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Review Article

Inflammatory Bowel Disease-Associated **Colorectal Cancer: Translational Risks from** Mechanisms to Medicines

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Abstract

The cumulative impact of chronic inflammation in patients with inflammatory bowel diseases predisposes to the development of inflammatory bowel disease-associated colorectal cancer [IBD-CRC]. Inflammation can induce mutagenesis, and the relapsing-remitting nature of this inflammation, together with epithelial regeneration, may exert selective pressure accelerating carcinogenesis. The molecular pathogenesis of IBD-CRC, termed the 'inflammation-dysplasia-carcinoma' sequence, is well described. However, the immunopathogenesis of IBD-CRC is less well understood. The impact of novel immunosuppressive therapies, which aim to achieve deep remission, is mostly unknown. Therefore, this timely review summarizes the clinical context of IBD-CRC, outlines the molecular and immunological basis of disease pathogenesis, and considers the impact of novel biological therapies.

Key Words: Colitis-associated cancer; cancer; biologics

Inflammatory bowel disease [IBD] describes chronic immunemediated conditions characterized by relapsing-remitting inflammation of the gastrointestinal tract. Ulcerative colitis [UC] and Crohn's disease [CD] are the dominant phenotypes and prevalence is estimated to be as high as 1 in 125 [0.8%] in countries such as the UK.1 While prevalence is rising throughout the world, the

greatest acceleration is observed in newly industrialized countries: since 1990 Africa, Asia and South America have seen an annual percentage change of +11.1% (95% confidence interval [CI] 4.8, 17.8) for CD and +14.9% [95% CI 10.4, 19.6] for UC.² With an ageing population compound prevalence suggests that IBDassociated colorectal cancer [herein IBD-CRC] could become an emerging global issue.

Abbreviations: AOM, azoxymethane; APC, adenomatous polyposis coli; CARD9, caspase recruitment domain containing protein 9; CRC, colorectal cancer; CD, Crohn's disease; CIB, chronic inflammatory burden; DSS, dextran sulphate sodium; Fgl2, fibrinogen-like protein 2; FMT, faecal microbial transplantation; iNOS, inducible nitric oxide synthase; IBD, inflammatory bowel disease; IBD-CRC, inflammatory bowel disease-associated colorectal cancer; IFN, interferon; IL, interleukin; IR, incidence rates; IEC, intestinal epithelial cell; JAK, Janus Kinase; MIF, macrophage migration inhibitory factor: MAdCAM-1, mucosal addressin cell adhesion molecule-1: M-cell, microfold cell; MSI, microsatellite instability; NF-κB, nuclear factor kappa-light-chain-enhancer of activated B cells; NIK, NF-κB-inducing kinase signalling; Nucleotide-binding; NOD, Oligomerisation Domain; PSC, primary sclerosing cholangitis; S1P, Sphingosine-1-Phosphate; SGPL1, S1P lyase1; SIR, standardised incidence ratios; STAT, signal transducer and activator of transcription; SphK1, sphingosine-kinase 1; S-CRC, sporadic CRC; TNF, tumour necrosis factor; TLR, toll-like receptors; TNBS, trinitrobenzene sulfonic acid; UC, ulcerative colitis.

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1. Introduction

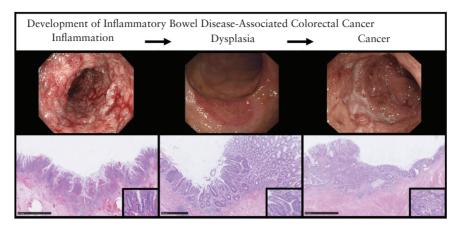


Figure 1. Clinical photographs and photomicrographs of IBD-CRC. Endoscopic images and histopathology photomicrographs illustrate the development of inflammatory bowel disease-associated colorectal cancer [IBD-CRC] through the 'inflammation-dysplasia-carcinoma' sequence. Histopathology photomicrographs provided by Lothian NRS Bioresource [M.J.A.] and colonoscopy images provided by Edinburgh IBD Unit [S.D.].

Over the past two decades, we have defined IBD-CRC through the inflammation–dysplasia–carcinoma sequence [Figure 1]. However, many fundamental questions remain, including elucidation of disease immunopathogenesis. The impact of potent immunosuppressive therapies in IBD, which aim to achieve 'deep remission', is mostly unknown and their subsequent effect on IBD-CRC is yet to be established. This timely review summarizes the epidemiological and clinical context of IBD-CRC, outlines molecular and immunological disease pathogenesis, and considers the impact of novel biological therapies.

2. Patients with IBD are at Increased Risk for Developing CRC with a Poor Prognosis

A 2012 meta-analysis of population-based cohort studies [n = 10,385 patients] reported that patients with UC have an increased risk of developing CRC [standardised incidence ratios (SIR) 2.4, 95% CI 2.1, 2.7], especially if they are male, have extensive colitis and are young when diagnosed with UC.³ A more recent study by Olén and colleagues [n = 96 447 UC and n = 949 207 control patients] reported that, while the incidence of UC-CRC may be decreasing in Scandinavian countries, patients with UC have a 1.7-fold increased risk for incident CRC compared with matched controls.⁴

For CD, Canavan and colleagues published a meta-analysis $[n = 11\ 840\ patients]$ that reported the relative risk for developing CRC in those with colonic disease is 4.5 [95% CI 1.3, 14.9], with a cumulative risk of 2.9% [95% CI 1.5, 5.3] 10 years after diagnosis.⁵ A prospective cohort study from Hong Kong $[n = 2621\ patients]$ reported that patients with CD have an increased risk of anorectal cancer [SIR 4.11, 95% CI 1.84, 9.14].⁶ A more recent study by Olén and colleagues $[n = 47\ 0.35\ CD\ and\ n = 463,187\ matched\ reference individuals] also demonstrated increased CRC incidence in CD: hazard ratio [HR] 1.40 [95% CI 1.27, 1.53].⁷$

There is also an increased IBD-CRC risk for paediatric patients; a 2018 review [n = 271 patients] concluded that, while rare, CRC is the most common fatal malignancy in paediatric IBD patients.⁸

IBD-CRC confers a poor prognosis. A large meta-analysis [n = 3472 patients] reported that patients with IBD-CRC have poorer overall survival compared to patients with sporadic[S]-CRC [HR 1.24 95% CI 1.19, 1.29].⁹ These patients were more likely to have proximal tumours [odds ratio [OR] 2.52, 95% CI 1.35, 4.72) and poorer histopathological differentiation [OR 1.59, 95% CI

1.26, 1.99]. Olén and colleagues also reported patients with UC have a 1.6-fold increased risk of death from cancer, compared with S-CRC.⁴ Similarly, patients with CD have increased mortality compared with matched controls [HR 1.42, 95% CI 1.16, 1.75], when adjusted for tumour stage.⁷ Reported differences between IBD-CRC and S-CRC prognosis are probably due to differences in tumour biology [Table 1].¹⁰

3. IBD-CRC Develops from Dysplasia and Inflammation is a Critical Initiating Factor

With an increasing global prevalence of IBD, and patients living longer, it is important to consider the cumulative impact that multiple occurrences of acute and chronic inflammation have on the development of IBD-CRC. Clinicians strive to modulate natural disease progression at a very early stage, often using potent agents to achieve early mucosal healing. The 'top down' or 'treat to target' approach aims to reduce the risk of hospitalizations, future use of biologics and surgery. While lower colectomy rates are desirable, preservation of damaged colorectum, particularly in the setting of potent immunomodulation, is unknown and could result in an increased incidence of IBD-CRC. Understanding the molecular and immunological pathogenesis of IBD-CRC is therefore important for clinicians and scientists to develop new therapies that achieve deep remission and reduce IBD-CRC risk.

