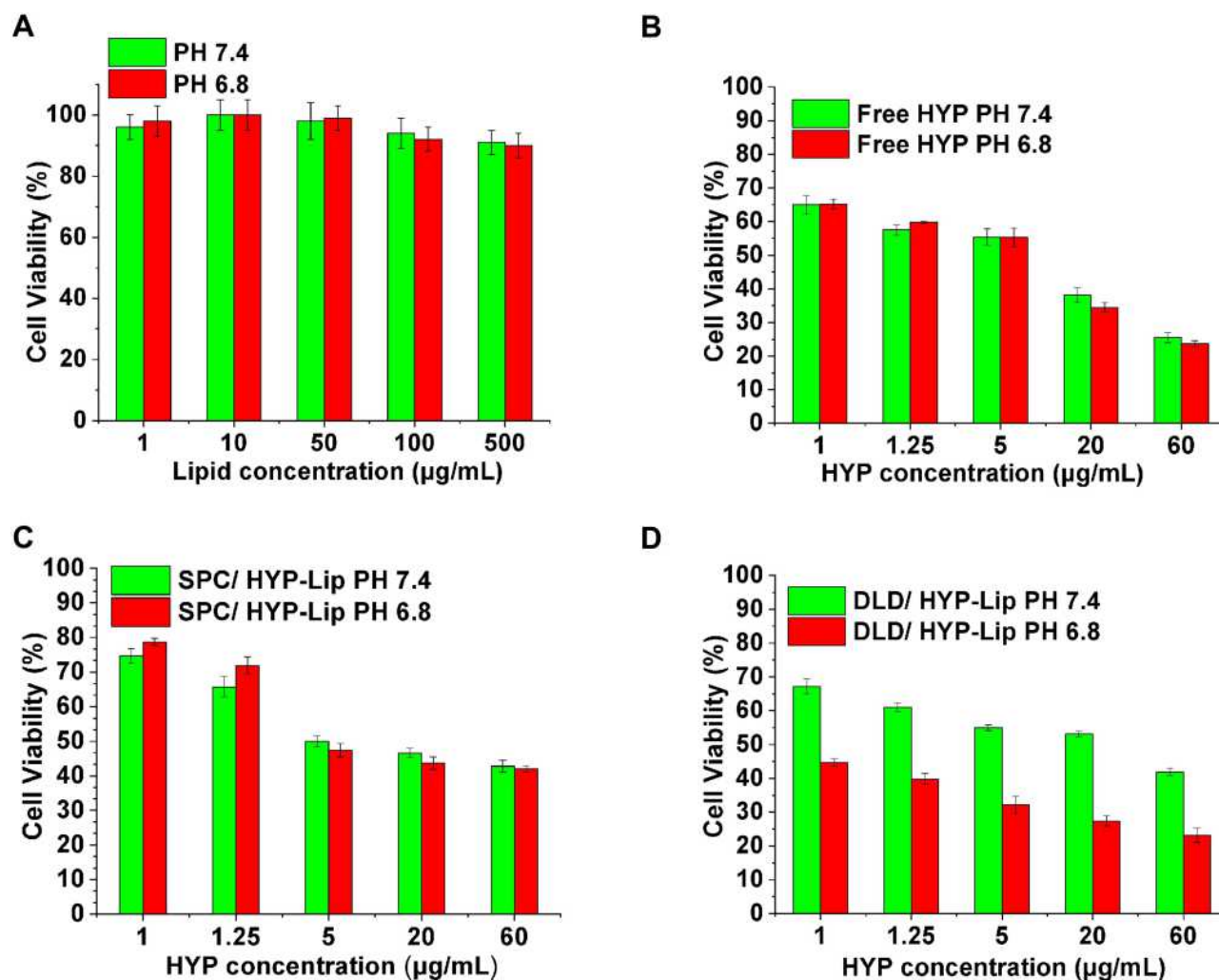


Figure 3 Cytotoxicity of SPC solution (A), HYP solution (B), SPC/HYP-Lip (C), and DLD/HYP-Lips (D) (pH 7.4 and pH 6.8) on CBRH 7919 (means \pm SD, n=3).

selectively accumulated into mitochondria and that SPC/HYP-Lips (no yellow fluorescence) did not.

