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## EXTENDED REPORT

# Therapeutic interleukin-6 blockade reverses transforming growth factor-beta pathway activation in dermal fibroblasts: insights from the faSScinate clinical trial in systemic sclerosis

Christopher P Denton,<sup>1</sup> Voon H Ong,<sup>1</sup> Shiwen Xu,<sup>1</sup> Haiyin Chen-Harris,<sup>2</sup> Zora Modrusan,<sup>2</sup> Robert Lafyatis,<sup>3</sup> Dinesh Khanna,<sup>4</sup> Angelika Jahreis,<sup>2</sup> Jeffrey Siegel,<sup>2</sup> Thierry Sornasse<sup>2</sup>

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<sup>1</sup>Division of Medicine, Royal Free Campus, University College London, London, UK

<sup>2</sup>Genentech, South San Francisco, California, USA

<sup>3</sup>University of Pittsburgh Medical Center, Pittsburgh, Pennsylvania, USA

<sup>4</sup>University of Michigan Ankle Scleroderma Program, Ann Arbor, Michigan, USA

**Correspondence to**

Professor Christopher P Denton, University College London Division of Medicine, Royal Free Campus, London, NW3 2PF, UK; [c.denton@ucl.ac.uk](mailto:c.denton@ucl.ac.uk)

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**ABSTRACT**

**Objectives** Skin fibrosis mediated by activated dermal fibroblasts is a hallmark of systemic sclerosis (SSc), especially in the subset of patients with diffuse disease. Transforming growth factor-beta (TGFβ) and interleukin-6 (IL-6) are key candidate mediators in SSc. Our aim was to elucidate the specific effect of IL-6 pathway blockade on the biology of SSc fibroblasts in vivo by using samples from a unique clinical experiment—the faSScinate study—in which patients with SSc were treated for 24 weeks with tocilizumab (TCZ), an IL-6 receptor-α inhibitor.

**Methods** We analysed the molecular, functional and genomic characteristics of explant fibroblasts cultured from matched skin biopsy samples collected at baseline and at week 24 from 12 patients receiving placebo (n=6) or TCZ (n=6) and compared these with matched healthy control fibroblast strains.

**Results** The hallmark functional and molecular-activated phenotype was defined in SSc samples and was stable over 24 weeks in placebo-treated cases. RNA sequencing analysis robustly defined key dysregulated pathways likely to drive SSc fibroblast activation in vivo. Treatment with TCZ for 24 weeks profoundly altered the biological characteristics of explant dermal fibroblasts by normalising functional properties and reversing gene expression profiles dominated by TGFβ-regulated genes and molecular pathways.

**Conclusions** We demonstrated the exceptional value of using explant dermal fibroblast cultures from a well-designed trial in SSc to provide a molecular framework linking IL-6 to key profibrotic pathways. The profound impact of IL-6R blockade on the activated fibroblast phenotype highlights the potential of IL-6 as a therapeutic target in SSc and other fibrotic diseases.

**Trial registration number** NCT01532869; Post-results.

**INTRODUCTION**

Systemic sclerosis (SSc) is characterised by fibrosis of the skin and internal organs, autoimmunity and vasculopathy.<sup>1</sup> Although the biological mechanisms underlying SSc are not fully understood, interplay between vasculopathy, autoimmunity and fibrosis may perpetuate a state of aberrant wound repair.<sup>2</sup> Disease modification remains elusive, and treatment

focuses on complications such as interstitial lung disease, pulmonary artery hypertension and peripheral vasculopathy.<sup>3</sup> Clinical heterogeneity is a hallmark of SSc, and patients with severe progressive diffuse skin involvement have poor outcomes; more effective treatment for this subset of patients is an important unmet medical need.<sup>4</sup>

Fibroblasts play a central role in the biology of the disease<sup>5</sup> by interacting with endothelial cells<sup>6</sup> and leucocytes<sup>7</sup> in a complex biological network involving cytokines and adhesion molecules,<sup>8</sup> resulting in excess deposition of extracellular matrix proteins and in tissue stiffening. Transforming growth factor-beta (TGFβ) plays a central role in fibrosis<sup>9</sup> by promoting the differentiation of fibroblasts into myofibroblasts and by inducing profibrotic molecules and proliferation.<sup>10</sup> Interleukin-6 (IL-6) also plays an important role in SSc<sup>11</sup> by regulating the function of immune and non-immune cells. Dermal fibroblasts from patients with SSc express increased levels of IL-6<sup>12</sup>; elevated levels of IL-6 are associated with early disease,<sup>13</sup> and increased serum IL-6 level predicts higher mortality risk, worse skin involvement and increased pulmonary decline.<sup>14 15</sup> IL-6 is known to induce TGFβ production by cardiac fibroblasts<sup>16</sup> and to enhance TGFβ-signalling in dermal fibroblasts<sup>17</sup> and cardiac fibroblasts.<sup>18</sup> Conversely, TGFβ regulates the expression of IL-6 by lung fibroblasts<sup>19</sup> and airway smooth muscle cells.<sup>20</sup>

To elucidate the specific effect of IL-6 on the biology of fibroblasts in vivo, we took advantage of a unique clinical experiment—the faSScinate study—in which SSc patients were treated for 24 weeks with tocilizumab (TCZ), an IL-6 receptor-α blocking antibody. The faSScinate study was a randomised, double-blind, placebo (PBO)-controlled phase 2 study of TCZ in adult SSc patients with <5 years' disease duration, a modified Rodnan skin score (mRSS) between 15 and 40 units and active progressive disease according to specified clinical or laboratory features. No concomitant immunosuppressive medication was permitted at study entry or during the first 24 weeks of the trial.<sup>21</sup> In this study, 43 patients with SSc received weekly TCZ 162 mg subcutaneously and 44 patients with SSc received weekly PBO subcutaneously. As previously reported, primary clinical data



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from the faSScinate trial showed a trend of benefit in favour of TCZ for the primary endpoint or mRSS and a strong trend at 48 weeks together with congruent benefit in exploratory endpoints including lung function. Based on these results, a phase 3 trial of TCZ is under way (ClinicalTrials.gov, NCT02453256).

Here we define the molecular and functional phenotype of explant dermal fibroblasts from a representative subset of patients with SSc enrolled into faSScinate and demonstrate a profound reversal of the hallmark profibrotic properties of these cells after treatment with TCZ for 24 weeks compared with controls from the PBO arm of the trial.

