



Maternal Separation Induced Visceral Hypersensitivity Evaluated via Novel and Small Size Distention Balloon in Post-weaning Mice

Enfu Tao^{1,2}, Gao Long¹, Ting Yang¹, Bo Chen¹, Rui Guo¹, Diya Ye¹, Marong Fang³ and Mizu Jiang^{1*}

¹ Endoscopy Center and Gastrointestinal Laboratory, Children's Hospital, Zhejiang University School of Medicine, National Clinical Research Center for Child Health, National Children's Regional Medical Center, Hangzhou, China, ² Department of Pediatrics, Wenling Maternal and Child Health Care Hospital, Wenling, China, ³ Institute of Neuroscience and Gastrointestinal Laboratory, Children's Hospital, Zhejiang University School of Medicine, Hangzhou, China

OPEN ACCESS

Edited by:

Carlos Henrique Xavier, Universidade Federal de Goiás, Brazil

Reviewed by:

Natalia Lima Machado, Beth Israel Deaconess Medical Center and Harvard Medical School, United States Luiz Gonzaga Silva Junior, Federal University of Juiz de Fora, Brazil

> *Correspondence: Mizu Jiang mizu@zju.edu.cn

Specialty section:

This article was submitted to Autonomic Neuroscience, a section of the journal Frontiers in Neuroscience

Received: 28 October 2021 Accepted: 20 December 2021 Published: 28 January 2022

Citation:

Tao E, Long G, Yang T, Chen B, Guo R, Ye D, Fang M and Jiang M (2022) Maternal Separation Induced Visceral Hypersensitivity Evaluated via Novel and Small Size Distention Balloon in Post-weaning Mice. Front. Neurosci. 15:803957. doi: 10.3389/fnins.2021.803957 Early life stress (ELS) disposes to functional gastrointestinal diseases in adult, such as irritable bowel syndrome (IBS). Maternal separation (MS) is a well-known animal model of IBS and has been shown to induce visceral hypersensitivity in adult rats and mice. However, to the best of our knowledge, it has not been reported whether MS induces visceral hypersensitivity in young mice, such as the post-weaning mice. Moreover, the method for evaluation of visceral sensitivity also has not been described. Accordingly, the present study aims to evaluate the visceral sensitivity caused by MS in post-weaning mice and develop a novel and small size distention balloon for assessment of visceral sensitivity of such mice. Male pups of C57BL/6 mice were randomly divided into two groups, MS (n = 12) and non-separation (NS) (n = 10). MS pups were separated from the dams through postnatal days (PND) 2 to 14, while NS pups were undisturbed. After, all pups stayed with respective dams and were weaned at PND 22. Visceral sensitivity was evaluated by colorectal distention (CRD) with a novel and small size distention balloon at PND 25. The threshold of abdominal withdrawal reflex (AWR) scores were significantly lower in MS than NS. In addition, AWR scores at different pressures of CRD were significantly higher in MS than NS. The results demonstrate that MS induced visceral hypersensitivity in post-weaning mice. The designed small size distention balloon for evaluation of visceral sensitivity is of significance to further study the pathophysiology of IBS from early life to adulthood.

Keywords: maternal separation, visceral hypersensitivity, distention balloon, irritable bowel syndrome, mice

INTRODUCTION

Irritable bowel syndrome (IBS) is a functional gastrointestinal disorder and newly called "disorders of the gut-brain interaction" based on Rome IV classification (Drossman, 2016). Although the mechanism underlying the pathogenesis of IBS is not well understood, brain-gut axis dysfunction and visceral hypersensitivity are two of the main characteristics of IBS, which may relate to psychological distress (anxiety, depression, impulsiveness, anger) in children (Iovino et al., 2009).

1

A large number of studies have shown that early life stress (ELS) can result in persistent changes in the central stress response systems, visceral hypersensitivity, and increase predisposition to IBS (Burke et al., 2017; Ju et al., 2020; Rincel and Darnaudéry, 2020).

Maternal separation (MS), which mimics the ELS, is a wellknown animal model of IBS, and it has been shown to induce visceral hypersensitivity in adult rats (Botschuijver et al., 2019; Fan et al., 2020) and mice (Riba et al., 2017, 2018). Furthermore, MS disrupts intestinal homeostasis (Wong et al., 2019), activates the immune system (Barreau et al., 2008; O'Malley et al., 2013), alters gut microbiota (O'Mahony et al., 2009), and induces lasting alterations in neuronal circuits, neurotransmitter systems, neuronal architecture, and plasticity that are further associated with emotional and cognitive information processing (Caliskan et al., 2020). These alterations derived from MS are based on the fact that the perinatal period is a crucial window for the intestinal barrier, the immune system, and the gut microbiota to mature to establish an appropriate complex relationship (Renz et al., 2011). However, the precise and dynamic process of the changes that underlie the development of IBS from MS pups to adults is still poorly understood. Whether ELS will induce IBS in early life, such as childhood, is not elucidated. Accordingly, it is of significance to evaluate IBS-like phenotype including visceral hypersensitivity caused by MS in young mice, such as the period after weaning.

Colorectal distention (CRD) is a widely used method for assessing visceral hypersensitivity (Larsson et al., 2003; Christianson and Gebhart, 2007; Annahazi et al., 2009; Huang et al., 2021). However, CRD for evaluation of visceral sensitivity has not been reported in post-weaning mice. One of the main reasons may be that there is no suitable distention balloon for CRD of such mice. Therefore, it is urgent to develop a tool for CRD in post-weaning mice.

Accordingly, the present study aims to evaluate the visceral sensitivity caused by MS in post-weaning mice and develop a novel and small size distention balloon for assessment of visceral sensitivity of such mice.

