



# **Practical Significance of Biomarkers in Axial Spondyloarthritis: Updates on Diagnosis, Disease Activity, and Prognosis**

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**Abstract:** Axial spondyloarthritis (axSpA) is a chronic inflammatory disease that can lead to ankylosis by secondary ossification of inflammatory lesions, with progressive disability and a significant impact on quality of life. It is also a risk factor for the occurrence of comorbidities, especially cardiovascular diseases (CVDs), mood disorders, osteoporosis, and malignancies. Early diagnosis and treatment are needed to prevent or decrease functional decline and to improve the patient's prognosis. In respect of axSpA, there is an unmet need for biomarkers that can help to diagnose the disease, define disease activity and prognosis, and establish personalized treatment approaches. The aim of this review was to summarize the available information regarding the most promising biomarkers for axSpA. We classified and identified six core categories of biomarkers: (i) systemic markers of inflammation; (ii) molecules involved in bone homeostasis; (iii) HLA-B27 and newer genetic biomarkers; (iv) antibody-based biomarkers; (v) microbiome biomarkers, few of them are used in clinical practice; however, we believe that these studies provide useful data that could aid in better disease management.

**Keywords:** axial spondyloarthritis; diagnosis; disease activity; prognosis; systemic markers of inflammation; molecules involved in bone homeostasis; HLA-B27; newer genetic biomarkers; antibody-based biomarkers; microbiome biomarkers

## 1. Introduction

## 1.1. Axial Spondyloarthritis: Past, Present, and Future

Spondyloarthritis (SpA) comprises a group of chronic immune-mediated inflammatory conditions with overlapping clinical and genetic features but with substantial heterogeneity, as inflammation can affect peripheral and axial joints and also result in extraarticular manifestations (enthesitis, dactylitis, uveitis, inflammatory bowel disease, psoriasis) [1–3].

SpA comprises a spectrum of diseases with significant overlaps between SpA phenotypes, which can be classified according to the Assessment of Spondyloarthritis International Society (ASAS) 2009 classification criteria. These depend on where patients primarily experience symptoms in axial SpA (axSpA) with predominantly axial manifestations, including non-radiographic (nr-axSpA) and radiographic axSpA (r-axSpA) or ankylosing spondylitis (AS) and peripheral SpA (pSpA) with predominant peripheral involvement (psoriatic arthritis, reactive arthritis, enteropathic arthritis, juvenile-onset SpA, and undifferentiated SpA) [4]. While the spectrum of axSpA extends from early or mild disease through severe or late disease with erosive damage in the sacroiliac joints, this arbitrary division of nr-axSpA and r-axSpA is becoming less clinically relevant.



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**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). The symptoms of axSpA create a significant burden for both patients and society. The major disease feature is the progression to ankylosis by secondary ossification of inflammatory lesions, with progressive disability and a significant impact on quality of life.

AxSpA is also a risk factor for the occurrence of comorbidities, especially cardiovascular diseases (CVDs), mood disorders, osteoporosis, and malignancies, which shows that axSpA has a systemic character and influences patient health in a complex manner [2]. All these characteristics affect quality of life and, because all comorbidity types correlate with axSpA, early diagnosis and treatment are needed to prevent or decrease functional decline and improve the patient's prognosis.

## 1.2. AxSpA and Biomarkers

According to the World Health Organization, a biomarker is a chemical, physical, or biological indicator of disease presence, activity, or severity, which can provide vital information. In respect of axSpA, there is an unmet need for biomarkers that can help to diagnose the disease, define disease activity and prognosis, and establish personalized treatment approaches [5].

Despite the progress made in the concept of axSpA, its identification remains challenging, frequently resulting in a diagnostic delay for patients. Ideally, the diagnosis should be made before the development of radiographic sacroiliitis, but currently the most sensitive imaging diagnosis method is magnetic resonance imaging (MRI), which is costly and not widely available [2].

AxSpA disease activity is monitored using the following scores: Bath Ankylosing Spondylitis Disease Activity Index (BASDAI), Bath Ankylosing Spondylitis Functional Index (BASFI), and Ankylosing Spondylitis Disease Activity Score (ASDAS), which may be subjective. Disease activity is also monitored by measuring of one of the most common inflammatory biomarkers, C-reactive protein (CRP), which is insensitive for this disease [6].

Although the range of effective modern drugs for axSpA has expanded in recent decades, therapeutic decisions and the response to specific therapies remain challenges in current practice, and further research is therefore required. Adequate evaluation of disease activity, radiographic progression, response to treatment and identification of negative prognostic factors can improve the therapeutic decision. In addition, data regarding the axSpA prognosis obtained in the early disease stages will improve therapeutic management [2].

The aim of our review was to summarize available information regarding the most promising biomarkers for axSpA. We performed a PubMed, Embase, and Medline search for articles published in the period of 2017-2022, with "axial spondyloarthritis" and "biomarkers" as search terms. We further refer to six core categories of biomarkers: (i) systemic markers of inflammation; (ii) molecules involved in bone homeostasis; (iii) HLA-B27 and newer genetic biomarkers; (iv) antibody-based biomarkers; (v) microbiome biomarkers; and (vi) miscellaneous biomarkers (Table 1, Figure 1). Furthermore, understanding the association between biomarkers and diagnosis, disease activity, structural damage, and treatment response, we followed the same pattern when analyzing available data.

Table 1. Biomarkers significance in axial spondyloarthritis.

Biomarker	Predictability	Study type	Reference
Anti-CD74 antibodies	Positive association with diagnosis and disease activity.	Cross-sectional study	2021, Marwa Mahmoud Abdelaziz [7]
	Positive association with radiographic progression.	Prospective study	2021, Lan Do [8]
	Positive association with diagnosis.	Prospective study	2019, Nelly R. Ziade [9]
	Negative association with diagnosis.	Prospective study	2018, Janneke J. de Winter [10]
	Positive association with diagnosis.	Prospective study	2020, Chao-Jun Hu [11]

Biomarker	Predictability	Study type	Reference
Bone morphogenetic protein 7 (BMP-7), sclerostin, dicckopf-1 (DKK-1)	Positive association with disease association	Prospective multicenter study	2021, Elise Descamps [12]
C-reactive protein (CRP)	Positive association with cardiovascular risk	Retrospective analysis	2022, Luca Navarini [13]
	Positive association with treatment response	Prospective study	2019, Xenofon Baraliakos [14]
	Positive association with treatment response	Case-control study	2020, Xu Yuansheng [15]
	Positive association with treatment response	Cross-sectional study	2018, Björn Sundström [16]
C-C motif chemokine ligand 11 (CCL11)	Positive association with radiographic progression	Cross-sectional study	2018, Dong Hyun Sohn [17]
CD163, CD206	Positive association with disease activity, treatment response	Prospective study	2018, Line Dam Heftdal [18]
CD27-CD38 <sup>low</sup> CD21 <sup>low</sup> B cells	Positive association with disease activity	Cross-sectional study	2021, Rick Wilbrink [19]
circRNA hsa_circ_0079787	Positive association with diagnosis, disease activity	Cross-sectional study	2020, Qing Luo [20]
Circulating microRNAs	Positive association with	Prospective study	2020, Qing Luo [20]
	Positive association with diagnosis and disease activity	Cross-sectional study	2018, Carlos Perez-Sanchez [21]
Complement component C3	Positive association with disease activity and cardiovascular risk	Cross-sectional study	2020, Ivan Arias de la Rosa [22]
CRP to albumin ratio (CAR)	Positive association with disease activity	Retrospective study	2021, Zheng Zhong [23]
	Positive association with disease activity	Cross-sectional study	2021, Melih Pamukcu [24]
Dickkopf homologue-1 (Dkk-1)	Positive association with radiographic progression	Prospective study	2019, Zhonghai Zhao [25]
Bone morphogenetic proteins (BMPs) and Dkk-1	Positive association with disease activity	Prospective study	2018, Hsien-Tzung Liao [26]
Drug-neutralizing Antibodies against TNF-α	Positive association with treatment response	Cross-sectional study	2020, Krasimir Kraev [27]
ERAP1 (rs2287987, rs30187, rs27044), ERAP2 (rs2248374)	Positive association with diagnosis	Case-control study	2019, Andrzej Wiśniewski [28]
Erythrocyte sedimentation	Positive association with	Retrospective study	2018, Kwi Young Kang [29]
ESR + Interleukin (IL)-6	Positive association with treatment response	Prospective study	2019, Yidian Dong [30]
Eukaryotic translation elongation factor 1 epsilon 1 ( <i>EEF1E1</i> )	Positive association with diagnosis	Prospective study	2019, Xutao Fan [31]
Fecal calprotectina and anti-Saccharaomyces cerevisiae antibodies (ASCA)	Positive association with disease activity	Cross-sectional study	2019, Tor Olofsson [32]
Fibrinogen-to-albumin ratio (FAR)	Positive association with disease activity	Retrospective study	2020, Meng Liu [33]

# Table 1. Cont.

Biomarker	Predictability	Study type	Reference
Galectin-3	Positive association with disease activity	Prospective study	2019, Ming-Yu Cao [34]
Galectin-3, soluble suppression-of- tumorogenity-2 (sST2) (cardiac fibrosis), hsCRP	Positive association with cardiovascular risk	Prospective study	2018, Demet Ozkaramanli Gur [35]
H-ficolin, L-ficolin	Positive association with diagnosis	Cross-sectional cohort	2020, Anne Troldborg [36]
Hepatocyte growth factor (s-HGF)	Positive association with radiographic progression Positive association with radiographic progression	Prospective cohort study Prospective study	2021, Anna Deminger [37] 2019, Liset Torres [38]
HLA B-27 antigen	Positive association with	Prospective study	2019, Nelly Ziade [39]
	Positive association with diagnosis	Prospective study	2018, Chong Seng Edwin Lim [40]
	Positive association with diagnosis	Prospective study	2021, Henriëtte M Y de Jong [41]
Interleukin (IL)-31	Positive association with radiographic progression	Longitudinal prospective cohort study	2018, Nicolas Rosine [42]
IL-33	Positive association with disease activity and treatment response	Prospective study	2021, Milena Iwaszko [43]
IL-12B and IL-6R	Positive association with diagnosis and prognosis	Prospective cohort study	2018, Wen-Feng Ruan [44]
IL-22	Positive association with diagnosis	Retrospective study	2022, Michal Sagiv [45]
IL-17 IL-23 and IL-17	Positive association with	Cross-sectional study	2020, Dalia S. Saif [46]
1L-23 and 1L-17	Positive association with diagnosis	Case-control study	2018, Jacob Sode [47]
Leptin	Positive association with radiographic progression	Longitudinal	2017, Ji-Heg Park [48]
	Positive association with radiographic progression	Prospective study	2019, Judith Rademacher [49]
Leptin + high molecular weight adiponectin + VEGF	Positive association with radiographic progression	Cross-sectional study	2019, Judith Rademacher [49]
LINC00311	Positive association with diagnosis and disease activity	Prospective study	2019, Hongfa Zhong [50]
Lipocalin 2 (LCN 2), Oncostatin M (OSM)	Positive association with disease activity and treatment response	Longitudinal observational study	2021, Florence W.L Tsui [51]
Metalloproteinase (MMP)-degraded collagen type I (C1M), type II (C2M), type III (C3M), and type IV (C4M2)	Positive association with disease activity	Prospective cohort study	2019, Markéta Husšáková [52]
Neutrophil extracellular trap (NET)	Positive association with disease activity and treatment response	Cross-sectional study, case report	2020, Patricia Ruiz-Limon [53]