Our current understanding of S-CRC is defined through the sequential histological and genetic changes known as the adenomacarcinoma sequence [Figure 2A]. In contrast, IBD-CRC develops through the 'inflammation–dysplasia–carcinoma' sequence [Figure 2B]. Here, low-grade dysplasia develops on a background of mucosa that has been genetically altered by chronic inflammation and is at increased risk of malignant progression. Inflammation can induce mutations and the relapsing–remitting nature of this inflammation with proliferative epithelial regeneration exerts selective pressure that accelerates evolution.¹¹ Increased reactive oxygen species production and lipid peroxidation and decreased antioxidant capacity with increased oxidative DNA damage in IBD are likely mechanisms that drive mutagenesis.^{12,13}

Mutations that contribute to IBD-CRC pathogenesis are similar to those implicated in S-CRC; however, the order that mutations are accrued is often described as 'reversed'. Early loss of *TP53* function is a hallmark of IBD-CRC, with mutations observed in
 Table 1.
 IBD-CRC is distinct from S-CRC. This table illustrates the key epidemiological, pathophysiological and clinical differences between sporadic [S-CRC] and inflammatory bowel disease-associated colorectal cancer [IBD-CRC]

	S-CRC	IBD-CRC
Epidemiology		
Disease burden	10% of global cancer diagnoses. ¹⁰⁰	IBD patients have a higher incidence of CRC, possibly >60%. ^{4,7,101}
Sex	Male preponderance. ^{100,102}	Male preponderance. ³
Age	Older age of onset [>50 years old]; an increasing incidence in younger patients. ¹⁰²	Younger age of IBD onset. ³ Paediatric IBD-CRCs can develop. ⁸ Patients with IBD are living longer. ¹
Risk factors	Diet, smoking, obesity, family CRC history, <i>H. pylori</i> , alcohol [J-shaped association likely], colonic polyps and others. ¹⁰³⁻¹⁰⁶	Extensive colitis, increased duration of disease, family history of S-CRC, PSC. ¹⁰⁷⁻¹⁰⁹
Disease pathogenesis		
Pre-malignant lesion	Adenomatous polyps [polypoid/sessile]. ¹¹⁰	Flat dysplasia. Genetic aberrations seen [<i>TP53</i> mutations] in histologically normal mucosa. ^{11,14,17}
Molecular sequence	Mostly adenoma-carcinoma sequence [slow]. ¹¹⁰	Inflammation-dysplasia-carcinoma sequence [fast]. ¹¹
Genetic aberrations	Chromosomal instability, microsatellite in- stability and CpG island methylator phenotype [CIMP] pathways [not mutually exclusive]. Early and more frequent <i>APC</i> mutations. Late and frequent <i>TP53</i> mutations. ^{14,15,111,112}	Mutation sequence is 'reversed': early <i>TP53</i> mutation, late and infrequent loss of <i>APC</i> , earlier MSI, later <i>KRAS</i> mutations. ¹⁴⁻¹⁶
Contribution of inflammation and regeneration to the initiation of cancer	Promotes cancer progression. ¹¹³	Drives mutagenesis and selects for mutagenic clones. ¹¹⁻¹³
Contribution of inflammation to the progression to cancer	Tumour-promoting inflammation is critical for most cancers, including colorectal cancer. ¹¹³	Critical pathways signal through NF-κB and IL-6/STAT3. ^{26-32,35-40,42,46,114} Th17 cells and associated cytokines are generally pathogenic. ^{30,43,45,47-49,52,53,55,56,58,115}
Clinical features		
Endoscopic characteristics	Commonly raised/polypoidal lesions; some sessile.	Flat dysplasia. Synchronous and recurrent tumours.
Histological characteristics	Majority adenocarcinoma. Comparatively favourable differentiation; fewer contain mucinous/signet ring cell morphology. ¹¹⁶	Majority adenocarcinoma. Mucinous/signet ring cell differentiation is more common. ^{10,116}
Mortality and prognosis	Prognosis is improving, especially if diagnosed early. ¹⁰²	Poor prognosis compared with S-CRC [2-fold]. ¹¹⁷ Increase in recurrence [3-fold]. ¹⁰ Poor prognosis associated with PSC, male sex, extensive colitis and early age of diagnosis. ^{4,7,9,10,107,108,118}

Abbreviations: APC [adenomatous polyposis coli]; IBD-CRC [inflammatory bowel disease-associated colorectal cancer]; IL-6 [interleukin-6]; KRAS [Kirsten rat sarcoma viral oncogene homologue]; MSI [microsatellite instability]; NF-κB [nuclear factor kappa-light-chain-enhancer of activated B cells]; PSC [primary sclerosing cholangitis]; qFIT [quantitative faecal immunohistochemical test]; S-CRC [sporadic colorectal cancer]; UC [ulcerative colitis]; STAT3 [signal transducer and activator of transcription 3].

diploid, non-dysplastic epithelial cells, and they precede TP53 loss of heterozygosity.14 In contrast, S-CRC TP53 mutations occur late.14 KRAS activating mutations are important alternative gatekeeper mutations that occur later and less frequently in IBD-CRC $[{\sim}20\%$ of cases].15 Hypermethylation of the tumour suppressor gene MLH1 also occurs earlier in IBD-CRC, and this contributes to microsatellite instability [MSI] in an important subset of cancers, similar in frequency to that observed in S-CRC.¹⁶ Dysregulation of the Wnt signalling pathway plays an important role in IBD-CRC, with ~55% of dysplastic lesions and up to 100% of cancers expressing nuclear β-catenin.¹⁷ However, unlike the S-CRC pathway, which has early loss of APC function leading to aberrant Wnt signalling, APC function is lost late in IBD-CRC and only occurs in <50% of cases.^{15,18} This may be explained by inflammation-driven upregulation of β -catenin in IBD-CRC, which can induce APC mutation-independent Wnt signalling.19

Throughout the IBD colon, genetic and epigenetic abnormalities develop in histologically normal mucosa and can expand to form pre-malignant patches.²⁰ IBD patients have dysplastic lesions with increased chromosomal instability compared with sporadic adenomas.²¹ Chronic inflammation is the underlying mechanism that leads to telomere shortening, and thus chromosomal instability,²² in pre-malignant IBD mucosa.²³ This induces senescence, which acts as a tumour-suppressor mechanism to prevent progression past low-grade dysplasia. Mutant intestinal epithelial cells [IECs] eventually escape senescence and progress—this is associated with telomere lengthening and loss of *TP53* function.

The mutational landscape of cancer is diverse; our recent mutational analysis of 34 IBD-CRCs identified six distinct mutational signatures.²⁴ In S-CRC with MSI, patients have a better prognosis by at least 15%, probably due to a cumulative mutational burden with resulting anti-tumour immune cell responses.²⁵ In IBD-CRC, proximal 2134

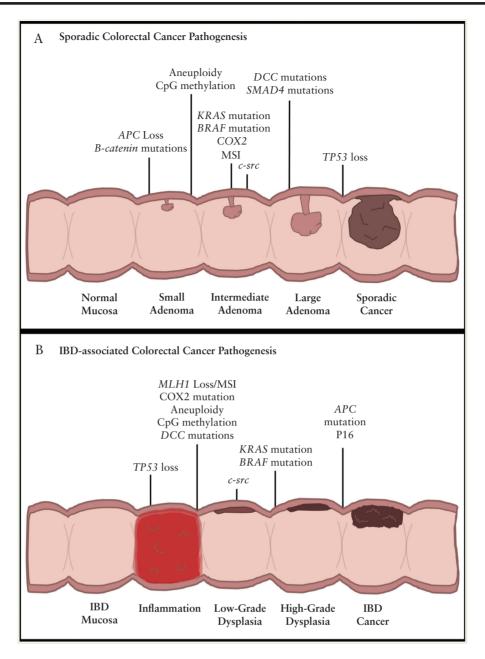


Figure 2. Contemporary model for the molecular pathogenesis of [A] sporadic and [B] IBD-associated colorectal cancer. Sporadic colorectal cancer [S-CRC] develops through the 'adenoma-dysplasia-carcinoma sequence' whereas inflammatory bowel disease-associated colorectal cancer develops through the 'inflammation-dysplasia-carcinoma' sequence. The figure illustrates genetic mutations that can contribute to cancer development.

tumours have high mutational rates, are associated with MSI [especially loss of *MLH1* and defects in DNA *POLE* proofreading function], and have a higher predicted neo-epitope load,²⁴ suggesting increased immunogenicity. It is unknown if or how chronic inflammation in IBD influences the development of IBD-CRC mutational signatures or molecular phenotypes.

4. Dysregulation of Critical Immune-Mediated Pathways in IBD-CRC

4.1. NF- κ B and IL-6/STAT3 signalling pathways promote IBD-CRC

The two most comprehensively studied pro-inflammatory and pro-tumour pathways in IBD-CRC are the nuclear factor kappa-light-chain-enhancer of activated B cells [NF- κ B] and interleukin [IL]-6/signal transducer and activator of transcription [STAT]3 signalling pathways. These pathways are well established²⁶⁻²⁸ [Figure 3].