HYP Content in Mitochondria

Mitochondria are the main site for cells to perform aerobic respiration and adenosine triphosphate (ATP) are formed by oxidative phosphorylation.⁵⁴ The mitochondrion is also an organelle that is capable of reproducing in eukaryotic cells. Mitochondria control apoptosis and many lethal signaling pathways focus on mitochondria.⁵⁵ The discovery of the mechanism of mitochondrial regulation of apoptosis has made it a broad prospect for anti-tumor targets.^{56–58} Studies had shown that the occurrence and development of tumors were apoptosis-related, and mitochondria had a regulatory effect on apoptosis.^{59,60}

Figure 4B and C, respectively, indicated the cumulative amount of the drug in the exfoliated mitochondria of CBRH-7919 cells, which was expressed by the amount of

fluorescence intensity. After the cells were incubated with DLD/HYP-Lips for a period of time, the fluorescence intensity in the mitochondria was significantly higher than that in the SPC/HYP-Lips group. The fluorescence intensity was (65.16 ± 5.07), which was approximately 3.4 times higher than that of the SPC/HYP-Lips group (18.23 ± 1.59). The above data indicate that DLD-Lips can effectively target mitochondria and increase the accumulation of drugs in the mitochondria.

The content of HYP in the mitochondria of tumor tissue was also determined, results (Figure S2) showed that the content of HYP in tumor tissue mitochondria in the DLD/HYP-Lip administration group was significantly higher than saline group, HYP and SPC/HYP-Lip group. The results indicate that DLD/HYP-Lip can be enriched in the mitochondria of the tumor tissue, which enhances the toxicity of DLD/HYP-Lip to tumor tissue. The experiment content and results are shown in Supporting Information.