## METHODS

### Sample collection and analysis

Culture of dermal explant fibroblasts from patients with SSc enrolled in the faSScinate study (ClinicalTrials.gov, NCT01532869) and matched controls was performed as described, as was protein quantification using Western blot analysis.<sup>15</sup> Cell migration was assessed using scratch wound assay,<sup>22</sup> and contractility assays were performed on 3D collagen gel lattices.<sup>23</sup> Gene expression patterns were analysed using Ingenuity Pathway Analysis (IPA; Qiagen). Additional methodology is described in online supplementary appendix S1.

### Statistical analysis

For functional assays, means were compared using Tukey's multiple comparison test. Correlations were calculated using the Spearman  $\rho$ . The Benjamini and Hochberg False Discovery Rate method was used to correct for multiple comparisons. All statistical calculations were conducted in JMP V.11.1 (SAS Institute).

## RESULTS

### SSc explant dermal fibroblasts present hallmark functional properties of activated fibrotic cells

Dermal fibroblasts were cultured from fresh skin biopsy samples collected at baseline and at week 24 from consecutive PBO-treated and TCZ-treated patients enrolled in the faSScinate study (online supplementary table S1).<sup>21</sup> Confirming earlier reports,<sup>22</sup> dermal fibroblasts isolated from patients with SSc at baseline produced significantly higher levels of connective tissue growth factor (CTGF), collagen alpha 1 (Col1) and alpha-smooth muscle actin ( $\alpha$ SMA) than normal fibroblasts (NFs) (NF vs SSc;  $p < 0.0001$  for all three factors) (figure 1A–C). Migratory capacity and contractility activity were also enhanced compared with NF (NF vs PBO baseline and NF vs TCZ baseline,  $p < 0.0001$  for both) (figure 1D,E). These results indicate that the SSc explant dermal fibroblasts obtained in this study have the hallmark functional characteristics of activated fibrotic fibroblasts.

### SSc dermal fibroblasts collected at baseline: typical profibrotic signature dominated by genes regulated by TGF $\beta$

Using RNA sequencing (RNA-Seq), we compared the expression profiles of SSc fibroblasts collected at baseline (SSc) with those of NF. Figure 2 depicts the expression values of NF and SSc baseline fibroblasts for 578 protein-coding genes significantly differentially expressed (criteria: fold difference  $\geq 1.5$ ; median reads per kilobase of transcript per million mapped reads  $\geq 1$ , adjusted  $p \leq 0.05$ ). Of the 578 selected genes, 145 are known to be regulated by TGF $\beta$  in human dermal or lung fibroblasts,<sup>20 24–26</sup> indicating that this pathway is dominant in these cells (figure 2). Of note, baseline erythrocyte sedimentation rate (ESR) and C reactive protein and IL-6 levels were not associated with distinct

baseline gene expression profiles (online supplementary table S2).

### TCZ treatment for 24 weeks attenuates the functional properties of SSc explant fibroblasts

Comparison of functional measures between fibroblasts collected at baseline and week 24 from patients treated with TCZ showed a significant decrease in protein production, in migration (increase in gap size) and in contractility (increase in gel weight) (figure 3), indicating that therapeutic treatment with TCZ results in a significant attenuation of the hallmark functional properties of SSc fibroblasts. In contrast, the phenotype of SSc fibroblasts was stable in PBO-treated patients.

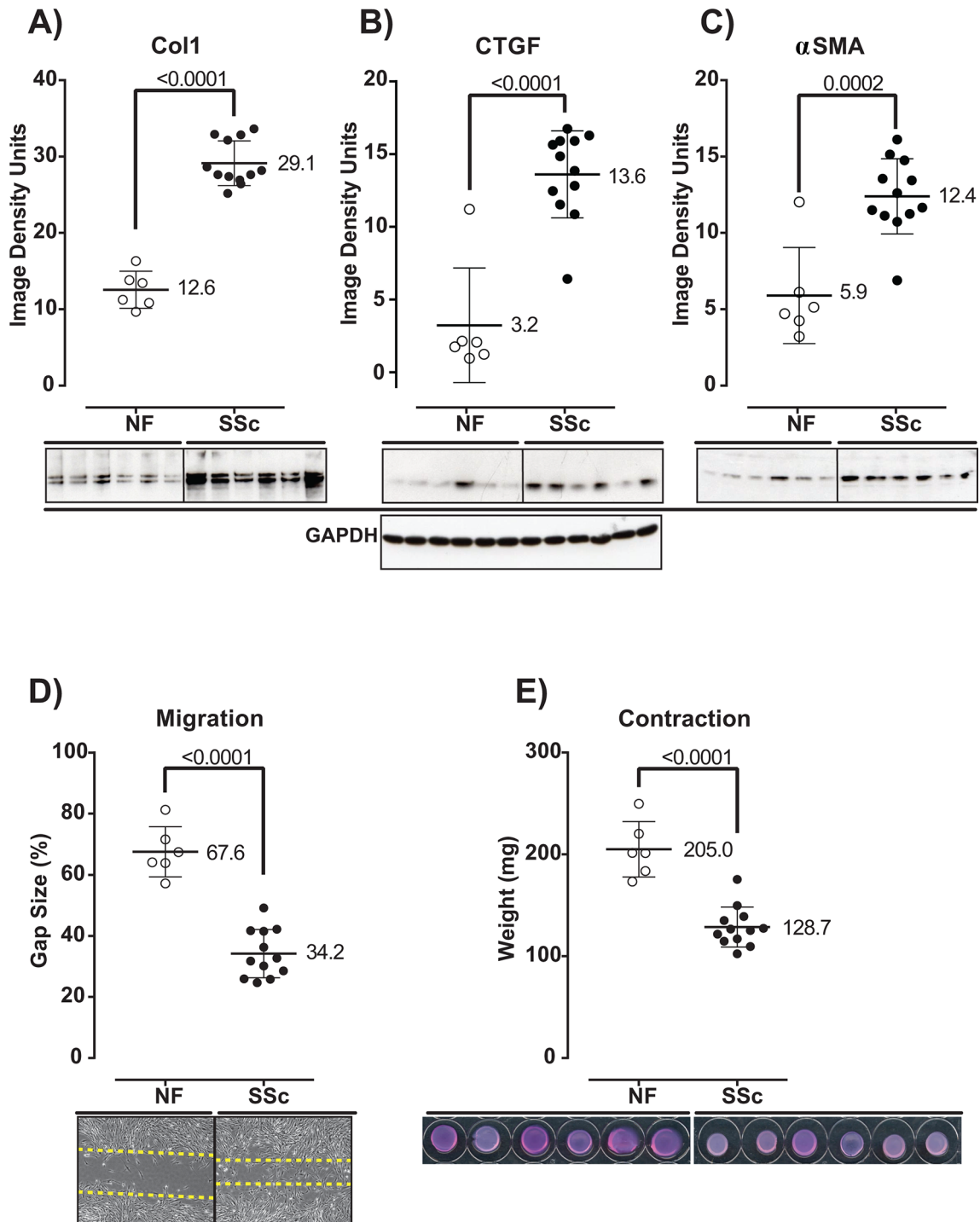
### Profibrotic expression profile of SSc dermal fibroblasts: normalised in TCZ-treated patients and stable in PBO-treated patients