## Table 1. Cont.

Biomarker	Predictability	Study type	Reference
Omentin-1	Positive association with cardiovascular risk	Prospective study	2020, Fernanda Genre [54]
Osteoprotegerin (OPG) and sclerostin (SCL)	Positive association with cardiovascular risk	Prospective study	2018, Fernanda Genre [55]
Pentraxin 3	Positive association with disease activity	Cross-sectional study	2018, Renato Nisihara [56]
Procollagen I N-terminal peptide	Positive association with disease activity	Cross-sectional study	2022, Xuegang Li [57]
Sclerostin and antisclerostin antibody	Positive association with gastrointestinal risk	Prospective study	2018, Michele Maria Luchetti [58]
	Positive association with diagnosis	Case-control study	2018, Perrota Fabio Massimo [59]
	Positive association with radiographic progression	Prospective study	2019, Judith Rademacher [49]
Semaphorin 3A (Sema 3A)	Positive association with disease activity	Prospective study	2019, Hsien-Tzung Liao [60]
	Positive association with treatment response	Prospective study	2017, Fabio Massimo Perrotta [61]
Serum amyloid A1 (SAA1)	Positive association with diagnosis	Prospective study	2020, Shijia Liu [62]
Serum calprotectin	Positive association with disease activity	Prospective study	2020, Matthias Jarlborg [63]
	Positive association with treatment response	Prospective study	2019, Hua Hu [64]
	Positive association with cardiovascular risk	Prospective study	2018, Fernanda Genre [65]
	Positive association with radiographic progression	Prospective study	2022, Judith Rademacher [66]
Serum miR-214	Positive association with disease activity	Prospective study	2019, Hyun Yi Kook [67]
Tenascin-C (TNC)	Positive association with disease activity	Prospective study	2020, Kristyna Bubova [68]
	Positive association with disease activity	Prospective study	2018, Latika Gupta [69]
Vaspin	Positive association with cardiovascular risk	Prospective study	2021, Javier Rueda-Gotor [70]
Vimentin	Positive association with radiographic progression	Prospective study	2019, Anne Sofie Siebuhr [71]
Visfatin	Positive association with cardiovascular risk	Prospective study	2021, Rabia Aydogan Baykara [72]
	Positive association with disease activity	Prospective study	2022, Judith Rademacher [66]

Table 1. Cont.



Figure 1. Biomarkers significance in axial spondiloarthtitis.

## 2. Systemic Markers of Inflammation

## 2.1. Acute-Phase Proteins

**C-reactive protein and erythrocyte sedimentation rate (ESR),** which are acute-phase proteins, are the most widely used inflammatory indicators in active disease status and have been extensively studied in respect of AS. Nevertheless, as diagnostic biomarkers, they have low sensitivity and specificity, being increased only in 40-50% of patients with AS [23]. The ASDAS-CRP score, which is used to assesses AS activity, and its subjective criteria include CRP. This score is the preferred tool for disease activity assessment in the decision-making process for axSpA treatment modification in clinical routine [73]. In addition, one of the ASAS classification criteria is high CRP values. The CRP metabolite (CRPM) is correlated with disease activity, being significantly increased in AS compared with nr-axSpA [13,71]. One recent study shows that CRP may predict spinal immobility development in AS patients [16]. Moreover, the CRP to albumin ratio (CAR), a novel inflammatory biomarker, is positively correlated with BASDAI and BASFI, being a reliable marker for assessing disease activity in patients with axSpA [23,24]. CRP levels can be predictive for axSpA treatment response [14], whereas CRP gene rs3091244 polymorphism

has been found to be a reliable biomarker for a suitable response to etanercept treatment in AS [15]. Navarini L. et al. showed that persistent elevated CRP levels can be an indicator for axSpA patients at higher risk of CVD identification [13]. ESR can predict the clinical response of TNF inhibitor treatment within AS patients. In a recent study, the cut-off value of baseline ESR was 47.00 mm/h (sensitivity 57.1% and specificity 94.9%), with AUC = 0.819 [30]. In a prospective study, ESR was associated with trabecular bone score, which may be useful as a tool for assessing osteoporosis for AS [29].

**Serum amyloid A protein (SAA)** is another inflammatory biomarker that is more sensitive than other acute-phase proteins, but with lower availability and higher cost [74]. The key role of SAA in different inflammatory rheumatic conditions including axSpA is its involvement in inflammasome cascade activation and recruitment of interleukin (IL)-17-producing T helper cells [75]. SAA levels are higher in AS as compared to healthy controls and the use of a combination of SAA and CRP detection may potentially increase the likelihood of a correct and prompt diagnosis, which would enable early treatment [62]. Although there is a positive association with systemic inflammation in axSpA, and thus with disease activity, its association with treatment response is inconclusive [76].

**Pentraxin 3** is a long pentraxin overexpressed during the acute-phase response, playing significant roles in the innate immune response and inflammation, as well as in tissue damage and remodeling. Its direct implications in axSpA and many other rheumatic disorders have been extensively studied [77]; unfortunately, some studies have shown no association with disease activity and acute inflammation in axSpA [56,78].

**Fibrinogen-to-albumin ratio (FAR)** is a novel inflammatory index that is inexpensive and easily measurable, and fibrinogen is also considered an acute-phase response protein able to reflect the systemic inflammatory status [79]. Fibrinogen is an important factor involved in the coagulation system and albumin (ALB) is considered an objective indicator to foreshadow nutritional status [80]. It seems that FAR may be used as an inflammatory parameter for monitoring disease activity in AS; in a retrospective study on 135 consecutive AS patients, FAR levels were higher than in controls, with significantly higher concentrations in active disease and a positive correlation with disease activity (BASDAI score) [33].

**Lipocalin-2 (LCN-2)** is an acute-phase protein released in response to microbial triggers, which has pro- and anti-inflammatory properties [81]. In addition, **Oncostatin M** (**OSM**) is an acute-phase protein, which may be associated with lack of response to TNF inhibitors in inflammatory bowel disease (p < 0.00001) [82]. Both tests are involved in inflammation and bone remodeling [83]. Interestingly, persistently elevated LCN2 and OSM levels were associated with sacroiliac joint inflammation (p = 0.0005, respectively p = 0.005) in a longitudinal observational study of axSpA [51]. Moreover, clinically and serologically active patients displayed constantly raised LCN2 and/or OSM and back pain scores (> 4) after treatments [51].

#### 2.2. Adipokines

Adipokines are proteins produced within white adipose tissue, with significant functions in lipid metabolism, insulin sensitivity, and energy balance regulation, and important roles in response to both infection and systemic inflammation. Due to their immunomodulatory effects, adipokines have been extensively studied in the pathophysiology of chronic inflammatory diseases, including axSpA [84].

**Leptin** is the best-known proinflammatory adipokine that can increase the direct production of several proinflammatory cytokines and may also induce the production of reactive oxygen species in macrophages, neutrophils, and endothelial cells, as well as potentiating interferon  $\gamma$  (IFN $\gamma$ )-induced expression of nitric oxide synthetase [85]. The role of leptin in bone metabolism is complex: in the central pathway, leptin binds to its hypothalamic receptor, then inhibits osteoblast differentiation and enhances osteoclast activation by increasing sympathetic activity, while in its peripheral pathway, leptin increases bone formation and inhibits bone resorption by acting directly on osteoblasts and

osteoclasts. In a longitudinal prospective study conducted in an AS population, Park J.-H. and colleagues found an increase in leptin levels that was significant from baseline to two-year follow-up and was associated with radiographic progression as reflected by the Modified Spine Assessment for Ankylosing Spondylitis Stock (mSASSS)  $\geq 2$ . These data suggest that high serum leptin levels may reflect structural damage proportional to disease progression [48]. Another study demonstrated that biomarker combination (leptin with high molecular weight adiponectin and vascular endothelial growth factor (VEGF)) with clinical parameters (age, symptom duration, HLA-B27-positive, BASDAI, BASFI, ESR) can improve the prediction of radiographic spinal progression in SpA [49]. A recent study in AS patients reported changes in leptin and visfatin levels over the first two years and a significant association with radiographic progression after four years [66]; however, several other studies showed inconclusive results when assessing leptin's role as a predictor of structural damage in axSpA patients [76]. Furthermore, a negative association with disease activity indices was described in respect of axSpA [76].

Visfatin, recognized as nicotinamide phosphoribosyltransferase and pre-B cell colony-enhancing factor (PBEF), is an adipokine involved in inflammation, atherosclerosis, and glucose metabolism [85]. Baykara et al. demonstrated that adipokine is significantly higher in patients with AS than in controls and correlates with BASDAI, BASFI, and carotid intima-media thickness; moreover, visfatin has been recognized to be a predictor of AS activity [72].

**Omentin-1**, known as **intellectin1**, is a novel pleiotropic adipokine involved in the inflammatory response, but is also described as an anti-atherosclerotic and cardioprotective molecule [86]; additionally, it plays an important role in bone homeostasis through the anti-osteoblastic-inhibiting and pro-osteoclastic effects of activated macrophages [87]. It seems that low omentin-1 levels are associated with metabolic dysfunction and high prevalence of cardiovascular involvement in axSpA patients [54].

**Vaspin**, an adipokine associated with CVD and systemic inflammation, was also studied in respect of axSpA. A recent study conducted on 510 patients diagnosed with axSpA suggested that vaspin is associated with cardiovascular risk and may influence the atherosclerotic process in axSpA [70].

## 2.3. Cytokines

Cytokines are small proteins predominantly produced by T helper cells (Th) and macrophages [88], and they are extensively involved in the pathobiology of different inflammatory rheumatic conditions including axSpA [89]. There are two distinct inflammatory pathways in axSpA, the tumor necrosis factor (TNF)- $\alpha$  axis and the interleukin (IL)-23/IL-17A axis, which have major implications for specific pathogenic events (inflammation and ossification) occurring locally (axial as well as peripheral, comprising synovitis and enthesitis) and also systemically (ocular, intestinal, cutaneous, etc.). Numerous studies have demonstrated that genetically determined high activities of the TNF- $\alpha$ , IL-23/IL17, and NFkB pathways are associated with increased risk of SpA [47,90]. Saif D. et al. showed that IL-17A is a valuable biomarker for detecting early axSpA changes in asymptomatic and nr-axSpA in psoriatic patients [46].

**TNF-** $\alpha$  is highly expressed in axSpA, especially in the sacroiliac joint biopsies of AS patients, providing a potential link between inflammation and abnormal bone homeostasis; however, different studies have shown inconclusive associations with disease activity indices and inconclusive associations with treatment responses. Thus, it seems that TNF- $\alpha$  is not a reliable biomarker of disease activity in axSpA [76].

