



Complete Genome Sequences of Mycobacteriophages OKaNui and DroogsArmy

[®]Kayla M. Fast,^a Kadaisha G. Johnson,^a Kaitlyn N. Mayfield,^a Leah A. Stephens,^a T. Hunter Reid,^a Emma D. Ryan,^a Tracy W. Keener,^a [®]Michael W. Sandel^a

^aDepartment of Biological and Environmental Sciences, University of West Alabama, Livingston, Alabama, USA

ABSTRACT Mycobacteriophages OKaNui and DroogsArmy were isolated from soil using the bacterial host *Mycobacterium smegmatis* mc²155, which belongs to the phylum *Actinobacteria*. OKaNui was discovered in east Mississippi and DroogsArmy in west Alabama in the United States. The genomes of OKaNui and DroogsArmy were 51,424 bp and 53,254 bp long, respectively.

A mong biological agents, bacteriophages (phages) are the most populous and Aubiquitous in the environment (1). The phages named OKaNui and DroogsArmy were isolated from the bacterial host *Mycobacterium smegmatis* mc²155. Most *Mycobacteria* species are saprophytic and reside in the soil or water or on plants (2). Although *M. smegmatis* is a nonpathogenic bacterium, an understanding of phage infection in this strain may contribute to advances in phage therapy for other *Mycobacteria* species. *M. smegmatis* may be used as a delivery system for phages intended for infection of *M. avium* and *M. tuberculosis* (3). Phages isolated from *M. smegmatis* have be used in the experimental treatment of closely related hosts, including *M. ulcerans* (4). OKaNui was discovered in Meridian, Mississippi, in moist soil in a shaded area, and DroogsArmy was found in Lisman, Alabama, in a dark-colored, moist soil (Table 1).

The soil samples were inoculated with the host bacterium and incubated at 37°C with shaking in 7H9 liquid medium (5). A filtrate was then plated on a lawn of M. smegmatis mc²155 along with a negative control (5). The formation of plaques following incubation at 37°C were indicative of phage presence. Isolates were purified using serial dilutions and a filtered high-titer lysate (HTL) collected from webbed plates (5). Phage genomic DNA was extracted using the Wizard DNA cleanup system with modified protocols (Promega, Madison, WI) (5). DNA libraries were built and pooled for sequencing using the NEBNext Ultra II FS kit with dual-indexed barcoding (New England BioLabs, Ipswich, MA). Sequencing was performed using the Illumina MiSeq platform at the Pittsburgh Bacteriophage Institute. The genome lengths, G+C content, and coverage depths are listed in Table 1. OKaNui yielded ~316,000 single-end 150-base reads, and DroogsArmy yielded \sim 410,000 reads. Both genomes displayed defined ends with 10-bp overhangs (CGGCCGGTAA). Assembly was performed using Newbler 2.9 with default settings (6). A single contig for each genome was produced and used to determine the genome ends; the beginning of each genome was chosen based on similar genomes. These were then checked for completeness and accuracy using Consed 2.0 (7).

Genome annotation was performed using the Phage Evidence Collection and Annotation Network (PECAAN; https://discover.kbrinsgd.org/) and DNA Master 5.23.3 (cobamide2.bio.pitt.edu/computer.htm). We used the following programs to determine gene presence, functions, and start sites: NCBI BLAST, Phamerator (https://phamerator .org/), PhagesDB BLAST, HHPred 3.0, Starterator (https://github.com/SEA-PHAGES/ Citation Fast KM, Johnson KG, Mayfield KN, Stephens LA, Reid TH, Ryan ED, Keener TW, Sandel MW. 2020. Complete genome sequences of mycobacteriophages OKaNui and DroogsArmy. Microbiol Resour Announc 9:e00791-20. https://doi.org/10.1128/MRA .00791-20.

Editor Simon Roux, DOE Joint Genome Institute

Copyright © 2020 Fast et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Kayla M. Fast, kfast@uwa.edu.

Received 8 July 2020 Accepted 21 July 2020 Published 13 August 2020

						Location				Closest relative	
Bacteriophage isolate	Accession no.	Genome size (bp)	No. of protein CDSs ^a	No. of tRNAs	G+C content (%)	County, state	Coordinates	Read coverage (×)	Phage life cycle	Isolate (GenBank accession no.)	Nucleotide identity (coverages [%])
OKaNui	MT490373.1	51,424	87	0	63.9	Lauderdale County, MS	32.59, -88.19	874	Lysogenic	Kingmustik0402 (MK359301.1)	99.2, 99.0
DroogsArmy	MT553337.1	53,254	83	1	63.0	Choctaw County, AL	32.28, -88.28	1,097	Lysogenic	Timshel (NC_041983.1)	98.9, 95.0

TABLE 1 Characteristics and accession numbers of genomes

^a CDSs, coding DNA sequences.

starterator), and GeneMark 3.25 (8–11). Detection and trimming of tRNAs were performed in ARAGORN 1.2.38 and tRNAscan-SE 2.0 (12, 13). The complete numbers of protein coding genes and tRNAs are reported in Table 1.

When plated with the host, OKaNui produced relatively large plaques with a halo, and DroogsArmy produced small plaques. Both OKaNui and DroogsArmy were identified as lysogenic phages because they produced turbid plaques, and the genomes contained genes for lysogeny (14). Transmission electron microscopy and genome analysis indicated that the phages share the *Siphoviridae* morphotype (5). Based on nucleotide similarity and synteny across the genome, OKaNui has been placed in the A4 subcluster of mycobacteriophages and DroogsArmy in A7 (15).