Figure 4 depicts expression levels for the PBO and TCZ groups at week 24 compared with the matching sample at baseline. In the PBO-treated group (figure 4), no significant difference was detected (see also online supplementary figure S1), indicating that baseline expression profile is sustained over  $\geq 24$  weeks in the absence of immunosuppressants. In contrast, in the TCZ group (figure 4), we identified 2136 significantly differentially expressed genes; of those, 411 were also differentially expressed in the SSc/NF comparison. Remarkably, the expression pattern of the 411 shared genes in the TCZ group was opposite to that observed in the SSc versus NF comparison (figure 4). Of note, 167 and 1725 genes were significantly differentially expressed in SSc/NF and TCZ week 24/baseline comparisons, respectively. For most genes, differential expression trends were similar to those observed in the 411 shared genes (online supplementary figure S2). Thus, gene expression data confirm that therapeutic treatment with TCZ for 24 weeks profoundly alters the SSc-associated molecular phenotype of explanted dermal fibroblasts. Considering this striking biological effect of TCZ, we explored the correlation between gene expression and mRSS across all patients and time points. Of the 411 genes analysed, 35 had a nominal correlation  $p \leq 0.05$  (online supplementary table S3), and only one, PRKCE, remained significant after correction for multiple testing. Treatment with TCZ resulted in a significant upregulation of PRKCE expression that was underexpressed in SSc dermal fibroblasts at baseline, as previously reported in SSc lung fibroblasts.<sup>27</sup>

### Treatment with TCZ for 24 weeks: reversal of activation status of key pathological pathways associated with SSc

We used the IPA (Qiagen) to assign biological functions to networks of all significantly differentially expressed genes (2303 genes) from the SSc versus NF and the TCZ week 24 versus baseline comparisons (online supplementary figure S1). Of the 609 significantly enriched pathways, we selected the top 50 pathways with the highest combined absolute Z scores (figure 5) and grouped them into 10 major biological nodes; annotations for the selected pathways are provided in (online supplementary table S4). Most pathways defined by genes that were only differentially expressed in one comparison overlapped with the 50 pathways.

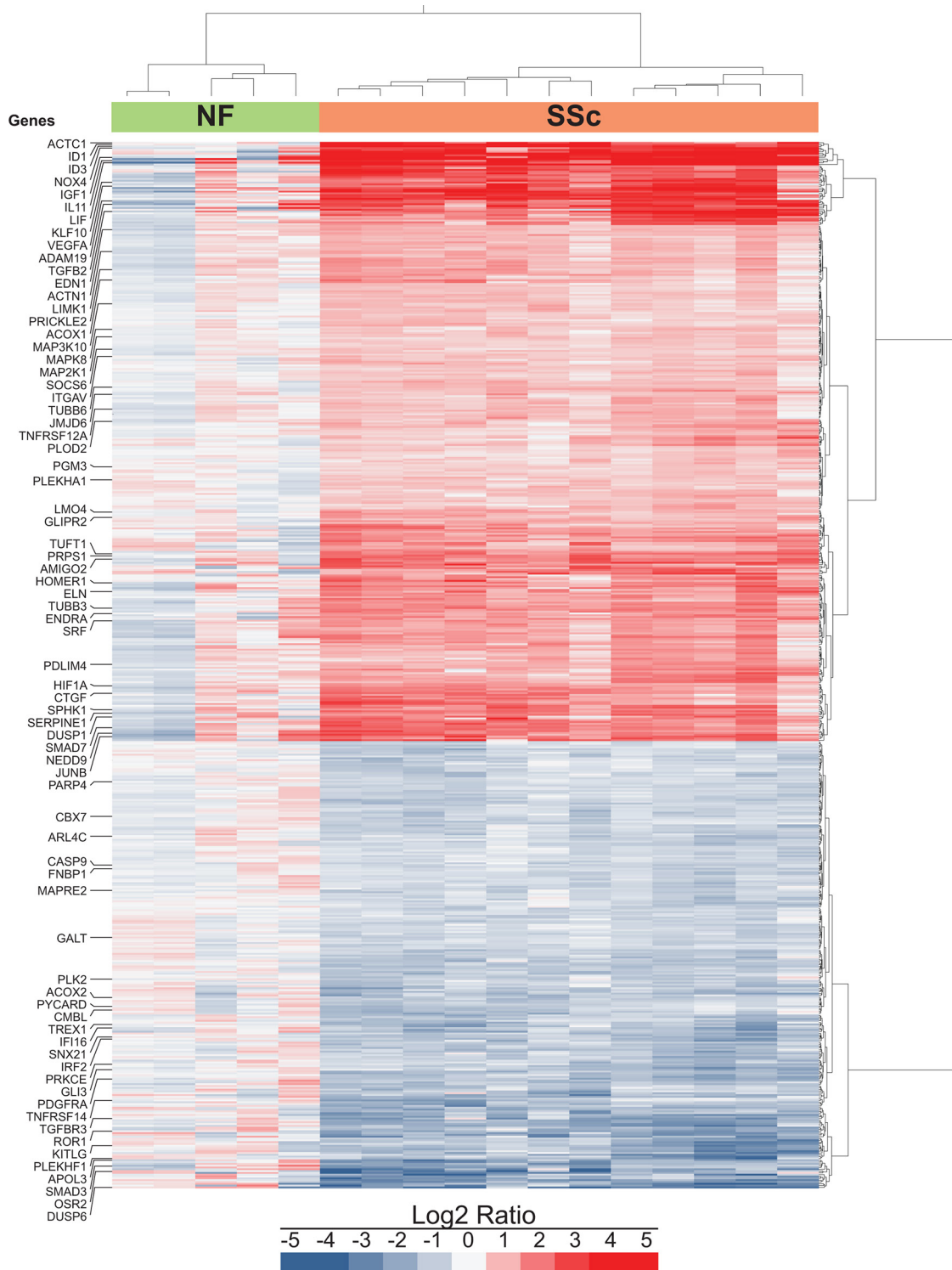
The fibrosis node is dominated by TGF $\beta$ -related pathways but also includes a pathway related to the activity of lysophosphatidic acid and to the antifibrotic transcription factor KLF2. The contraction node regroups 11 pathways related to cellular contractility machinery. The migration node includes four pathways positively



