REVIEW ARTICLE



Taurine and oxidative stress in retinal health and disease

Vanessa Castelli¹ / Antonella Paladini¹ / Michele d'Angelo¹ / Marcello Allegretti² Flavio Mantelli² | Laura Brandolini² | Pasquale Cocchiaro² | Annamaria Cimini^{1,3} Giustino Varrassi⁴

¹Department of Life, Health and Environmental Sciences, University of L'Aquila, L'Aquila, Italy

²Dompé Farmaceutici SpA, L'Aquila, Italy

³Sbarro Institute for Cancer Research and Molecular Medicine and Center for Biotechnology, Temple University, Philadelphia, PA, USA

⁴Paolo Procacci Foundation, Roma, Italy

Correspondence

Giustino Varrassi, Paolo Procacci Foundation, Via Tacito 7, 00193 Roma, Italy. Email: giuvarr@gmail.com

Vanessa Castelli, Department of Life, Health and Environmental Sciences, University of L'Aquila, L'Aquila, Italy. Email: vanessa.castelli@univaq.it

Abstract

Retinal disorders are leading causes of blindness and are due to an imbalance between reactive oxygen species and antioxidant scavenger (in favor of pro-oxidant species) or a disruption of redox signaling and control. Indeed, it is well known that oxidative stress is one of the leading causes of retinal degenerative diseases. Different approaches using nutraceuticals resulted in protective effects in these disorders. This review will discuss the impact of oxidative stress in retinal neurodegenerative diseases and the potential strategies for avoiding or counteracting oxidative damage in retinal tissues, with a specific focus on taurine. Increasing data indicate that taurine may be effective in slowing down the progression of degenerative retinal diseases, thus suggesting that taurine can be a promising candidate for the prevention or as adjuvant treatment of these diseases. The mechanism by which taurine supplementation acts is mainly related to the reduction of oxidative stress. In particular, it has been demonstrated to improve retinal reduced glutathione, malondialdehyde, superoxide dismutase, and catalase activities. Antiapoptotic effects are also involved; however, the protective mechanisms exerted by taurine against retinal damage remain to be further investigated.

KEYWORDS

antioxidants, oxidative stress, retina, retinal disorders, taurine

1 | INTRODUCTION

Oxidative stress (OS) plays a major role in the neurodegenerative process.^{1,2} Retinal cell survival involves redox signaling and a balance between reactive oxygen species (ROS) and antioxidant scavengers to counteract OS injury.³⁻⁵ The retina is susceptible to OS due to its elevated oxygen consumption and exposure to visible light, which can potentiate cellular damage caused by ROS.⁶ Elevated OS levels determine dramatic changes that lead to visual impairment. Age-related macular degeneration (AMD), diabetic

retinopathy (DR), and glaucoma are ocular disorders that can lead to visual loss, and for which the involvement of ROS has been evoked.⁷ Moreover, OS is thought to induce a deficiency of cone photoreceptors in rare inherited retinopathies.⁸ High ROS levels can cause lipid peroxidation which is found at elevated amounts in the photoreceptor cell membrane.⁹ It is noteworthy that products of the oxidation of docosahexaenoic acid (DHA)-containing lipids (CEP-EPs) are observed at elevated levels both in the eyes and in serum of AMD patients compared with age-matched controls^{9,10} and are reported as activators of Toll-like receptor 2

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2021 The Authors. CNS Neuroscience & Therapeutics Published by John Wiley & Sons Ltd.

WILFY-CNS Neuroscience & Therapeutics

(TLR2) in AMD and in other retinal diseases where ROS exert a role in pathology.¹¹ The retina is endowed with an efficient innate immune system that activates three essential pathways: migration of microglia, stimulation of the complement system, and inflammasome assembly in the retinal pigment epithelium (RPE).¹² For this response, retinal cells are endowed with a variety of immune receptors and mediators such as microbial sensors (TLRs), NOD-like receptors-NLRs, RIG-1 like helicases, cytokines, chemokines, and complement components; all these players are in charge to help the cells to eliminate the insult.¹³ Under OS, the activation of this immune pathway aims to repair tissue homeostasis. Still, under continual stress, the inflammatory system's chronic hyperactivation can determine dramatic tissue changes and damage, resulting in irreversible retinal pathologies, including AMD or DR.^{14,15}

The role of ROS as a crucial cause of pathogenic inflammation in chronic disorders has been validated.^{2,16} In fact, it has been reported that pro-inflammatory cytokines, including TNF- α , interleukin-1 β , or interferon- γ , determine ROS increase in RPE cells. Indeed, these pro-inflammatory cytokines appear to increase in patients' eyes affected by glaucoma, AMD, DR, or retinal vein occlusion.^{3,17}

Taurine (2-aminoethanesulfonic acid) is a non-essential amino acid, mostly consumed with the diet.¹⁸ It is highly present in the eyes, but its physiological role is still uncertain.¹⁹ Although its role in the retina is unclear, numerous events have been attributed to taurine, involving osmoregulation, antioxidant defense, stress responses, and protein stabilization.²⁰ Taurine deficiency leads to photoreceptor degeneration but also to RGC loss. Cone photoreceptors and RGCs appear as the most sensitive cells to taurine deficiency.²¹

It has been reported that taurine levels in animals decrease with aging, and specific electroretinogram changes in rats can be associated with these decreased tissue levels, suggesting that the retina has a reduced capability to counteract the OS.²² Exogenous taurine administration may be helpful in counteracting and preventing age-related alterations in the retina.²²

Numerous potential targets have been proposed for the neuroprotective effects of taurine, including the restoration of the expression of anti- and pro-apoptotic proteins, its high antioxidant activities, the reduction of calcium influx through voltage-gated calcium channels, and the reduction of glutamate-induced excitotoxicity.^{19,23-25}

Based on the shreds of evidence exposed, this review discusses the role of OS in retinal disorders and the approaches for preventing or counteracting oxidative injuries in retinal tissues, with a particular focus on a promising candidate for their prevention, the taurine.

2 | THE ORGANIZATION OF THE RETINA

As an extension of the central nervous system, the retina displays similarities to the brain and spinal cord in terms of functionality, anatomy, and immunology.²⁶ For instance, the eye shows unique structures and different surface molecules and cytokines and has a specialized immune system similar to those of the brain and spinal

cord.²⁶ Moreover, in terms of anatomy both the retina and the brain present a barrier that impede circulating pathogens or toxins that could induce infections, but at the same time regulate the passage of vital nutrients.²⁶ Blood flow alterations in the brain, due to ischemia for example, are strictly related to blood flow alterations occurring in the eye following by visual impairments.^{27,28}

Visual processing starts in the retina: a thin, multilayered tissue composed of light-sensitive neurons lining the back of the vertebrate eye.²⁹ The retina elaborates the light produced from visual images through transduction (transferring energy from one form to another) and transfers these data to the brain for the perceptual appreciation of the images.³⁰ All vertebrate retinas are composed of three layers of nerve cell bodies, the outer nuclear layer (ONL), the inner nuclear layer (INL), and the ganglion cell layer; and two layers of synapses.^{31,32} ONL is composed of nuclei of photoreceptor cells, which are of two types: rods and cones; the rods and cones synapse with the bipolar cells are in the second layer. INL contains the nuclei and cell bodies of the bipolar, horizontal, and amacrine cells, as well as Müller glial cells.³² Bipolar cells spread their extremities to transmit with both the first and third layers. Müller cells are the principal glial cell of the retina and stretch radially across the thickness of the retina. They are responsible for the homeostatic and metabolic support of retinal neurons³¹ (Figure 1).