In summary, inhibition of the canonical NF- κ B pathway abrogates tumorigenesis in the azoxymethane [AOM]/dextran sulphate sodium [DSS] mouse model by two main mechanisms: [1] IKK β deletion in myeloid cells reduces both intestinal inflammation and tumour size through decreased production of pro-inflammatory cytokines (IL-1 α , IL-1 β , IL-6, KC, MIP-2, tumour necrosis factor [TNF]- α , COX-2 and ICAM), and [2] IKK β deletion in IECs reduces tumour incidence and is associated with apoptosis; however, it does not reduce inflammation.²⁹ Non-canonical NF- κ B signalling, mediated by NF- κ B-inducing kinase signalling [NIK], contributes to intestinal homeostasis through maintenance and differentiation

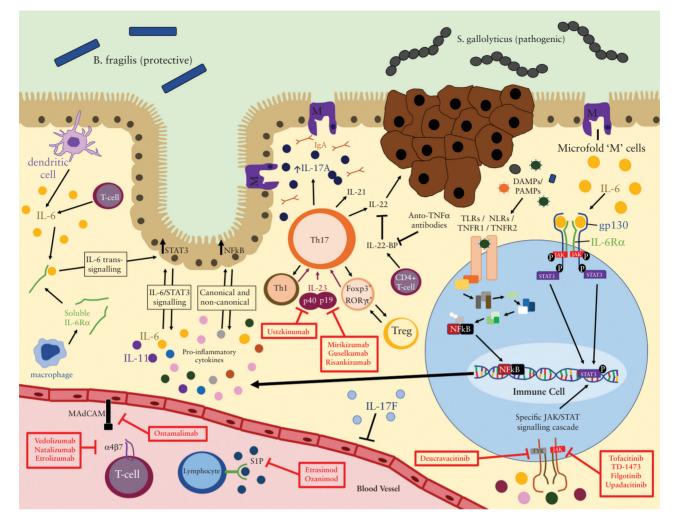


Figure 3. Immunopathogenesis of inflammatory bowel disease-associated colorectal cancer [IBD-CRC]—a perfect storm? There is dysregulation of critical immune-mediated pathways in IBD-CRC, such as NFκB and IL-6/STAT3 signalling, and Th17 cell responses. The impact of novel therapeutic immunomodulators requires careful consideration.

of microfold [M]-cells [specialized epithelial cells of mucosalassociated lymphoid tissue], and local and systemic IL-17A and IgA production.³⁰ Mice with intact NIK are protected against colitis; however, constitutively activated NIK signalling worsens colitis and is associated with increased IL-17A production and ectopic colonic M-cells.³⁰ NF- κ B signalling can also be driven by genetic aberrations: mutant p53 augments and prolongs the response of IECs to low levels of inflammatory cytokines, resulting in chronic NF- κ B activation, which promotes persistent tissue damage and inflammation.³¹ Mutant p53 mice exposed to DSS are prone to colitisassociated cancer: the gain-of-function mutation is associated with flat dysplastic lesions that progress to cancer, similar to those seen in IBD-CRC.³¹

TNF- α is the quintessential pro-inflammatory cytokine and can bind to either of its receptors [TNFR1 or TNFR2] and induce inflammation through either canonical or non-canonical NF- κ B signalling pathways. There are data suggesting TNF- $\alpha\alpha$ can enhance Wnt signalling through NF- κ B actvation³² and promote mucosal regenerative healing through colonic epithelial stem and progenitor cell populations.³³ A protective role for TNF is perhaps controversial. Nonetheless, this is important to consider in IBD-CRC as IEC p53 stabilization post-immune activation is dependent on TNFR1/2 and inducible nitric oxide synthase [iNOS].³⁴ TNF- α -induced iNOS activates a p53-dependent pathway of IEC apoptosis, and this may hypothetically be prevented in patients receiving anti-TNF treatment. This could mean that without p53 wild-type function, such as during early IBD-CRC, damaged IECs evade apoptosis and thus have selective advantage.³⁴

NF-κB signalling in immune and epithelial cells upregulates IL-6 and constitutively activates STAT3 in human tumours, maintaining NF-κB signalling.³⁵ Thus, IL-6/STAT-3 and NF-κB signalling are not mutually exclusive. IL-6/STAT3-deficient mice treated with AOM/ DSS have a reduced tumour burden compared with wild-type mice.^{36,37} This is because IL-6, produced mainly by bone marrowderived myeloid cells, such as macrophages and dendritic cells [along with some T-cells], increases IEC proliferation and resistance to apoptosis through the STAT3-dependent pathway.^{36,37} Furthermore, STAT3 acts as a critical mediator for stimulating cell survival [*bcl-x*, *survivin*, *Hspa1a*] and proliferation [cdk4/cyclinD1, cdc2/cyclinB1, cMyc, RegIIIb/PAP], through G1 and G2/M phases of the cell cycle to promote carcinogenesis.³⁷

Increased IL-6 expression in UC is also associated with reduced nuclear expression of *MSH3*, and this increases with duration of disease, as well as when dysplasia then cancer develops; expression in UC-CRC is higher than in S-CRC.³⁸ This suggests a link between IL-6 signalling and MSI in IBD-CRC. Trans-IL-6 signalling, in which IL-6 binds to soluble IL-6R and dimerizes with gp130 on cells that do not express IL-6R, is also important; macrophage-derived IL-6/soluble IL-6R α is particularly important.^{39,40}

STAT3 signalling can also be induced by other cytokines, such as IL-11, which may be more potent.⁴¹ STAT3 is important to Th17 cell function as FAM64A, a multifunctional protein involved in cell cycle progression, drives the IL-6/STAT signalling pathway and induces Th17 differentiation in AOM/DSS-induced murine colitis.⁴²

4.2. Th17 cells and associated cytokines promote IBD-CRC

Patients with IBD have increased numbers of Th17 cells and associated cytokines [IL-17, IL-21 and IL-22] in their intestinal mucosa and peripheral blood, compared with healthy controls.⁴³ It is important to differentiate IL-17A [which promotes inflammation and tumorigenesis] from IL-17F [which is protective against IBD-CRC, possibly by inhibiting angiogenesis].^{44,45} IL-6/STAT3 signalling is also involved in the induction of T-cell RORyt expression, which is a key transcription factor of Th17 cells.⁴⁶ However, Th17 cells demonstrate functional plasticity and can convert into interferon [IFN]-y producing Th1 cells or regulatory T-cells.⁴⁷

IL-23 is produced by many antigen presenting cells and plays an important role in maintaining the Th17 cell phenotype.⁴⁸ In IBD-CRC, data suggest Baft-dependent IL-23*IL-6*CD4*Th17 cells, rather than RORγt-dependent Th17 cells, mediate downstream effects of IL-23.⁴⁹ IL-23 and IL-12 are part of the IL-12 family of cytokines, both share the p40 subunit, and they heterodimerize with p19 or p35, respectively.^{50,51} p47phox is a protein of NADPH oxidase that regulates induction of the TLR9-induced IL-12/Th1 axis. In AOM/DSS-treated mice, IL-12p35^{-/-} mice have reduced colitis but increased susceptibility to CRC, whereas p47phox^{-/-} mice have worsened colitis but reduced tumour growth.⁵² Therefore, tilting the IL-23/IL-12 balance toward IL-12 might reduce tumorigenesis in IBD-CRC. However, this is not viable as this would probably worsen IBD symptoms.⁵²

IL-22 is a pleiotropic cytokine, part of the IL-10 family, that is produced by mature Th17 cells through IL-23-mediated STAT3 activation.⁴⁷ In Rag2-/- mice with Helicobacter-associated colitis, IL-22 induces iNOS within IECs, which induces DNA damage and dysplasia.53 Patients with IBD have increased CD4+ T-cells that produce high levels of IL-22 binding protein [IL-22BP]: IL-22BP is a soluble IL-22 receptor, without a transmembrane/intracellular domain, that binds to and neutralizes IL-22. The anti-inflammatory effects of TNF- α antibodies have been associated with reduced levels of IL-22BP.⁵⁴ IL-22BP is highly expressed in dendritic cells and during NLRP3 or NLRP6 inflammasome activation, such as in IBD; inflammasome activation can lead to IL-18-dependent IL-22BP downregulation. IL-22BP-/- mice also show strongly accelerated tumour growth.55 Pleiotropic effects are likely because IL-22 is initially protective in inflammation, but induces tumorigenesis if uncontrolled during restitution of inflammation.55 Therefore, dysregulation of the IL-22/ IL-22BP axis may play a pivotal role in IBD-CRC development, perhaps in the context of anti-TNF therapy.