**Figure 1** Explant dermal fibroblasts present hallmark functional properties of activated fibrotic fibroblasts. Production of Col1 (A), CTGF (B) and  $\alpha$ SMA (C); migration activity (D); and contractility (E) by normal and SSc dermal fibroblasts collected at baseline were analysed by western blot densitometry of cell lysates (A–C), scratch assay: percentage of remaining gap size (D) and 3D gel contraction assay: weight of the lattice plug (E), respectively. Representative western blot images are presented for each protein analysed (A–C, lower panel); western blot of the reference protein glyceraldehyde 3-phosphate dehydrogenase (GAPDH) is shown for the corresponding samples. Representative images of scratch assay performed over an incubation period of 48 hours are shown (D, lower panel). Representative images of gel contraction assay performed over an incubation period of 24 hours are shown (E, lower panel). BL, baseline; Col1, collagen alpha 1; CTGF, connective tissue growth factor; NFs, normal fibroblasts;  $\alpha$ SMA, alpha-smooth muscle actin; SSc, systemic sclerosis.

related to cell movement. The growth node contains three pathways positively related to proliferation and cell survival. The five pathways grouped under the cardiovascular node are generally associated with endothelial and vascular smooth muscle cell biology but are also known to be positively related to fibrosis.

Although the main function of the two pathways grouped under the coagulation node is blood clotting, these pathways play an important role in linking vascular events to fibrotic events in SSc. The activation node regroups eight pathways representing major signalling molecules and transcription factors positively

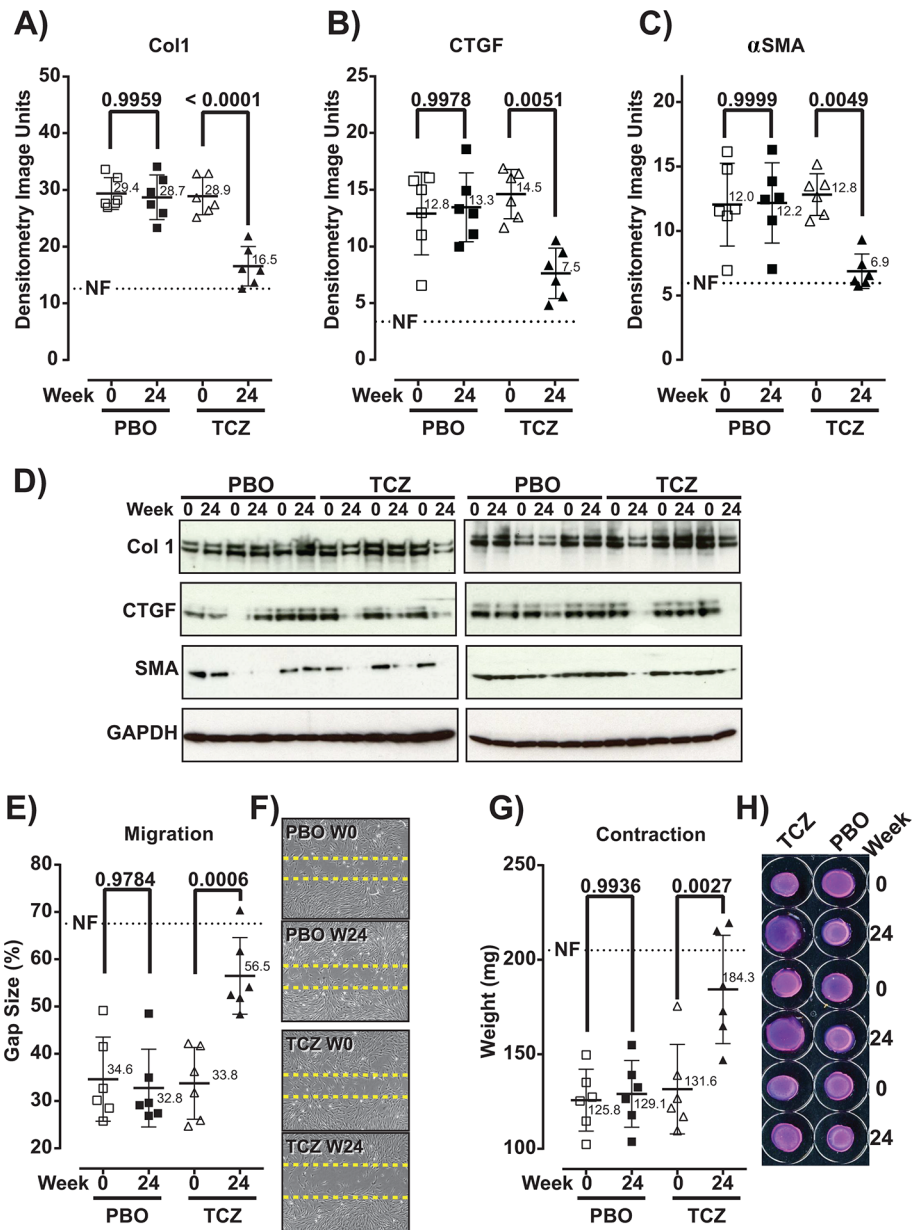


**Figure 2** Genes differentially expressed by SSc dermal fibroblasts collected at baseline compared with normal dermal fibroblasts represent a typical profibrotic signature dominated by genes regulated by TGF $\beta$ . Gene expression of normal, SSc PBO and SSc TCZ dermal fibroblasts collected at baseline and normalised to the average NF expression. Protein coding RNA showing a median RPKM  $\geq 1$ , 1.5-fold overexpressed or underexpressed compared with NF, with an adjusted  $p \leq 0.05$ , was selected as significantly differentially expressed ( $n=578$ ). Genes and samples were clustered using the Ward method. BL, baseline; NFs, normal fibroblasts; PBO, placebo; RPKM, reads per kilobase of transcript per million mapped reads; SSc, systemic sclerosis; TCZ, tocilizumab; TGF $\beta$ , transforming growth factor beta.