Rods work mostly in dim light and create no-colored images. Cones operate in well-light circumstances and permit the perception of colors and for high-awareness vision utilized for tasks like reading. The third type of light-sensitive cells, the ganglion cells, is essential for the entrainment of circadian rhythms and reflexive reactions.^{31,33}

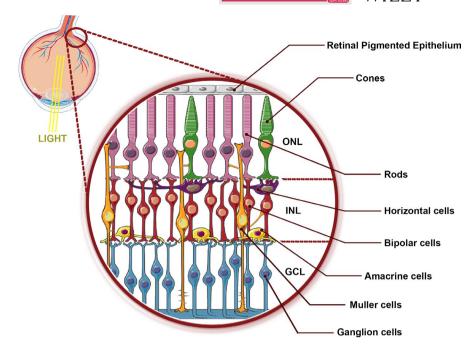
Neural signals from the rods and cones are processed by other neurons, whose output brings action potentials in RGCs whose axons generate the optic nerve.³⁴ The axons of RGCs in the retina's third layer convey the visual data as coded by the retina to the following synapse point in the visual pathway through the optic nerve.³⁵ Amacrine and horizontal cells situated in the INL participate in visual information processing through lateral contacts.³⁶ These lateral contacts regulate data transmission through the retina synaptic layers, between the first and second layer and the second and third layer. This intricate system of neurons collects the transduced visual data and processes them by compression, encoding, convergence, and integration.³⁰

3 | OXIDATIVE STRESS IN RETINAL DEGENERATIVE DISEASES

Aging, gene alterations, and excessive exposure to exogenous oxidative stressors (eg, a light exposure) increase oxidative stress in the eye. The relationship between oxidative stress and retinal disorders has been established.^{6,7,22} Oxidative stress shows a crucial role in the onset and progression of retinal disorders, comprising DR, AMD, and glaucoma.^{6,8}

405

FIGURE 1 Organization of the mature retina. ONL: outer nuclear layer; INL: inner nuclear layer; and GCL: ganglion cell layer. In the ONL resides the rod and cone photoreceptor cells. The INL contains the amacrine cells, the horizontal cells, and the bipolar cells. The Müller cell body is in this layer, while the processes of the develop into the contiguous layers, expanding all through the thickness of the retina. The GCL mostly contains of the ganglion cells which send their axons out of the eye via the optic disk



3.1 | Diabetic retinopathy

DR is one of the leading causes of visual impairment and is one of the most common microvascular complications of diabetes.³⁷ The polyol pathway is one of the high glucose-induced metabolic alterations in DR. It converts the glucose in sorbitol, and the reaction is catalyzed by aldose reductase. Sorbitol is then oxidized to produce fructose by sorbitol dehydrogenase. Enhanced polyol pathway in diabetes increases the OS because aldose reductase needs NADPH.³⁸ This event may reduce the availability of NADPH for stimulating the intracellular antioxidant. GSH.^{39,40} The production of advanced glycation end products (AGEs) is another pathway causing the detrimental consequences of glucose. AGEs are generated from potent glycating dicarbonyl elements, that is, methylglyoxal and glyoxal.⁴¹ Chronic hyperglycemia supports enzymatic and non-enzymatic glycation, inducing altered functions and degradation of intracellular and extracellular proteins by chemical rearrangement and cross-linking. AGEs are created on amino groups of proteins, lipids, and DNA, leading to intramolecular and intermolecular cross-links.⁴¹ In diabetic conditions, the accumulation of AGEs and its receptor, RAGE, are increased in retinal capillary cells.⁴² In the later phases of DR, AGEs are irreversibly generated and deposited in the retinal microvasculature.^{42,43} AGEs enhance nitrative stress in the capillary cells and trigger apoptosis events leading to retinal capillary cell death and pathological consequences.44,45

The activation of protein kinase C (PKC) pathway is also involved in the pathogenesis of DR.⁴⁶⁻⁴⁸ High glucose levels enhance ROS levels, and the synthesis of diacylglycerol, inducing PKC activation.⁴⁹ Activated PKC can determine numerous alterations typical of DR, including enhancing vessel permeability, blood flow, endothelial proliferation, apoptosis, altered hormone and growth factor receptor recycling, increased neovascularization, and regulating various factors as vascular endothelial growth factor, insulin-like growth factor-1, and transforming growth factor $\beta.^{44,50}$ Inhibition of PKC by PKC β specific inhibitor (LY53331) was able to avoid diabetes-induced OS. 44,51 Ohshiro and colleagues showed that lack of PKC β isoform in mice protected them from diabetes-induced OS. 52 These studies linked OS and PKC supported PKC's role in ROS-mediated diabetic problems.

In diabetes, glucose oxidation is enhanced, generating an elevated voltage gradient across the mitochondrial membrane.¹⁷ One of the ROS-induced impairments in mitochondria is the reduced antioxidant defense that may increase retinal cells' sensitivity to OS. The isoform of SOD in the mitochondria, MnSOD, and GSH are inhibited in diabetic patients and high glucose-cultured retinal mitochondria.⁵³ Mitochondrial impairment also involves injury to mitochondrial DNA,² which also occurs in the diabetic retina.⁵⁴ Enhanced swelling of mitochondrial lipid membranes is detected in the retina of diabetic mice.⁵⁴ The inner mitochondrial membrane includes numerous soluble proteins, including cytochrome c. The release of cytochrome c from mitochondria to the cytoplasm and Bax translocation from the cytosol to mitochondria are enhanced in capillary cells and in the retina in diabetic conditions, events that lead to apoptosis.⁵⁵ Therefore, it is clear that OS can regulate mitochondria activity, causing higher apoptosis in retinal microvasculature; further investigations to define the role of OS-induced mitochondrial impairments in DR are necessary. Antioxidants may act at various levels; they may prevent ROS formation or scavenge free radicals or enhance antioxidant defenses.

3.2 | Age-related macular degeneration

AMD is a multifactorial disorder,⁵⁶ and its pathogenesis remains unclear. Evidence indicates that an intricate interaction of WILFY-CNS Neuroscience & Therapeutics

environmental, genetic, and metabolic factors contributes to the pathology of AMD.⁵⁷ AMD is the main reason for visual impairment in the elderly. AMD initially alters the RPE and gradually leads to secondary loss of photoreceptor cells.^{58,59} It is characterized by the degeneration of the macula, described by high number of cone photoreceptors responsible for visual perception and color vision. In AMD, three stages have been recognized: the early stage, the intermediate stage, and the late stage (based on the most recent Three Continent AMD Consortium Severity Scale⁶⁰ and Clinical Classification⁶¹). The early stage (mild and moderate or severe) is marked by the development of several small drusen or a few medium-sized drusen⁶²; the intermediate stage is characterized by some pigmentary abnormalities and large drusen; and the late stage with several large drusen is characterized by two forms: non-exudative ("dry form") and an exudative/neovascular ("wet form").⁶³ The "dry form" is characterized by atrophic alterations in the macula and, clinically, has better conservation of visual acuity than "wet form".⁶² "Wet" AMD is characterized by atypical blood vessels in the choriocapillaris, which is the formation of new through Bruch's membrane. These vessels lead to bleeding and leakage into the macula and ultimately induce irreversible damage to photoreceptors if untreated.⁶² The "wet form" leads to significant incidence of substantial visual impairment.⁶⁴ End-stage macular degeneration is the last and irreversible stage, and the patients show visual loss and cannot be longer treated with surgery or ocular injections.