IL-21 is a multifunctional cytokine produced mainly by T-cell subsets such as follicular helper T-cells and Th17 cells. Some studies suggest IL-21-deficient mice are protected from DSS and trinitrobenzene sulfonic acid [TNBS]-induced colitis and this is likely because they are unable to upregulate Th17 responses.⁵⁶ However, other studies have suggested that IL-21 signalling, through IL-21R, is protective in

DSS-treated mice due to downregulation of Th1 and upregulation of Th2, Th17 and Treg responses.⁵⁷ Studies have reported a reduced tumour burden in AOM/DSS-treated IL-21^{-/-} mice,⁵⁸ and the underlying mechanism may be due to a reduced number of infiltrating T-cells, reduced STAT3 signalling and thus reduced IL-17A and IL-6.⁵⁸

4.3. Recent advances in the immunopathogenesis of IBD-CRC

There remains a paucity of data characterizing the immune cell landscape in human IBD-CRC. From the studies that exist, IBD-CRC has a lower number of immune cells expressing CD3, CD8, Foxp3 or PD-L1;⁵⁹ increased CD3⁺ and CD8⁺ lymphocytes are associated with improved prognosis.⁵⁹ There is a need to comprehensively characterize the cells and cytokines that define the inflammation–dysplasia–carcinoma sequence.

The role of macrophages in IBD-CRC is poorly defined. Macrophages and mast cells infiltrate the colonic submucosa in a stage-dependent manner in the progression from inflammation to dysplasia to cancer.⁶⁰ Macrophage migration inhibitory factor [MIF] mediates macrophage and T-cell recruitment and MIF-/- mice treated with AOM/DSS have an increased tumour burden, associated with lower levels of macrophages.⁶¹ Fibrinogen-like protein 2 [Fgl2] may be important for macrophage recruitment with/without polarization as Fgl2 loss induces M1-polarized and suppresses M2-polarized macrophages; Fgl2 may therefore reduce inflammation and IBD-CRC.62 In contrast, TGFB promotes macrophage recruitment through expression of CCR2 in the tumour microenvironment, and myeloid-cell TGF_{β2} expression worsens AOM/DSS-induced tumorigenesis. Conditional TGFB2 knock-out mice have reduced IL-6 and TNF-α expression, and increased numbers of Foxp3⁺ T-regulatory cells [Tregs] in the early stages of carcinogenesis.⁶² This suggests a pathogenic role of TGF^β signalling via macrophages in IBD-CRC. This hypothesis is strengthened by a study that reported conjugated linoleic acid ameliorated DSS-induced murine colitis through a macrophage PPARy receptor-dependent pathway, whereas PPARy activation induced TGFB production by macrophages and T-cells that increased tumourigenesis in AOM/DSS-induced colitis.63

A novel subset of Foxp3*RORyt* T-cells have been described and were thought to represent an intermediate stage during differentiation between immunosuppressive Tregs and proinflammatory Th17 cells. However, these cells can also be stable and functional [regulatory] in the intestine. Patients with IBD have increased expression of this unique T-cell subset, associated with IBD-dysplasia.⁶⁴⁻⁶⁶ A recent study using AOM/DSS-treated mice reported that Foxp3*RORyt* T-cells reduce the expression of FoxO3 in tumour infiltrating dendritic cells—FoxO3 is a transcription factor that controls the production of IL-6 by antigen presenting cells. This results in aberrant IL-6 signalling that upregulates STAT3 and induces proliferation of dysplastic cells.⁶⁶

Barcode sequence analysis using 16S [MiSeq for bacteria] and ITS2 [pyrosequencing for fungi] reported a difference in bacterial, but not fungal, microbiome populations in IBD-CRC patients compared with both healthy controls and S-CRC.⁶⁷ Compared with S-CRC, IBD-CRC patients have increased abundance of the family Enterobacteriaceae family and genus *Sphingomonas* and reduced abundance of the genera *Fusobacterium* and *Ruminococcus*.⁶⁷ The mechanistic, clinical and therapeutic consequences of microbial dysbiosis in IBD-CRC are poorly understood. Some bacteria, such as *Bacteroides fragilis*, are protective^{68,69} whereas others such as *Streptococcus gallolyticus* are pathogenic.⁷⁰ Faecal microbial transplantation [FMT] is being trialled as a therapy for IBD, with variable success. Transplantation of carcinogenic bacteria may occur during FMT; robust screening and appropriate follow-up is needed to minimize FMT-associated IBD-CRC.⁷¹

Antimicrobial peptides, such as cathelicidin/LL-37, are essential for maintaining intestinal homeostasis. It is unsurprising that AOM/DSS-treated cathelicidin-related antimicrobial peptide knockout mice have an increased tumour burden.⁷² This field is in its infancy and further work is required to determine whether bacteria and/or defence peptides play a significant role in the development of IBD-CRC.

Zhong and colleagues recently reviewed conflicting data on caspase recruitment domain-containing protein 9 [CARD9], an adaptor protein that can mediate inflammation.⁷³ The pro-tumour role for CARD9 may be through IL-1 β -mediated STAT3 activation; however, whether CARD9 contributes to inflammasome-mediated cytokine production and whether intestinal fungi promote or prevent IBD-CRC are undetermined.⁷³

The immunomodulatory role of the appendix has also recently attracted interest, as appendicectomy has been shown to induce clinical improvement in UC.⁷⁴ However, appendicectomy is also associated with an increased risk of developing IBD-dysplasia and IBD-CRC.⁷³ While trials are ongoing,⁷⁶ mechanistic data elucidating the impact of appendicectomy on the gut microbiota and immune cell responses are currently lacking.

5. Immunosuppression can be a Double-Edged Sword in IBD-CRC

The pro- or anti-tumour effects of 5-aminosalicylate, traditional immunomodulators [e.g. thiopurine] and anti-TNF therapy in IBD have been extensively discussed elsewhere.⁷⁷⁻⁷⁹ IBD Cancer and Serious Infection in Europe [I-CARE] is an ongoing prospective, longitudinal, observational, multicentre [n = 16 countries] cohort study that aims to determine the risk of developing cancer or serious infections in IBD patients receiving immunosuppressive and biological therapies [NCT02377258]. Newly identified signalling pathways that can be manipulated to ameliorate inflammation may have unintended carcinogenic effects. This section explores the potential proor anti-tumour effects of the latest targeted IBD biological and small molecule therapies.

5.1. Therapeutic manipulation of the IL-12/IL-23 axis

Ustekinumab is a humanized monoclonal antibody that binds to the p40 subunit that comprises both IL-12 and IL-23.80 The impact of p40 neutralization on IBD-CRC development is mostly unknown; however, the impact of neutralizing IL-12 and IL-23 activity can be considered separately. IL-12 induces anti-tumour immunity [involving IFN-y, CD4+ and CD8+ T-cells]⁸¹ whereas IL-23 can promote carcinogenesis involving IL-17-associated pathways.52 Teng and colleagues investigated the impact of IL-23 and IL-12 on methylcholanthrene-induced p53 mutant cancers in murine models and reported that IL-23p19 inhibition reduced the malignant potential of colonic lesions whereas IL-12/23p40 inhibition enhanced tumour outgrowth.82 Therefore, neutralizing p40 may have some theoretical or potential pro-tumour effects in humans. A randomized control trial involving 961 patients with moderate-to-severe UC reported one case of colonic and one case of rectal cancer in patients receiving ustekinumab [n = 825] over 52 weeks, compared with zero CRC in patients receiving placebo [n = 319].⁸³ An observational cohort study has started recruiting patients to assess the long-term safety of ustekinumab compared with other biologics in CD; the primary outcome is incidence of malignancy with a time frame of up to 12 years [NCT04372108].