associated with cellular activation. The two pathways grouped in the hormones node are thought to contribute to the pathobiology of SSc, but the exact mechanisms by which this occurs remain ill defined. The inflammation node includes two pathways related to the innate immune system. Three of the four pathways included

in the morphogenesis node belong to the bone morphogenetic protein family, and the fourth pathway in this node is related to neuregulin biology. Overall, for the baseline SSc explant dermal fibroblasts (SSc vs NF comparison), the functions and activation status of the top 50 selected pathways depict a highly activated





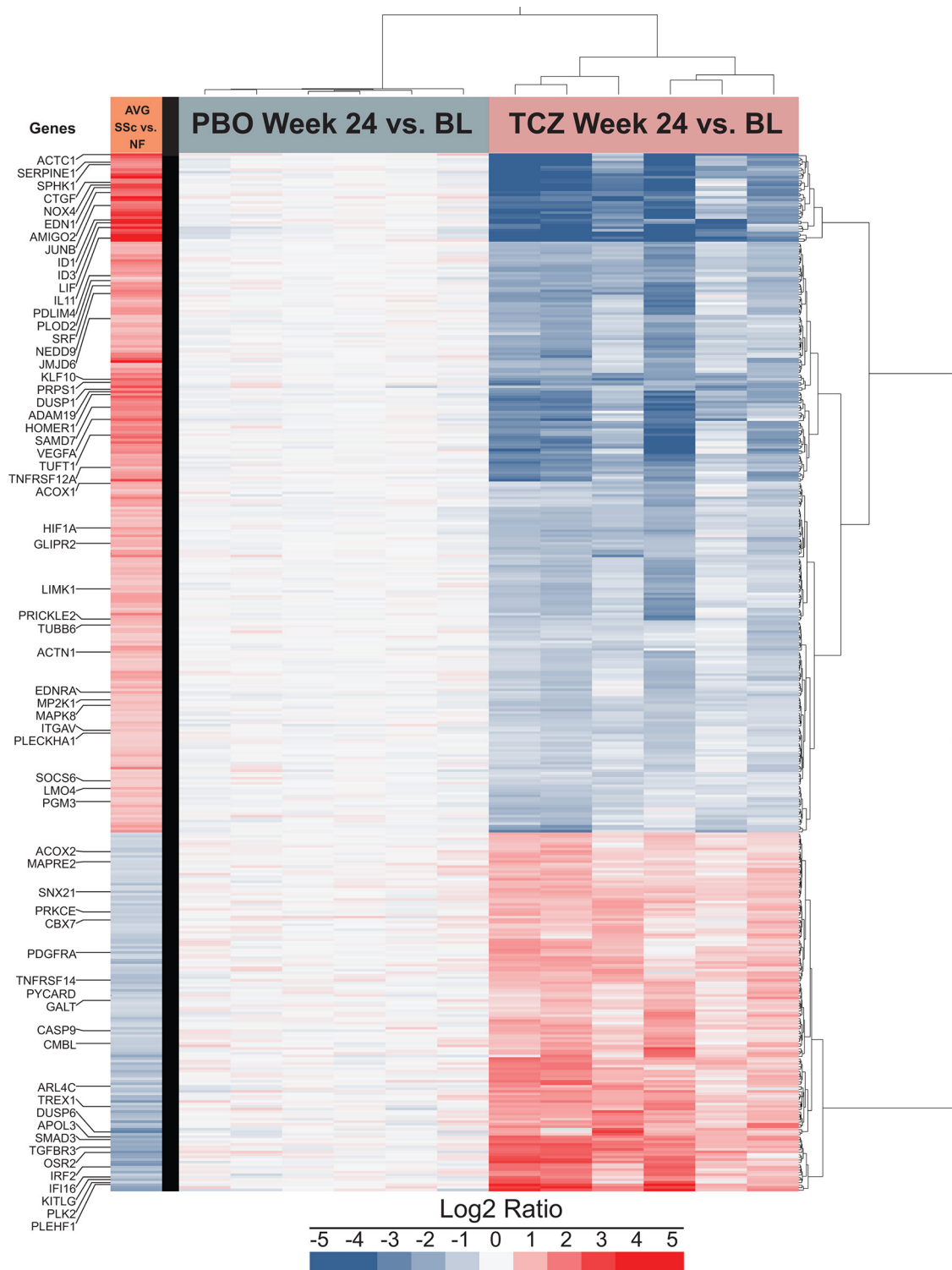
**Figure 3** Hallmark functional properties of SSc explant dermal fibroblasts are attenuated after treatment of patients with TCZ for 24 weeks. Comparison of functional properties of explant dermal fibroblasts from PBO-treated and TCZ-treated patients collected at baseline (week 0) and at week 24. Production of Col1 (A), CTGF (B) and αSMA (C); migration activity (E); and contractility (G) were analysed by western blot densitometry of cell lysates (A–C), scratch assay: percentage of remaining gap size (E) and 3D gel contraction assay: weight of the lattice plug (G), respectively. Representative western blot images are presented for each protein analysed (D, lower panel). Western blot of the reference protein GAPDH is shown for the corresponding samples. Representative images of scratch assay performed over an incubation period of 48 hours are shown (F). Representative images of gel contraction assay performed over an incubation period of 24 hours are shown (H). For each graph, the average measure of NFs is depicted as a dotted line. BL, baseline; Col1, collagen alpha 1; CTGF, connective tissue growth factor; NFs, normal fibroblasts; PBO, placebo; αSMA, alpha-smooth muscle actin; SSc, systemic sclerosis; TCZ, tocilizumab.

fibrotic, motile and contractile phenotype dominated by TGFβ superfamily member-regulated biology, where positive regulators (n=47) are activated and negative regulators (n=3) are predicted to be inhibited. In contrast, for the dermal fibroblasts collected from patients treated with TCZ for 24 weeks (TCZ week 24 vs baseline comparison), the same 50 pathways are predicted to have the opposite activation status. Notably, the level of inhibition observed with TCZ generally matches the levels of activation observed at baseline. Thus, in keeping with the functional assay and the gene expression data, the pathway analysis results indicate that the 24-week treatment of patients with TCZ almost completely

reverses the SSc-associated fibrotic phenotype of explant dermal fibroblasts.