Different pathways are involved in AMD, including OS, apoptosis, the formation of drusen and RPE aberration, immune system activation, senescent failure of homeostatic control, and Bruch's membrane defects.⁶⁴ During aging, antioxidant decreases, and ROS level increases, supporting OS.^{2,65} Furthermore, glutathione S-transferase-1 expression level,⁶⁶ macular carotenoids level,⁶⁷ and vitamin E level⁶⁸ are reduced. On the other hand, lipid peroxidation is enhanced,⁶⁹ and lipofuscin,^{70,71} altered mitochondrial DNA in the retina,⁷² and advanced lipid peroxidation and glycation end products⁷³ are enhanced. To date, there is no therapy accessible for the "dry type" AMD. In the Age-Related Eye Disease Study (AREDS), nutraceuticals (AREDS and AREDS2), comprising vitamins C and E, β -carotene, and zinc, counteracted the disease progression from intermediate to advanced AMD by about 25%.⁷⁴ However, AREDS and AREDS2 supplements do not prevent AMD onset. AREDS investigators followed participants for an additional five years (ten year in total).75-77

For the "wet-type," the anti-vascular endothelial growth factor (VEGF) antibody is generally used as standard therapy able to ameliorate patients' visual function.^{78,79} The route of administration is intravitreal injection⁸⁰; but this procedure is invasive and is related to the possibility of infection.^{81,82} Also, the anti-VEGF antibody is costly; consequently, it is crucial to develop new therapeutic approaches for this pathology.

On light of the pathways involved in AMD, it has been suggested that antioxidant supplement may counteract cellular damage in the retina by reacting with free radicals that are produced in the process of light absorption, thus reducing the risk and progression of AMD .⁸³

3.3 | Glaucoma

Another disorder that leads to irreversible blindness is glaucoma. Open-angle glaucoma (OAG) is the most common form. OAG is characterized by degeneration of the trabecular meshwork (TM) which rises the intraocular pressure, which in turn, lead to altered axons of RGCs forming the optical nerve, and then progressive concentric damage of the RGCs.⁸⁴⁻⁸⁶ Degeneration of these cells results in a typical form of the optic disk (cupping) and visual impairment. Furthermore, it is characterized by retinal nerve fiber layer variations and typical visual field defects.⁸⁷ The biological mechanism of glaucoma is poorly recognized, and the factors supporting its progress have not been entirely described.^{84,88} One of the leading risk factors for glaucoma advancement, and the only adaptable factor, is elevated intraocular pressure.⁸⁹ Even if the mechanism is still uncertain, early neuroinflammation is indicated as an underlying trigger of glaucoma pathology.^{90,91} Like AMD, glaucoma is also associated with OS.^{92,93} Indeed, it has been reported that the progressive loss of TM cells in glaucoma patients may be attributed to the long-term effects of oxidative injury induced by free radicals.^{94,95} This hypothesis was then supported by in vitro and in vivo studies. Human TM in vitro upon hydrogen peroxide showed loss of cellular integrity and reduced cell adhesion.⁹⁶ In vivo, calf TM treated with hydrogen peroxide showed altered the mechanism of drainage of the aqueous humor from the anterior chamber of the eye.⁹⁷ Combined treatment with trophic and antioxidant factors was able to prevent the RGCs death in rats with elevated IOP.98 Also, it has been reported that oxidative damage to DNA is considerably higher in the TM of glaucomatous patients compared to controls.⁹⁹ Additionally, in patients, IOP increase and visual impairment are proportional to the amount of oxidative DNA damage affecting TM cells.¹⁰⁰ In addition, the plasma level of glutathione, an important antioxidant, resulted reduced in glaucoma patients.^{101,102} On light of the exposed evidence, antioxidant therapies could help counteract or reduce this pathology.

4 | TAURINE IN THE RETINA AND RETINAL DISORDERS

In the retina of mammals, taurine is the most copious amino acid during development and adulthood.²⁵ Moreover, the retina appears to be the taurine richest organ,¹⁹ with concentrations higher than any other ocular structures or the brain,¹⁰³ reaching up to 50 mmol/g tissue in rats. Retinal taurine is provided by Müller cells and RPE, which generally collect taurine and transfer it to photoreceptor cells.¹⁰⁴ Photoreceptor cells are significantly rich in taurine, and all retinal cells use taurine from the extracellular environment. Highand low-affinity Na⁺ and Cl⁻-dependent taurine transporters have been reported in the retina. Also, it has been reported that taurine treatment can avoid or counteract retinal neurodegeneration.²³ Thus, photoreceptors require a sufficient amount of extracellular taurine, depending on their transporter for osmoregulation.^{20,105} However, its function in the retina is still uncertain.

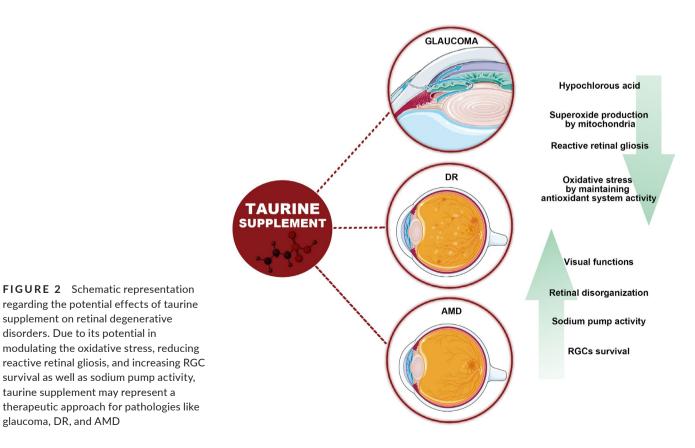
In humans, chronic parenteral nutrition absent of taurine resulted in reduced plasma taurine concentrations and atypical electroretinograms in children and conceivably in adults.^{25,106}

A research group investigated if light exposure exacerbated retinal neuronal loss induced by taurine depletion.¹⁰⁷ As a model, they used albino rats receiving β -alanine (which causes taurine depletion) in the drinking water, and after one month of treatment, 50% of the rats were subjected to white light (3000 lux).¹⁰⁷ The results indicated that light exposure under taurine depletion increased photoreceptor degeneration, suggesting that taurine is essential for retinal survival and for light-induced photoreceptor degeneration of the ration.¹⁰⁷ Consequently, the taurine supplement may counteract degeneration of the retina, particularly S-cone degeneration or can be useful for the treatment of pathologies for which light may represent an etiologic feature.

One of the main cytoprotective effects of taurine includes its antioxidant activity, mediated by three different processes. First, taurine counteracts the neutrophil oxidant, hypochlorous acid. The product of the reaction between taurine and hypochlorous acid, taurine chloramine, also hinders with the inflammatory pathway.^{108,109} Second, taurine reduces the production of superoxide by mitochondria.^{25,110} Third, mitochondrial ROS can impair antioxidant enzyme activity in balancing OS.^{3,110} Because some antioxidant enzymes are receptive to oxidative injury, taurine may counteract OS by avoiding this enzyme impairment. As mentioned above, taurine deficiency leads to photoreceptor degeneration and also to retinal ganglion cell loss^{21,111}; thus, taurine therapy may exert an essential role in preventing retinal degeneration¹⁰⁶ (Figure 2).