**IL-6**, the second most important inflammatory cytokine in rheumatic conditions, has higher levels in AS patients than in controls, particularly in the early stages of disease, in serum, cartilage, synovial fluid, and sacroiliac joint biopsy specimens; however, studies have also shown an inconclusive association with disease activity scores, but a positive association with structural damage, as high IL-6 levels seem to predict damage in axSpA [76]. Dong, Y. et al showed that IL-6 can predict the clinical response of TNF inhibitor treat-

ment in AS patients (the cut-off value of baseline was 9.05 pg/mL, sensitivity 76.2% and specificity 94.6%, with AUC = 0.78) [30]. Interestingly, IL-6 is associated with MRI changes; lower IL-6 serum levels relate to lower MRI inflammation scores. High baseline levels of Il-6 were highly predictive of a structural response to TNF inhibitors as assessed with the mSASSS [76].

**IL-17** has an inconclusive association with axSpA activity scores but decreases following treatment response [76].

**IL-31** serum levels correlate with sCD40L levels and mSASSS < 1, suggesting that high IL-31 is associated with reduced structural damage in early axSpA [42].

**IL-33**, a recently identified member of the IL-1 family, could be a potential candidate biomarker of disease susceptibility and anti-TNF treatment response in AS patients (p = 0.046, pc = NS, OR = 1.98, CI95% = 0.99–3.98) [43].

**IL-22** is a cytokine related to the IL-23/IL-17 axis and could be an independent biomarker for the differentiation of axSpA patients from non-inflammatory patients [45].

Finally, **IL-12B (rs6871626) and IL-6R (rs4129267)** gene polymorphisms could serve as promising biomarkers for diagnosis and prognosis in AS patients [44].

It is widely accepted that chronic joint inflammation in axSpA is characterized by the infiltration of activated macrophages; the **haptoglobin–hemoglobin receptor CD163** and the **mannose receptor CD206** are strongly expressed on M2c and M2a macrophages, respectively. In a prospective study, sCD206 levels were significantly higher in patients with early disease and definite radiological sacroiliitis and was significantly increased in those with a response to TNF inhibitors (p < 0.0001); this was not the case for sCD163 [18].

**CD27-negative B cells expressing low levels of CD21 (CD21low B cells)** are implicated in autoimmune diseases with strong B cell involvement. A recently published article emphasizes that CD27-CD38lowCD21low B cells in axSpA patients correlate positively with age and ESR. In addition, axSpA patients with and without extra-skeletal manifestations showed different frequencies of CD27-CD38lowCD21low B cells, with higher levels in those with extra-skeletal involvement [19].

#### 2.4. Miscellaneous

**Calprotectin**, a DAMP (damage-associated molecular pattern) protein, is a calciumand zinc-binding protein mainly found in the cytosol of monocytes, macrophages, and neutrophil granulocytes [91]. Recent advances suggest calprotectin as a new biomarker in inflammatory rheumatic diseases as an alternative to acute-phase proteins, since its serum levels may reflect the local activation of inflammatory innate immune cells involved in enthesitis and arthritis [92]. Serum calprotectin levels are elevated in axSpA and progressively decrease under specific medication (TNF- and IL-17A-inhibitors) [93]; in a prospective study on 451 axSpA patients, calprotectin serum levels were higher than in healthy controls and significantly associated with ASDAS as well as the presence of coxitis, suggesting calprotectin's role in disease activity [63]. Furthermore, different papers reported raised fecal calprotectin concentrations correlating with worse disease activity (ASDAS-CRP) and visual analog scale (VAS) physical function scores [32,94]. In addition, it seems that calprotectin correlates with lipid parameters (high-density lipoprotein cholesterol, atherogenetic index) and CRP levels in axSpA patients, with subsequent implications on atherosclerosis development [65]. Calprotectin also positively correlates with CRP, ESR, BASDAI, BASFI, patient global assessment, and VAS scores, reflecting disease severity in AS patients. Moreover, those patients with elevated calprotectin levels present with higher IL-1 $\beta$ , IL-17, and TNF- $\alpha$  levels [64]. Baseline calprotectin levels correlated with mSASSS and with new syndesmophyte formation after two years according to a recent study [66]. Thus, it has been strongly demonstrated that calprotectin positively associates with disease activity indices and structural damage in axSpA, while its levels decrease following treatment response; in addition, high calprotectin predicts damage in axSpA [76].

**Neutrophil extracellular trap (NET)** is described as extracellular deoxyribonucleic acid (DNA) fibers made of histone and cytoplasmic granule proteins, involved in numerous

processes, which can capture pathogens, degrade toxic bacterial factors, and kill bacteria [95]. NETosis is a mechanism of the innate immune defense playing an important role in diverse rheumatic diseases; NET-associated proteins can participate in autoimmune diseases by inducing autoantibodies against self-antigens. In a recent prospective study, neutrophils showed increased spontaneous NET formation, high expression of NET-associated signaling components, nuclear translocation of peptidilarginine deiminase 4, and elevated H3 histone citrullination in AS. In addition, patients had altered circulating levels of cell-free NETose-derived products (DNA, nucleosomes, and elastase), which positively correlated with CRP, ESR, and IL-1 $\beta$  concentrations. Furthermore, cell-free nucleosome levels were positively correlated with TNF- $\alpha$  concentration and cell-free elastase levels with CRP concentration. Interestingly, significant decreases in circulating levels of DNA, nucleosomes, and cell-free elastases were reported after six months of infliximab in a small cohort of AS patients (p < 0.05) [53].

**Galectin-3** is a soluble  $\beta$ -galactoside-binding lectin with at least one evolutionarily conserved carbohydrate-recognition domain; it performs important functions in biological activities, including cell growth, apoptosis, pre-mRNA splicing, differentiation, transformation, angiogenesis, inflammation, fibrosis, and host defense. It also seems that galectin-3 may be used as a diagnostic or prognostic biomarker for certain types of heart disease, kidney disease, and cancer [96]. In a study of 112 patients with AS and 130 healthy controls, serum galectin-3 levels were higher in AS and positively correlated with CRP, ESR, ASDAS index, and global pain index. Thus, inflammation and immune dysfunction can increase serum galectin-3 levels in AS patients, contributing to disease progression [34].

# 3. Molecules Involved in Bone Homeostasis

Understanding the pathogenesis of AS plays a key role in preventing new bone formation, one of the main causes of disability and poor quality of life. Numerous cellular elements (osteocytes, chondrocytes, immune cells, etc.), inflammatory cytokines, and cellular pathways are involved in new bone formation in AS [97]. The role of the Wnt/ $\beta$ -catenin pathway and its inhibitors, sclerostin and Dikkopf, has been evaluated in AS pathogenesis to identify a possible link with bone formation.

Sclerostin is a small glycoprotein encoded by the SOST gene primarily expressed by mature osteocytes, and is a critical regulator of bone formation [97]; it binds to the lipoprotein receptor-related protein and antagonizes the canonical Wnt signaling that normally activates TNF- $\alpha$  by a positive feedback loop [98]. Thus, SOST downregulates osteoprotegerin (OPG) in a dose-dependent manner, leading to an increase in the receptor activator of nuclear factor-kB ligand (RANKL)/OPG mRNA ratio, which exerts catabolic action through the promotion of osteoclast formation and activity [99]. Different studies have demonstrated lower sclerostin serum levels associated with radiographic progression in AS patients compared to healthy controls [59,100]. However, a negative association with disease activity indices was reported in axSpA [75]. The presence of circulating immunoglobulin G (IgG) antibodies that bind sclerostin (Scl-Ab) has been referred to as the cause of lower sclerostin serum levels, playing a presumed role in articular disease development. Furthermore, Scl-Ab is an emerging anti-osteoporosis treatment due to its enhanced osteogenic effects, leading to an increase in bone formation and a decrease in bone resorption [101]. In addition, high levels of Scl-Ab are associated with inflammatory bowel disease in axSpA patients [58].

**Bone morphogenetic protein-7 (BMP-7)** is an anti-inflammatory growth factor belonging to the transforming growth factor beta (TGF- $\beta$ ) superfamily and plays an important role in various biological processes, such as skeletal morphogenesis, embryogenesis, hematopoiesis, and neurogenesis [102]. In a prospective, multicenter French study, serum BMP-7 changes over 5 years were associated with inflammation and were elevated in active disease. In that study, serum sclerostin levels also increased significantly over time, but to a lesser extent than for serum BMP-7 [12].

The extracellular matrix (ECM) is a combination of collagens, glycosaminoglycans, and other molecules, but also fibers that fill intercellular spaces that undergo substantial changes during inflammation, which reflect pathological events, such as the influx of inflammatory cells. Matrix metalloproteinases (MMPs) can degrade ECM proteins, resulting in the release of protease-specific metabolites that can be detected as serum biomarkers of ECM tissue turnover, which would reflect local pathogenic processes. High serum MMP-3, MMP-8, and MMP-9 levels are classically reported in patients with AS and high disease activity and structural progression [103]. Type I, II, III, and IV collagens are found in the ECMs of various joint tissues (articular cartilage and hyaline, tendons, bones, and connective tissue) and can be degraded by proteases. The MMP-mediated metabolite of type I collagen (C1M) shows soft tissue destruction, and it is increased in AS [52]. C1M has been significantly associated with mSaSSS progression [104]. C2M is an MMP-mediated metabolite of type II collagen, which reflects cartilage destruction, while MMP-mediated metabolite of type III collagen (C3M) and MMP-mediated metabolite of type IV collagen (C4M2) reflect soft tissue degradation. Collagen degradation metabolite levels are higher in axSpA and these metabolites could be axSpA disease activity biomarkers. In a cross-sectional study, metabolite levels were higher in axSpA (both nr-axSpA and AS) compared to asymptomatic controls, and all metabolites except C2M were higher in AS compared to nr-axSpA. C1M, C3M, and C4M2 were associated with life quality levels, function index, and disease activity indices (ASDAS-CRP) in nr-axSpA and AS patients [52].

**Procollagen I N-terminal peptide (PINP)** is one of the most sensitive biomarkers and is particularly useful for monitoring bone formation and antiresorptive therapies. PINP is released into circulation and offers some practical benefits, including its low diurnal variability and room-temperature stability [105]. The International Federation of Clinical Chemistry and Laboratory Medicine and the International Osteoporosis Foundation recently recommended using PINP as a reference marker for bone formation in studies concerning fracture risk assessment and treatment response [106]. PINP can be useful as a marker of objective inflammation in AS, and correlates with MRI-determined SIJ inflammatory scores in AS patients but not in those with nr-axSpA [57].

**Type I and II C-terminal telopeptides (CTX-I and CTX-II)** are markers of osteoclast activity reflecting bone degradation. Different studies have shown higher serum and urinary CTX-I levels in AS patients, while CTX-II correlates with structural damage as supported by MRI inflammation scores in the sacroiliac joint and/or lumbar spine in axSpA, being considered a biomarker of radiographic progression. Remarkably, CTX-II is able to predict radiological damage and progression and correlates with changes in the mSASSS score in two-year follow-up analyses. Thus, there is a positive association of CTX-II with structural damage in axSpA, higher levels predicting further damage; CTX-II levels also predict patients at risk of developing syndesmophytes and radiographic spinal progression. In addition, CTX-II serum levels significantly correlate with disease activity and decline after TNF inhibitors, being a biomarker of treatment response in axSpA patients [76].