MATERIALS AND METHODS

Animal Preparation

Specific pathogen-free (SPF) C57BL/6 mice were used in the experiment. Protocols for animal research were approved by the Zhejiang University Ethics Committee for Animal Research. C57BL/6 male and female mice (8 weeks old) were bred for mating at the ratio of 1:2. They were housed (cage size $318 \times 202 \times 135$ mm) and maintained on a 12-h light-dark cycle (light turned on at 9:00 am and turned off at 9:00 pm automatically) with access to food and water *ad libitum*, at a room temperature of 25–26°C, under a relative humidity of $50 \pm 5\%$. Ventilation rate reached 8–15 times/h and the ammonia concentration was less than 20 ppm. Mice received irradiated standard chow and wood chip bedding. The bedding was changed twice weekly. After mating, gestational mice were housed individually. Gestational mice delivering on the same day

were included in the experiments. To avoid the effects of stress on dams, litters were not disturbed at the first day after delivery. On postnatal day (PND) 2, four litters were standardized to five to six male pups per litter. Female pups were killed and not used for following experiments to avoid female hormonal cycle interference. Then male neonatal mice were randomly assigned to the MS group (n = 12) and non-separation (NS) group (n = 10). During grouping, all male pups were labeled by cutting off one of the rear claws, with left for MS, and right for NS. The procedure of animal preparation is summarized in **Figure 1**.

Maternal Separation Paradigms

Maternal separation was performed as previously described (Riba et al., 2018; Wong et al., 2019) with some modifications. Briefly, on PND 2–14, MS pups were removed from dams daily for 3 h consecutively (9:00 am to 12:00 am) and were individually placed into a plastic box (size $180 \times 130 \times 55$ mm) with the wood chip bedding in a separated room with ambient temperature $26 \pm 2^{\circ}$ C. The bedding was changed once weekly. After separation, MS pups were returned to dams and left undisturbed, whereas NS pups were nursed as usual. All pups were weaned on PND 22. After that, five to six per cage of MS and NS mice were housed (cage size $318 \times 202 \times 135$ mm) independently.

Body Weight Measurements

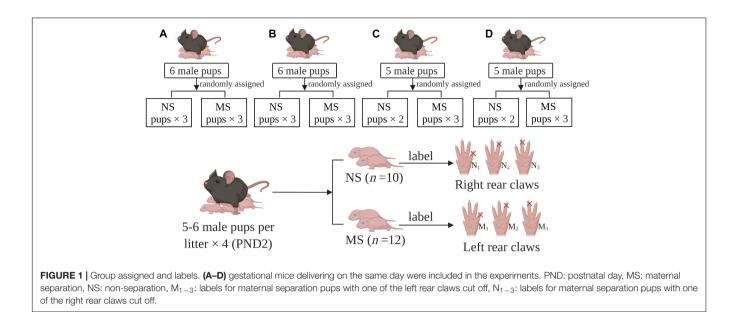
Both MS and NS mice were measured for body weight daily by electronic scale from PND 2 to 25 before separation, and body weight was recorded in a pre-made form.

Novel and Small Size Distention Balloon

The distention balloon was made from a little finger of a small size latex glove (2 cm in length) placed into Teflon tubes (10 cm length; outer diameter, 1 mm; internal diameter, 0.6 mm) purchased from Taobao, and were tied with 6-0 silk thread at a distance of 1, 1.5, and 2 cm, respectively, from the tip of glove finger. Before the latex glove was placed into Teflon tubes, at the front of the Teflon tube, two side holes were sheared with ophthalmic scissors at an oblique angle on both sides of the tube to increase the uniformity of balloon inflation. This step was critical in that it helped avoidance of glove tip obstructing the tube. The balloon was about 3 mm in diameter, 1 cm long, and was subsequently sealed using parafilm in a manner of winding at a distance of 2 cm from the tip of the glove finger. The end of the Teflon tube was also wrapped by parafilm to increase the diameter size to connect with a three-way tube. The two ends of the tube connected 10-ml syringes and pressure gauge, respectively. In this way, the pressure of the distention balloon could be monitored in real time (Figure 2).

Abdominal Withdrawal Reflex

Abdominal withdrawal reflex score was evaluated by colorectal distention (CRD) on mice between 10:00 am and 4:00 pm at PND 25 according to the previous study with some modifications (Al-Chaer et al., 2000; Yu et al., 2012; Zhang et al., 2020). Briefly, mice were allowed free access to food and water before the experiment. Anuses were lubricated with Vaseline ahead of



the experiment. The distention balloon was introduced into the rectum at 1.0 cm from the anus and fixed at the base of the tail under 1.5% isoflurane (RWD Life Science Co., Ltd. Shenzhen, China) anesthesia. Then, mice were placed in a hollow cylinder made from transparent acrylic (cylinder size 6 cm long, 3 cm inner diameter, purchased from Taobao) and allowed to recover 30 min fully from the anesthesia and adaption of the balloon. One end of the cylinder was fixed and the other was a lid (3 cm diameter). There was a hole (1.5 cm diameter) in the middle of both ends. The hole in the fixed end was used for ventilation, while the hole in the lid was used for pulling out the tail and then to connect with a three-way tube. In this way, the mice could not move backward and forward but could move up and down, such as lifting of abdomen, body arching, and lifting of pelvic structures, and remained in a relative quiescent state. All the mice were evaluated with the same size balloon.