RGC degeneration appears in several retinal disorders leading to visual loss, either as a primary process like in glaucoma or secondary to photoreceptor loss.⁸⁶ However, to date, there is no available treatment directly targeting RGCs neuroprotection. As we mentioned above, taurine seems to be essential for photoreceptors' survival since its deprivation is related to retinal death. Froger et al¹¹¹ studied the taurine effect on RGCs in vitro models and various animal models of RGC degeneration. Taurine protective effects were evaluated in vitro on primary RCG cultures in serum-free conditions and on N-methyl-D-aspartate (NMDA)treated retinal explants from adult rats. In vivo, two glaucomatous models (mice and rats with vein occlusion) and a model of RP with secondary RGC degeneration (P23H rats) were used. Taurine was administered in the drinking water for 6 days.¹¹¹ Notably, taurine significantly improved RGC survival (+68%) in vitro and partly prevented NMDA-induced RGC excitotoxicity. In vivo, taurine administration was also able to increase RGC densities in both animal models compared to control groups. This study indicated that enriched taurine nutrition could directly maintain RGC survival, reducing the OS, positively affecting retinal degenerative disorders.¹¹¹

Furthermore, it has been reported that taurine supplementation may be protective and promising therapeutic approach for retinopathies with a chronic cycle, for example, retinitis pigmentosa, an



WII FY-CNS Neuroscience & Therapeutics

inherited disorder characterized by a progressive degeneration of rod photoreceptors.¹¹² Indeed, in a mouse model of N-methyl-Nnitrosourea (MNU)-induced retinal degeneration, intravenous taurine therapy broadly improved the retinal taurine concentration. Morphological experiments revealed that taurine ameliorated the retinal disorganization in the MNU-induced animals. Furthermore, taurine was able to ameliorate the vision loss in the MNU-induced animals, as demonstrated by functional analyses (ie, electroretinogram and optokinetic test). Immunostaining analyses showed that taurine ameliorated both M-cone and S-cone populations in the degenerative retinas. Regarding the mechanism, the OS and photoreceptor apoptosis in the degenerative retina were strongly reduced by taurine.¹¹³

In another interesting study, Arfuzir et al¹¹⁴ evaluated taurine neuroprotective properties against glaucoma. In particular, they used endothelin-1 (ET-1)-induced retinal and optic nerve damage. ET-1 was administered intravitreally to Sprague-Dawley rats, and taurine was injected as pre-, co-, or post-treatment.¹¹⁴ This study suggested that the treatment with taurine, particularly in a preventive regimen, prevented apoptosis of retinal cells induced by ET-1 and prevented the changes in the morphology of the retina and optic nerve. The protective effect of taurine was also associated with reduced retinal OS. In particular, taurine was able to improve retinal reduced glutathione, malondialdehyde, superoxide dismutase, and catalase activities.¹¹⁴

Another group examined the consequences and the main taurine mechanisms on hyperglycemia-induced variations of Müller cells' glutamate degradation and uptake. A decreased capability of Müller cells to eliminate glutamate from the extracellular space is critical in the disruption of glutamate homeostasis that appears in the diabetic retina. Taurine substantially reduced the high glucose-induced reductions in glutamate uptake and counteracted OS induced by high glucose, increasing the antioxidant enzyme events. These results indicate that taurine might control Müller cells' glutamate uptake and degradation under diabetic conditions through its antioxidant activity.¹¹⁵

Recently, Fan et al investigated the role and mechanisms of taurine supplementation (intraperitoneally or intragastrically) in early diabetic retinas using an eight-week-old streptozotocin (STZ)-induced diabetic rats. Taurine protected retinal cone cells as well as RGCs from diabetic attacks by activating retinal taurine transporter, reducing reactive retinal gliosis, enhancing retinal synaptic connections, and reducing retinal cell apoptosis.¹¹⁶

Another study examined the chronic taurine treatment vs. a mixture of vitamin E and selenium on biochemical retinal alterations caused by diabetes at different disease stages. STZ-diabetic rats were treated for 4 months, and taurine was able to significantly reduce retinal OS and to enhance sodium pump activity in experimental diabetes in a dose- and time-dependent manner.¹¹⁷ Overall, these findings strengthen the hypothesis that taurine could represent a novel approach for DR.

A recent study evaluated taurine's effects in a family with taurine deficiency (homozygous amino acid substitution in the third transmembrane domain of the taurine transporter SLC6A6). In particular, the authors evaluated taurine levels in the blood and analyzed the fundus and macular with optical coherence tomography after 2 years of taurine supplementation. Interestingly, the retinal degeneration was counteracted, and the vision was clinically ameliorated mostly in the youngest patients (6 years old).¹¹⁸

5 | DISCUSSION AND CONCLUSIONS

It is well known that OS is one of the leading causes of neurodegeneration and retinal degenerative disorders. Indeed, different approaches using antioxidants resulted in protective effects in these disorders.¹¹⁹⁻¹²¹ Researchers focused on the effects of taurine in retinal disorders. In this review, we reported evidence of the protective role of taurine against retinal functional and morphological injuries in animal and in in vitro retinal disease models, thus implying that taurine may have a therapeutic potential in the treatment of retinal and degenerative disorders. Indeed, increasing data indicate that taurine may be effective in slowing down the progression of retinal diseases, thus suggesting that taurine can be a promising candidate for the prevention or as adjuvant treatment of these diseases. The mechanism by which taurine supplementation acts is mainly reducing the OS, even also antiapoptotic effects are involved; however, the protective effects of taurine against retinal damage still unclear. Further investigations should focus on the way in which taurine protects cells against oxidative stress and toxicity at cellular level and determine whether other treatments can trigger these neuroprotective pathways. Along with other antioxidant molecules, taurine should therefore be strongly reconsidered as a potential treatment for retinal diseases.

6 | METHODS

Extensive bibliographic research was conducted using the PubMed National Library of Medicine (NIH), Web of Science platform, Google Scholar, and Clinical Key databases. Examples of the search terms used were "Taurine" "oxidative stress", "retina", "therapies", "in vitro", "in vivo" "health retina" "retinal degeneration". For screening, a restriction was made to those articles published in the last 10 years and preferably in English. Priority was given to prospective studies and reviews with a clear and well-described methods section. In addition, a secondary search of the bibliography of the articles finally selected was carried out to detect possible omissions. For the analysis of all relevant publications, consensus meetings were held with all the authors.

ACKNOWLEDGEMENTS

The editorial activities have been generously supported by Paolo Procacci Foundation.

CONFLICTS OF INTEREST

The authors declare they have no conflict of interest with the material reported in this article.

AUTHOR CONTRIBUTIONS

VC wrote the manuscript with support from AC and GV. AP, MA, FM, LB, and PC substantively revised it. MdA prepared all the figures. All authors contributed to the final manuscript. All authors have read and agreed to the published version of the manuscript.

DATA AVAILABILITY STATEMENT

Not applicable.

ORCID

Vanessa Castelli D https://orcid.org/0000-0002-7005-3218 Michele d'Angelo https://orcid.org/0000-0002-3693-840X Annamaria Cimini https://orcid.org/0000-0002-2737-7970