Vimentin, also known as fibroblast intermediate filament (IF), is the major IF protein of mesenchymal cells, which are an important part of the cytoskeleton [107]. A specific **MMP-mediated and citrullinated fragment of vimentin** (VICM) could be an antigen for anticitrullinated protein antibodies that have been found in patients with r-axSpA. VICM is a measure of macrophage activity and citrullination and is higher in inflammatory diseases [71]. It seems that VICM is associated with disease activity as supported by a positive association with systemic inflammation; moreover, it is a potential biomarker of radiographic progression in r-axSpA [108], although certain studies have failed to demonstrate any relationship with mSASSS progression [71].

**Osteoprotegerin** is a member of the TNF receptor family, but it is secreted and acts like a cytokine. It is the decoy receptor for receptor activator of nuclear factor-kappaB ligand (RANKL), another cytokine that activates osteoclasts and causes bone resorption. Some studies have found elevated serum levels of osteoprotegerin [55,109] and an association with disease activity [55]. In a prospective study, osteoprotegerin levels were also

associated with some metabolic syndrome features, being associated with an increased risk of CVD [55].

**Osteocalcin** is the main non-collagenous protein of bone tissue produced exclusively by osteoblasts, being a relevant marker of bone formation. Several studies have reported that high serum levels in AS are associated with radiographic progression, predicting structural damage [110,111]. However, one recent study failed to demonstrate any association between osteocalcin and ASDAS, BASDAI, BASFI and CRP levels in axSpA [112].

**Dickkopf-1 (DKK-1) protein** is a member of the Dickkopf family, composed of two cysteine-rich domains and a signal peptide sequence. DKK-1 has been acknowledged as a soluble inhibitor of the Wnt/ $\beta$ -catenin signaling pathway involved in bone formation through its regulation of the proliferation and differentiation of osteoblasts and osteoclasts [113]. Thus, DKK-1 participates in bone resorption or new bone formation, which are both important milestones in AS. In a recent meta-analysis, serum levels of DKK-1 were not significantly different in AS vs. healthy controls, but serum DKK-1 levels in patients with CRP > 10 mg/L and mSASSS > 30 were significantly lower than in healthy controls [114]. A prospective study showed correlation between Dkk-1 and abnormal findings in respect of MRI [25], while another study demonstrated that BMPs and Dkk-1 play an important role in the pathogenesis of abnormal bone remodeling in AS. Furthermore, Dkk-1 is higher in those taking NSAIDs regularly and bone morphogenetic proteins BMP-2/Dkk-1 significantly correlate with disease duration [26].

**Semaphorins**, which are proteins initially described as regulators of nervous system development, are involved in processes such as the regulation of immunity, angiogenesis, bone remodeling, apoptosis, and cell migration and invasion [115]. Liao H.T. et al. found increased levels of **semaphorin 3A** in AS patients compared to healthy controls and demonstrated that semaphorin 3A concentrations > 2 ng/ml are better for predicting a higher BASDAI > 4 than ESR or CRP. Furthermore, significant correlations between higher semaphorin 3A levels and the presence of uveitis, Schober's test, and interstitial lung disease were reported in axSpA. Semaphorin 3A is also a suitable indicator for monitoring disease activity and functional status during anti-TNF treatment in axSpA (p = 0.013) [60]. However, Perrotta F.M. et al. found no differences between SA patients and healthy controls, but serum semaphorin 3A levels correlated positively with ESR values (p = 0.049) and disease activity assessed by the physician VAS (p < 0.01) [61].

Chemokines play an important role in immune functions and inflammatory responses, and facilitate leukocyte migration and trafficking [115].

C-C motif chemokine ligand 11 (CCL11), known as an eosinophil chemotactic protein (eotaxin-1), is a chemokine involved in innate immunity and inflammatory responses. CCL11 is associated with leukocyte recruitment in inflammation sites by binding to the receptor CCR3, and has been described in several chronic inflammatory diseases, brain disorders, aging, and allergic conditions [116]. In addition, inflammation-driven CCL11 expression has been shown in bone tissue as well as CCL11's role in osteoclast migration and resorption [117]. Osteoblasts are also able to overexpress CCL11 during inflammation. Furthermore, together with its cognate receptor CCR3, CCL11 correlates with disturbed bone remodeling, and stimulates migration of osteoclast precursors and bone resorption [117]. Although the clinical significance of CCL11 in bone metabolism in AS patients has not been completely investigated, it seems that CCL11 serum levels are significantly positively correlated with mSASSS and syndesmophyte numbers [17]. Eotaxin-11 is also overexpressed in human atherosclerotic lesions [118].

#### 4. Antibody-Based Biomarkers

**CD74**, an integral transmembrane molecule participating in intracellular sorting of MHC class II molecules, is involved in diverse biological processes, including the correct folding and transport of MHC class II molecules, the differentiation of B cells, and the inflammatory response as part of the macrophage migratory inhibitory factor [11]. The presence of **anti-CD74 autoantibodies** is higher in AS patients than in healthy con-

trols [7,8,10,11]; additionally, a significant correlation between positive **anti-CD74 IgG antibodies** in patients and disease activity was demonstrated in a cross-sectional study; based on their sensitivity and specificity, anti-CD74 IgGs were suggested as promising diagnostic tools to support the clinical diagnosis of axSpA [7].

Although **anti-CD74 IgA** is elevated in patients with early axSpA, this increase is not specific enough to produce significant diagnostic value in patients under 45 years old with low back pain [10]. Anti-CD74 IgA serum levels were increased in AS and independently associated with radiological changes in the spine, suggesting that anti-CD74 IgA can play an important role in AS pathobiology [8]. Some studies show significant association with BASDAI and BASFI scores [11].

**Drug-induced neutralizing antibodies against** TNF- $\alpha$  blockers. Biologics are widely used to control disease, usually resulting in rapid and persistent remission; unfortunately, a significant proportion of patients do not achieve remission or fail to maintain the therapeutic response, experiencing disease flare-ups [27]. In recent years, particular attention has been paid to the formation of neutralizing antibodies during treatment with biologics, resulting in a decrease in treatment efficacy and/or safety issues. Drug-induced neutralizing antibodies against TNF- $\alpha$  blockers (adalimumab and etanercept) have a negative effect on the progression of inflammatory joint disease and can be used as reliable biomarkers with which to evaluate the effectiveness of treatments (p < 0.005) [27].

#### 5. Human Leukocyte Antigen B27 (HLA-B27) and Newer Genetic Biomarkers

HLA-B27 is an MHC class I antigen and a pivotal factor of the WHO classification criteria for axSpA. HLA-B27 testing is one of the most widely used genetic tests for any common disease, being demonstrated in up to 90-95% of AS cases compared to the general population, in which HLA-B27 is found in only 8% of cases [4]. HLA-B27 prevalence varies within different continents and ethnic/racial populations [39,40,119]. Some studies have demonstrated that HLA-B27 can predict treatment response and patients with positive HLA-B27 have a better response to TNF inhibitors than negative HLA-B27 patients [120]. HLA-B27 may also be valuable in predicting clinical manifestations, especially acute anterior uveitis; HLA-B27-positive patients have higher risk of developing anterior acute uveitis [121]. Additionally, HLA-B27-positive patients have a significantly younger age at onset, more uveitis, and a higher frequency of involvement of peripheral joints and hips than HLA-B27-negative patients [122], with a recognized progression to clinical axSpA within 1 year of follow-up as recently demonstrated [41]. However, only 2-5% of HLA-B27-positive to the disease [123].

**Circular RNA (circRNA)** comprises a different class of noncoding RNA produced; it is a functional molecule with important roles in regulating transcription and splicing, microRNA sponging, and modulating protein–protein interactions. CircRNA can be classified into exonic RNAs (ecircRNAs), formed by exons, circular intronic RNAs (ciRNAs), formed by introns, and exon-intron circRNAs (ElciRNAs), formed by both exons and introns [124]. More studies have indicated that circRNAs may be associated with the risk of nerve diseases, atherosclerotic vascular diseases, cancer, and autoimmune diseases, suggesting it is a novel diagnostic and prognostic biomarker candidate, as well as a new therapeutic target for different disorders [125–128]. Recent studies reported that circRNA plays an important role in axSpA; the expression of **hsa\_circ\_0079787** was significantly reduced in the peripheral blood of axSpa patients as compared to healthy controls and SLE patients [129]. Moreover, Hsa\_circ\_0079787 was negatively correlated with BASDAI and positively correlated with the platelet count (PLT) and the lymphocyte-to-monocyte ratio. The ROC curve analysis indicated that hsa\_circ\_0079787 (0.715) and the combination hsa\_circ\_0079787-PLT-mean platelet volume (MPV)-plateletcrit (PCT) (0.835) had significant diagnostic value for axSpa. In addition, hsa\_circ\_0079787-HLA-B27 for axSpa vs. healthy controls was superior to that of HLA-B27 [130].

**MiR-214**, a vertebrate-specific miR precursor, is expressed in several human cells and associated with multiple diseases [131], including playing a crucial role in skeletal disorders; increased osteoclastic miR-214 is associated with both elevated serum exosomal miR-214 and reduced bone formation in elderly women with fractures [132]. In a prospective study, miR-214 levels were associated with disease activity in AS patients [67].

Endoplasmic reticulum aminopeptidase 1 (ERAP 1) is an aminopeptidase whose main function is to cut endoplasmic reticulum (ER) peptides to a length suitable for binding and presentation by MHC class I molecules, including HLA-B27 [133]. Various studies have demonstrated that ERAP1 is the second strongest locus associated with AS in Caucasians, just after HLA-B27 [134]. The genetic interaction between ERAP1 and HLAB27 found in AS reflects that peptide cleavage and presentation contribute to disease susceptibility [135]. A second member of the aminopeptidase family present in ER is **endoplasmic reticulum** aminopeptidase 2 (ERAP2), which significantly influences the HLA-B27:05-bound peptidome. ERAP 2 destroys some ligands and decreases the abundance of many more ligands with N-terminal basic residues, while increasing the nanometer abundance [136]. A casecontrol study, which evaluated the role of four single-nucleotide polymorphisms in the ERAP1 (rs2287987, rs30187, rs27044) and ERAP2 (rs2248374) genes and their haplotypes, demonstrated the association between these proteins and AS. The minor T allele and homozygous TT genotype of rs30187 were the most associated with AS and significantly increased disease risk, while the minor C allele of rs2287987 had a protective effect. Unlike ERAP1, no effect of rs2248374 in ERAP2 was described in relation to the disease [28].

Long Intergenic Non-Protein-Coding RNA 311 (LINC00311) is an RNA gene affiliated with the lncRNA class; it has been described as a critical regulator in human disease development and progression due to its role in regulating gene expression at multiple levels [137]. One study showed that lncRNA LINC00311 is involved in osteoporosis by regulating the differentiation and proliferation of osteoclasts [138]. Zhong H. and Zhong M. found that LINC00311 was upregulated in AS patients compared to healthy controls and patients with low back pain, suggesting its involvement in AS pathogenesis. They analyzed the diagnostic value of LINC00311 and showed that plasma LINC00311 may be a potential diagnostic marker for AS (AUC = 0.9041). They also reported an association with disease activity (BASDAI, ASDAS, CRP, and ESR). Interestingly, LINC00311 levels in plasma decreased after treatment, reflecting treatment outcome, and it was observed that high levels of LINC00311 showed a significantly higher rate of re-hospitalization, reflecting recurrence [50].