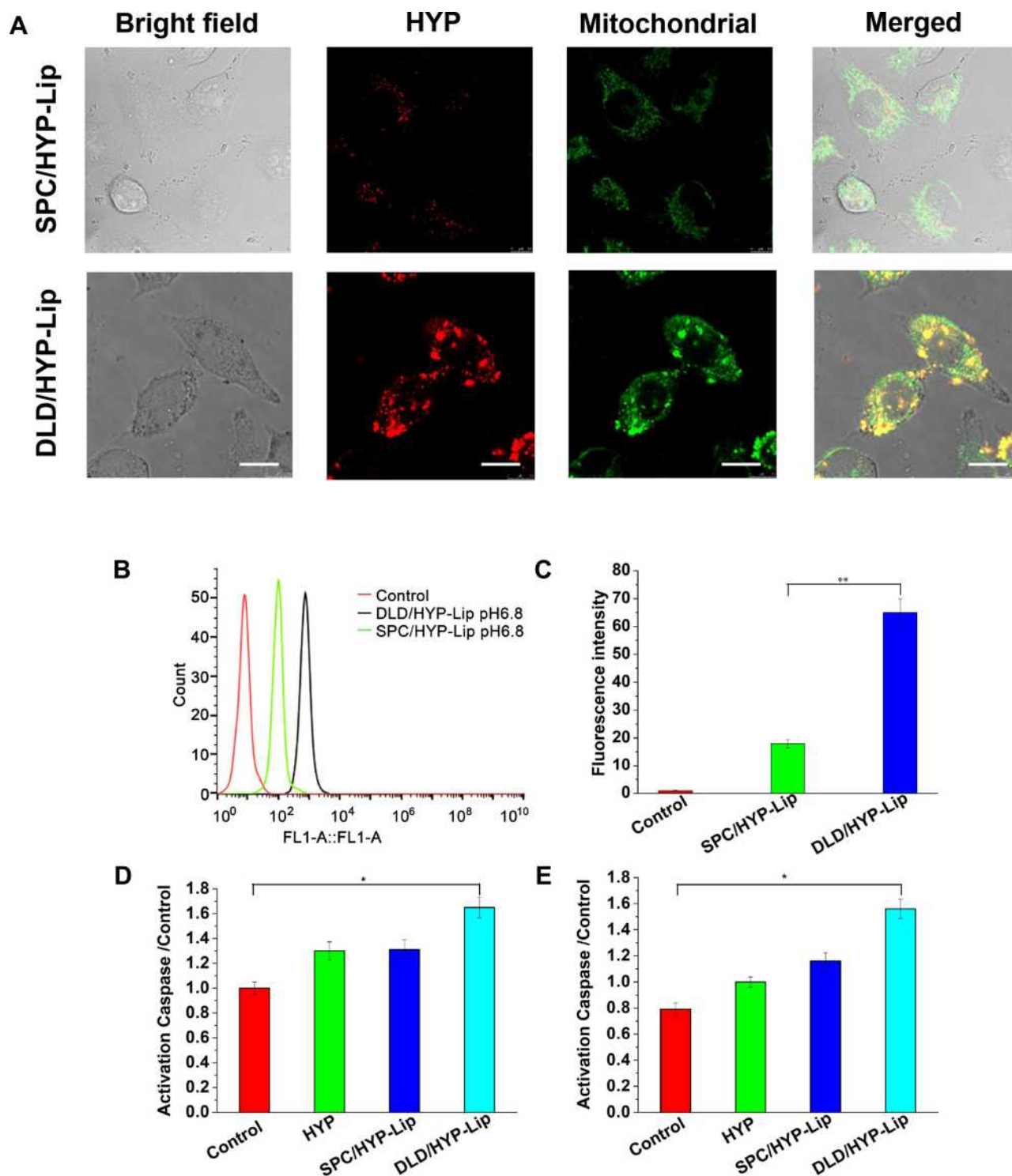


Figure 4 Mitochondrial targeting of liposomal formulations. Mitochondrial localization of DLD/HYP-Lips in CBRH-7919 cells by confocal laser scanning microscopy. CBRH-7919 cells were incubated with SPC/HYP-Lips and DLD/HYP-Lips for 12 h and then stained with Mitotracker Green FM. Yellow spots in the merged pictures denote the co-localization of the liposomes within mitochondrial compartments. The scale bar represents 25 μ m (A). The cumulative amount of HYP in mitochondria of CBRH-7919 cells at pH 6.8 measured by flow cytometry (B and C). Activity ratios of caspase 9 (D) and caspase 3 (E). Data are presented as means \pm SD (n = 3). *p < 0.05, **p < 0.01.

Activity Determination of Caspase 3 and Caspase 9

The activation of apoptosis-related enzymes is an essential factor in the process of cell apoptosis, and caspase 9 and caspase 3 are usually used as downstream markers of tumor cell apoptosis.^{61–63} Figure 4D and E, respectively, reflect the caspase 9 and caspase 3 activities of CBRH-7919 cells treated with different HYP preparations. After treatment with DLD/HYP-Lips, the caspase 9 activity ratio in CBRH-7919 cells was 1.68 times higher than that in the control group, while the caspase 3 activity ratio increased by 1.97 times compared to the control group. Compared with other HYP preparations, DLD/HYP-Lips had the most significant effect of inducing caspase 9 and caspase 3 activation. These data show that cell death induced by DLD/HYP-Lip is mediated by mitochondrial-dependent apoptosis pathways.

Cell Apoptosis

The early and late apoptotic cells were counted to prove that HYP induced apoptosis of liver cancer CBRH-7919 cells

(Figure 5A–E). When HYP, SPC/HYP-Lip, and DLD/HYP-Lip (6.0 mg/mL HYP) were used, the apoptosis induction rate was 24.44%, 10.28%, and 41.38%, respectively. In addition, blank DLD-Lip induced apoptosis at a rate of 1.52%, which was similar to the control group (0.96%). Results indicate that blank DLD-Lips have no direct effect on cell apoptosis. In addition, DLD concentrate liposomes in cells, exert cytotoxic function, and promoted cell apoptosis, suggesting that DLD/HYP-Lip-induced cell death is mediated by a mitochondrial-dependent apoptosis pathway.

In vivo Antitumor Efficacy

In order to confirm the feasibility of DLD/HYP-Lips for cancer therapy in vivo, tumor xenograft models were used to evaluate the antitumor efficacy of DLD/HYP-Lips in vivo. The anti-tumor effect was reflected by measuring the changes in tumor volume of tumor-bearing nude mice in each test group within 25 days (Figure 6A). The tumors of nude mice in the saline injection group grew very fast. Both HYP and SPC/HYP-Lip inhibited tumor growth to a certain extent,