Targeted anti-IL-23 therapies are thus being explored for IBD as, theoretically, targeted IL-23 blockade therapy may ameliorate inflammation and reduce the risk of IBD-CRC.⁸⁴ Anti-IL-23 therapies against the p19 subunit are being trialled for CD patients compared with ustekinumab, including risankizumab [NCT04524611], mirikizumab [NCT03926130] and guselkumab [NCT03466411]. All trials have relatively short follow-up periods, which limits their usefulness for inferring overall IBD-CRC risk.

5.2. Therapies targeting leukocyte trafficking

 $\alpha 4\beta 7$ is an integrin [a transmembrane protein that facilitates cell adhesion] expressed on lymphocytes and is associated with increased responsiveness to pro-inflammatory cytokines IL-6, IL-7 and IL-21.⁸⁵ $\alpha 4\beta 7$ allows peripheral lymphocytes to bind with mucosal addressin cell adhesion molecule-1 [MAdCAM-1] on intestinal endothelial cells, which allows lymphocytes to undergo diapedesis into the lamina propria.

Vedolizumab is the first humanized, gut-selective antibody used to treat IBD that blocks α4β7 integrin-expressing lymphocytes from trafficking from the systemic circulation into the lamina propria.86 Data generally support no increased risk of malignancy in patients receiving vedolizumab: a retrospective analysis of 1087 patients reported only one case of IBD-CRC.87 However, median follow-up in this study was only 302 days, which is too short to determine the true risk of malignancy.⁸⁷ These findings are supported by other data, such as a study that reports four CRC cases in a population of >2800 patients, which translates to 0.1/100 person years, and is no different to the background IBD risk [2.1/1000 person years, 95% CI 1.3, 3.2].88 A recent retrospective cohort study reported no increased risk of new or recurrent cancer among patients with IBD and a history of cancer who were treated with vedolizumab or anti-TNF therapy, compared with patients who did not receive immunosuppression [follow-up median 6.2 person years].89 Caution may be warranted, especially in patients with concurrent primary sclerosing cholangitis [PSC]. A retrospective observational cohort study [median follow-up of 19 months] reported that, of 75 patients with IBD and PSC treated with vedolizumab, nine developed digestive neoplasia [seven of which were colorectal cancers].⁹⁰ While there are no published data for similar therapies, there is an ongoing large interventional trial investigating etrolizumab in UC [9 years follow-up] that may reveal data regarding IBD-CRC risk [NCT02118584].

Preliminary data suggest MAdCAM antibodies are efficacious in IBD, especially UC.⁹¹ No studies have assessed the impact of MAdCAM antibodies on IBD-CRC development; however, MAdCAM-1 expression is reduced in colonic adenocarcinomas.⁹² This could suggest that blocking MAdCAM may be advantageous for tumour development, warranting further investigation. There is an ongoing safety extension study investigating ontamalimab [MAdCAM-1 inhibitor] for the treatment of moderate to severe IBD [NCT03283085].

5.3. Small molecule therapies

Compared with monoclonal antibodies, small molecules are attractive as they have no inherent immunogenicity [they are synthetic drugs rather than proteins], can be administered orally and have relatively stable and predictable pharmacokinetics. Small molecules include janus kinase [JAK] inhibitors, tyrosine kinase inhibitors and sphingosine-1-phosphate [S1P] receptor antagonists.

Cytokines principally impact immune cell function by signalling through JAK/STAT pathways. As previously discussed, targeting these pathways could theoretically improve colitis and reduce the risk of IBD-CRC. With JAK inhibitors, the extent of signal inhibition is related to the target of the small molecule [i.e. pan vs selective JAK inhibition]. Most data relate to tofacitinib: data from 1157 patients who received tofacitinib [a JAK3-specific inhibitor, with lesser activity against JAK1/JAK2] reported that 11 patients developed malignancy [excluding non-melanoma skin cancer], one of which was colorectal adenocarcinoma-the risk of malignancy was not significant [incidence rates (IR) 0.7, 95% CI 0.3, 1.2].93 However, we must be cautious as these data are only over 4.4 years. Another study reported that 2/1124 patients receiving tofacitinib developed CRC [IR 0.08, 95% CI 0.01, 0.27], with evaluation up to 6.8 years; however, one cancer possibly developed prior to tofacitinib therapy.94 Data from other JAK inhibitors are on the horizon for IBD, such as TD-1473 [NCT03920254], filgotinib [NCT02914600, NCT02914535] and upadacitinib [NCT03345823, NCT02782663, NCT03006068]. Tyrosine kinase inhibitors may also be therapeutic in IBD, and deucravacitinib [BMS-986165] is an allosteric inhibitor of tyrosine kinase 2 under investigation for CD [NCT03599622] and UC [NCT03934216, NCT04613518]. Long-term follow-up studies using these patient cohorts will be important to determine IBD-CRC risk.

Sphingolipids are ubiquitous bioactive molecules that form part of the cell membrane and play a role in a multitude of cell functions such as migration, proliferation and apoptosis. S1P is the final product derived from sphingolipids and can activate STAT3 and NF-kB. STAT3-induced S1P receptor expression is important for persistent STAT3 activation [creating a positive feedback loop in immune and tumour cells] during carcinogenesis.95 S1P also has immunoregulatory activity as T-cells require S1P signalling to egress from the thymus and from peripheral lymphoid organs.96 A protumour role for S1P, and its regulatory enzyme sphingosine-kinase 1 [SphK1], has therefore been hypothesized. In a small [n = 20 patients] translational study, biopsies from curative surgical resections reported higher expression of phosphorylated SphK1 in IBD-CRC compared with S-CRC, which suggests the S1P pathway is especially important for IBD-related malignancy.97 Given the outlined mechanism of action, inhibiting S1P may reduce both IBD and IBD-CRC. Carcinogenesis may be triggered by S1P lyase1 [SGPL1], which is responsible for the irreversible degradation of S1P. In an AOM/ DSS model that utilized isogenic bone marrow transplantation of inducible SGPL1 knockout mice, immune-cell SGPL1 knockout was associated with colitis and pathological crypt remodelling with extracellular S1P signalling, which caused delayed tumour formation. However, tissue-SGPL1 knockout reduced immune activity and induced immediate tumorigenesis which was associated with an IL-12 to IL-23 shift.98 This suggests that understanding the difference between tissue vs immune cell S1P lyase activity is important for optimizing S1P blockade therapy to treat IBD and reduce IBD-CRC risk. There are ongoing safety and efficacy trials for the S1P receptor modulators, such as etrasimod [NCT03950232] and ozanimod [NCT0253112].

6. Conclusions

Patients with IBD have an increased risk for developing CRC. The inflammation–dysplasia–carcinoma sequence of IBD-CRC is distinct from the sporadic normal–adenoma–adenocarcinoma sequence and

confers a poorer prognosis. While key inflammatory pathways have been described, the immune cell landscape of IBD-CRC remains poorly characterized, although new data suggest Th17 cells and macrophages play important roles.

The selection pressure exerted by efficacious therapeutic agents is unknown and, although the overall inflammatory burden is lessened, the potential disruption to immune cancer surveillance is yet to be fully appreciated. Indeed, it will be some time before we see the full effect of these therapies on the incidence of IBD-CRC. More sensitive tools to detect early dysplasia are needed, and may include non-invasive stool testing for DNA methylation markers and improved computer-aided identification of dysplasia during surveillance procedures.⁹⁹

In the clinic, preventing the burden of inflammation is probably the most important factor for minimizing IBD-CRC. With the growing therapeutic arsenal, caution is warranted that immunosuppression can be a double-edged sword, and it will be some time before we see the full effect of these therapies on the incidence of IBD-CRC.

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Conflicts of Interest

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Author Contributions

S.D., M.J.A. and R.J.P. contributed to review conceptualization and design. R.J.P. wrote the first draft of the manuscript and drafted Figures and Tables. M.J.A. and A.M.D.C. provided histopathology photomicrographs [via Lothian NRS Bioresource] and S.D. provided endoscopy images [via Edinburgh IBD Unit]. R.J.P., M.J.A., A.M.D.C. and S.D. performed literature searches, reviewed articles for inclusion, and reviewed, edited and approved the final manuscript [including Figures and Tables]. Medical Writer or Editor – none.

Data Availability

No new data were generated or analysed in support of this research.