**Eukaryotic Translation Elongation Factor 1 Epsilon 1 (EEF1E1)**, also known as **aminoacyl-tRNA synthetase-interacting multifunctional protein 3 (aiMP3/p18)**, is a proteincoding gene and is involved in AS by influencing aminoacyl-tRNA biosynthesis. Fan X. et al. suggest that EEF1E1 may be an underlying novel and important genetic biomarker for the diagnosis of AS. EEF1E1 can be controlled to promote the development of various types of cancer [139] and may also function as a tumor suppressor [140]. They also identified another crucial gene for the diagnosis of AS: serpin family a member 1 (SERPINA1). SER-PINA 1 is a gene that encodes alpha-1-antitrypsin (AAT) and can be correlated with AS by participating in platelet degranulation. The AAT concentration was found to be higher in patients with active-phase AS than in patients in remission/partial remission [141]. In their study, EEF1E1 and SERPINA1 levels were significantly increased and the average expression log r ratios of single-nucleotide polymorphism (SNP) sites in these genes were significantly higher in AS patients than in healthy controls [31].

**Soluble suppression of tumorigenicity 2 (sST2)** is the circulating form of the ST2L cellular receptor, expressed by cardiomyocytes and vascular endothelial cells together with its interleukin-33 ligand after cardiovascular injury [142]. It is a marker of poor prognosis in chronic inflammatory conditions [143] and a novel biomarker of cardiac fibrosis [144]. The prevalence of cardiovascular association in AS varies depending on the diagnostic method used and the pathology described, ranging from 10% to 40% [145]. Demet Ozkaramanli

Gur et al. reported that sST2 can represent the link between inflammation and fibrosis in AS [35].

## 6. The Microbiome as a Biomarker

The occurrence of a joint lesion resembling reactive arthritis in a patient with bacterial dysentery prompted researchers to investigate the link between the gut and arthritis [146]. Intestinal microbial dysbiosis occurs as a common component in various inflammatory disorders, including SpA [147]. The complexity and dynamic nature of the gut microbiome in axial spondyloarthritis are under research. The gut is involved in maintaining homeostasis between the microbiota and the host, leading to the effective coordination of various immune cells [148]. Innate cells that appear to be implicated in the pathogenesis of SpA can control intestinal homeostasis by inducing apoptotic cell death and the deletion of T cells specific to activated commensal bacteria [149]. Intestinal dysbiosis is associated with worse axSpA disease activity and physical function, apparently regardless of intestinal inflammation and treatments [150]. There is no consensus regarding the bacterial species that are involved in the development of SpA. Some studies have demonstrated that microorganisms may be used as new biomarkers to characterize the pathogenesis of AS. A biomarker commonly used in the diagnosis of inflammatory bowel disease is the anti-Saccharomyces cerevisiae antibodies (ASCAs), yeast organisms found mainly in the small intestine. Olofsson T. et al. reported in a cross-sectional study that anti-Saccharomyces cerevisiae antibodies titers were elevated vs. controls, but seropositivity was not associated with disease subtype or status [32]. Another study shows distinct microbial colonization in the terminal ileum with Lachnospiraceae, Ruminococcaceae, and Prevotellaceae in AS patients vs. control [150]. In addition, Gill T. et al. found that *Prevotella* and *Blautia* (Lewis rats) and Akkermansia muciniphila, rc4-4, Lachnospira, and Lachnospiraceae (Fischer rats) are closely related to dysregulated inflammatory pathways in SpA [151].

#### 7. Miscellaneous Biomarkers

The complement system is a part of the innate immune system that participates not only in the inflammatory response, coagulation, or fibrinolysis but also in the development and progression of cardiometabolic diseases [152]. axSpA is characterized by an increased risk of cardiovascular comorbidities. Therefore, the identification of biomarkers that can be used as a screening strategy to select those patients who require a closer evaluation is of considerable interest to clinicians. **Complement C3** is a potential predictor of cardiovascular events; elevated C3 levels have been closely linked to insulin resistance, abdominal obesity, and hypertension [153]. The activated form of the C3 complement can act as a hormone involved in lipid storage and energy homeostasis [154]. In a cross-sectional, Iván Arias de la Rosa et al. reported that axSpA patients with high levels of C3 complement had an increased prevalence of cardiometabolic risk factors and higher disease activity. In addition, complement C3 levels allowed the identification of insulin-resistant patients [22].

**Tenascin-C (TNC)** is a large glycoprotein of the extracellular matrix consisting of the N-terminal TNC assembly domain, 14.5 repeats similar to epidermal growth factor, repeated type III fibronectin domains, and fibrinogen-like C-terminal globular domain. TNC reflects tissue damage during the inflammatory process and also active tissue remodeling [155]. In a prospective study, it was reported that serum TNC levels are significantly higher in patients with axSpA, particularly in those with radiographic signs of advanced disease, compared to healthy subjects [68]. In addition, another study showed that TNC levels were higher in AS than in healthy controls. Gupta L et al. found that serum TNC levels had a better correlation with ESR and CRP in patients with early disease duration ( $\leq$  5 years), and TNC decreased treatment levels in those who responded to treatment vs. those without a response [69].

Hepatocyte growth factor (HGF) is a multifunctional cytokine involved in embryogenesis, organogenesis, wound healing, and tissue repair and is also involved in tumorigenesis and cancer invasion [156]. HGF is expressed most by stromal cells and binds to the c-MET receptor tyrosine kinase. HGF-MET signaling can lead to a variety of cellular responses, such as morphogenesis, proliferation, cell survival, tissue regeneration, and protection against inflammatory diseases [157]. C-MET is also expressed on osteoblasts and osteoclasts; thus, HGF may regulate bone metabolism [158]. In a prospective study, HGF levels were increased in AS male patients and correlated with high mSASSS and low BMD. Moreover, HGF was positively associated with risk factors for osteoproliferation, including age, CRP, and smoking [38]. Another prospective cohort study demonstrated that elevated s-HGF levels were associated with the development of new syndesmophytes in men with AS [37].

L- and H-ficolin are proteins of the lectin pathway and play important roles in innate immunity participating in the clearance of apoptotic cells through complement activation [159]. In a cross-sectional cohort, H- and L-ficolin levels were increased in patients with axSpA compared to controls; in addition, the diagnostic specificity of HLA-B27 could be improved by combining it with increased concentrations of H- or L-ficolin [36].

#### 8. Conclusions

Despite advances in biomarker research over recent years and continued efforts to validate new biomarkers for patients with axSpA, only a few are currently indicated in routine clinical practice, and the "classical" biomarkers still help treatment decisions via assessment of disease activity, prognosis, and response to treatment. Indeed, validation of a new biomarker usually entails a long process, but implementation requires a standardized methodology, ideal cost–benefit ratio, and definite efficiency for diagnosis, therapy, and prognosis. While higher costs and accessibility issues can limit their use, additional costs derived from a delayed diagnosis, therapeutic failure, as well as poor prognosis justify the investment.

Several newer biomarkers such as anti-CD74 antibodies, circulating microRNAs, CAR, FAR, ERAP1, EEF1E1, H-ficolin, L-ficolin, LINC00311, pentraxin 3, serum calprotectin, and tenascin-C have already expressed relevant associations with diagnosis and disease activity across multiple studies in axial spondyloarthritis. Additionally, CRP, ESR, CD163, CD206, drug-neutralizing antibodies against TNF- $\alpha$ , lipocalin 2, oncostatin M, NET, semaphorin 3A, and serum calprotectin properly assess response to different therapeutic agents and, thus, increase the biological therapies quality. In addition, since the sizable cardiovascular risk is widely recognized in patients with axSpA, different biomarkers are suggested, especially CRP, complement component C3, omentin-1, osteoprotegerin, sclerostin, serum calprotectin, vaspin, and visfatin serum levels for the early identification and improvement in cardiovascular outcomes in such patients.

Future studies are needed to investigate the roles and limits of the most promising serum biomarkers in providing personalized care for patients with axSpA.in the era of biologic and targeted synthetic DMARDs.