The AWR scores were graded on a scale of 0–4 (Figure 3): 0, no behavioral response to CRD; 1, brief head movement followed by immobility; 2, contraction of abdominal muscles; 3, lifting of abdomen; 4, body arching and lifting of pelvic structures (Gosselin et al., 2010). For measuring the pressure threshold of AWR scores 1, 2, 3, and 4, the colorectal balloon was progressively inflated, from 0 mmHg of pressure to the maximum pressure, and different degrees of pain behaviors (0-4) displayed and the realtime pressure was recorded in a pre-made form. When recording, inflation of the balloon was ceased but the pressure was retained. In this manner, the pressure threshold of AWR scores 1, 2, 3, and 4 was determined and recorded continuously. After 5 min of rest, the repeated measurement was performed. Then the mice had 5 min of rest for measuring the AWR. For measuring the AWR at different pressures of the balloon, the balloon was rapidly inflated to constant pressure (10, 20, 30, 40, 50, 60, 70, and 80 mmHg). Each pressure was maintained for 5 s, and afterwards, the balloon was rapidly and completely deflated. After 20 s of rest, the next pressure was performed. In this way, AWR at the pressures of 10-80 mmHg was determined and recorded in sequence. Likewise, after 5 min of rest, the repeated measurement was performed. Pressures are always presented from lowest to highest. All measurements were carried out by two blinded observers. They were blinded to the experimental group (NS or MS group). The average value of the measurements (the pressure threshold of AWR scores 1, 2, 3, and 4 and AWR score at the pressure of 10, 20, 30, 40, 50, 60, 70, and 80 mmHg) was recorded, respectively.

Statistical Analysis

The distribution of data was analyzed by Kolmogorov–Smirnov test. Data were shown to fit a normal distribution and were expressed as mean \pm SD. The sample size was analyzed using Power Analysis and Sample Size (PASS 15.0). Differences between two groups were determined by two-way repeated measures ANOVA with Sidak's multiple comparisons test. All data were analyzed by IBM Statistical Package for the Social Sciences (SPSS), version 23 (IBM SPSS Statistics). A *P*-value < 0.05 was considered statistically significant.

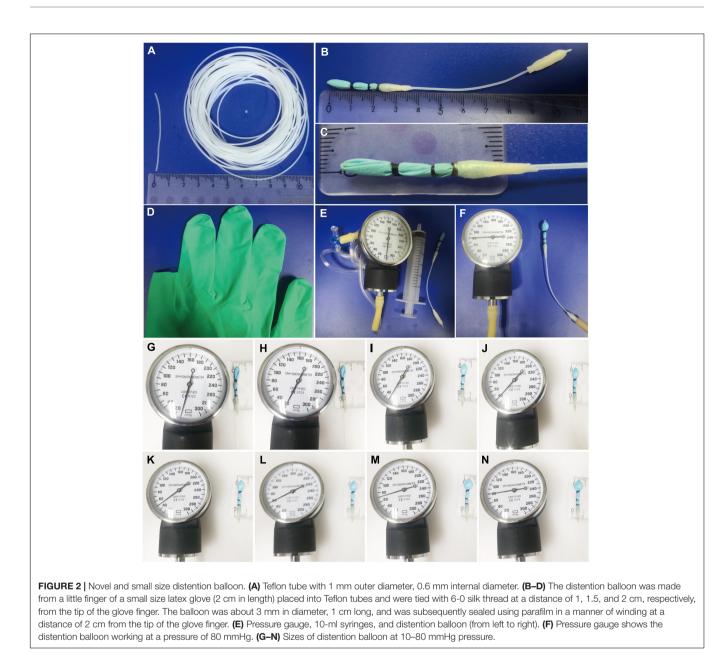
RESULTS

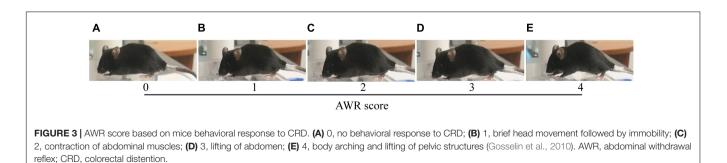
Body Weights

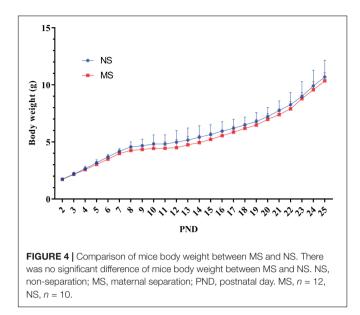
There was no significant difference of mice body weight between MS and NS [two-way repeated measures ANOVA (ELS × PND) with Sidak's multiple comparisons test, no interaction: F(23,483) = 0.55, P = 0.96; also no significant main effect of ELS: F(1,21) = 1.73, P = 0.20; however, it had a significant main effect of PND: F(23,483) = 672.5, P < 0.0001] (**Figure 4**).

Abdominal Withdrawal Reflex Threshold

The CRD threshold of ARW scores 1 (18.83 \pm 3.54 vs. 31.3 \pm 5.07 mmHg, *P* < 0.0001) (**Figure 5A**), 2 (27.25 \pm 3.49 vs. 42.05 \pm 6.15 mmHg, *P* < 0.0001) (**Figure 5B**), 3 (38.33 \pm 5.29 vs. 58.25 \pm 8.93 mmHg, *P* < 0.0001) (**Figure 5C**), and 4 (52.33 \pm 7.72 vs. 77.6 \pm 6.29 mmHg, *P* < 0.0001) (**Figure 5D**)







was significantly lower in MS compared with NS, respectively, [two-way repeated measures ANOVA (ELS × pressure) with Sidak's multiple comparisons test, interaction: F(3,60) = 16.64, P < 0.0001; and it had significant main effect of ELS: F(1,20) = 63.80, P < 0.0001; also significant main effect of pressure: F(3,60) = 617, P < 0.0001].

