

# Complete genome sequence of bacteriocin-producing *Ligilactobacillus salivarius* B4311 isolated from fecal samples of broiler chicken with anti-listeria activity

Subin Han<sup>#</sup>, Arxel G. Elnar<sup>#</sup>, Chiwoong Lim and Geun-Bae Kim\*

Department of Animal Science and Technology, Chung-Ang University, Anseong 17546, Korea



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<sup>#</sup>These authors contributed equally to this work.

## \*Corresponding author

Geun-Bae Kim

Department of Animal Science and Technology, Chung-Ang University, Anseong 17546, Korea.

Tel: +82-31-670-3027

E-mail: kimgeun@cau.ac.kr

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## ORCID

Subin Han

<https://orcid.org/0000-0001-6658-2822>

Arxel G. Elnar

<https://orcid.org/0000-0002-2716-4924>

Chiwoong Lim

<https://orcid.org/0000-0002-6272-4464>

Geun-Bae Kim

<https://orcid.org/0000-0001-8531-1104>

## Competing interests

No potential conflict of interest relevant to this article was reported.

## Abstract

*Ligilactobacillus* is a genus of Gram-positive lactobacilli commonly found in the intestinal tracts of vertebrates. It has been granted a Qualified Presumption of Safety (QPS) status from the European Food Safety Authority (EFSA). One specific strain, *Ligilactobacillus salivarius* B4311, was isolated from fecal samples of broiler chickens from a farm associated with Chung-Ang University (Anseong, Korea). This strain was observed to have inhibitory effects against *Listeria monocytogenes*. In this paper, we present the complete genome sequence of *Lig. salivarius* B4311. The whole genome of strain B4311 comprises 2,071,255 bp assembled into 3 contigs representing a chromosome, *repA*-type megaplasmid, and small plasmid. The genome contains 1,963 protein-coding sequences, 22 rRNA genes, and 78 tRNA genes, with a guanine + cytosine (GC) content of 33.1%. The megaplasmid of strain B4311 was found to contain the bacteriocin gene cluster for salivaricin P, a two-peptide bacteriocin belonging to class IIb.

**Keywords:** *Ligilactobacillus salivarius*, Probiotics, Bacteriocin, *Listeria monocytogenes*

Bacteriocin production in lactic acid bacteria (LAB) has been consistently gaining attention owing to its potential as a viable alternative to antibiotics. Bacteriocins are ribosomally-synthesized peptides secreted by the producing strain. These peptides can have either a narrow or broad spectrum of activity, which indirectly determines the niche of the producing strain. The production of bacteriocins is generally viewed as a positive trait as it enables the producing strain to hinder potential competitors in the immediate environment as well as inhibit potentially harmful microorganisms [1]. The proteinaceous nature of bacteriocins renders them suitable for human use as they can be inactivated by digestive proteases. With the rapid development of antimicrobial drug resistance in microorganisms [2], research efforts focused on developing alternative solutions must be prioritized.

Commonly associated with vertebrate hosts, *Ligilactobacillus* is a genus of lactic acid bacteria composed of members that are homofermentative, non-motile, and urease-positive. Their ability to

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**Availability of data and material**

Upon reasonable request, the datasets of this study can be available from the corresponding author.

**Authors' contributions**

Conceptualization: Kim GB.  
Data curation: Han S, Elnar AG.  
Formal analysis: Elnar AG, Kim GB.  
Methodology: Elnar AG, Kim GB.  
Software: Elnar AG, Lim C, Kim GB.  
Validation: Kim GB.  
Investigation: Elnar AG.  
Writing - original draft: Han S, Elnar AG.  
Writing - review & editing: Han S, Elnar AG, Lim C, Kim GB.

**Ethics approval and consent to participate**

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survive in gastric acid conditions and their Qualified Presumption of Safety (QPS) status from the European Food Safety Authority (EFSA) [3] make them popular choices for probiotics. Furthermore, the production of various antimicrobial salivaricins among strains of *Lig. salivarius* is well accounted for the development of probiotic strains [4]. In the present study, we report the genome analysis of a bacteriocin-producing *Ligilactobacillus salivarius* (formerly *Lactobacillus salivarius*) strain B4311, which was isolated from fecal samples collected from broiler chickens.