REFERENCES

- 1. Rekatsina M, Paladini A, Piroli A, Zis P, Pergolizzi JV, Varrassi G. Pathophysiology and therapeutic perspectives of oxidative stress and neurodegenerative diseases: a narrative review. *Adv Ther.* 2020;37(1):113-139. https://doi.org/10.1007/s12325-019-01148-5
- Castelli V, Benedetti E, Antonosante A, et al. Neuronal cells rearrangement during aging and neurodegenerative disease: metabolism, oxidative stress and organelles dynamic. *Front Mol Neurosci*. 2019;12:132. https://doi.org/10.3389/fnmol.2019.00132
- Nita M, Grzybowski A. The role of the reactive oxygen species and oxidative stress in the pathomechanism of the age-related ocular diseases and other pathologies of the anterior and posterior eye segments in adults. Oxid Med Cell Longev. 2016;2016:3164734. https://doi.org/10.1155/2016/3164734
- Jones DP. Redefining oxidative stress. Antioxid Redox Signal. 2006;8(9-10):1865-1879. https://doi.org/10.1089/ ars.2006.8.1865.
- Wu L, Zhang K, Sun L, Bai J, Zhang M, Zheng J. Laminin degradation by matrix metalloproteinase 9 promotes ketamine-induced neuronal apoptosis in the early developing rat retina. CNS Neurosci Ther. 2020;26(10):1058-1068. https://doi.org/10.1111/ cns.13428.
- Masuda T, Shimazawa M, Hara H. Retinal diseases associated with oxidative stress and the effects of a free radical scavenger (Edaravone). Oxid Med Cell Longev. 2017;2017:1-14. https://doi. org/10.1155/2017/9208489.
- Nishimura Y, Hara H, Kondo M, Hong S, Matsugi T. Oxidative stress in retinal diseases. Oxid Med Cell Longev. 2017;2017:1-2. https://doi.org/10.1155/2017/4076518
- B Domènech E, Marfany G. The relevance of oxidative stress in the pathogenesis and therapy of retinal dystrophies. *Antioxidants* (*Basel*). 2020;9(4):347. https://doi.org/10.3390/antiox9040347
- Shindou H, Koso H, Sasaki J, et al. Docosahexaenoic acid preserves visual function by maintaining correct disc morphology in retinal photoreceptor cells. J Biol Chem. 2017;292(29):12054-12064. https://doi.org/10.1074/jbc.M117.790568
- Wong BH, Chan JP, Cazenave-Gassiot A, et al. Mfsd2a is a transporter for the essential ω-3 fatty acid docosahexaenoic acid (DHA) in eye and is important for photoreceptor cell development. *J Biol Chem.* 2016;291(20):10501-10514. https://doi.org/10.1074/jbc. M116.721340
- Mulfaul K, Ozaki E, Fernando N, et al. Toll-like receptor 2 facilitates oxidative damage-induced retinal degeneration. *Cell Rep.* 2020;30(7):2209-2224.e5. https://doi.org/10.1016/j. celrep.2020.01.064
- 12. Akhtar-Schäfer I, Wang L, Krohne TU, Xu H, Langmann T. Modulation of three key innate immune pathways for the

most common retinal degenerative diseases. *EMBO Mol Med.* 2018;10(10):e8259. https://doi.org/10.15252/emmm.20170 8259

- Detrick B, Hooks JJ. The RPE cell and the immune system. In: Klettner AK, Dithmar S, eds. *Retinal Pigment Epithelium in Health* and Disease. Berlin: Springer International Publishing; 2020:101-114. https://doi.org/10.1007/978-3-030-28384-1_6
- Birch DG, Liang FQ. Age-related macular degeneration: a target for nanotechnology derived medicines. *Int J Nanomedicine*. 2007;2(1):65-77. https://doi.org/10.2147/nano.2007.2.1.65
- Rashid K, Akhtar-Schaefer I, Langmann T. Microglia in retinal degeneration. Front Immunol. 2019;10:1975. https://doi. org/10.3389/fimmu.2019.01975
- Manoharan S, Guillemin GJ, Abiramasundari RS, Essa MM, Akbar M, Akbar MD. The role of reactive oxygen species in the pathogenesis of alzheimer's disease, Parkinson's disease, and Huntington's disease: a mini review. Oxid Med Cell Longev. 2016;2016:8590578. https://doi.org/10.1155/2016/8590578
- Burgos-Morón E, Abad-Jiménez Z, de Marañón AM, et al. Relationship between oxidative stress, ER stress, and inflammation in type 2 diabetes: the battle continues. J Clin Med. 2019;8(9):1385. https://doi.org/10.3390/jcm8091385
- Yuan Y-S, Zhou X-J, Tong Q, et al. Change in plasma levels of amino acid neurotransmitters and its correlation with clinical heterogeneity in early Parkinson's disease patients. CNS Neurosci Ther. 2013;19(11):889-896. https://doi.org/10.1111/cns.12165
- Ripps H, Shen W. Review: taurine: a "very essential" amino acid. Mol Vis. 2012;18:2673-2686.
- El-Sherbeny A, Naggar H, Miyauchi S, et al. Osmoregulation of taurine transporter function and expression in retinal pigment epithelial, ganglion, and Müller cells. *Invest Ophthalmol Vis Sci.* 2004;45(2):694-701. https://doi.org/10.1167/iovs.03-0503.
- Gaucher D, Arnault E, Husson Z, et al. Taurine deficiency damages retinal neurones: cone photoreceptors and retinal ganglion cells. *Amino Acids*. 2012;43(5):1979-1993. https://doi.org/10.1007/ s00726-012-1273-3
- Militante J, Lombardini JB. Age-related retinal degeneration in animal models of aging: possible involvement of taurine deficiency and oxidative stress. *Neurochem Res.* 2004;29(1):151-160. https:// doi.org/10.1023/b:nere.0000010444.97959.1b
- Hadj-Saïd W, Fradot V, Ivkovic I, Sahel J-A, Picaud S, Froger N. Taurine promotes retinal ganglion cell survival through GABAB receptor activation. Adv Exp Med Biol. 2017;975(Pt 2):687-701. https://doi.org/10.1007/978-94-024-1079-2_54
- Sun M, Xu C. Neuroprotective mechanism of taurine due to up-regulating calpastatin and down-regulating calpain and Caspase-3 during focal cerebral ischemia. *Cell Mol Neurobiol*. 2008;28(4):593-611. https://doi.org/10.1007/s10571-007-9183-8
- Schaffer S, Kim HW. Effects and mechanisms of taurine as a therapeutic agent. *Biomol Ther* (*Seoul*). 2018;26(3):225-241. https://doi. org/10.4062/biomolther.2017.251
- London A, Benhar I, Schwartz M. The retina as a window to the brain—from eye research to CNS disorders. *Nat Rev Neurol.* 2013;9(1):44-53. https://doi.org/10.1038/nrneurol.2012.227
- Lee J-Y, Castelli V, Bonsack B, et al. Eyeballing stroke: Blood flow alterations in the eye and visual impairments following transient middle cerebral artery occlusion in adult rats. *Cell Transplant*. 2020;29:963689720905805. https://doi.org/10.1177/09636 89720905805
- Nguyen H, Lee JY, Sanberg PR, Napoli E, Borlongan CV. Eye opener in stroke. Stroke. 2019;50(8):2197-2206. https://doi.org/10.1161/ STROKEAHA.119.025249
- Hunter JJ, Merigan WH, Schallek JB. Imaging retinal activity in the living eye. Annu Rev Vis Sci. 2019;5:15-45. https://doi.org/10.1146/ annurev-vision-091517-034239