Abdominal Withdrawal Reflex Versus Pressure

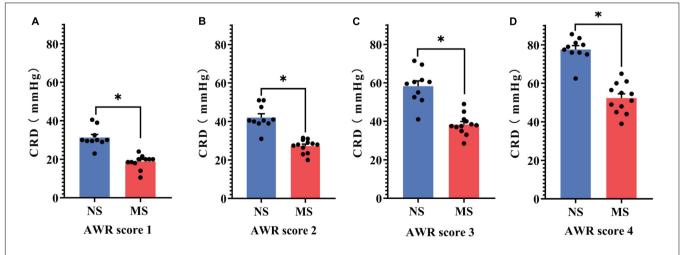
The ARW scores at 20 mmHg (1.50 ± 1.0 vs. 0.25 ± 0.35 , P < 0.0001), 30 mmHg (2.54 ± 0.92 vs. 0.75 ± 0.72 , P < 0.0001), 40 mmHg (3.13 ± 0.74 vs. 1.6 ± 0.57 , P < 0.0001), 50 mmHg (3.67 ± 0.44 vs. 2.3 ± 0.15 , P < 0.0001), and 60 mmHg (3.96 ± 0.14 vs. 3.0 ± 0.47 , P = 0.0005) were significantly higher in MS compared with NS, respectively, [two-way repeated measures

ANOVA (ELS × pressure) with Sidak's multiple comparisons test, interaction: F(7,140) = 11.20, P < 0.0001; and it had significant main effect of ELS: F(1,20) = 36.85, P < 0.0001; also significant main effect of pressure: F(7,140) = 236.1, P < 0.0001]. However, there was no difference of AWR score at 10, 70, and 80 mmHg pressure of CRD between MS and NS (P > 0.05). These results are shown in **Figure 6**.

DISCUSSION

This study evaluated visceral hypersensitivity induced by MS in post-weaning mice and developed a novel and small size distention balloon for CRD to evaluate visceral sensitivity in such mice. To our knowledge, the study is the first time to report that MS induced visceral hypersensitivity in post-weaning mice and also is the first time to develop a novel distention balloon for CRD to evaluate visceral sensitivity in such mice.

Visceral hypersensitivity as one of the major characteristics of IBS had been widely studied in rats with the model of MS (Hyland et al., 2009; Gosselin et al., 2010; Enck et al., 2016; Botschuijver et al., 2017). However, it remains difficult to study this model to mice because MS led to only mild behavioral anomalies in adult mice (Millstein and Holmes, 2007). Moloney et al. (2012) developed a mouse model of MS stress-induced visceral hyperalgesia as measured by CRD (Moloney et al., 2012); from then on, MS of mice model were extensively conducted to study the mechanism underlying the pathogenesis of IBS, such as intestinal dysbiosis, intestinal dysfunction, intestinal permeability alteration, and IBS-like anxiety and depression behaviors (De Palma et al., 2015; Li et al., 2017; Riba et al., 2017; Rincel et al., 2019; Zhou et al., 2020). Indeed, compared with the rat model, the mouse model of MS had the advantages of its small size, relatively high reproductivity, inexpensive, and can be genetically manipulated (Kong et al., 2013; Shaqour et al., 2021). Because early postnatal life is a critical period during which both brain





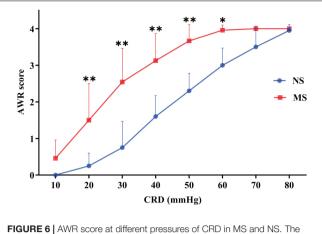


FIGURE 6 [AWR score at different pressures of CRD in MS and NS. The ARW scores at 20, 30, 40, 50, and 60 mmHg were significantly higher in MS compared with NS, respectively. However, there was no difference of AWR score at 10, 70, and 80 mmHg pressure of CRD between MS and NS (P > 0.05). CRD, colorectal distention; AWR, abdominal withdrawal reflex; MS, maternal separation; NS, non-separation, *P < 0.001, **P < 0.0001. MS, n = 12, NS, n = 10.

and gut undergo important maturation (Borre et al., 2014; Sharon et al., 2016), this period constitutes a critical window of sensitivity to stress (Rincel and Darnaudéry, 2020). Numerous studies had shown that exposure to adverse stress during childhood has been repeatedly associated with increased vulnerability to both psychiatric and gastrointestinal disorders such as IBS (Chitkara et al., 2008; Nemeroff, 2016). Moreover, a recent study focused on the effect of MS on young rodents and the process of the changes from early life to adulthood (Yi et al., 2017). Indeed, dynamic investigation of the period during early life to adult may, at least partly, contribute to exploring the underlying mechanism of pathogenesis of IBS. Accordingly, with the development of transgenic mice, optogenetic technology, tracing techniques, and so on (Ritsma et al., 2014; Boesmans et al., 2015; Chalazonitis et al., 2020; Liu et al., 2020), the novel distention balloon may be proved useful to explore the potential mechanism of pathogenesis of IBS from early life to adulthood in the future.

The underlying mechanism of MS-induced visceral hypersensitivity has been widely studied in adult rats and mice (Tang et al., 2017; Fan et al., 2020; Chen et al., 2021; Huang et al., 2021). It was reported that sensitization of corticotropin-releasing factor (CRF) neurons in the hypothalamic paraventricular nucleus (PVN) plays an important role in the pathogenesis of visceral pain (Zhang et al., 2016). A recent study suggested that a priori, PVNCRF-projecting GABAergic neurons in bed nucleus of stria terminalis (BNST)anterior ventral (AV) region participated in the occurrence of visceral hypersensitivity induced by MS in adult mice, providing new insight into the neural circuit mechanism of chronic visceral pain. Moreover, MS induced the activation of PVN mast cells, which secreted numerous proinflammatory mediators that might involve in neighboring CRF neuronal activity, ultimately directly inducing visceral hypersensitivity in adulthood rats (Chen et al., 2021), implying central immunoinflammatory process also taking part in the MS-induced visceral