### DISCUSSION

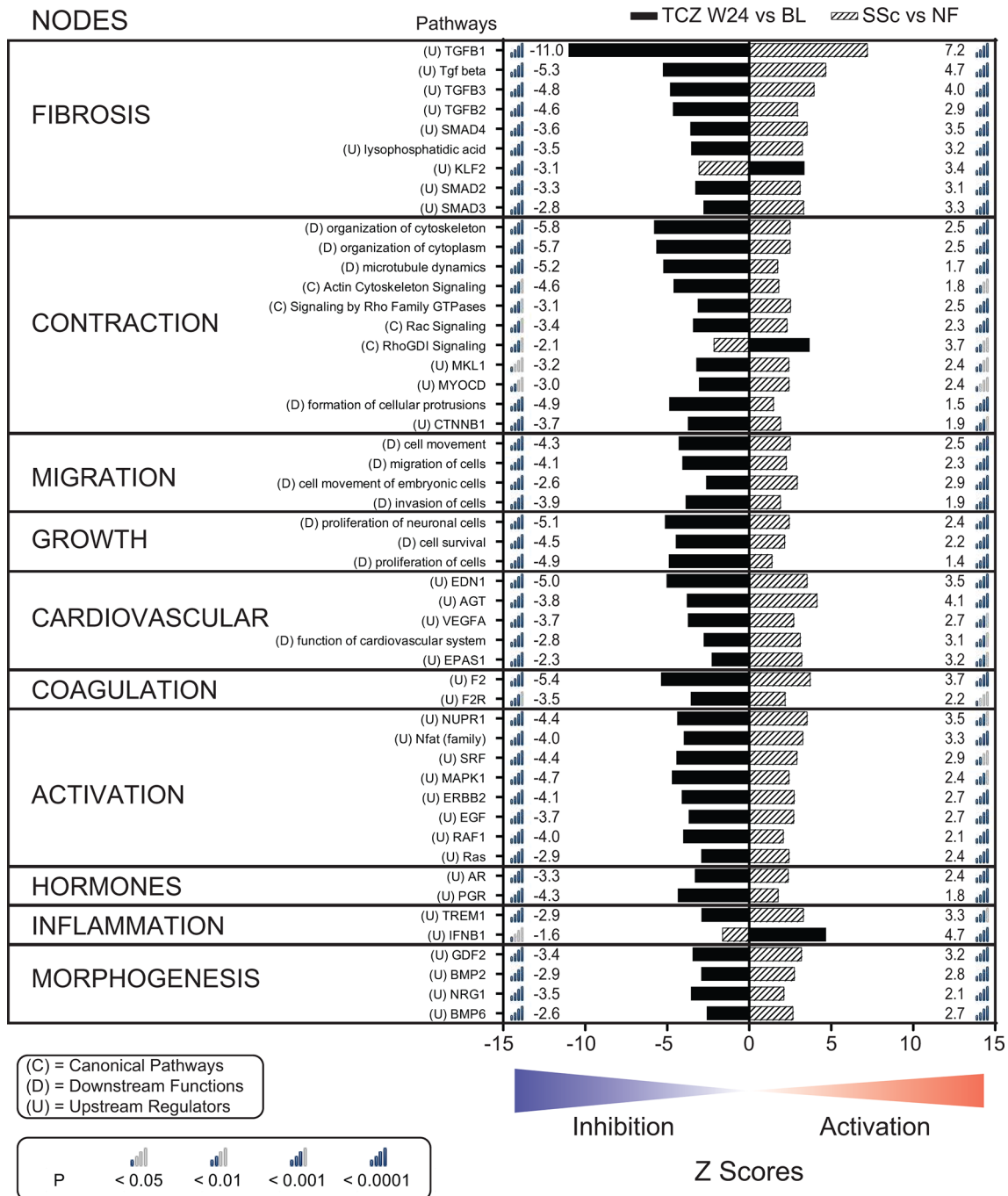
This study provides exceptional insight into the molecular pathology of dermal fibroblasts in severe SSc and unique insight into the likely key pathways and mediators involved in SSc pathogenesis, building on the original observations of LeRoy.<sup>28</sup> By linking the ex vivo data to promising clinical responses in a PBO-controlled trial of TCZ, our results also elucidate new and



**Figure 4** The profibrotic expression profile of SSc dermal fibroblasts tends to be normalised by 24-week treatment of patients with TCZ and tends to be stable in PBO-treated patients. Gene expression levels of SSc PBO and SSc TCZ dermal fibroblasts were measured at week 24 and normalised to matching baseline expression levels. Protein coding RNA showing a median RPKM  $\geq 1$ , 1.5-fold overexpressed or underexpressed at week 24 compared with baseline with an adjusted  $p \leq 0.05$  was marked as significantly differentially expressed ( $n=2136$ ). For clarity, only the subset of 411 genes differentially expressed in both SSc versus NF and in TCZ week 24 versus baseline is shown. Genes and samples were clustered using the Ward method. The first column represents the average differential expression of SSc baseline samples compared with NF samples. AVG, average; BL, baseline; NF, normal fibroblast; PBO, placebo; RPKM, reads per kilobase of transcript per million mapped reads; SSc, systemic sclerosis; TCZ, tocilizumab.

important mechanisms by which TCZ may exert an antifibrotic effect and build on data from whole-skin gene expression analysis from the faSSinate phase 2 study.<sup>21</sup> Although it was demonstrated

that treatment with TCZ for 24 weeks resulted in a specific down-regulation of genes associated with M2 macrophages and with the IL-6 pathway, little specific effect of TCZ treatment could be