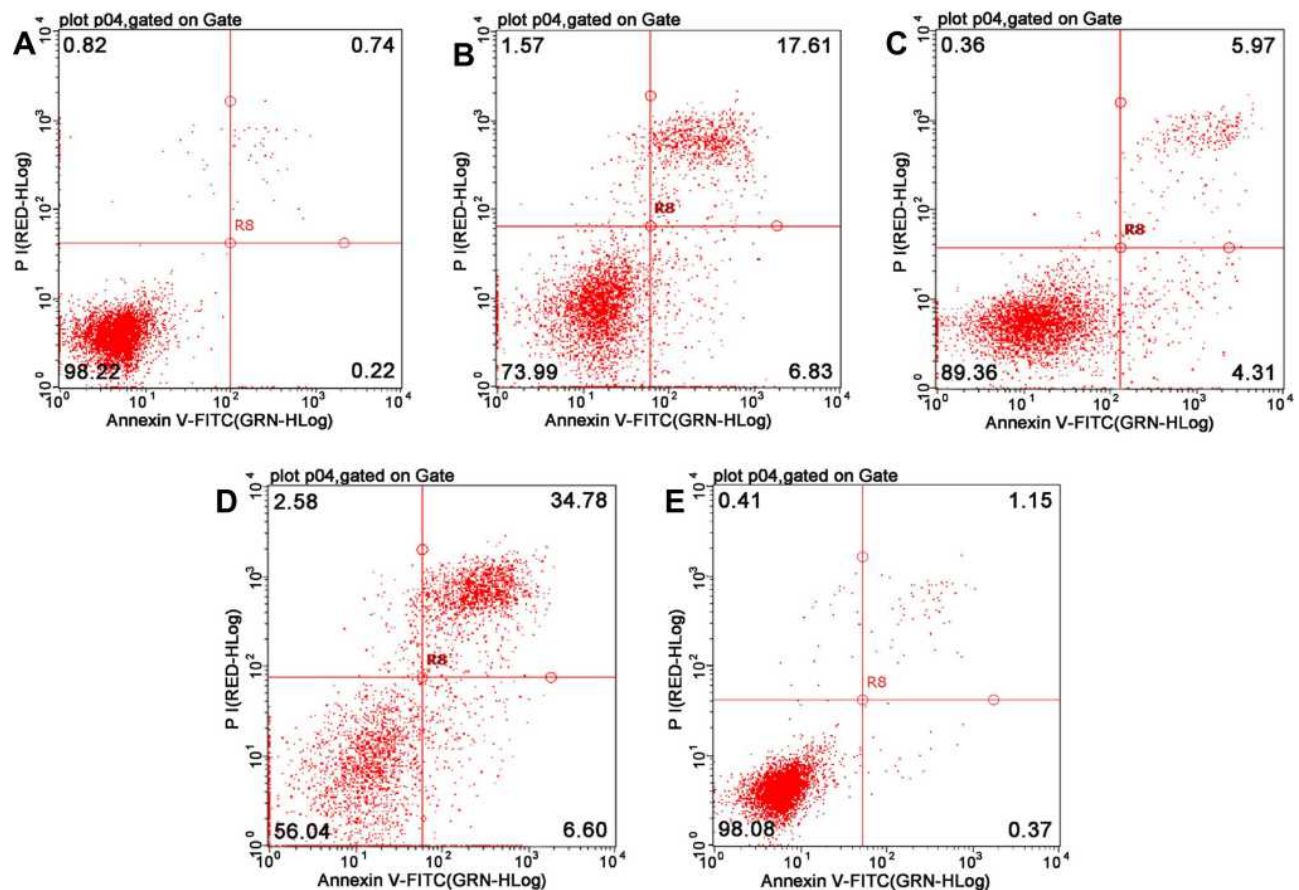


Figure 5 Cell apoptosis of CBRH-7919 cells (A–E) treated with the culture medium, HYP, SPC/HYP-Lip, DLD/HYP-Lip, and DLD-Lip for 12 h and determined using flow cytometry.