hypersensitivity. A recent study also using MS paradigm revealed that MS-induced visceral pain and visceral hypersensitivity were associated with the underfunction of small-conductance calcium-activated potassium channel subtype 2 (SK2) in the spinal dorsal horn (DH). The results showed that specific ion channel was also implicated in MS-induced visceral hypersensitivity. Furthermore, Piezo2 was reported as an important mechanogated ion channel that was involved in light touch sensitivity and inflammatory allodynia (Yang et al., 2016; Szczot et al., 2018). It was reported that in IBS patients, the expression of Piezo2 significantly correlated to the visceral sensitivity (Bai et al., 2017), indicating that Piezo2 was a candidate biomarker for visceral hypersensitivity in IBS. In rats, Piezo2 knock-down in dorsal root ganglion (DRG) attenuated visceral sensation to innocuous stimuli in control rats and both innocuous and noxious stimuli with neonatal irritation (Yang et al., 2016). These results implied the potential mechanism of Piezo2 in the visceral hypersensitivity of IBS. However, whether Piezo2 involve in MS-induced visceral hypersensitivity is not clear. Accordingly, it is worth of further study. In addition, some important signaling pathways also played a role in MS-induced visceral hypersensitivity. One study demonstrated that MS caused visceral hypersensitivity by activating brain-derived neurotrophic factor (BDNF)-tyrosine kinase receptor B (TrkB)protein kinase M ζ (PKM ζ) signaling in the thoracolumbar spinal cord of adult rats (Fan et al., 2020). In addition, sensitization and upregulation of microglial toll-like receptor 4 (TLR4) signaling activity in the PVN (Tang et al., 2017), ephrin-B2/EphB1 spinal signaling pathway (Theofanous et al., 2020), and nerve growth factor (NGF)-mediated tropomyosin receptor kinase A (TrkA) signaling (Wong et al., 2019) were involved in MS-induced visceral hypersensitivity. Moreover, various important signaling molecule dysfunctions were also involved in MS-induced visceral hypersensitivity, such as elevated expression of Nesfatin-1 in the dorsal raphe nucleus (DRN) (Zhang et al., 2018). Together, these studies focused the mechanism of MS-induced visceral hypersensitivity in adulthood. However, it remains unclear when these alterations are initial and whether these alterations occur earlier, such as post-weaning. Therefore, our novel balloon will be helpful to explore the possible mechanism of MS-induced visceral hypersensitivity in early life, contributing to new treatment for IBS.

Colorectal distention is a well-documented method to assist in assessing visceral hypersensitivity (Larsson et al., 2003; Christianson and Gebhart, 2007; Annahazi et al., 2009), and different diameters and materials of the balloon were used for rats and mice (Million et al., 2006; Grabauskas et al., 2020). However, the diameters of the balloon are too large to be used for post-weaning mice, whose body weight are around 10-13 g (Figure 4). Furthermore, the distention pressure of the balloon cannot be monitored in real time (Million et al., 2006; Grabauskas et al., 2020). Thus, we develop a mini-balloon of 3 mm diameter. More importantly, the pressure of our balloon could be monitored in real time (Figure 2). In addition, our balloon is also characterized by the simplicity of the process, flexibility of distention, and the cheap cost, as it was made of latex glove and Teflon tubes (Figure 2). Moreover, using the novel balloon, both AWR threshold and AWR scores at different pressures

can be measured. Therefore, our novel balloon may be more accurate and sensitive to evaluate visceral sensitivity compared with reported balloon (Million et al., 2006; Grabauskas et al., 2020), as our novel balloon can measure visceral sensitivity at low and a series of pressures (**Figures 5, 6**). We found that compared with NS, MS resulted in visceral hypersensitivity as measured by CRD using the novel distention balloon (**Figures 5A-D, 6**). The result was consistent with a previous study in a rat model of MS, which demonstrated that MS induced visceral hypersensitivity from childhood to adulthood (Yi et al., 2017).

We paid attention to the impact of MS on body weight of the mice. On the one hand, we aimed to investigate whether MS might lead to poor weight gain in MS pups. On the other hand, a significant difference in body weight may have an impact on response to CRD with the same size balloon thereby influencing the results of CRD. We found that MS did not affect the body weight of MS pups as measured at PND 2-25 (Figure 4). Our results indicated that the MS protocol used in this study did not affect body weight, which was consistent with the results of the previous studies (George et al., 2010; Gracia-Rubio et al., 2016). However, one study indicated that MS pups got lower weight during the neonatal period in rats, and various factors have been reported to be associated with the weight of MS pups, such as breastfeeding time, mother's touching, and caloric expenditure to keep warm (Yi et al., 2017). Inconsistent results may attribute to different species, increased care behavior of dams in mice after neonatal separation, and the efforts we paid to maintain warm ambient temperature.

Our study has some limitations. For example, we did not check the influence of MS on gastrointestinal parameters in postweaning mice and also did not evaluate whether MS-induced visceral hypersensitivity at post-weaning mice persisted into adulthood. Also, we did not explore the underlying mechanisms in the visceral hypersensitivity observed in the results.

In conclusion, maternal separation induced visceral hypersensitivity in post-weaning mice, which was evaluated via a novel and small size distention balloon in this study.