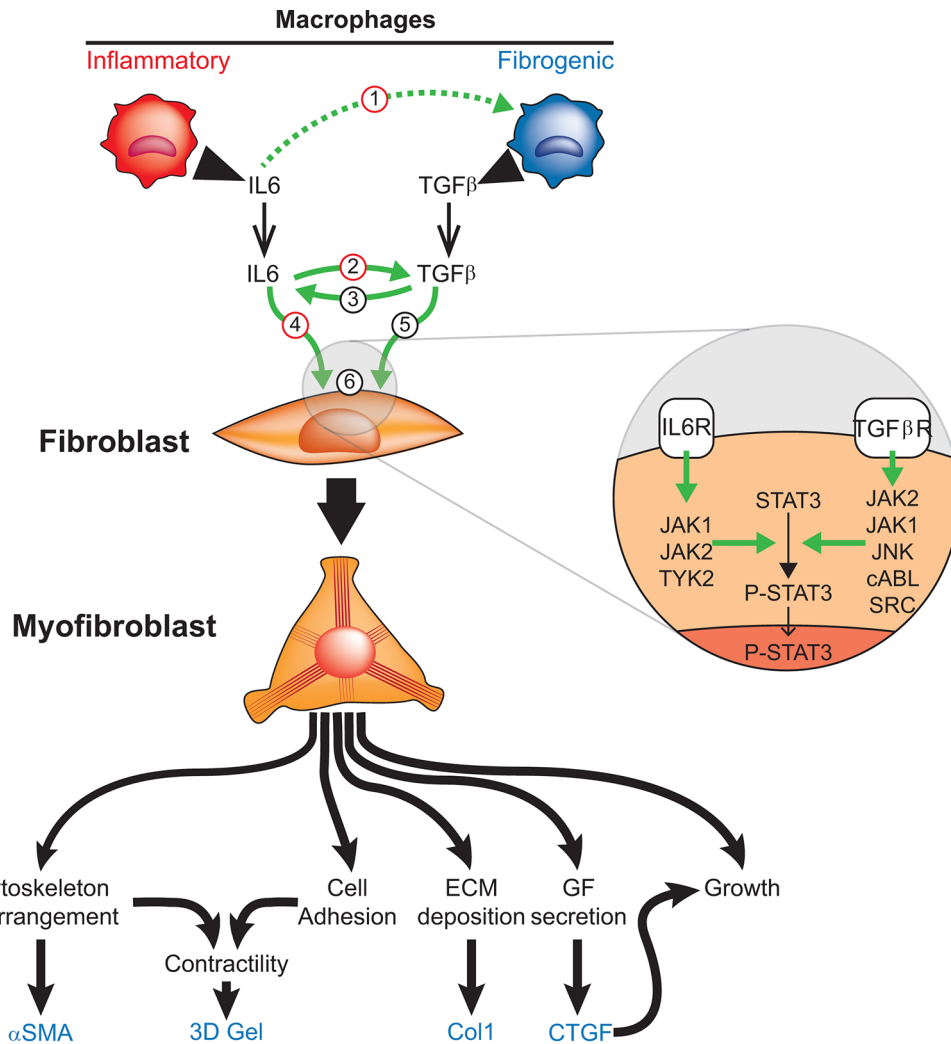


**Figure 5** Treatment with TCZ for 24 weeks normalises the activation status of molecular and functional pathways associated with the biology of SSc fibroblasts. Activation status Z scores for the top 50 associated pathways grouped according to functional nodes. Genes significantly differentially expressed in the SSc versus NF (hatched bars) and the TCZ W24 versus baseline (solid black bars) comparisons (online supplementary figure S1) were analysed with IPA (Qiagen). Differential gene expression data for all differentially expressed genes are available in online supplementary appendix S2. BL, baseline; EGF, epidermal growth factor; IPA, Ingenuity Pathway Analysis; MAPK1, mitogen-activated protein kinase 1; NF, normal fibroblast; SSc, systemic sclerosis; TCZ, tocilizumab; TGFβ1, tumour necrosis factor beta 1; VEGFA, vascular endothelial growth factor A; W24, week 24.

detected on genes associated with fibrosis at the whole-skin biopsy level. Considering the multicellular complexity and relative heterogeneity of whole-skin biopsy samples used in this study, we hypothesised that the effect of TCZ on fibroblasts might be masked by the gene expression of other cellular compartments. The clinical homogeneity of the faSScinate patients and the access to samples collected from the same patients before and after 24 weeks of treatment with TCZ or PBO allowed us to use relatively small numbers of samples to generate robust and reliable results. As a result, we are able to confirm and expand on the description of the

profibrotic properties of SSc dermal fibroblasts at the functional and genomic levels, establish that the profibrotic phenotype of SSc dermal fibroblasts is stable over ≥24 weeks in patients treated with PBO in the absence of immunosuppressant (cyclophosphamide, mycophenolate mofetil or methotrexate) and demonstrate that treatment of SSc patients with TCZ for 24 weeks profoundly alters the biological characteristics of dermal fibroblasts.

The baseline functional and genomic properties of the dermal explant fibroblasts analysed in this study closely match those described in previous publications.<sup>23 24 29</sup> Specifically, the baseline



**Figure 6** Potential mechanisms of interaction between IL-6 and TGFβ leading to fibrosis in SSc. (1) Inflammatory macrophage-produced IL-6 enhances fibrogenic macrophage function (potential TCZ target). (2) IL-6 induces TGFβ production and enhances its signalling in fibroblasts (potential TCZ target). (3) TGFβ enhances IL-6 production by fibroblasts. (4) IL-6 directly activates fibroblasts to produce profibrotic proteins such as Col1 (potential TCZ target). (5) TGFβ directly induces fibroblast differentiation into myofibroblasts. (6) Intracellularly, IL-6 classic signalling and TGFβ alternative signalling converge towards the activation of STAT3 (STAT3 → P-STAT3) resulting in aberrant expression of STAT3-dependent genes. αSMA, alpha-smooth muscle actin; Col1, collagen alpha 1; CTGF, connective tissue growth factor; ECM, extracellular matrix; GF, growth factor; IL-6, interleukin-6; JAK, Janus-activated kinase; JNK, Jun N-terminal kinase; P-STAT, phosphorylated STAT; STAT, signal transducer and activator of transcription; TGFβ, transforming growth factor beta; TGFβR, transforming growth factor beta receptor; TYK, tyrosine kinase.

SSc dermal fibroblasts produced elevated levels of profibrotic proteins and of migration and contractility activities and differentially expressed a distinct set of fibrosis-related genes with a high prevalence of TGFβ-regulated genes. Considering that the faSScinate patients were experiencing active progressive disease at study entry,<sup>21</sup> our observation is consistent with the work of Sargent *et al*,<sup>24</sup> who reported that the differential expression of genes regulated by TGFβ is associated with more severe disease when detected at the whole-biopsy level. Over the years, the study of explant dermal fibroblasts *in vitro* has provided valuable insight into the pathobiology of SSc.<sup>6,7,22,28,30,31</sup> The fact that the phenotypic abnormalities of SSc dermal fibroblasts persist *in vitro* over multiple passages has been critical to this line of investigation. However, it is important to note that dermal fibroblasts are heterogeneous and that they manifest different properties as a function of their microenvironment.<sup>32</sup> The potential selection pressure imposed by isolation, plating and passage procedures makes it likely that only a subset of fibroblasts is accessible to *in vitro* studies. These may represent fibroblasts that are more

motile, better able to acclimate to the culture conditions or have higher growth potential, thus perhaps representing stem cell population(s). For any of these reasons, we might have selected a fibroblast subpopulation in the explant culture conditions more sensitive to the effects of IL-6 inhibition.