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References

 Jones GR, Lyons M, Plevris N, *et al.* IBD prevalence in Lothian, Scotland, derived by capture–recapture methodology. *Gut* 2019;68:1953–60.

- Ng SC, Shi HY, Hamidi N, *et al*. Worldwide incidence and prevalence of inflammatory bowel disease in the 21st century: a systematic review of population-based studies. *Lancet* 2017;390:2769–78.
- Jess T, Rungoe C, Peyrin-Biroulet L. Risk of colorectal cancer in patients with ulcerative colitis: a meta-analysis of population-based cohort studies. *Clin Gastroenterol Hepatol* 2012;10:639–45.
- Olén O, Erichsen R, Sachs MC, *et al*. Colorectal cancer in ulcerative colitis: a Scandinavian population-based cohort study. *Lancet* 2020;395:123–31.
- Canavan C, Abrams KR, Mayberry J. Meta-analysis: colorectal and small bowel cancer risk in patients with Crohn's disease. *Aliment Pharmacol Ther* 2006;23:1097–104.
- So J, Tang W, Leung WK, et al. Cancer risk in 2621 Chinese patients with inflammatory bowel disease: a population-based cohort study. Inflamm Bowel Dis 2017;23:2061–8.
- Olén O, Erichsen R, Sachs MC, et al. Colorectal cancer in Crohn's disease: a Scandinavian population-based cohort study. Lancet Gastroenterol Hepatol 2020;5:475–84.
- Aardoom MA, Joosse ME, de Vries ACH, Levine A, de Ridder L. Malignancy and mortality in pediatric-onset inflammatory bowel disease: a systematic review. *Inflamm Bowel Dis* 2018;24:732–41.
- Ou B, Zhao J, Guan S, Lu A. Survival of colorectal cancer in patients with or without inflammatory bowel disease: a meta-analysis. *Dig Dis Sci* 2016;61:881–9.
- Renz BW, Thasler WE, Preissler G, et al. Clinical outcome of IBDassociated versus sporadic colorectal cancer: a matched-pair analysis. J Gastrointest Surg 2013;17:981–90.
- 11. Baker AM, Cross W, Curtius K, *et al.* Evolutionary history of human colitis-associated colorectal cancer. *Gut* 2019;68:985–95.
- Pereira C, Grácio D, Teixeira JP, Magro F. Oxidative stress and DNA damage: implications in inflammatory bowel disease. *Inflamm Bowel Dis* 2015;21:2403–17.
- Pereira C, Coelho R, Grácio D, et al. DNA damage and oxidative DNA damage in inflammatory bowel disease. J Crohns Colitis 2016;10:1316–23.
- Brentnall TA, Crispin DA, Rabinovitch PS, *et al.* Mutations in the p53 gene: an early marker of neoplastic progression in ulcerative colitis. *Gastroenterology* 1994;107:369–78.
- Robles AI, Traverso G, Zhang M, et al. Whole-exome sequencing analyses of inflammatory bowel disease-associated colorectal cancers. *Gastroenterology* 2016;150:931–43.
- 16. Fleisher AS, Esteller M, Harpaz N, et al. Microsatellite instability in inflammatory bowel disease-associated neoplastic lesions is associated with hypermethylation and diminished expression of the DNA mismatch repair gene, hMLH1. Cancer Res 2000;60:4864–8.
- Claessen MM, Schipper ME, Oldenburg B, Siersema PD, Offerhaus GJ, Vleggaar FP. WNT-pathway activation in IBD-associated colorectal carcinogenesis: potential biomarkers for colonic surveillance. *Cell Oncol* 2010;**32**:303–10.
- Chakrabarty S, Varghese VK, Sahu P, et al. Targeted sequencing-based analyses of candidate gene variants in ulcerative colitis-associated colorectal neoplasia. Br J Cancer 2017;117:136–43.
- Ma B, Hottiger MO. Crosstalk between Wnt/β-catenin and NF-κB signaling pathway during inflammation. Front Immunol 2016;7:378.
- Choi CR, Bakir IA, Hart AL, Graham TA. Clonal evolution of colorectal cancer in IBD. Nat Rev Gastroenterol Hepatol 2017;14:218–29.
- Wanders LK, Cordes M, Voorham Q, et al. IBD-associated dysplastic lesions show more chromosomal instability than sporadic adenomas. *Inflamm Bowel Dis* 2020;26:167–80.
- O'Sullivan JN, Bronner MP, Brentnall TA, et al. Chromosomal instability in ulcerative colitis is related to telomere shortening. Nat Genet 2002;32:280–4.
- Risques RA, Lai LA, Himmetoglu C, et al. Ulcerative colitis-associated colorectal cancer arises in a field of short telomeres, senescence, and inflammation. *Cancer Res* 2011;71:1669–79.
- Din S, Wong K, Mueller MF, et al. Mutational analysis identifies therapeutic biomarkers in inflammatory bowel disease-associated colorectal cancers. *Clin Cancer Res* 2018;24:5133–42.
- Popat S, Hubner R, Houlston RS. Systematic review of microsatellite instability and colorectal cancer prognosis. J Clin Oncol 2005;23:609–18.

- 26. Yao D, Dong M, Dai C, Wu S. Inflammation and inflammatory cytokine contribute to the initiation and development of ulcerative colitis and its associated cancer. *Inflamm Bowel Dis* 2019;25:1595–602.
- Luo C, Zhang H. The role of proinflammatory pathways in the pathogenesis of colitis-associated colorectal cancer. *Mediators Inflamm* 2017;2017:5126048.
- Waldner MJ, Neurath MF. Mechanisms of immune signaling in colitisassociated cancer. Cell Mol Gastroenterol Hepatol 2015;1:6–16.
- Greten FR, Eckmann L, Greten TF, et al. IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. Cell 2004;118:285–96.
- Ramakrishnan SK, Zhang H, Ma X, *et al.* Intestinal non-canonical NFκB signaling shapes the local and systemic immune response. *Nat Commun* 2019;10:660.
- Cooks T, Pateras IS, Tarcic O, *et al.* Mutant p53 prolongs NF-κB activation and promotes chronic inflammation and inflammation-associated colorectal cancer. *Cancer Cell* 2013;23:634–46.
- Schwitalla S, Fingerle AA, Cammareri P, et al. Intestinal tumorigenesis initiated by dedifferentiation and acquisition of stem-cell-like properties. Cell 2013;152:25–38.
- Bradford EM, Ryu SH, Singh AP, et al. Epithelial TNF receptor signaling promotes mucosal repair in inflammatory bowel disease. J Immunol 2017;199:1886–97.
- Goretsky T, Dirisina R, Sinh P, et al. p53 mediates TNF-induced epithelial cell apoptosis in IBD. Am J Pathol 2012;181:1306–15.
- Lee H, Herrmann A, Deng JH, et al. Persistently activated Stat3 maintains constitutive NF-kappaB activity in tumors. Cancer Cell 2009;15:283–93.
- 36. Grivennikov S, Karin E, Terzic J, et al. IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. Cancer Cell 2009;15:103–13. Erratum in: Cancer Cell 2009;15:241.
- Bollrath J, Phesse TJ, von Burstin VA, et al. gp130-mediated Stat3 activation in enterocytes regulates cell survival and cell-cycle progression during colitis-associated tumorigenesis. Cancer Cell 2009;15:91–102.
- Munakata K, Koi M, Kitajima T, et al. Inflammation-associated microsatellite alterations caused by MSH3 dysfunction are prevalent in ulcerative colitis and increase with neoplastic advancement. Clin Transl Gastroenterol 2019;10:e00105.
- Rose-John S. IL-6 trans-signaling via the soluble IL-6 receptor: importance for the pro-inflammatory activities of IL-6. Int J Biol Sci 2012;8:1237–47.
- 40. Matsumoto S, Hara T, Mitsuyama K, et al. Essential roles of IL-6 transsignaling in colonic epithelial cells, induced by the IL-6/soluble-IL-6 receptor derived from lamina propria macrophages, on the development of colitis-associated premalignant cancer in a murine model. J Immunol 2010;184:1543–51.
- Putoczki TL, Thiem S, Loving A, et al. Interleukin-11 is the dominant IL-6 family cytokine during gastrointestinal tumorigenesis and can be targeted therapeutically. Cancer Cell 2013;24:257–71.
- 42. Xu ZS, Zhang HX, Li WW, et al. FAM64A positively regulates STAT3 activity to promote Th17 differentiation and colitis-associated carcinogenesis. Proc Natl Acad Sci U S A 2019;116:10447–52.
- 43. Jiang W, Su J, Zhang X, et al. Elevated levels of Th17 cells and Th17related cytokines are associated with disease activity in patients with inflammatory bowel disease. Inflamm Res 2014;63:943–50.
- 44. Yang XO, Chang SH, Park H, et al. Regulation of inflammatory responses by IL-17F. J Exp Med 2008;205:1063–75.
- Hyun YS, Han DS, Lee AR, Eun CS, Youn J, Kim HY. Role of IL-17A in the development of colitis-associated cancer. *Carcinogenesis* 2012;33:931–6.
- 46. Zhou L, Ivanov II, Spolski R, et al. IL-6 programs T(H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. Nat Immunol 2007;8:967–74.
- Gálvez J. Role of Th17 cells in the pathogenesis of human IBD. ISRN Inflamm 2014;2014:928461.
- Gaudino SJ, Kumar P. Cross-talk between antigen presenting cells and T cells impacts intestinal homeostasis, bacterial infections, and tumorigenesis. *Front Immunol* 2019;10:360.
- Punkenburg E, Vogler T, Büttner M, et al. Batf-dependent Th17 cells critically regulate IL-23 driven colitis-associated colon cancer. Gut 2016;65:1139–50.