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# References

- 1. Firestein, G.S.; Budd, R.C.; Koretzky, G.; Gabriel, S.E.; McInnes, I.B.; O'Dell, J.R. *Firestein & Kelley's Textbook of Rheumatology*, 11th ed.; Elsevier: Amsterdam, The Netherlands, 2020; Volume 2, p. 1307.
- Cardelli, C.; Monti, S.; Terenzi, R.; Carli, L. One year in review 2021: Axial spondyloarthritis. *Clin. Exp. Rheumatol.* 2021, 39, 1272–1281. [CrossRef] [PubMed]
- Walsh, J.A.; Magrey, M. Clinical Manifestations and Diagnosis of Axial Spondyloarthritis. J. Clin. Rheumatol. 2021, 27, e547–e560. [CrossRef] [PubMed]
- 4. Ancuta, C.; Pomirleanu, C. *Principii de Diagnostic si Tratament in Reumatologie*, 2nd ed.; Universitatea de Medicină și Farmacie "Grigore T. Popa" din Iași: Iasi, Romania, 2019; p. 177.
- 5. Califf, R.M. Biomarker definitions and their applications. *Exp. Biol. Med.* **2018**, 243, 213–221. [CrossRef] [PubMed]
- Abdal, S.J.; Yesmin, S.; Shazzad, M.N.; Azad, M.A.K.; Shahin, M.A.; Choudhury, M.R.; Islam, M.N.; Haq, S.A. Development of a Bangla version of the Bath Ankylosing Spondylitis Disease Activity Index (BASDAI) and the Bath Ankylosing Spondylitis Functional Index (BASFI). *Int. J. Rheum. Dis.* 2021, 24, 74–80. [CrossRef]
- Abdelaziz, M.M.; Gamal, R.M.; Ismail, N.M.; Lafy, R.A.; Hetta, H.F. Diagnostic value of anti-CD74 antibodies in early and late axial spondyloarthritis and its relationship to disease activity. *Rheumatology* 2021, 60, 263–268. [CrossRef]
- 8. Do, L.; Granåsen, G.; Hellman, U.; Lejon, K.; Geijer, M.; Baraliakos, X.; Witte, T.; Forsblad-d'Elia, H. Anti-CD74 IgA autoantibodies in radiographic axial spondyloarthritis: A longitudinal Swedish study. *Rheumatology* **2021**, *60*, 4085–4093. [CrossRef]
- 9. Ziade, N.R.; Mallak, I.; Merheb, G.; Ghorra, P.; Baerlecken, N.; Witte, T.; Baraliakos, X. Added Value of Anti-CD74 Autoantibodies in Axial SpondyloArthritis in a Population with Low HLA-B27 Prevalence. *Front. Immunol.* **2019**, *10*, 574. [CrossRef]
- 10. de Winter, J.J.; van de Sande, M.G.; Baerlecken, N.; Berg, I.; Ramonda, R.; van der Heijde, D.; van Gaalen, F.A.; Witte, T.; Baeten, D.L. Anti-CD74 antibodies have no diagnostic value in early axial spondyloarthritis: Data from the spondyloarthritis caught early (SPACE) cohort. *Arthritis Res. Ther.* **2018**, *20*, 38. [CrossRef]
- 11. Hu, C.J.; Li, M.T.; Li, X.; Peng, L.Y.; Zhang, S.Z.; Leng, X.M.; Su, J.M.; Zeng, X.F. CD74 auto-antibodies display little clinical value in Chinese Han population with axial spondyloarthritis. *Medicine* **2020**, *99*, 23433. [CrossRef]
- 12. Descamps, E.; Molto, A.; Borderie, D.; Lories, R.; Richard, C.M.; Pons, M.; Roux, C.; Briot, K. Changes in bone formation regulator biomarkers in early axial spondyloarthritis. *Rheumatology* **2021**, *60*, 1185–1194. [CrossRef]
- 13. Navarini, L.; Currado, D.; Marino, A.; Di Donato, S.; Biaggi, A.; Caso, F.; Costa, L.; Tasso, M.; Ruscitti, P.; Pavlych, V.; et al. Persistence of C-reactive protein increased levels and high disease activity are predictors of cardiovascular disease in patients with axial spondyloarthritis. *Sci. Rep.* **2022**, *12*, 7498. [CrossRef] [PubMed]
- 14. Baraliakos, X.; Szumski, A.; Koenig, A.S.; Jones, H. The role of C-reactive protein as a predictor of treatment response in patients with ankylosing spondylitis. *Semin. Arthritis Rheum.* **2019**, *48*, 997–1004. [CrossRef] [PubMed]
- 15. Xu, Y.; Jiang, W.; Zhang, H. Association between C-reactive protein gene variant and treatment efficacy of etanercept in ankylosing spondylitis patients receiving hip arthroplasty. *J. Clin. Lab. Anal.* **2020**, *34*, e23343. [CrossRef] [PubMed]
- 16. Sundström, B.; Ljung, L.; Wållberg-Jonsson, S. Exercise habits and C-reactive protein may predict development of spinal immobility in patients with ankylosing spondylitis. *Clin. Rheumatol.* **2018**, *37*, 2881–2885. [CrossRef] [PubMed]
- 17. Sohn, D.H.; Jeong, H.; Roh, J.S.; Lee, H.N.; Kim, E.; Koh, J.H.; Lee, S.G. Serum CCL11 level is associated with radiographic spinal damage in patients with ankylosing spondylitis. *Rheumatol. Int.* **2018**, *38*, 1455–1464. [CrossRef]
- Heftdal, L.D.; Loft, A.G.; Hendricks, O.; Ashouri Christiansen, A.; Schiøttz-Christensen, B.; Arnbak, B.; Jurik, A.G.; Østgård, R.; Winding Deleuran, B.; Møller, H.J.; et al. Divergent effects on macrophage biomarkers soluble CD163 and CD206 in axial spondyloarthritis. *Scand. J. Clin. Lab. Investig.* 2018, *78*, 483–489. [CrossRef]
- 19. Wilbrink, R.; Spoorenberg, A.; Arends, S.; van der Geest, K.S.M.; Brouwer, E.; Bootsma, H.; Kroese, F.G.M.; Verstappen, G.M. CD27-CD38lowCD21low B-Cells Are Increased in Axial Spondyloarthritis. *Front. Immunol.* **2021**, *12*, 686273. [CrossRef]
- 20. Luo, Q.; Fu, B.; Zhang, L.; Guo, Y.; Huang, Z.; Li, J. Expression and clinical significance of circular RNA hsa\_circ\_0079787 in the peripheral blood of patients with axial spondyloarthritis. *Mol. Med. Rep.* **2020**, *22*, 4197–4206. [CrossRef]
- Perez-Sanchez, C.; Font-Ugalde, P.; Ruiz-Limon, P.; Lopez-Pedrera, C.; Castro-Villegas, M.C.; Abalos-Aguilera, M.C.; Barbarroja, N.; Arias-de la Rosa, I.; Lopez-Montilla, M.D.; Escudero-Contreras, A.; et al. Circulating microRNAs as potential biomarkers of disease activity and structural damage in ankylosing spondylitis patients. *Hum. Mol. Genet.* 2018, 27, 875–890. [CrossRef]
- Arias de la Rosa, I.; Font, P.; Escudero-Contreras, A.; López-Montilla, M.D.; Pérez-Sánchez, C.; Ábalos-Aguilera, M.C.; Ladehesa-Pineda, L.; Ibáñez-Costa, A.; Torres-Granados, C.; Jimenez-Gomez, Y.; et al. Complement component 3 as biomarker of disease activity and cardiometabolic risk factor in rheumatoid arthritis and spondyloarthritis. *Ther. Adv. Chronic. Dis.* 2020, 11, 2040622320965067. [CrossRef]
- Zhong, Z.; Huang, Y.; Liu, Y.; Chen, J.; Liu, M.; Huang, Q.; Zheng, S.; Guo, X.; Deng, W.; Li, T. Correlation between C-Reactive Protein to Albumin Ratio and Disease Activity in Patients with Axial Spondyloarthritis. *Dis. Markers* 2021, 2021, 6642486. [CrossRef] [PubMed]
- 24. Pamukcu, M.; Duran, T.I. Could C-Reactive Protein/Albumin Ratio be an Indicator of Activation in Axial Spondyloarthritis? J. Coll. Physicians Surg. Pak. 2021, 30, 537–541. [PubMed]

- Zhao, Z.; Wang, G.; Wang, Y.; Yang, J.; Wang, Y.; Zhu, J.; Huang, F. Correlation between magnetic resonance imaging (MRI) findings and the new bone formation factor Dkk-1 in patients with spondyloarthritis. *Clin. Rheumatol.* 2019, *38*, 465–475. [CrossRef] [PubMed]
- 26. Liao, H.T.; Lin, Y.F.; Tsai, C.Y.; Chou, T.C. Bone morphogenetic proteins and Dickkopf-1 in ankylosing spondylitis. *Scand. J. Rheumatol.* **2018**, *47*, 56–61. [CrossRef]
- Kraev, K.; Geneva-Popova, M.; Popova, V.; Popova, S.; Maneva, A.; Batalov, A.; Stankova, T.; Delcheva, G.; Stefanova, K. Drug-neutralizing Antibodies against TNF-α blockers as Biomarkers of Therapy Effect Evaluation. *Folia Med.* 2020, 62, 282–289. [CrossRef]
- 28. Wiśniewski, A.; Kasprzyk, S.; Majorczyk, E.; Nowak, I.; Wilczyńska, K.; Chlebicki, A.; Zoń-Giebel, A.; Kuśnierczyk, P. ERAP1-ERAP2 haplotypes are associated with ankylosing spondylitis in Polish patients. *Hum. Immunol.* **2019**, *80*, 339–343. [CrossRef]
- Kang, K.Y.; Chung, M.K.; Kim, H.N.; Hong, Y.S.; Ju, J.H.; Park, S.H. Severity of Sacroiliitis and Erythrocyte Sedimentation Rate are Associated with a Low Trabecular Bone Score in Young Male Patients with Ankylosing Spondylitis. *J. Rheumatol.* 2018, 45, 349–356. [CrossRef]
- Dong, Y.; Guo, J.; Bi, L. Baseline Interleukin-6 and Erythrocyte Sedimentation Rate Can Predict Clinical Response of TNF Inhibitor Treatment in Patients with Ankylosing Spondylitis. Ann. Clin. Lab. Sci. 2019, 49, 611–618.
- Fan, X.; Qi, B.; Ma, L.; Ma, F. Screening of underlying genetic biomarkers for ankylosing spondylitis. *Mol. Med. Rep.* 2019, 19, 5263–5274. [CrossRef]
- Olofsson, T.; Lindqvist, E.; Mogard, E.; Andréasson, K.; Marsal, J.; Geijer, M.; Kristensen, L.E.; Wallman, J.K. Elevated faecal calprotectin is linked to worse disease status in axial spondyloarthritis: Results from the SPARTAKUS cohort. *Rheumatology* 2019, 58, 1176–1187. [CrossRef]
- 33. Liu, M.; Huang, Y.; Huang, Z.; Zhong, Z.; Deng, W.; Huang, Z.; Huang, Q.; Li, T. The role of fibrinogen to albumin ratio in ankylosing spondylitis: Correlation with disease activity. *Clin. Chim. Acta* 2020, *505*, 136–140. [CrossRef]
- Cao, M.Y.; Wang, J.; Gao, X.L.; Hu, Y.B. Serum galectin-3 concentrations in patients with ankylosing spondylitis. *J. Clin. Lab. Anal.* 2019, 33, e22914. [CrossRef] [PubMed]
- Ozkaramanli Gur, D.; Ozaltun, D.N.; Guzel, S.; Sarifakioglu, B.; Akyuz, A.; Alpsoy, S.; Aycicek, O.; Baykiz, D. Novel imaging modalities in detection of cardiovascular involvement in ankylosing spondylitis. *Scand. Cardiovasc. J.* 2018, 52, 320–327. [CrossRef] [PubMed]
- Troldborg, A.; Thiel, S.; Mistegaard, C.E.; Hansen, A.; Korsholm, T.L.; Stengaard-Pedersen, K.; Loft, A.G. Plasma levels of Hand L-ficolin are increased in axial spondyloarthritis: Improvement of disease identification. *Clin. Exp. Immunol.* 2020, 199, 79–87. [CrossRef]
- Deminger, A.; Klingberg, E.; Nurkkala, M.; Geijer, M.; Carlsten, H.; Jacobsson, L.T.H.; Forsblad-d'Elia, H. Elevated serum level of hepatocyte growth factor predicts development of new syndesmophytes in men with ankylosing spondylitis. *Rheumatology* 2021, 60, 1804–1813. [CrossRef] [PubMed]
- Torres, L.; Klingberg, E.; Nurkkala, M.; Carlsten, H.; Forsblad-d'Elia, H. Hepatocyte growth factor is a potential biomarker for osteoproliferation and osteoporosis in ankylosing spondylitis. *Osteoporos. Int.* 2019, 30, 441–449. [CrossRef] [PubMed]
- Ziade, N.; Abi Karam, G.; Merheb, G.; Mallak, I.; Irani, L.; Alam, E.; Messaykeh, J.; Menassa, J.; Mroue', K.; Uthman, I.; et al. HLA-B27 prevalence in axial spondyloarthritis patients and in blood donors in a Lebanese population: Results from a nationwide study. *Int. J. Rheum. Dis.* 2019, 22, 708–714. [CrossRef]
- Lim, C.S.E.; Sengupta, R.; Gaffney, K. The clinical utility of human leucocyte antigen B27 in axial spondyloarthritis. *Rheumatology* 2018, 57, 959–968. [CrossRef]
- 41. de Jong, H.M.Y.; de Winter, J.J.H.; van der Horst-Bruinsma, I.E.; van Schaardenburg, D.J.; van Gaalen, F.A.; van Tubergen, A.M.; Weel, A.E.A.M.; Landewé, R.B.M.; Baeten, D.L.P.; van de Sande, M.G.H. Progression from subclinical inflammation to overt SpA in first degree relatives of SpA patients is associated with HLA-B27: The Pre-SpA cohort. *Arthritis Care Res.* 2021, 0, 1–9.
- Rosine, N.; Etcheto, A.; Hendel-Chavez, H.; Seror, R.; Briot, K.; Molto, A.; Chanson, P.; Taoufik, Y.; Wendling, D.; Lories, R.; et al. Increase In II-31 Serum Levels Is Associated With Reduced Structural Damage In Early Axial Spondyloarthritis. *Sci. Rep.* 2018, *8*, 7731. [CrossRef]
- 43. Iwaszko, M.; Wielińska, J.; Świerkot, J.; Kolossa, K.; Sokolik, R.; Bugaj, B.; Chaszczewska-Markowska, M.; Jeka, S.; Bogunia-Kubik, K. IL-33 Gene Polymorphisms as Potential Biomarkers of Disease Susceptibility and Response to TNF Inhibitors in Rheumatoid Arthritis, Ankylosing Spondylitis, and Psoriatic Arthritis Patients. *Front. Immunol.* 2021, 12, 631603. [CrossRef] [PubMed]
- 44. Ruan, W.F.; Xie, J.T.; Jin, Q.; Wang, W.D.; Ping, A.S. The Diagnostic and Prognostic Role of Interleukin 12B and Interleukin 6R Gene Polymorphism in Patients With Ankylosing Spondylitis. *J. Clin. Rheumatol.* **2018**, *24*, 18–24. [CrossRef] [PubMed]
- Sagiv, M.; Adawi, M.; Awisat, A.; Shouval, A.; Peri, R.; Sabbah, F.; Rosner, I.; Kessel, A.; Slobodin, G. The association between elevated serum interleukin-22 and the clinical diagnosis of axial spondyloarthritis: A retrospective study. *Int. J. Rheum. Dis.* 2022, 25, 56–60. [CrossRef] [PubMed]
- Saif, D.S.; El Tabl, M.A.; Afifi, N.; Abdallah, M.S.; El Hefnawy, S.M.; Hassanein, S.A. Interleukin-17A biomarker as a predictor for detection of early axial spondyloarthritis changes in patients with psoriasis. *Int. J. Rheum. Dis.* 2020, 23, 1664–1669. [CrossRef]
- 47. Sode, J.; Bank, S.; Vogel, U.; Andersen, P.S.; Sørensen, S.B.; Bojesen, A.B.; Andersen, M.R.; Brandslund, I.; Dessau, R.B.; Hoffmann, H.J.; et al. Genetically determined high activities of the TNF-alpha, IL23/IL17, and NFkB pathways were associated with increased risk of ankylosing spondylitis. *BMC Med. Genet.* **2018**, *19*, 165. [CrossRef]