WILFY-CNS Neuroscience & Therapeutics

- Masland RH. The neuronal organization of the retina. Neuron. 2012;76(2):266-280. https://doi.org/10.1016/j. neuron.2012.10.002
- Mahabadi N, Al Khalili Y. Neuroanatomy, retina. In: StatPearls. StatPearls Publishing; 2020. http://www.ncbi.nlm.nih.gov/books/ NBK545310/. Accessed October 16, 2020.
- Remington LA. Retina. In: Clinical Anatomy and Physiology of the Visual System. Amsterdam: Elsevier; 2012:61-92. https://doi. org/10.1016/B978-1-4377-1926-0.10004-9
- Kandel ER, Schwartz JH, Jessell TM. Principles of Neural Science, 4th Ed. New York, NY: McGraw-Hill; 2000.
- Demb JB, Singer JH. Functional circuitry of the retina. Annu Rev Vis Sci. 2015;1:263-289. https://doi.org/10.1146/annurev-visio n-082114-035334
- Gupta M, Bordoni B. Neuroanatomy, visual pathway. In: StatPearls. StatPearls Publishing; 2020. http://www.ncbi.nlm.nih.gov/books/ NBK553189/. Accessed October 16, 2020.
- Asari H, Meister M. The projective field of retinal bipolar cells and its modulation by visual context. *Neuron*. 2014;81(3):641-652. https://doi.org/10.1016/j.neuron.2013.11.029
- Zhang W, Liu H, Al-Shabrawey M, Caldwell RW, Caldwell RB. Inflammation and diabetic retinal microvascular complications. J Cardiovasc Dis Res. 2011;2(2):96-103. https://doi. org/10.4103/0975-3583.83035
- Yan L-J. Redox imbalance stress in diabetes mellitus: role of the polyol pathway. *Animal Model Exp Med*. 2018;1(1):7-13. https://doi. org/10.1002/ame2.12001
- Miwa K, Nakamura J, Hamada Y, et al. The role of polyol pathway in glucose-induced apoptosis of cultured retinal pericytes. Diabetes Res Clin Pract. 2003;60(1):1-9. https://doi.org/10.1016/ s0168-8227(02)00248-6
- 40. Furukawa A, Koriyama Y. A role of heat shock protein 70 in photoreceptor cell death: potential as a novel therapeutic target in retinal degeneration. *CNS Neurosci Ther.* 2016;22(1):7-14. https://doi. org/10.1111/cns.12471
- Glomb MA, Monnier VM. Mechanism of protein modification by glyoxal and glycolaldehyde, reactive intermediates of the Maillard reaction. J Biol Chem. 1995;270(17):10017-10026. https://doi. org/10.1074/jbc.270.17.10017
- Stitt AW. The role of advanced glycation in the pathogenesis of diabetic retinopathy. *Exp Mol Pathol*. 2003;75(1):95-108. https:// doi.org/10.1016/s0014-4800(03)00035-2
- Mohamed AK, Bierhaus A, Schiekofer S, Tritschler H, Ziegler R, Nawroth PP. The role of oxidative stress and NF-kappaB activation in late diabetic complications. *BioFactors*. 1999;10(2–3):157-167. https://doi.org/10.1002/biof.5520100211
- Kowluru RA. Effect of advanced glycation end products on accelerated apoptosis of retinal capillary cells under in vitro conditions. *Life Sci.* 2005;76(9):1051-1060. https://doi.org/10.1016/j. lfs.2004.10.017
- Cowell RM, Russell JW. Nitrosative injury and antioxidant therapy in the management of diabetic neuropathy. J Investig Med. 2004;52(1):33-44. https://doi.org/10.1136/jim-52-01-24
- Caldwell RB, Bartoli M, Behzadian MA, et al. Vascular endothelial growth factor and diabetic retinopathy: role of oxidative stress. *Curr Drug Targets*. 2005;6(4):511-524. https://doi. org/10.2174/1389450054021981
- Safi SZ, Qvist R, Kumar S, Batumalaie K, Ismail ISB. Molecular mechanisms of diabetic retinopathy, general preventive strategies, and novel therapeutic targets. *Biomed Res Int.* 2014;2014:801269. https://doi.org/10.1155/2014/801269
- Aiello LP. The potential role of PKC β in diabetic retinopathy and macular edema. Surv Ophthalmol. 2002;47:S263-S269. https://doi. org/10.1016/S0039-6257(02)00391-0

- Volpe CMO, Villar-Delfino PH, Dos Anjos PMF, Nogueira-Machado JA. Cellular death, reactive oxygen species (ROS) and diabetic complications. *Cell Death Dis.* 2018;9(2):119. https://doi. org/10.1038/s41419-017-0135-z
- Antonetti DA, Klein R, Gardner TW. Diabetic retinopathy. N Engl J Med. 2012;366(13):1227-1239. https://doi.org/10.1056/NEJMr a1005073
- 51. Wu Y, Wu G, Qi X, et al. Protein kinase C beta inhibitor LY333531 attenuates intercellular adhesion molecule-1 and monocyte chemotactic protein-1 expression in the kidney in diabetic rats. J Pharmacol Sci. 2006;101(4):335-343. https://doi.org/10.1254/ jphs.fp0050896
- Ohshiro Y, Ma RC, Yasuda Y, et al. Reduction of diabetes-induced oxidative stress, fibrotic cytokine expression, and renal dysfunction in protein kinase Cbeta-null mice. *Diabetes*. 2006;55(11):3112-3120. https://doi.org/10.2337/db06-0895
- Barot M, Gokulgandhi MR, Mitra AK. Mitochondrial dysfunction in retinal diseases. *Curr Eye Res.* 2011;36(12):1069-1077. https://doi. org/10.3109/02713683.2011.607536
- Kanwar M, Chan P-S, Kern TS, Kowluru RA. Oxidative damage in the retinal mitochondria of diabetic mice: possible protection by superoxide dismutase. *Invest Ophthalmol Vis Sci.* 2007;48(8):3805-3811. https://doi.org/10.1167/iovs.06-1280
- Wang C, Youle RJ. The role of mitochondria in apoptosis*. Annu Rev Genet. 2009;43:95-118. https://doi.org/10.1146/annurevgenet-102108-134850
- Rein DB, Wittenborn JS, Zhang X, et al. Forecasting age-related macular degeneration through the year 2050: the potential impact of new treatments. Arch Ophthalmol. 2009;127(4):533-540. https://doi.org/10.1001/archophthalmol.2009.58
- 57. Nowak JZ. Age-related macular degeneration (AMD): pathogenesis and therapy. *Pharmacol Rep.* 2006;58(3):353-363.
- Gehrs KM, Anderson DH, Johnson LV, Hageman GS. Agerelated macular degeneration-emerging pathogenetic and therapeutic concepts. Ann Med. 2006;38(7):450-471. https://doi. org/10.1080/07853890600946724
- Boulton M, Dayhaw-Barker P. The role of the retinal pigment epithelium: topographical variation and ageing changes. *Eye (Lond)*. 2001;15(Pt 3):384-389. https://doi.org/10.1038/eye.2001.141
- Brandl C, Breinlich V, Stark KJ, et al. Their dependency on age, sex, and smoking: results from the German KORA Study. *PLoS One.* 2016;11(11):e0167181. https://doi.org/10.1371/journ al.pone.0167181
- Ferris FL, Wilkinson CP, Bird A, et al. Clinical classification of age-related macular degeneration. *Ophthalmology*. 2013;120(4):844-851. https://doi.org/10.1016/j.ophtha.2012.10.036
- Fine SL, Berger JW, Maguire MG, Ho AC. Age-related macular degeneration. N Engl J Med. 2000;342(7):483-492. https://doi. org/10.1056/NEJM200002173420707
- Brandl C, Zimmermann ME, Günther F, et al. On the impact of different approaches to classify age-related macular degeneration: Results from the German AugUR study. *Sci Rep.* 2018;8(1):8675. https://doi.org/10.1038/s41598-018-26629-5
- Michalska-Małecka K, Kabiesz A, Nowak M, Śpiewak D. Age related macular degeneration – challenge for future: pathogenesis and new perspectives for the treatment. *Eur Geriatr Med.* 2015;6(1):69-75. https://doi.org/10.1016/j.eurger.2014.09.007
- Golden TR, Hinerfeld DA, Melov S. Oxidative stress and aging: beyond correlation. *Aging Cell*. 2002;1(2):117-123. https://doi. org/10.1046/j.1474-9728.2002.00015.x
- 66. Maeda A, Crabb JW, Palczewski K. Microsomal glutathione S-transferase 1 in the retinal pigment epithelium: protection against oxidative stress and a potential role in aging. *Biochemistry*. 2005;44(2):480-489. https://doi.org/10.1021/bi048016f