- Oppmann B, Lesley R, Blom B, *et al.* Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. *Immunity* 2000;13:715–25.
- Wolf SF, Temple PA, Kobayashi M, et al. Cloning of cDNA for natural killer cell stimulatory factor, a heterodimeric cytokine with multiple biologic effects on T and natural killer cells. J Immunol 1991;146:3074–81.
- Richter C, Herrero San Juan M, Weigmann B, et al. Defective IL-23/ IL-17 axis protects p47phox^{-/-} mice from colon cancer. Front Immunol 2017;8:44.
- 53. Wang C, Gong G, Sheh A, et al. Interleukin-22 drives nitric oxidedependent DNA damage and dysplasia in a murine model of colitisassociated cancer. Mucosal Immunol 2017;10:1504–17.
- Pelczar P, Witkowski M, Perez LG, et al. A pathogenic role for T cellderived IL-22BP in inflammatory bowel disease. Science 2016;354:358–62.
- 55. Huber S, Gagliani N, Zenewicz LA, *et al*. IL-22BP is regulated by the inflammasome and modulates tumorigenesis in the intestine. *Nature* 2012;**491**:259–63.
- Fina D, Sarra M, Fantini MC, et al. Regulation of gut inflammation and th17 cell response by interleukin-21. Gastroenterology 2008;134:1038–48.
- 57. Wang Y, Jiang X, Zhu J, et al. IL-21/IL-21R signaling suppresses intestinal inflammation induced by DSS through regulation of Th responses in lamina propria in mice. *Sci Rep* 2016;6:31881.
- Stolfi C, Rizzo A, Franzè E, et al. Involvement of interleukin-21 in the regulation of colitis-associated colon cancer. J Exp Med 2011;208:2279–90.
- Soh JS, Jo SI, Lee H, et al. Immunoprofiling of colitis-associated and sporadic colorectal cancer and its clinical significance. Sci Rep 2019;9:6833.
- 60. Khan MW, Keshavarzian A, Gounaris E, et al. PI3K/AKT signaling is essential for communication between tissue-infiltrating mast cells, macrophages, and epithelial cells in colitis-induced cancer. Clin Cancer Res 2013;19:2342–54.
- Pacheco-Fernández T, Juárez-Avelar I, Illescas O, et al. Macrophage migration inhibitory factor promotes the interaction between the tumor, macrophages, and T cells to regulate the progression of chemically induced colitis-associated colorectal cancer. Mediators Inflamm 2019;2019:2056085.
- Zhu Y, Zhou J, Feng Y, *et al.* Control of intestinal inflammation, colitisassociated tumorigenesis, and macrophage polarization by fibrinogen-like protein 2. *Front Immunol* 2018;9:87.
- Moreira TG, Horta LS, Gomes-Santos AC, *et al.* CLA-supplemented diet accelerates experimental colorectal cancer by inducing TGF-β-producing macrophages and T cells. *Mucosal Immunol* 2019;12:188–99.
- 64. Ueno A, Jijon H, Chan R, et al. Increased prevalence of circulating novel IL-17 secreting Foxp3 expressing CD4⁺ T cells and defective suppressive function of circulating Foxp3⁺ regulatory cells support plasticity between Th17 and regulatory T cells in inflammatory bowel disease patients. *Inflamm Bowel Dis* 2013;19:2522–34.
- 65. Yang BH, Hagemann S, Mamareli P, et al. Foxp3⁺ T cells expressing RORγt represent a stable regulatory T-cell effector lineage with enhanced suppressive capacity during intestinal inflammation. *Mucosal Immunol* 2016;9:444–57.
- 66. Rizzo A, Di Giovangiulio M, Stolfi C, et al. RORγt-expressing tregs drive the growth of colitis-associated colorectal cancer by controlling IL6 in dendritic cells. *Cancer Immunol Res* 2018;6:1082–92.
- Richard ML, Liguori G, Lamas B, et al. Mucosa-associated microbiota dysbiosis in colitis associated cancer. Gut Microbes 2018;9:131–42.
- Chan JL, Wu S, Geis AL, et al. Non-toxigenic Bacteroides fragilis (NTBF) administration reduces bacteria-driven chronic colitis and tumor development independent of polysaccharide A. Mucosal Immunol 2019;12:164–77.
- 69. Lee YK, Mehrabian P, Boyajian S, *et al*. The protective role of *Bacteroides fragilis* in a murine model of colitis-associated colorectal cancer. *mSphere* 2018;3:e00587-18.
- Zhang Y, Weng Y, Gan H, Zhao X, Zhi F. Streptococcus gallolyticus conspires myeloid cells to promote tumorigenesis of inflammatory bowel disease. Biochem Biophys Res Commun 2018;506:907–11.
- Drewes JL, Corona A, Sanchez U, et al. Transmission and clearance of potential procarcinogenic bacteria during fecal microbiota transplantation for recurrent Clostridioides difficile. JCI Insight 2019;4:e130848.