REFERENCES

- Al-Chaer, E. D., Kawasaki, M., and Pasricha, P. J. (2000). A new model of chronic visceral hypersensitivity in adult rats induced by colon irritation during postnatal development. *Gastroenterology* 119, 1276–1285. doi: 10.1053/gast. 2000.19576
- Annahazi, A., Gecse, K., Dabek, M., Ait-Belgnaoui, A., Rosztoczy, A., Roka, R., et al. (2009). Fecal proteases from diarrheic-IBS and ulcerative colitis patients exert opposite effect on visceral sensitivity in mice. *Pain* 144, 209–217. doi: 10.1016/j.pain.2009.04.017
- Bai, T., Li, Y., Xia, J., Jiang, Y., Zhang, L., Wang, H., et al. (2017). Piezo2: a candidate biomarker for visceral hypersensitivity in irritable bowel syndrome? *J. Neurogastroenterol. Motil.* 23, 453–463. doi: 10.5056/jnm16114
- Barreau, F., Salvador-Cartier, C., Houdeau, E., Bueno, L., and Fioramonti, J. (2008). Long-term alterations of colonic nerve-mast cell interactions induced by neonatal maternal deprivation in rats. *Gut* 57, 582–590. doi: 10.1136/gut. 2007.126680
- Boesmans, W., Hao, M. M., and Vanden Berghe, P. (2015). Optical tools to investigate cellular activity in the intestinal wall. J. Neurogastroenterol. Motil. 21, 337–351. doi: 10.5056/jnm15096

The development of the distention balloon will be helpful to understand the mechanism of pathogenesis of IBS from early life to adulthood.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

The animal study was reviewed and approved by the Zhejiang University Ethics Committee for Animal Research.

AUTHOR CONTRIBUTIONS

ET and MJ designed the present study and critically revised the manuscript for important intellectual content. ET and GL performed the experiments. ET, GL, and TY drafted the manuscript. TY, BC, RG, DY, and MF analyzed the data. All authors have read and approved the manuscript.

FUNDING

This work was supported by a grant from the Key Research and Development Project of Zhejiang Province (2021C03064).

ACKNOWLEDGMENTS

Figure 1 was created with BioRender.com. We thank Yue Du from the Institute of Immunology, Zhejiang University School of Medicine for the gift of the small size latex glove.

- Borre, Y. E., O'keeffe, G. W., Clarke, G., Stanton, C., Dinan, T. G., and Cryan, J. F. (2014). Microbiota and neurodevelopmental windows: implications for brain disorders. *Trends Mol. Med.* 20, 509–518. doi: 10.1016/j.molmed.2014.05.002
- Botschuijver, S., Roeselers, G., Levin, E., Jonkers, D. M., Welting, O., Heinsbroek, S. E. M., et al. (2017). Intestinal fungal dysbiosis is associated with visceral hypersensitivity in patients with irritable bowel syndrome and rats. *Gastroenterology* 153, 1026–1039. doi: 10.1053/j.gastro.2017.06.004
- Botschuijver, S., Van Diest, S. A., Van Thiel, I. A. M., Saia, R. S., Strik, A. S., Yu, Z., et al. (2019). Miltefosine treatment reduces visceral hypersensitivity in a rat model for irritable bowel syndrome via multiple mechanisms. *Sci. Rep.* 9:12530. doi: 10.1038/s41598-019-49096-y
- Burke, N. N., Finn, D. P., Mcguire, B. E., and Roche, M. (2017). Psychological stress in early life as a predisposing factor for the development of chronic pain: clinical and preclinical evidence and neurobiological mechanisms. *J. Neurosci. Res.* 95, 1257–1270. doi: 10.1002/jnr.23802
- Caliskan, G., Muller, A., and Albrecht, A. (2020). Long-term impact of early-life stress on hippocampal plasticity: spotlight on astrocytes. *Int. J. Mol. Sci.* 21:4999. doi: 10.3390/ijms21144999
- Chalazonitis, A., Li, Z., Pham, T. D., Chen, J., Rao, M., Lindholm, P., et al. (2020). Cerebral dopamine neurotrophic factor is essential for enteric neuronal

development, maintenance, and regulation of gastrointestinal transit. J. Comp. Neurol. 528, 2420-2444. doi: 10.1002/cne.24901