410

CNS Neuroscience & Therapeutics

- Beatty S, Koh H, Phil M, Henson D, Boulton M. The role of oxidative stress in the pathogenesis of age-related macular degeneration. Surv Ophthalmol. 2000;45(2):115-134. https://doi. org/10.1016/s0039-6257(00)00140-5
- Friedrichson T, Kalbach HL, Buck P, van Kuijk FJ. Vitamin E in macular and peripheral tissues of the human eye. *Curr Eye Res.* 1995;14(8):693-701. https://doi.org/10.3109/0271368950 8998497
- Castorina C, Campisi A, Di Giacomo C, Sorrenti V, Russo A, Vanella A. Lipid peroxidation and antioxidant enzymatic systems in rat retina as a function of age. *Neurochem Res.* 1992;17(6):599-604. https://doi.org/10.1007/BF00968789
- 70. Wolg G, Lipofuscin and macular degeneration. *Nutr Rev.* 2003;61(10):342-346. https://doi.org/10.1301/nr.2003. oct.342-346.
- Reeg S, Grune T. Protein oxidation in aging: does it play a role in aging progression? Antioxid Redox Signal. 2015;23(3):239-255. https://doi.org/10.1089/ars.2014.6062
- Jarrett SG, Lewin AS, Boulton ME. The importance of mitochondria in age-related and inherited eye disorders. *Ophthalmic Res.* 2010;44(3):179-190. https://doi.org/10.1159/000316480
- Glenn JV, Stitt AW. The role of advanced glycation end products in retinal ageing and disease. *Biochim Biophys Acta*. 2009;1790(10):1109-1116. https://doi.org/10.1016/j. bbagen.2009.04.016
- 74. Age-Related Eye Disease Study Research Group. A randomized, placebo-controlled, clinical trial of high-dose supplementation with vitamins C and E, beta carotene, and zinc for age-related macular degeneration and vision loss: AREDS report no. 8. Arch Ophthalmol. 2001;119(10):1417-1436. https://doi.org/10.1001/ archopht.119.10.1417
- Chew EY, Clemons TE, Agrón E, et al. Ten-year follow-up of age-related macular degeneration in the age-related eye disease study. JAMA Ophthalmol. 2014;132(3):272. https://doi.org/10.1001/ jamaophthalmol.2013.6636
- 76. van Asten F, Chiu C-Y, Agrón E, et al. No CFH or ARMS2 Interaction with Omega-3 fatty acids, low versus high zinc, or β-carotene versus lutein and zeaxanthin on progression of age-related macular degeneration in the age-related eye disease study 2. *Ophthalmology*. 2019;126(11):1541-1548. https://doi. org/10.1016/j.ophtha.2019.06.004
- 77. Glaser TS, Doss LE, Shih G, et al. The association of dietary lutein plus zeaxanthin and B vitamins with cataracts in the age-related eye disease study. Ophthalmology. 2015;122(7):1471-1479. https:// doi.org/10.1016/j.ophtha.2015.04.007
- Yang S, Zhao J, Sun X. Resistance to anti-VEGF therapy in neovascular age-related macular degeneration: a comprehensive review. *Drug Des Devel Ther*. 2016;10:1857-1867. https://doi.org/10.2147/ DDDT.S97653
- Boyle J, Vukicevic M, Koklanis K, Itsiopoulos C. Experiences of patients undergoing anti-VEGF treatment for neovascular age-related macular degeneration: a systematic review. *Psychol Health Med.* 2015;20(3):296-310. https://doi.org/10.1080/13548 506.2014.936886
- Nikkhah H, Karimi S, Ahmadieh H, et al. Intravitreal injection of anti-vascular endothelial growth factor agents for ocular vascular diseases: clinical practice guideline. J Ophthalmic Vis Res. 2018;13(2):158. https://doi.org/10.4103/jovr.jovr_50_18
- Pilli S, Kotsolis A, Spaide RF, et al. Endophthalmitis associated with intravitreal anti-vascular endothelial growth factor therapy injections in an office setting. *Am J Ophthalmol.* 2008;145(5):879-882. https://doi.org/10.1016/j.ajo.2007.12.036
- Scott IU, Flynn HW. Reducing the risk of endophthalmitis following intravitreal injections. *Retina*. 2007;27(1):10-12. https://doi. org/10.1097/IAE.0b013e3180307271

- Evans JR, Lawrenson JG. Antioxidant vitamin and mineral supplements for preventing age-related macular degeneration. *Cochrane Database Syst Rev.* 2017;7:CD000253. https://doi. org/10.1002/14651858.CD000253.pub4
- Weinreb RN, Aung T, Medeiros FA. The pathophysiology and treatment of glaucoma: a review. JAMA. 2014;311(18):1901-1911. https://doi.org/10.1001/jama.2014.3192
- Mélik Parsadaniantz S, Réaux-le Goazigo A, Sapienza A, Habas C, Baudouin C. Glaucoma: a degenerative optic neuropathy related to neuroinflammation? *Cells.* 2020;9(3):535. https://doi. org/10.3390/cells9030535
- Wu Y, Zhan Z, Quan Y, et al. SP1-mediated upregulation of LINGO-1 promotes degeneration of retinal ganglion cells in optic nerve injury. CNS Neurosci Ther. 2020;26(10):1010-1020. https:// doi.org/10.1111/cns.13426
- Costagliola C, dell'Omo R, Romano MR, Rinaldi M, Zeppa L, Parmeggiani F. Pharmacotherapy of intraocular pressure: part I. Parasympathomimetic, sympathomimetic and sympatholytics. *Expert Opin Pharmacother*. 2009;10(16):2663-2677. https://doi. org/10.1517/14656560903300103
- Cherecheanu AP, Garhofer G, Schmidl D, Werkmeister R, Schmetterer L. Ocular perfusion pressure and ocular blood flow in glaucoma. *Curr Opin Pharmacol.* 2013;13(1):36-42. https://doi. org/10.1016/j.coph.2012.09.003
- Mahabadi N, Foris LA, Tripathy K. Open angle glaucoma. In: StatPearls. StatPearls Publishing; 2020. http://www.ncbi.nlm.nih. gov/books/NBK441887/. Accessed October 19, 2020.
- Soto I, Howell GR. The complex role of neuroinflammation in glaucoma. Cold Spring Harb Perspect Med. 2014;4(8):a017269. https:// doi.org/10.1101/cshperspect.a017269.
- Williams PA, Marsh-Armstrong N, Howell GR. Lasker/irrf initiative on astrocytes and glaucomatous neurodegeneration participants. neuroinflammation in glaucoma: a new opportunity. *Exp Eye Res.* 2017;157:20-27. https://doi.org/10.1016/j.exer.2017.02.014
- McMonnies C. Reactive oxygen species, oxidative stress, glaucoma and hyperbaric oxygen therapy. J Optom. 2018;11(1):3-9. https://doi.org/10.1016/j.optom.2017.06.002
- Uchida K, Himori N, Hashimoto K, et al. The association between oxidative stress and corneal hysteresis in patients with glaucoma. *Sci Rep.* 2020;10(1):545. https://doi.org/10.1038/s41598-020-57520-x
- Alvarado J, Murphy C, Polansky J, Juster R. Age-related changes in trabecular meshwork cellularity. *Invest Ophthalmol Vis Sci.* 1981;21(5):714-727.
- Alvarado J, Murphy C, Juster R. Trabecular meshwork cellularity in primary open-angle glaucoma and nonglaucomatous normals. *Ophthalmology.* 1984;91(6):564-579. https://doi.org/10.1016/ s0161-6420(84)34248-8
- 96. Zhou L, Li Y, Yue BY. Oxidative stress affects cytoskeletal structure and cell-matrix interactions in cells from an ocular tissue: the trabecular meshwork. J Cell Physiol. 1999;180(2):182-189. https:// doi.org/10.1002/(SICI)1097-4652(199908)180:2<182:AID-JCP6>3.0.CO;2-X
- Kahn MG, Giblin FJ, Epstein DL. Glutathione in calf trabecular meshwork and its relation to aqueous humor outflow facility. *Invest Ophthalmol Vis Sci.* 1983;24(9):1283-1287.
- Ko ML, Hu DN, Ritch R, Sharma SC. The combined effect of brain-derived neurotrophic factor and a free radical scavenger in experimental glaucoma. *Invest Ophthalmol Vis Sci.* 2000;41(10):2967-2971.
- Izzotti A, Saccà SC, Cartiglia C, De Flora S. Oxidative deoxyribonucleic acid damage in the eyes of glaucoma patients. Am J Med. 2003;114(8):638-646. https://doi.org/10.1016/s0002 -9343(03)00114-1