Data availability. The complete genome sequences of OKaNui and DroogsArmy are available from GenBank under the accession numbers MT490373.1 and MT553337.1, respectively. The raw Illumina reads for OKaNui and DroogsArmy are available on NCBI's Sequence Read Archive under accession numbers SRX8622883 and SRX8622882, respectively.

ACKNOWLEDGMENTS

This material is based upon work supported by the National Science Foundation under grant number 1911457. Any opinions, findings, and conclusions or recommendations expressed in this material are those of the authors and do not necessarily reflect the views of the National Science Foundation. This project was supported by the Howard Hughes Medical Institute Science Education Alliance–Phage Hunters Advancing Genomics and Evolutionary Science program (www.seaphages.org) and the University of West Alabama 2018–2020 Phage Hunters.

We thank Daniel A. Russell and Rebecca A. Garlena for conducting genome sequencing, quality control, and assembly at the University of Pittsburgh. We thank Kimberly Lackey with the Optical Analysis Facility at the University of Alabama for assisting with transmission electron microscopy.

REFERENCES

- 1. Hatfull GF. 2008. Bacteriophage genomics. Curr Opin Microbiol 11: 447–453. https://doi.org/10.1016/j.mib.2008.09.004.
- 2. Caballero B, Trugo LC, Finglas PM (ed). 2003. Encyclopedia of food sciences and nutrition. Academic Press, San Diego, CA.
- Broxmeyer L, Sosnowska D, Miltner E, Chacón O, Wagner D, McGarvey J, Barletta RG, Bermudez LE. 2002. Killing of Mycobacterium avium and Mycobacterium tuberculosis by a mycobacteriophage delivered by a nonvirulent mycobacterium: a model for phage therapy of intracellular bacterial pathogens. J Infect Dis 186:1155–1160. https://doi.org/10.1086/ 343812.
- Trigo G, Martins TG, Fraga AG, Longatto-Filho A, Castro AG, Azeredo J, Pedrosa J. 2013. Phage therapy is effective against infection by Mycobacterium ulcerans in a murine footpad model. PLoS Negl Trop Dis 7:e2183. https://doi.org/10.1371/journal.pntd.0002183.
- Poxleitner M, Pope W, Jacobs-Sera D, Sivanathan V, Hatfull G. 2018. Phage discovery guide. Howard Hughes Medical Institute, Chevy Chase, MD.
- Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, Berka J, Braverman MS, Chen Y-J, Chen Z, Dewell SB, Du L, Fierro JM,

Gomes XV, Godwin BC, He W, Helgesen S, Ho CH, Irzyk GP, Jando SC, Alenquer MLI, Jarvie TP, Jirage KB, Kim J-B, Knight JR, Lanza JR, Leamon JH, Lefkowitz SM, Lei M, Li J, Lohman KL, Lu H, Makhijani VB, McDade KE, McKenna MP, Myers EW, Nickerson E, Nobile JR, Plant R, Puc BP, Ronan MT, Roth GT, Sarkis GJ, Simons JF, Simpson JW, Srinivasan M, Tartaro KR, Tomasz A, Vogt KA, Volkmer GA, Wang SH, Wang Y, Weiner MP, Yu P, Begley RF, Rothberg JM. 2005. Genome sequencing in microfabricated high-density picolitre reactors. Nature 437:376–380. https://doi.org/10 .1038/nature03959.

- Gordon D, Abajian C, Green P. 1998. Consed: a graphical tool for sequence finishing. Genome Res 8:195–202. https://doi.org/10.1101/gr .8.3.195.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/ S0022-2836(05)80360-2.
- Russell DA, Hatfull GF. 2017. PhagesDB: the actinobacteriophage database. Bioinformatics 33:784–786. https://doi.org/10.1093/bioinformatics/btw711.
- 10. Söding J, Biegert A, Lupas AN. 2005. The HHpred interactive server for

protein homology detection and structure prediction. Nucleic Acids Res 33:W244-W248. https://doi.org/10.1093/nar/gki408.

- 11. Borodovsky M, McIninch JD. 1993. Recognition of genes in DNA sequence with ambiguities. Biosystems 30:161–171. https://doi.org/10 .1016/0303-2647(93)90068-N.
- 12. Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. https://doi.org/10.1093/nar/gkh152.
- Lowe TM, Chan PP. 2016. tRNAscan-SE On-line: integrating search and context for analysis of transfer RNA genes. Nucleic Acids Res 44: W54–W57. https://doi.org/10.1093/nar/gkw413.
- Dedrick RM, Marinelli LJ, Newton GL, Pogliano K, Pogliano J, Hatfull GF. 2013. Functional requirements for bacteriophage growth: gene essentiality and expression in mycobacteriophage Giles. Mol Microbiol 88: 577–589. https://doi.org/10.1111/mmi.12210.
- 15. Pope WH, Bowman CA, Russell DA, Jacobs-Sera D, Asai DJ, Cresawn SG, Jacobs WR, Jr, Hendrix RW, Lawrence JG, Hatfull GF, Science Education Alliance Phage Hunters Advancing Genomics and Evolutionary Science, Phage Hunters Integrating Research and Education, Mycobacterial Genetics Course. 2015. Whole genome comparison of a large collection of mycobacteriophages reveals a continuum of phage genetic diversity. Elife 4:e06416. https://doi.org/10.7554/eLife.06416.