We used IPA (Qiagen) to associate gene expression with biological pathways, focusing on the molecular and cellular biology of relevance to fibroblasts. The top 50 pathways associated with SSc differentially expressed genes could be grouped into 10 core biological nodes that covered two levels of biology: biology intrinsic to fibroblasts (fibrosis, contraction, migration, growth and activation) and biology extrinsic to fibroblasts (cardiovascular, coagulation, hormones, inflammation and morphogenesis). The pathways forming the intrinsic group reflect canonical features of fibrotic fibroblasts<sup>5</sup> with a dominant contribution by TGFβ. This dominance of TGFβ-related biology is apparent in the fibrosis node and in the contraction node, where activation of Rho GTPases can be triggered by TGFβ in a SMAD-independent process,<sup>33</sup> and in the activation node, where Ras, RAF1 and



ERK are part of one branch of SMAD-independent TGF $\beta$  signalling.<sup>34</sup> Conversely, the pathways forming the extrinsic group represent biological processes that link inflammation and vasculopathy to fibrosis. For example, not only are the cardiovascular pathways EDN1 and AGT potent mediators of vasoconstriction,<sup>35 36</sup> they are profibrotic mediators relevant to the pathobiology of SSc.<sup>37 38</sup> This is notable considering the prominent vasculopathy of SSc and the putative benefit for patients with SSc of drugs that target these pathways. Similarly, in addition to playing a key role in response to angiogenesis, vascular endothelial growth factor A (VEGFA) and the VEGFA-inducing transcription factor endothelial PAS domain-containing protein 1 amplify fibrosis,<sup>39</sup> creating a link between response to hypoxia and fibrosis. Although excessive inhibition of angiogenesis could be detrimental, our data support normalisation of the activation status of cardiovascular-related pathways rather than complete blockade of these pathways, which might be detrimental for wound healing or tissue repair.

Patient treatment with TCZ for 24 weeks profoundly affected the properties of explant dermal fibroblasts. It is not possible to determine whether treatment with TCZ modified the phenotype of resident cells or the cellular composition of the dermis; both these mechanisms are non-exclusive. Independent of the exact mechanism, the effect of IL-6R $\alpha$  blockade on biological pathways dominated by TGF $\beta$ -related biology is striking and highlights the interaction between IL-6 and TGF $\beta$ . Although TGF $\beta$  is the quintessential profibrotic factor,<sup>9</sup> IL-6 has also been shown to induce the production of fibrotic markers (CTGF,  $\alpha$ SMA and collagen 1) by human fibroblasts *in vitro*.<sup>15</sup> Hence, IL-6 and TGF $\beta$  may form a self-sustaining loop leading to fibrosis that could be interrupted by blockade of either or both cytokines. This is supported by recent work demonstrating that STAT3, the main downstream signalling element for IL-6, is an important TGF $\beta$ -dependent molecular checkpoint of fibrosis in SSc.<sup>40</sup> Altogether, these results and our observations at the whole-biopsy level<sup>21</sup> suggest a potential interaction network between IL-6 and TGF $\beta$  involving fibroblasts and macrophages that leads to fibrosis in SSc (figure 6). Thus, blocking IL-6 signalling with TCZ may represent a novel approach to control both the TGF $\beta$  pathway and the inflammatory pathways, supporting further exploration of IL-6 and related mediators as potential therapeutic targets in SSc and other inflammation-driven fibrotic diseases.

A unique strength of this study is longitudinal sampling of well-characterised and otherwise untreated SSc patients with clinically active disease receiving TCZ or PBO treatment. The combination of clinical homogeneity of patients, repeat sampling at two time points and quality of characterisation assays allowed us to use relatively small numbers of samples to generate robust and reliable results. In addition, the combination of well-characterised functional assays with gene expression analysis using RNA-Seq provides a rare window into the biology of active SSc. However, this study does not address the apparent discrepancy between biological and clinical effects of TCZ on skin fibrosis. Although additional studies are needed to fill this gap, we hypothesise that TCZ primarily affects *de novo* fibrosis and has little impact on the reduction of established fibrosis. Hence, in established skin fibrosis, the clinical effect of TCZ would be substantially delayed after its biological effect.

In conclusion, this study in the field of SSc translational research sheds new light on SSc biology by leveraging the conduct of a well-designed clinical trial in SSc and rigorous functional and molecular analysis of skin biopsy samples to elucidate the links between IL-6 biology and other key pathways and mediators in determining the hallmark profibrotic phenotype of

SSc dermal fibroblasts. Moreover, our findings strongly support further exploration of IL-6 and related mediators as potential therapeutic targets in SSc and other fibrotic diseases.

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**Contributors** All authors were involved in drafting the manuscript or revising it critically for important intellectual content, made substantial contributions to the conception or design of the study or the acquisition, analysis or interpretation of data, approved the final version of the manuscript to be published and agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of the work are appropriately investigated and resolved.

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**Competing interests** CPD reports personal fees from Roche/Genentech, Actelion, GlaxoSmithKline, Sanofi-Aventis, Inventiva and Boehringer-Ingelheim; grants and personal fees from Bayer; and grants from CSL Behring during the conduct of the study. VHO and SX have nothing to disclose. HC-H is an employee of Genentech. ZM, JS and TS are employees of and own stock in Genentech. RL reports personal fees from Biocron and Merck, grants from Elpidera and grants and personal fees from PRISM Biolab outside of the submitted work. DK reports personal fees from Actelion, Cytari, EMD Serono, Gilead, GlaxoSmithKline, Sanofi-Aventis, Corbus, ChemomAb, Eicos and UCB Pharma; grants from Bristol-Myers Squibb, the National Institutes of Health and Pfizer; grants and personal fees from Bayer, Roche/Genentech and Boehringer-Ingelheim during the conduct of the study; and personal fees from AstraZeneca outside the submitted work. AJ is an employee of Genentech, owns stock and stock options in Roche and has a patent issued for tocilizumab.

**Patient consent** Obtained.

**Ethics approval** The study protocol was approved by the institutional review boards or ethics committees before the study commenced. All subjects provided written consent. The study was conducted in accordance with the Declaration of Helsinki and with Good Clinical Practice. 'Control' skin biopsy samples were obtained with informed consent from healthy volunteers (London-Hampstead NRES Committee, MREC Reference 6398).

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