Strain B4311 was routinely cultured in de Mann, Rogosa, Sharpe ([MRS] BD Difco, Franklin Lakes, NJ, USA) broth supplemented with 0.05% L-cysteine, and incubated aerobically at 37°C for 24 h. Genomic DNA was extracted using the MagAttract HWM DNA Kit (Qiagen, Hilden, Germany) and quantified using Qubit ds DNA HS assay kit (Invitrogen, Waltham, MA, USA) with the Epoch™ Spectrometer (BioTek, Winooski, VT, USA). The genome was sequenced using the Pacific Biosciences (PacBio, Menlo Park, CA, USA) Sequel II platform. *De novo* assembly of the sequence reads was performed using the PacBio SMAR Analysis program (ver. 2.3.0). Functional annotation of the genome was performed using PRODIGAL ver. 2.6.2 [5] software and compared against protein databases (SwissProt, KEGG, SEED, EggNOG). Rapid annotation was employed using Subsystem Technology (RAST) with default parameters (<https://rast.nmpdr.org/>). Transfer RNAs (tRNA) and non-coding ribosomal RNAs (rRNA) were identified using tRNAscan-SE ver. 1.3.1 [6] and INFERNAL ver. 1.1.3 [7], respectively.

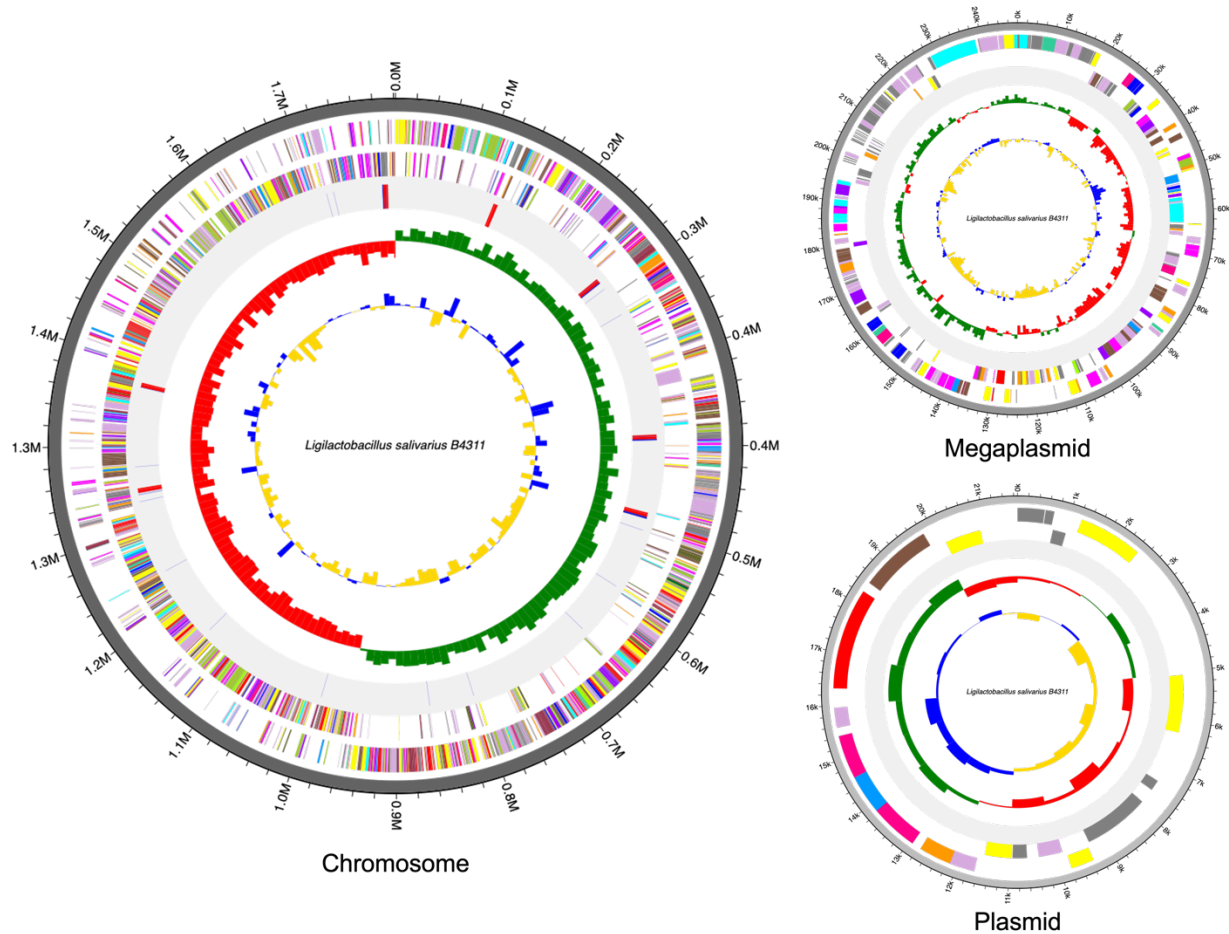
The complete genome of *Lig. salivarius* B4311 is 2,071,255 base pair (bp) which is assembled into three contigs: a single chromosome (1,801,655 bp), one megaplasmid (247,930 bp), and a small plasmid (21,670 bp) with a guanine + cytosine (GC) content of 33.1%. In addition, the genome contains 1,963 protein-coding sequences (CDS), 22 rRNA genes, and 78 tRNA genes. The genome features and circular maps of strain B4311 are presented in Table 1 and Fig. 1, respectively. Antimicrobial resistance genes, specifically for tetracycline and glycopeptides, were also detected via Resistance Gene Identifier ([RGI] <https://card.mcmaster.ca/home>). Among the 1,963 CDS, 1,241 were predicted with biological functions associated with cell cycle (n = 23), cell wall and motility (n = 116), cellular response (n = 69), DNA processing (n = 154), RNA processing (n = 119), protein processing (n = 202), defense mechanism (n = 31), energy production (n = 63), and transport and metabolism (n = 464). Additionally, 61 putative genes were detected with putative functions including stress response, DNA and RNA processing, antibiotic resistance, periplasm signaling, acetylation, amino acid transport, and production of enzymes including various hydrolases, methyltransferases, and transport proteins.