WILEY-CNS Neuroscience & Therapeutics

- 100. Saccà SC, Pascotto A, Camicione P, Capris P, Izzotti A. Oxidative DNA damage in the human trabecular meshwork: clinical correlation in patients with primary open-angle glaucoma. Arch Ophthalmol. 2005;123(4):458-463. https://doi.org/10.1001/archo pht.123.4.458
- Gherghel D, Griffiths HR, Hilton EJ, Cunliffe IA, Hosking SL. Systemic reduction in glutathione levels occurs in patients with primary open-angle glaucoma. *Invest Ophthalmol Vis Sci.* 2005;46(3):877. https://doi.org/10.1167/iovs.04-0777
- 102. Gherghel D, Mroczkowska S, Qin L. Reduction in blood glutathione levels occurs similarly in patients with primary-open angle or normal tension glaucoma. *Invest Ophthalmol Vis Sci.* 2013;54(5):3333. https://doi.org/10.1167/iovs.12-11256
- Pasantes-Morales H, Klethi J, Ledig M, Mandel P. Free amino acids of chicken and rat retina. *Brain Res.* 1972;41(2):494-497. https:// doi.org/10.1016/0006-8993(72)90523-9
- 104. Rascher K, Servos G, Berthold G, et al. Light deprivation slows but does not prevent the loss of photoreceptors in taurine transporter knockout mice. Vision Res. 2004;44(17):2091-2100. https://doi. org/10.1016/j.visres.2004.03.027
- 105. Schaffer S, Takahashi K, Azuma J. Role of osmoregulation in the actions of taurine. Amino Acids. 2000;19(3-4):527-546. https:// doi.org/10.1007/s007260070004
- 106. Froger N, Sahel J-A, Picaud S. Taurine deficiency and the eye. In: Handbook of Nutrition, Diet and the Eye. Elsevier; 2014:505-513. https://doi.org/10.1016/B978-0-12-401717-7.00051-4
- 107. García-Ayuso D, Di Pierdomenico J, Hadj-Said W, et al. Taurine Depletion causes ipRGC loss and increases light-induced photoreceptor degeneration. *Invest Ophthalmol Vis Sci.* 2018;59(3):1396. https://doi.org/10.1167/iovs.17-23258
- Kim C, Cha Y-N. Taurine chloramine produced from taurine under inflammation provides anti-inflammatory and cytoprotective effects. Amino Acids. 2014;46(1):89-100. https://doi.org/10.1007/ s00726-013-1545-6
- 109. Marcinkiewicz J, Grabowska A, Bereta J, Stelmaszynska T. Taurine chloramine, a product of activated neutrophils, inhibits in vitro the generation of nitric oxide and other macrophage inflammatory mediators. J Leukoc Biol. 1995;58(6):667-674. https://doi. org/10.1002/jlb.58.6.667
- Jong CJ, Azuma J, Schaffer S. Mechanism underlying the antioxidant activity of taurine: prevention of mitochondrial oxidant production. *Amino Acids*. 2012;42(6):2223-2232. https://doi. org/10.1007/s00726-011-0962-7
- Froger N, Cadetti L, Lorach H, et al. Taurine provides neuroprotection against retinal ganglion cell degeneration. *PLoS One*. 2012;7(10):e42017. https://doi.org/10.1371/journal.pone.0042017
- 112. Garcia-Delgado AB, Valdés-Sánchez L, Calado SM, Diaz-Corrales FJ, Bhattacharya SS. Rasagiline delays retinal degeneration in a

mouse model of retinitis pigmentosa via modulation of Bax/Bcl-2 expression. *CNS Neurosci Ther.* 2018;24(5):448-455. https://doi. org/10.1111/cns.12805

- 113. Tao Y, He M, Yang Q, et al. Systemic taurine treatment provides neuroprotection against retinal photoreceptor degeneration and visual function impairments. *Drug Des Devel Ther.* 2019;13:2689-2702. https://doi.org/10.2147/DDDT.S194169
- 114. Nor Arfuzir N, Agarwal R, lezhitsa I, Agarwal P, Sidek S, Ismail N. Taurine protects against retinal and optic nerve damage induced by endothelin-1 in rats via antioxidant effects. *Neural Regen Res.* 2018;13(11):2014. https://doi.org/10.4103/1673-5374.239450
- 115. Zeng K, Xu H, Chen K, et al. Effects of taurine on glutamate uptake and degradation in Müller cells under diabetic conditions via antioxidant mechanism. *Mol Cell Neurosci*. 2010;45(2):192-199. https:// doi.org/10.1016/j.mcn.2010.06.010
- 116. Fan Y, Lai J, Yuan Y, Wang L, Wang Q, Yuan F. Taurine protects retinal cells and improves synaptic connections in early diabetic rats. *Curr Eye Res.* 2020;45(1):52-63. https://doi.org/10.1080/02713 683.2019.1653927
- 117. Di Leo MAS, Ghirlanda G, Silveri NG, Giardina B, Franconi F, Santini SA. Potential therapeutic effect of antioxidants in experimental diabetic retina: a comparison between chronic taurine and vitamin e plus selenium supplementations. *Free Radical Res.* 2003;37(3):323-330. https://doi.org/10.1080/1071576021000055271
- 118. Ansar M, Ranza E, Shetty M, et al. Taurine treatment of retinal degeneration and cardiomyopathy in a consanguineous family with SLC6A6 taurine transporter deficiency. *Hum Mol Genet*. 2020;29(4):618-623. https://doi.org/10.1093/hmg/ddz303
- 119. Kowluru RA, Odenbach S. Effect of long-term administration of alpha-lipoic acid on retinal capillary cell death and the development of retinopathy in diabetic rats. *Diabetes*. 2004;53(12):3233-3238. https://doi.org/10.2337/diabetes.53.12.3233
- 120. Kowluru RA, Kern TS, Engerman RL. Abnormalities of retinal metabolism in diabetes or experimental galactosemia. IV. Antioxidant defense system. *Free Radic Biol Med.* 1997;22(4):587-592. https:// doi.org/10.1016/s0891-5849(96)00347-4
- 121. Harris A, Gross J, Moore N, et al. The effects of antioxidants on ocular blood flow in patients with glaucoma. Acta Ophthalmol. 2018;96(2):e237-e241. https://doi.org/10.1111/aos.13530

How to cite this article: Castelli V, Paladini A, d'Angelo M, et al. Taurine and oxidative stress in retinal health and disease. CNS Neurosci Ther. 2021;27:403–412. <u>https://doi.org/10.1111/</u> cns.13610