- Park, J.H.; Lee, S.G.; Jeon, Y.K.; Park, E.K.; Suh, Y.S.; Kim, H.O. Relationship between serum adipokine levels and radiographic progression in patients with ankylosing spondylitis: A preliminary 2-year longitudinal study. *Medicine* 2017, 96, e7854. [CrossRef]
- Rademacher, J.; Tietz, L.M. Added value of biomarkers compared with clinical parameters for the prediction of radiographic spinal progression in axial spondyloarthritis. *Rheumatology* 2019, *58*, 1556–1564. [CrossRef]
   The the test of the prediction of the predictin of the prediction of the predictin of the prediction of the p
- Zhong, H.; Zhong, M. LINC00311 is overexpressed in ankylosing spondylitis and predict treatment outcomes and recurrence. BMC Musculoskelet. Disord. 2019, 20, 278. [CrossRef]
- 51. Tsui, F.W.L.; Lin, A.; Sari, I.; Zhang, Z.; Tsui, H.W.; Inman, R.D. Serial Lipocalin 2 and Oncostatin M levels reflect inflammation status and treatment response in axial spondyloarthritis. *Arthritis Res. Ther.* **2021**, *23*, 141. [CrossRef]
- 52. Hušáková, M.; Bay-Jensen, A.C.; Forejtová, Š.; Zegzulková, K.; Tomčík, M.; Gregová, M.; Bubová, K.; Hořínková, J.; Gatterová, J.; Pavelka, K.; et al. Metabolites of type I, II, III, and IV collagen may serve as markers of disease activity in axial spondyloarthritis. *Sci. Rep.* 2019, 9, 11218. [CrossRef]
- 53. Ruiz-Limon, P.; Ladehesa-Pineda, M.L.; Castro-Villegas, M.D.C.; Abalos-Aguilera, M.D.C.; Lopez-Medina, C.; Lopez-Pedrera, C.; Barbarroja, N.; Espejo-Peralbo, D.; Gonzalez-Reyes, J.A.; Villalba, J.M.; et al. Enhanced NETosis generation in radiographic axial spondyloarthritis: Utility as biomarker for disease activity and anti-TNF-α therapy effectiveness. *J. Biomed. Sci.* 2020, 27, 54. [CrossRef]
- Genre, F.; Rueda-Gotor, J.; Remuzgo-Martínez, S.; Pulito-Cueto, V.; Corrales, A.; Mijares, V.; Lera-Gómez, L.; Portilla, V.; Expósito, R.; Mata, C.; et al. Omentin: A biomarker of cardiovascular risk in individuals with axial spondyloarthritis. *Sci. Rep.* 2020, 10, 9636. [CrossRef] [PubMed]
- Genre, F.; Rueda-Gotor, J.; Remuzgo-Martínez, S.; Corrales, A.; Ubilla, B.; Mijares, V.; Fernández-Díaz, C.; Portilla, V.; Blanco, R.; Hernández, J.L.; et al. Implication of osteoprotegerin and sclerostin in axial spondyloarthritis cardiovascular disease: Study of 163 Spanish patients. *Clin. Exp. Rheumatol.* 2018, *36*, 302–309. [PubMed]
- Nisihara, R.; Skare, T.L.; Zeni, J.O.; Rasera, H.; Lidani, K.; Messias-Reason, I. Plasma levels of pentraxin 3 in patients with spondyloarthritis. *Biomarkers* 2018, 23, 14–17. [CrossRef] [PubMed]
- Li, X.; Liang, A.; Chen, Y.; Lam, N.S.; Long, X.; Xu, X.; Zhong, S. Procollagen I N-terminal peptide correlates with inflammation on sacroiliac joint magnetic resonance imaging in ankylosing spondylitis but not in non-radiographic axial spondyloarthritis: A cross-sectional study. *Mod. Rheumatol.* 2022, *32*, 770–775. [CrossRef]
- Luchetti, M.M.; Ciccia, F.; Avellini, C.; Benfaremo, D.; Guggino, G.; Farinelli, A.; Ciferri, M.; Rossini, M.; Svegliati, S.; Spadoni, T.; et al. Sclerostin and Antisclerostin Antibody Serum Levels Predict the Presence of Axial Spondyloarthritis in Patients with Inflammatory Bowel Disease. *J. Rheumatol.* 2018, 45, 630–637. [CrossRef]
- 59. Perrotta, F.M.; Ceccarelli, F.; Barbati, C.; Colasanti, T.; De Socio, A.; Scriffignano, S.; Alessandri, C.; Lubrano, E. Serum Sclerostin as a Possible Biomarker in Ankylosing Spondylitis: A Case-Control Study. J. Immunol. Res. 2018, 2018, 9101964. [CrossRef]
- Liao, H.T.; Lin, Y.F.; Chou, C.T.; Tsai, C.Y. Semaphorin 3A in Ankylosing Spondylitis. J. Microbiol. Immunol. Infect. 2019, 52, 151–157. [CrossRef]
- Perrotta, F.M.; Ceccarelli, F.; Barbati, C.; Colasanti, T.; Montepaone, M.; Alessandri, C.; Valesini, G.; Lubrano, E. Assessment of semaphorin 3A and its role in bone remodelling in a group of ankylosing spondylitis patients. *Clin. Exp. Rheumatol.* 2017, 35, 313–316.
- 62. Liu, S.; Ji, W.; Lu, J.; Tang, X.; Guo, Y.; Ji, M.; Xu, T.; Gu, W.; Kong, D.; Shen, Q. Discovery of Potential Serum Protein Biomarkers in Ankylosing Spondylitis Using Tandem Mass Tag-Based Quantitative Proteomics. J. Proteome Res. 2020, 19, 864–872. [CrossRef]
- Jarlborg, M.; Courvoisier, D.S.; Lamacchia, C.; Martinez Prat, L.; Mahler, M.; Bentow, C.; Finckh, A.; Gabay, C.; Nissen, M.J. Serum calprotectin: A promising biomarker in rheumatoid arthritis and axial spondyloarthritis. *Arthritis Res. Ther.* 2020, 22, 105. [CrossRef] [PubMed]
- 64. Hu, H.; Du, F.; Zhang, S.; Zhang, W. Serum calprotectin correlates with risk and disease severity of ankylosing spondylitis and its change during first month might predict favorable response to treatment. *Mod. Rheumatol.* **2019**, *29*, 836–842. [CrossRef] [PubMed]
- Genre, F.; Rueda-Gotor, J.; Remuzgo-Martínez, S.; Corrales, A.; Mijares, V.; Expósito, R.; Mata, C.; Portilla, V.; Blanco, R.; Hernández, J.L.; et al. Association of circulating calprotectin with lipid profile in axial spondyloarthritis. *Sci. Rep.* 2018, *8*, 13728. [CrossRef] [PubMed]
- 66. Rademacher, J.; Siderius, M.; Gellert, L.; Wink, F.R.; Verba, M.; Maas, F.; Tietz, L.M.; Poddubnyy, D.; Spoorenberg, A.; Arends, S. Baseline serum biomarkers of inflammation, bone turnover and adipokines predict spinal radiographic progression in ankylosing spondylitis patients on TNF inhibitor therapy. *Semin. Arthritis Rheum.* 2022, 53, 151974. [CrossRef] [PubMed]
- Kook, H.Y.; Jin, S.H.; Park, P.R.; Lee, S.J.; Shin, H.J.; Kim, T.J. Serum miR-214 as a novel biomarker for ankylosing spondylitis. *Int. J. Rheum. Dis.* 2019, 22, 1196–1201. [CrossRef]
- Bubová, K.; Prajzlerová, K.; Hulejová, H.; Gregová, M.; Mintálová, K.; Hušáková, M.; Forejtová, Š.; Filková, M.; Tomčík, M.; Vencovský, J.; et al. Elevated Tenascin-C Serum Levels in Patients with Axial Spondyloarthritis. *Physiol. Res.* 2020, 69, 653–660. [CrossRef]
- 69. Gupta, L.; Bhattacharya, S.; Aggarwal, A. Tenascin-C, a biomarker of disease activity in early ankylosing spondylitis. *Clin. Rheumatol.* **2018**, *37*, 1401–1405. [CrossRef]
- 70. Rueda-Gotor, J.; López-Mejías, R.; Remuzgo-Martínez, S.; Pulito-Cueto, V.; Corrales, A.; Lera-Gómez, L.; Portilla, V.; González-Mazón, Í.; Blanco, R.; Expósito, R.; et al. Vaspin in atherosclerotic disease and cardiovascular risk in axial spondyloarthritis: A genetic and serological study. *Arthritis Res. Ther.* 2021, 23, 111. [CrossRef]