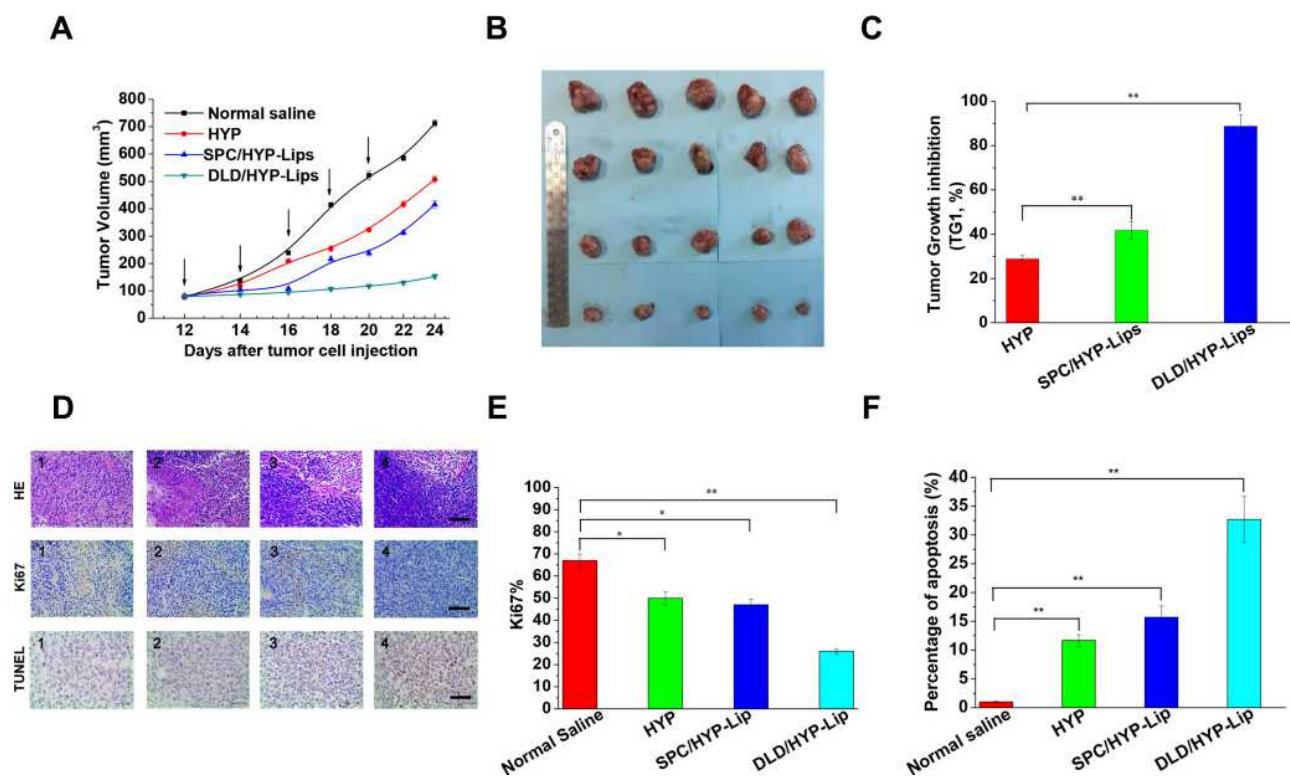


Figure 6 In vivo antitumor efficacy of various HYP formulations in CBRH-7919 cell tumor-bearing mice. Mice ($n = 5$) received injections of HYP (6.0 mg/kg) as indicated by the arrows. Tumor growth as a function of time (**A**); tumors were photographed at the end of the experiment (**B**). Tumor tissues were isolated and tumor growth inhibition (TGI, %) was calculated (**C**); morphological changes in HE stained sections of tumors (**D**); evaluation of tumor cell proliferation in vivo using Ki67 staining (proliferative cells shown in brown) (**E**). The Ki67 index was calculated as the ratio of proliferative cells to total cells in each field ($n = 5$) (**E**). Evaluation of tumor cell apoptosis in vivo using a TUNEL assay (apoptotic cells shown in brown) (**D**); percentage of apoptotic index was calculated as the ratio of apoptotic cells to total cells in each field ($n = 5$) (**F**). 1. Saline, 2. HYP, 3. SPC/HYP-Lips and 4. DLD/HYP-Lips treatment. Results are presented as means \pm SD, * $p < 0.05$, ** $p < 0.01$. Scale bars represent 250 μ m.

while DLD/HYP-Lip almost completely inhibited tumor growth, indicating that DLD/HYP-Lip had a significant anticancer effect. Photographs of the tumors at day 25 are shown in Figure 6B, as are the growth curves of the tumors.

After the experiment, the tumors in each drug group were stripped and weighed to calculate the tumor inhibition rate (Figure 6C). The tumor inhibition rates of HYP, SPC/HYP-Lip, and DLD/HYP-Lip were 28.78%, 41.71%, and 88.79%, respectively. The findings showed that DLD/HYP-Lip has the best anti-tumor effect.