- Chen, Z., Zhou, T., Zhang, Y., Dong, H., and Jin, W. (2021). Mast cells in the paraventricular nucleus participate in visceral hypersensitivity induced by neonatal maternal separation. *Behav. Brain Res.* 402:113113. doi: 10.1016/j.bbr. 2020.113113
- Chitkara, D. K., Van Tilburg, M. A., Blois-Martin, N., and Whitehead, W. E. (2008). Early life risk factors that contribute to irritable bowel syndrome in adults: a systematic review. *Am. J. Gastroenterol.* 103, 765–774; quiz 775. doi: 10.1111/j.1572-0241.2007.01722.x
- Christianson, J. A., and Gebhart, G. F. (2007). Assessment of colon sensitivity by luminal distension in mice. *Nat. Protoc.* 2, 2624–2631. doi: 10.1038/nprot.2007. 392
- De Palma, G., Blennerhassett, P., Lu, J., Deng, Y., Park, A. J., Green, W., et al. (2015). Microbiota and host determinants of behavioural phenotype in maternally separated mice. *Nat. Commun.* 6:7735. doi: 10.1038/ncomms8735
- Drossman, D. A. (2016). Functional gastrointestinal disorders: history, pathophysiology, clinical features and Rome IV. *Gastroenterology* S0016–5085, 223–227. doi: 10.1053/j.gastro.2016.02.032
- Enck, P., Aziz, Q., Barbara, G., Farmer, A. D., Fukudo, S., Mayer, E. A., et al. (2016). Irritable bowel syndrome. *Nat. Rev. Dis. Primers* 2:16014. doi: 10.1038/nrdp. 2016.14
- Fan, F., Tang, Y., Dai, H., Cao, Y., Sun, P., Chen, Y., et al. (2020). Blockade of BDNF signalling attenuates chronic visceral hypersensitivity in an IBS-like rat model. *Eur. J. Pain* 24, 839–850. doi: 10.1002/ejp.1534
- George, E. D., Bordner, K. A., Elwafi, H. M., and Simen, A. A. (2010). Maternal separation with early weaning: a novel mouse model of early life neglect. *BMC Neurosci.* 11:123. doi: 10.1186/1471-2202-11-123
- Gosselin, R. D., O'connor, R. M., Tramullas, M., Julio-Pieper, M., Dinan, T. G., and Cryan, J. F. (2010). Riluzole normalizes early-life stress-induced visceral hypersensitivity in rats: role of spinal glutamate reuptake mechanisms. *Gastroenterology* 138, 2418–2425. doi: 10.1053/j.gastro.2010.03.003
- Grabauskas, G., Wu, X., Gao, J., Li, J. Y., Turgeon, D. K., and Owyang, C. (2020). Prostaglandin E2, produced by mast cells in colon tissues from patients with irritable bowel syndrome, contributes to visceral hypersensitivity in mice. *Gastroenterology* 158, 2195–2207 e2196. doi: 10.1053/j.gastro.2020.02.022
- Gracia-Rubio, I., Moscoso-Castro, M., Pozo, O. J., Marcos, J., Nadal, R., and Valverde, O. (2016). Maternal separation induces neuroinflammation and long-lasting emotional alterations in mice. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 65, 104–117. doi: 10.1016/j.pnpbp.2015.09.003
- Huang, S. T., Song, Z. J., Liu, Y., Luo, W. C., Yin, Q., and Zhang, Y. M. (2021). BNST(AV) (GABA)-PVN(CRF) circuit regulates visceral hypersensitivity induced by maternal separation in Vgat-Cre mice. *Front. Pharmacol.* 12:615202. doi: 10.3389/fphar.2021.615202
- Hyland, N. P., Julio-Pieper, M., O'mahony, S. M., Bulmer, D. C., Lee, K., Quigley, E. M., et al. (2009). A distinct subset of submucosal mast cells undergoes hyperplasia following neonatal maternal separation: a role in visceral hypersensitivity? *Gut* 58, 1029–1030. doi: 10.1136/gut.2008.167882
- Iovino, P., Tremolaterra, F., Boccia, G., Miele, E., Ruju, F. M., and Staiano, A. (2009). Irritable bowel syndrome in childhood: visceral hypersensitivity and psychosocial aspects. *Neurogastroenterol. Motil.* 21, 940–e974. doi: 10.1111/j. 1365-2982.2009.01303.x
- Ju, T., Naliboff, B. D., Shih, W., Presson, A. P., Liu, C., Gupta, A., et al. (2020). Risk and protective factors related to early adverse life events in irritable bowel syndrome. *J. Clin. Gastroenterol.* 54, 63–69. doi: 10.1097/MCG. 000000000001153
- Kong, L. L., Wu, H., Cui, W. P., Zhou, W. H., Luo, P., Sun, J., et al. (2013). Advances in murine models of diabetic nephropathy. J. Diabetes Res. 2013:797548. doi: 10.1155/2013/797548
- Larsson, M., Arvidsson, S., Ekman, C., and Bayati, A. (2003). A model for chronic quantitative studies of colorectal sensitivity using balloon distension in conscious mice-effects of opioid receptor agonists. *Neurogastroenterol. Motil.* 15, 371–381. doi: 10.1046/j.1365-2982.2003.00418.x
- Li, B., Lee, C., Filler, T., Hock, A., Wu, R. Y., Li, Q., et al. (2017). Inhibition of corticotropin-releasing hormone receptor 1 and activation of receptor 2 protect against colonic injury and promote epithelium repair. *Sci. Rep.* 7:46616. doi: 10.1038/srep46616