- Yoshimura T, McLean MH, Dzutsev AK, et al. The antimicrobial peptide CRAMP is essential for colon homeostasis by maintaining microbiota balance. J Immunol 2018;200:2174–85.
- 73. Zhong X, Chen B, Liu M, Yang Z. The role of adaptor protein CARD9 in colitis-associated cancer. *Mol Ther Oncolytics* 2019;15:1–6.
- Stellingwerf ME, Sahami S, Winter DC, et al. Prospective cohort study of appendicectomy for treatment of therapy-refractory ulcerative colitis. Br J Surg 2019;106:1697–704.
- 75. Stellingwerf ME, de Koning MA, Pinkney T, Bemelman WA, D'Haens GR, Buskens CJ. The risk of colectomy and colorectal cancer after appendectomy in patients with ulcerative colitis: a systematic review and metaanalysis. J Crohns Colitis 2019;13:309–18.
- 76. Gardenbroek TJ, Pinkney TD, Sahami S, et al. The ACCURE-trial: the effect of appendectomy on the clinical course of ulcerative colitis, a randomised international multicenter trial (NTR2883) and the ACCURE-UK trial: a randomised external pilot trial (ISRCTN56523019). BMC Surg 2015;15:30.
- 77. de Boer NKH, Peyrin-Biroulet L, Jharap B, et al. Thiopurines in inflammatory bowel disease: new findings and perspectives. J Crohns Colitis 2018;12:610–20.
- Qiu X, Ma J, Wang K, Zhang H. Chemopreventive effects of 5-aminosalicylic acid on inflammatory bowel disease-associated colorectal cancer and dysplasia: a systematic review with meta-analysis. Oncotarget 2017;8:1031–45.
- 79. Axelrad J, Bernheim O, Colombel JF, et al.; New York Crohn's and Colitis Organization. Risk of new or recurrent cancer in patients with inflammatory bowel disease and previous cancer exposed to immunosuppressive and anti-tumor necrosis factor agents. Clin Gastroenterol Hepatol 2016;14:58–64.
- Reddy M, Davis C, Wong J, Marsters P, Pendley C, Prabhakar U. Modulation of CLA, IL-12R, CD40L, and IL-2Ralpha expression and inhibition of IL-12- and IL-23-induced cytokine secretion by CNTO 1275. *Cell Immunol* 2007;247:1–11.
- Nastala CL, Edington HD, McKinney TG, et al. Recombinant IL-12 administration induces tumor regression in association with IFN-gamma production. J Immunol 1994;153:1697–706.
- Teng MW, Vesely MD, Duret H, et al. Opposing roles for IL-23 and IL-12 in maintaining occult cancer in an equilibrium state. Cancer Res 2012;72:3987–96.
- Sands BE, Sandborn WJ, Panaccione R, et al.; UNIFI Study Group. Ustekinumab as induction and maintenance therapy for ulcerative colitis. N Engl J Med 2019;381:1201–14.
- Kashani A, Schwartz DA. The expanding role of anti-IL-12 and/or anti-IL-23 antibodies in the treatment of inflammatory bowel disease. *Gastroenterol Hepatol* 2019;15:255–65.
- Lord JD, Long SA, Shows DM, et al. Circulating integrin alpha4/beta7⁺ lymphocytes targeted by vedolizumab have a pro-inflammatory phenotype. Clin Immunol 2018;193:24–32.
- 86. Feagan BG, Greenberg GR, Wild G, et al. Treatment of active Crohn's disease with MLN0002, a humanized antibody to the α4β7 integrin. Clin Gastroenterol Hepatol 2008;6:1370–7.
- Meserve J, Aniwan S, Koliani-Pace JL, *et al.* Retrospective analysis of safety of vedolizumab in patients with inflammatory bowel diseases. *Clin Gastroenterol Hepatol* 2019;17:1533–40.e2.
- Colombel JF, Sands BE, Rutgeerts P, et al. The safety of vedolizumab for ulcerative colitis and Crohn's disease. Gut 2017;66:839–51.
- Vedamurthy A, Gangasani N, Ananthakrishnan AN. Vedolizumab or TNF-antagonist use and risk of new or recurrent cancer in patients with inflammatory bowel disease with prior malignancy: a retrospective cohort study. *Clin Gastroenterol Hepatol* 2020. In Press. doi: 10.1016/j. cgh.2020.10.007.
- Caron B, Peyrin-Biroulet L, Pariente B, *et al.* Vedolizumab therapy is ineffective for primary sclerosing cholangitis in patients with inflammatory bowel disease: a GETAID multicentre cohort study. *J Crohns Colitis* 2019;13:1239–47.
- Duijvestein M, D'Haens GR. Rational and clinical development of the anti-MAdCAM monoclonal antibody for the treatment of IBD. *Expert Opin Biol Ther* 2019;19:361–6.

- Svensson H, Olofsson V, Lundin S, et al. Accumulation of CCR4⁺CTLA-4 FOXP3⁺CD25[hi) regulatory T cells in colon adenocarcinomas correlate to reduced activation of conventional T cells. PLoS One 2012;7:e30695.
- Sandborn WJ, Panés J, D'Haens GR, et al. Safety of tofacitinib for treatment of ulcerative colitis, based on 4.4 years of data from global clinical trials. Clin Gastroenterol Hepatol 2019;17:1541–50.
- Lichtenstein GR, Rogler G, Ciorba MA, et al. Tofacitinib, an oral janus kinase inhibitor: analysis of malignancy (excluding nonmelanoma skin cancer) events across the ulcerative colitis clinical program. *Inflamm Bowel Dis* 2021;27:816–25.
- Lee H, Deng J, Kujawski M, et al. STAT3-induced S1PR1 expression is crucial for persistent STAT3 activation in tumors. Nat Med 2010;16:1421–8.
- Matloubian M, Lo CG, Cinamon G, *et al.* Lymphocyte egress from thymus and peripheral lymphoid organs is dependent on S1P receptor 1. *Nature* 2004;**427**:355–60.
- Yuza K, Nagahashi M, Shimada Y, et al. Upregulation of phosphorylated sphingosine kinase 1 expression in colitis-associated cancer. J Surg Res 2018;231:323–30.
- Schwiebs A, Herrero San Juan M, Schmidt KG, *et al.* Cancer-induced inflammation and inflammation-induced cancer in colon: a role for S1P lyase. Oncogene 2019;38:4788–803.
- Zhen Y, Luo C, Zhang H. Early detection of ulcerative colitis-associated colorectal cancer. *Gastroenterol Rep* (Oxf) 2018;6:83–92.
- 100. Sung H, Ferlay J, Siegel RL, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2021;71:209–49.
- Herrinton LJ, Liu L, Levin TR, Allison JE, Lewis JD, Velayos F. Incidence and mortality of colorectal adenocarcinoma in persons with inflammatory bowel disease from 1998 to 2010. *Gastroenterology* 2012;143:382–9.
- Siegel RL, Miller KD, Goding Sauer A, et al. Colorectal cancer statistics, 2020. CA Cancer J Clin 2020;70:145–64.
- 103. Johnson CM, Wei C, Ensor JE, *et al*. Meta-analyses of colorectal cancer risk factors. *Cancer Causes Control* 2013;**24**:1207–22.
- Butt J, Epplein M. *Helicobacter pylori* and colorectal cancer—A bacterium going abroad? *PLoS Pathog* 2019;15:e1007861.
- 105. McNabb S, Harrison TA, Albanes D, *et al.* Meta-analysis of 16 studies of the association of alcohol with colorectal cancer. *Int J Cancer* 2020;**146**:861–73.

- 106. Zauber AG, Winawer SJ, O'Brien MJ, et al. Colonoscopic polypectomy and long-term prevention of colorectal-cancer deaths. N Engl J Med 2012;366:687–96.
- 107. Sørensen JØ, Nielsen OH, Andersson M, et al. Inflammatory bowel disease with primary sclerosing cholangitis: a Danish population-based cohort study 1977-2011. Liver Int 2018;38:532–41.
- Boonstra K, Weersma RK, van Erpecum KJ, *et al.*; EpiPSCPBC Study Group. Population-based epidemiology, malignancy risk, and outcome of primary sclerosing cholangitis. *Hepatology* 2013;58:2045–55.
- 109. Nuako KW, Ahlquist DA, Mahoney DW, Schaid DJ, Siems DM, Lindor NM. Familial predisposition for colorectal cancer in chronic ulcerative colitis: a case-control study. *Gastroenterology* 1998;115:1079–83.
- Muto T, Bussey HJ, Morson BC. The evolution of cancer of the colon and rectum. *Cancer* 1975;36:2251–70.
- 111. Yaeger R, Shah MA, Miller VA, et al. Genomic alterations observed in colitis-associated cancers are distinct from those found in sporadic colorectal cancers and vary by type of inflammatory bowel disease. *Gastroenterology* 2016;151:278–87.e6.
- 112. Pino MS, Chung DC. The chromosomal instability pathway in colon cancer. *Gastroenterology* 2010;**13**8:2059–72.
- Lasry A, Zinger A, Ben-Neriah Y. Inflammatory networks underlying colorectal cancer. Nat Immunol 2016;17:230–40.
- 114. Pandey A, Shen C, Man SM. Inflammasomes in colitis and colorectal cancer: mechanism of action and therapies. *Yale J Biol Med* 2019;92:481–98.
- 115. Araki A, Jin L, Nara H, et al. IL-21 enhances the development of colitisassociated colon cancer: possible involvement of activation-induced cytidine deaminase expression. J Immunol 2019;202:3326–33.
- 116. Sideris GA, Sardeli A, Sachtouris G, Segkou I, Kondi-Pafiti A, Papaconstantinou I. Sporadic versus inflammatory bowel disease-related colorectal adenocarcinoma: lessons learned from a single institution experience. J BUON 2015;20:1178–85.
- 117. Sebastian S, Hernández V, Myrelid P, et al. Colorectal cancer in inflammatory bowel disease: results of the 3rd ECCO pathogenesis scientific workshop (I). J Crohns Colitis 2014;8:5–18.
- 118. Jewel Samadder N, Valentine JF, Guthery S, *et al.* Colorectal cancer in inflammatory bowel diseases: a population-based study in Utah. *Dig Dis Sci* 2017;**62**:2126–32.