After the tumor suppression experiment in tumor-bearing nude mice was completed, we excised the tumor tissue and performed immunohistochemical staining. As shown in Figure 6D, the HE experimental results showed that DLD/HYP-Lips caused the largest area of apoptosis in drug-resistant tumor tissue. The Ki67 immunohistochemical staining experiment showed that tumor-bearing nude mice injected with DLD/HYP-Lip showed a significantly reduced proliferation level of tumor cells (Figure 6E). The TUNEL test showed that tumor-bearing nude mice injected with DLD/HYP-Lip more significantly induced apoptosis of drug-resistant tumor

cells (Figure 6F). The reasons of enhanced efficacy of DLD/HYP-Lip may be as follows: (1) Negatively charged DLD/HYP-Lips in the circulation interact less with plasma proteins, so it could resist non-specific protein absorption and escape RES rapid elimination; (2) Suitable particle sizes of DLD/HYP-Lips could induce greater accumulation of HYP into tumor tissue (pH 6.8) via the EPR effect and charge-conversion increase cellular uptake of DLD/HYP-Lips to enhance cytotoxicity in liver cancer cells; (3) Apoptosis-inducing effect of DLD/HYP-Lips could increase anticancer efficacy.

In vivo Toxicity Assays on Normal Mice

Healthy tumor-free BALB/c mice were used to evaluate the toxicity of HYP, SPC/HYP-Lip, and DLD/HYP-Lip (6.0 mg/kg HYP) in vivo. The clinical toxicity behavior and body weight changes of the mice during the administration period were recorded. After the end of the final observation period, the organs were excised for HE staining experiments. Compared with the control group, there was no obvious dehydration, dyskinesia, muscle loss, anorexia, or other animal toxicity-

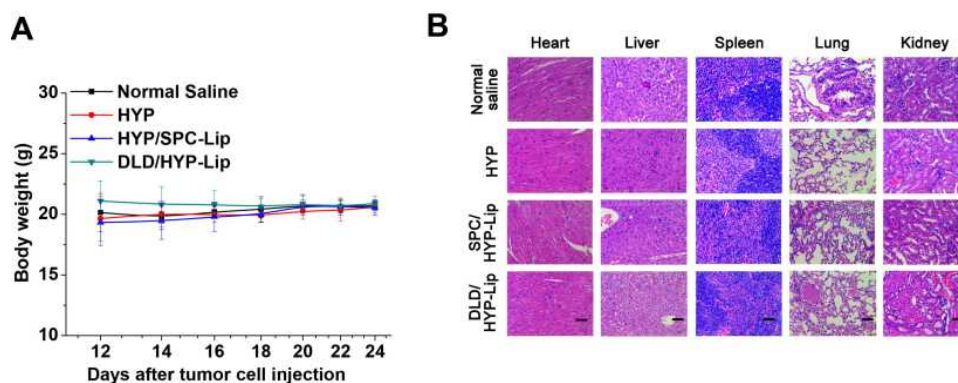


Figure 7 In vivo toxicity assays in normal mice. Body weight curves of mice receiving different treatments (HYP 6.0 mg/kg) (A). Morphological changes in HE stained of different organs of normal mice with different treatments (B). 1. Normal saline, 2. HYP, 3. SPC/HYP-Lip, 4. DLD/HYP-Lip. Scale bars represent 250 μ m.

related symptoms during the administration period. The body weight changes of tumor-bearing nude mice in each group are shown in Figure 7A. The body weight of mice in each administration group increased slightly, indicating that there was likely no serious systemic toxicity in each drug delivery system. The results of the HE staining experiment showed that the organs of the mice in each administration group did not show significant damage (Figure 7B).

Conclusions

In the present study, a dual-function tumor-targeting liposomes (DLD-HYP-Lips) has been developed, in which HYP was delivered to the mitochondria of tumor cells to enhance anticancer activity. Unlike most therapeutic liposomes that exerted function at the cellular level, our HYP loaded DLD-Lips target the mitochondria at the subcellular level for more effective treatment. The results showed that DLD-Lips can be oriented to mitochondria by modification with a pH-responsive phospholipid DSPE-Lys-DMA. Compared to free HYP and HYP-loaded traditional liposomes, HYP loaded DLD-Lips increased cytotoxicity of liver cancer CBRH-7919 cells and antitumor effect in vivo. This was due to increased cellular uptake and mitochondria-mediated apoptosis, which obviously decreased mitochondrial membrane potential, activation of caspase-9/3 and induced apoptosis. Therefore, DLD-Lip is a promising anticancer drug delivery system to cancer therapy both in vitro and in vivo.

Acknowledgment

This study was financially supported by the National Natural Science Foundation of China (No. 81703944), Heilongjiang Natural Science Foundation Project (YQ2019H031), Post-

doctoral Researchers Settled in Heilongjiang Scientific Research Startup Fund (2020), and Outstanding Innovative Talents Project from Heilongjiang University of Chinese medicine (2018).

Disclosure

The authors report no conflicts of interest in this work.

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