- Liu, Y., Hegarty, S., Winter, C., Wang, F., and He, Z. (2020). Viral vectors for neuronal cell type-specific visualization and manipulations. *Curr. Opin. Neurobiol.* 63, 67–76. doi: 10.1016/j.conb.2020.03.011
- Million, M., Wang, L., Wang, Y., Adelson, D. W., Yuan, P. Q., Maillot, C., et al. (2006). CRF2 receptor activation prevents colorectal distension induced visceral pain and spinal ERK1/2 phosphorylation in rats. *Gut* 55, 172–181. doi: 10.1136/ gut.2004.051391
- Millstein, R. A., and Holmes, A. (2007). Effects of repeated maternal separation on anxiety- and depression-related phenotypes in different mouse strains. *Neurosci. Biobehav. Rev.* 31, 3–17. doi: 10.1016/j.neubiorev.2006.05.003
- Moloney, R. D., O'leary, O. F., Felice, D., Bettler, B., Dinan, T. G., and Cryan, J. F. (2012). Early-life stress induces visceral hypersensitivity in mice. *Neurosci. Lett.* 512, 99–102. doi: 10.1016/j.neulet.2012.01.066
- Nemeroff, C. B. (2016). Paradise lost: the neurobiological and clinical consequences of child abuse and neglect. *Neuron* 89, 892–909. doi: 10.1016/j.neuron.2016. 01.019
- O'Mahony, S. M., Marchesi, J. R., Scully, P., Codling, C., Ceolho, A. M., Quigley, E. M., et al. (2009). Early life stress alters behavior, immunity, and microbiota in rats: implications for irritable bowel syndrome and psychiatric illnesses. *Biol. Psychiatry* 65, 263–267. doi: 10.1016/j.biopsych.2008. 06.026
- O'Malley, D., Cryan, J. F., and Dinan, T. G. (2013). Crosstalk between interleukin-6 and corticotropin-releasing factor modulate submucosal plexus activity and colonic secretion. *Brain Behav. Immun.* 30, 115–124. doi: 10.1016/j.bbi.2013. 01.078
- Renz, H., Brandtzaeg, P., and Hornef, M. (2011). The impact of perinatal immune development on mucosal homeostasis and chronic inflammation. *Nat. Rev. Immunol.* 12, 9–23. doi: 10.1038/nri3112
- Riba, A., Olier, M., Lacroix-Lamandé, S., Lencina, C., Bacquié, V., Harkat, C., et al. (2017). Paneth cell defects induce microbiota dysbiosis in mice and promote visceral hypersensitivity. *Gastroenterology* 153, 1594–1606.e1592. doi: 10.1053/ j.gastro.2017.08.044
- Riba, A., Olier, M., Lacroix-Lamandé, S., Lencina, C., Bacquié, V., Harkat, C., et al. (2018). Early life stress in mice is a suitable model for irritable bowel syndrome but does not predispose to colitis nor increase susceptibility to enteric infections. *Brain Behav. Immun.* 73, 403–415. doi: 10.1016/j.bbi.2018. 05.024
- Rincel, M., Aubert, P., Chevalier, J., Grohard, P. A., Basso, L., Monchaux De Oliveira, C., et al. (2019). Multi-hit early life adversity affects gut microbiota, brain and behavior in a sex-dependent manner. *Brain Behav. Immun.* 80, 179–192. doi: 10.1016/j.bbi.2019.03.006
- Rincel, M., and Darnaudéry, M. (2020). Maternal separation in rodents: a journey from gut to brain and nutritional perspectives. *Proc. Nutr. Soc.* 79, 113–132. doi: 10.1017/S0029665119000958
- Ritsma, L., Ellenbroek, S. I. J., Zomer, A., Snippert, H. J., De Sauvage, F. J., Simons, B. D., et al. (2014). Intestinal crypt homeostasis revealed at singlestem-cell level by in vivo live imaging. *Nature* 507, 362–365. doi: 10.1038/nature 12972
- Shaqour, B., Aizawa, J., Guarch-Pérez, C., Górecka, Ż, Christophersen, L., Martinet, W., et al. (2021). Coupling additive manufacturing with hot melt extrusion technologies to validate a ventilator-associated pneumonia mouse model. *Pharmaceutics* 13:772. doi: 10.3390/pharmaceutics13060772
- Sharon, G., Sampson, T. R., Geschwind, D. H., and Mazmanian, S. K. (2016). The central nervous system and the gut microbiome. *Cell* 167, 915–932. doi: 10.1016/j.cell.2016.10.027
- Szczot, M., Liljencrantz, J., Ghitani, N., Barik, A., Lam, R., Thompson, J. H., et al. (2018). PIEZO2 mediates injury-induced tactile pain in mice and humans. *Sci. Transl. Med.* 10:eaat9892. doi: 10.1126/scitranslmed.aat9892
- Tang, H. L., Zhang, G., Ji, N. N., Du, L., Chen, B. B., Hua, R., et al. (2017). Toll-like receptor 4 in paraventricular nucleus mediates visceral hypersensitivity induced by maternal separation. *Front. Pharmacol.* 8:309. doi: 10.3389/fphar.2017. 00309
- Theofanous, S. A., Florens, M. V., Appeltans, I., Denadai Souza, A., Wood, J. N., Wouters, M. M., et al. (2020). Ephrin-B2 signaling in the spinal cord as a player in post-inflammatory and stress-induced visceral hypersensitivity. *Neurogastroenterol. Motil.* 32:e13782. doi: 10.1111/nmo. 13782

- Wong, H. L. X., Qin, H. Y., Tsang, S. W., Zuo, X., Che, S., Chow, C. F. W., et al. (2019). Early life stress disrupts intestinal homeostasis via NGF-TrkA signaling. *Nat. Commun.* 10:1745. doi: 10.1038/s41467-019-09744-3
- Yang, J., Zhang, J., Yang, H., Li, K., Lei, X., and Xu, C. (2016). The potential role of Piezo2 in the mediation of visceral sensation. *Neurosci. Lett.* 630, 158–163. doi: 10.1016/j.neulet.2016.07.058
- Yi, L., Zhang, H., Sun, H., Zhou, L., Chen, Y., Xuan, L., et al. (2017). Maternal separation induced visceral hypersensitivity from childhood to adulthood. *J. Neurogastroenterol. Motil.* 23, 306–315. doi: 10.5056/jnm16089
- Yu, Y. B., Zuo, X. L., Zhao, Q. J., Chen, F. X., Yang, J., Dong, Y. Y., et al. (2012). Brain-derived neurotrophic factor contributes to abdominal pain in irritable bowel syndrome. *Gut* 61, 685–694. doi: 10.1136/gutjnl-2011-300265
- Zhang, G., Yu, L., Chen, Z. Y., Zhu, J. S., Hua, R., Qin, X., et al. (2016). Activation of corticotropin-releasing factor neurons and microglia in paraventricular nucleus precipitates visceral hypersensitivity induced by colorectal distension in rats. *Brain Behav. Immun.* 55, 93–104. doi: 10.1016/j.bbi.2015.12.022
- Zhang, H. A., Sang, N., Ge, X., Huang, Q., Li, X. L., and Sha, J. (2018). Nesfatin-1 in the dorsal raphe nucleus influences visceral sensitivity via 5-HT neurons in male maternally separated rats. *Sci. Rep.* 8:9334. doi: 10.1038/s41598-018-27592-x
- Zhang, Y., Zhang, H., Zhang, W., Zhang, Y., Wang, W., and Nie, L. (2020). LncRNA XIST modulates 5-hydroxytrytophan-induced visceral hypersensitivity by epigenetic silencing of the SERT gene in mice with diarrhea-predominant IBS. *Cell Signal*. 73:109674. doi: 10.1016/j.cellsig.2020.109674

Zhou, L., Wu, Z., Wang, G., Xiao, L., Wang, H., Sun, L., et al. (2020). Long-term maternal separation potentiates depressive-like behaviours and neuroinflammation in adult male C57/BL6J mice. *Pharmacol. Biochem. Behav.* 196:172953. doi: 10.1016/j.pbb.2020.17 2953

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Tao, Long, Yang, Chen, Guo, Ye, Fang and Jiang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.