*In silico* analysis of the B4311 genome using BAGEL4 online program (<http://bagel4.molgenrug.nl/>) revealed the presence of a bacteriocin gene cluster for salivaricin P, a family of two-peptide bacteriocins belonging to class IIb. This bacteriocin family was originally discovered

**Table 1. Genome features of *Ligilactobacillus salivarius* B4311**

Attribute	Value			
	Chromosome	Megaplasmid	Plasmid	Total
Size (bp)	1,801,655	247,930	21,670	2,071,255
GC content (%)	33.24	32.25	33.68	33.1
No. of contigs	1	1	1	3
Total genes	1,768	273	22	2,063
Protein-coding gene	1,668	273	22	1,963
tRNA	78	-	-	78
rRNA	22	-	-	22

bp, base pair; GC, guanine + cytosine.



**Fig. 1. Circular map of *Ligilactobacillus salivarius* B4311 genome.** Circles represent the following characteristics from the outermost circle to the center: (1) contig information, (2) coding sequences on forward strand, (3) coding sequences on reverse strand, (4) transfer RNAs (tRNAs) and ribosomal RNAs (rRNAs), (5) GC skew, and (6) GC ratio. G, guanine; C, cytosine; CDS, coding sequences.

from *Lig. salivarius* DPC6005 [6] and is commonly produced among strains of *Lig. salivarius* isolated from animals intestines [8]. The salivaricin P gene cluster is located in the *repA*-type megaplasmid. Although the presence of megaplasmids is considered a typical feature of *Lig. salivarius*, variations exist among megaplasmid-encoded traits, including contingency metabolism genes (i.e., assimilation of sugars) and the presence or absence of bacteriocin genes, which provides a competitive advantage.

The genetic architecture of the bacteriocin gene cluster (Fig. 2) revealed the presence of two open reading frames (ORFs) encoding the salivaricin P chain A and chain B. The two peptide chains share a homologous sequence. Located downstream of the genes for the bacteriocin peptides are two ORFs encoding a histidine kinase and *AbpR*, which function as regulator proteins [9]. These are followed by *AbpIM* which encodes an immunity protein. These five ORFs are flanked by two *comC* genes, which have been reported as competence-stimulating peptide precursors in streptococci [10]. At the 3' end of the gene cluster, two export proteins, *LanT* and *HlyD* were detected, encoding *AbpT* and *AbpD* bacteriocin export accessory proteins, respectively. Several ORFs encoding bacteriocin core peptides (i.e., lactacin F and plantaricins) were also detected. However, the similarity of these genes with the reference was poor, suggesting that the translated peptides might



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