- 71. Siebuhr, A.S.; Hušaková, M.; Forejtová, S.; Zegzulková, K.; Tomčik, M.; Urbanová, M.; Grobelná, K.; Gatterová, J.; Bay-Jensen, A.C.; Pavelka, K. Metabolites of C-reactive protein and vimentin are associated with disease activity of axial spondyloarthritis. *Clin. Exp. Rheumatol.* 2019, 37, 358–366.
- Baykara, A.R.; Küçük, A.; Tuzcu, A.; Tuzcu, G.; Cüre, E.; Uslu, A.U.; Omma, A. The relationship of serum visfatin levels with clinical parameters, flow-mediated dilation, and carotid intima-media thickness in patients with ankylosing spondylitis. *Turk. J. Med. Sci.* 2021, *51*, 1865–1874. [CrossRef]
- 73. Proft, F.; Muche, B.; Schmidt, M.; Spiller, L.; Rodriguez, V.R.; Rademacher, J.; Weber, A.-K.; Lüders, S.; Protopopov, M.; Spiller, I.; et al. FRI0194 Ankylosing spondylitis disease activity score (ASDAS) based on a quick quantitative crp assay performs similarly well to asdas based on conventional crp in patients with axial spondyloarthritis. *Ann. Rheum. Dis.* **2018**, *77*, 637–638.
- 74. Smoldovskaya, O.V.; Voloshin, S.A.; Novikov, A.A.; Aleksandrova, E.N.; Feyzkhanova, G.U.; Rubina, A.Y. Adaptation of Microarray Assay for Serum Amyloid A Analysis in Human Serum. *Mol. Biol.* **2022**, *56*, 336–342. [CrossRef] [PubMed]
- 75. Sorić Hosman, I.; Kos, I.; Lamot, L. Serum Amyloid A in Inflammatory Rheumatic Diseases: A Compendious Review of a Renowned Biomarker. *Front. Immunol.* 2021, *11*, 631299. [CrossRef] [PubMed]
- Lorenzin, M.; Ometto, F.; Ortolan, A.; Felicetti, M.; Favero, M.; Doria, A.; Ramonda, R. An update on serum biomarkers to assess axial spondyloarthritis and to guide treatment decision. *Ther. Adv. Musculoskelet. Dis.* 2020, 12, 1759720X20934277. [CrossRef]
- 77. Inoue, K.; Kodama, T.; Daida, H. Pentraxin 3: A novel biomarker for inflammatory cardiovascular disease. *Int. J. Vasc. Med.* 2012, 2012, 657025. [CrossRef]
- Özdemirel, A.E.; Güven, S.C.; Sunar, İ.; Sari Sürmeli, Z.; Doğanci, A.; Tutkak, H.; Yalçin Sayin, A.P.; Ataman, Ş. The Relationship between Plasma Pentraxin 3 and Serum Amyloid P Levels and Disease Activity in Ankylosing Spondylitis. *Int. J. Clin. Pract.* 2022, 2022, 7243399. [CrossRef]
- 79. Qi, Q.; Geng, Y.; Sun, M.; Chen, H.; Wang, P.; Chen, Z. Hyperfibrinogen Is Associated with the Systemic Inflammatory Response and Predicts Poor Prognosis in Advanced Pancreatic Cancer. *Pancreas* 2015, 44, 977–982. [CrossRef]
- 80. Keller, U. Nutritional Laboratory Markers in Malnutrition. J. Clin. Med. 2019, 8, 775. [CrossRef]
- Singh, V.; Yeoh, B.S.; Chassaing, B.; Zhang, B.; Saha, P.; Xiao, X.; Awasthi, D.; Shashidharamurthy, R.; Dikshit, M.; Gewirtz, A.; et al. Microbiota-inducible Innate Immune, Siderophore Binding Protein Lipocalin 2 is Critical for Intestinal Homeostasis. *Cell. Mol. Gastroenterol. Hepatol.* 2016, 2, 482–498. [CrossRef]
- 82. West, N.R.; Hegazy, A.N.; Owens, B.M.J.; Bullers, S.J.; Linggi, B.; Buonocore, S.; Coccia, M.; Görtz, D.; This, S.; Stockenhuber, K.; et al. Oncostatin M drives intestinal inflammation and predicts response to tumor necrosis factor-neutralizing therapy in patients with inflammatory bowel disease. *Nat. Med.* **2017**, *23*, 579–589. [CrossRef]
- Moschen, A.R.; Adolph, T.E.; Gerner, R.R.; Wieser, V.; Tilg, H. Lipocalin-2: A Master Mediator of Intestinal and Metabolic Inflammation. *Trends Endocrinol. Metab.* 2017, 28, 388–397. [CrossRef] [PubMed]
- 84. Mancuso, P. The role of adipokines in chronic inflammation. Immunotargets Ther. 2016, 5, 47–56. [CrossRef] [PubMed]
- 85. Saddi-Rosa, P.; Oliveira, C.S.; Giuffrida, F.M.; Reis, A.F. Visfatin, glucose metabolism and vascular disease: A review of evidence. *Diabetol. Metab. Syndr.* 2010, 2, 21. [CrossRef]
- Watanabe, T.; Watanabe-Kominato, K.; Takahashi, Y.; Kojima, M.; Watanabe, R. Adipose Tissue-Derived Omentin-1 Function and Regulation. *Compr. Physiol.* 2017, 7, 765–781. [PubMed]
- 87. Rao, S.S.; Hu, Y.; Xie, P.L.; Cao, J.; Wang, Z.X.; Liu, J.H.; Yin, H.; Huang, J.; Tan, Y.J.; Luo, J.; et al. Omentin-1 prevents inflammation-induced osteoporosis by downregulating the pro-inflammatory cytokines. *Bone Res.* **2018**, *6*, 9. [CrossRef]
- 88. Zhang, J.M.; An, J. Cytokines, inflammation, and pain. Int. Anesthesiol. Clin. 2007, 45, 27–37. [CrossRef]
- 89. Furst, D.E.; Louie, J.S. Targeting inflammatory pathways in axial spondyloarthritis. Arthritis Res. Ther. 2019, 21, 135. [CrossRef]
- 90. Jethwa, H.; Bowness, P. The interleukin (IL)-23/IL-17 axis in ankylosing spondylitis: New advances and potentials for treatment. *Clin. Exp. Immunol.* **2016**, *183*, 30–36. [CrossRef]
- 91. Walsham, N.E.; Sherwood, R.A. Fecal calprotectin in inflammatory bowel disease. Clin. Exp. Gastroenterol. 2016, 9, 21–29.
- 92. Romand, X.; Bernardy, C.; Nguyen, M.V.C.; Courtier, A.; Trocme, C.; Clapasson, M.; Paclet, M.H.; Toussaint, B.; Gaudin, P. Systemic calprotectin and chronic inflammatory rheumatic diseases. *Jt. Bone Spine* **2019**, *86*, 691–698. [CrossRef]
- Turina, M.C.; Sieper, J.; Yeremenko, N.; Conrad, K.; Haibel, H.; Rudwaleit, M.; Baeten, D.; Poddubnyy, D. Calprotectin serum level is an independent marker for radiographic spinal progression in axial spondyloarthritis. *Ann. Rheum. Dis.* 2014, 73, 1746–1748. [CrossRef] [PubMed]
- 94. Duran, A.; Kobak, S.; Sen, N.; Aktakka, S.; Atabay, T.; Orman, M. Fecal calprotectin is associated with disease activity in patients with ankylosing spondylitis. *Bosn. J. Basic Med. Sci.* 2016, 16, 71–74. [CrossRef] [PubMed]
- Li, T.; Zhang, Z.; Li, X.; Dong, G.; Zhang, M.; Xu, Z.; Yang, J. Neutrophil Extracellular Traps: Signaling Properties and Disease Relevance. *Mediat. Inflamm.* 2020, 2020, 9254087. [CrossRef]
- Dong, R.; Zhang, M.; Hu, Q.; Zheng, S.; Soh, A.; Zheng, Y.; Yuan, H. Galectin-3 as a novel biomarker for disease diagnosis and a target for therapy (Review). *Int. J. Mol. Med.* 2018, 41, 599–614. [CrossRef] [PubMed]
- Omran, A.; Atanasova, D.; Landgren, F.; Magnusson, P. Sclerostin: From Molecule to Clinical Biomarker. Int. J. Mol. Sci. 2022, 23, 4751. [CrossRef]
- Baron, R.; Rawadi, G. Targeting the Wnt/beta-catenin pathway to regulate bone formation in the adult skeleton. *Endocrinology* 2007, 148, 2635–2643. [CrossRef]

- 99. Ito, N.; Prideaux, M.; Wijenayaka, A.R.; Yang, D.; Ormsby, R.T.; Bonewald, L.F.; Atkins, G.J. Sclerostin Directly Stimulates Osteocyte Synthesis of Fibroblast Growth Factor-23. *Calcif. Tissue Int.* **2021**, *109*, 66–76. [CrossRef]
- 100. Appel, H.; Ruiz-Heiland, G.; Listing, J.; Zwerina, J.; Herrmann, M.; Mueller, R.; Haibel, H.; Baraliakos, X.; Hempfing, A.; Rudwaleit, M.; et al. Altered skeletal expression of sclerostin and its link to radiographic progression in ankylosing spondylitis. *Arthritis Rheum.* 2009, 60, 3257–3262. [CrossRef]
- Mihara, A.; Yukata, K.; Seki, T.; Iwanaga, R.; Nishida, N.; Fujii, K.; Nagao, Y.; Sakai, T. Effects of sclerostin antibody on bone healing. World J. Orthop. 2021, 12, 651–659. [CrossRef]
- 102. Aluganti Narasimhulu, C.; Singla, D.K. The Role of Bone Morphogenetic Protein 7 (BMP-7) in Inflammation in Heart Diseases. *Cells* 2020, *9*, 280. [CrossRef]
- 103. Mattey, D.L.; Packham, J.C.; Nixon, N.B.; Coates, L.; Creamer, P.; Hailwood, S.; Taylor, G.J.; Bhalla, A.K. Association of cytokine and matrix metalloproteinase profiles with disease activity and function in ankylosing spondylitis. *Arthritis Res. Ther.* 2012, 14, R127. [CrossRef] [PubMed]
- 104. Siebuhr, A.S.; van der Heijde, D.; Bay-Jensen, A.C.; Karsdal, M.A.; Landewé, R.; van Tubergen, A.; Ramiro, S. Is radiographic progression in radiographic axial spondyloarthritis related to matrix metalloproteinase degradation of extracellular matrix? *RMD Open* 2018, 4, 000648. [CrossRef] [PubMed]
- Kučukalić-Selimović, E.; Valjevac, A.; Hadžović-Džuvo, A. The utility of procollagen type 1 N-terminal propeptide for the bone status assessment in postmenopausal women. *Bosn. J. Basic Med. Sci.* 2013, 13, 259–265. [CrossRef] [PubMed]
- 106. Koivula, M.K.; Risteli, L.; Risteli, J. Measurement of aminoterminal propeptide of type I procollagen (PINP) in serum. *Clin. Biochem.* **2012**, *45*, 920–927. [CrossRef] [PubMed]
- Ivaska, J.; Pallari, H.M.; Nevo, J.; Eriksson, J.E. Novel functions of vimentin in cell adhesion, migration, and signaling. *Exp. Cell Res.* 2007, 313, 2050–2062. [CrossRef]
- 108. Bay-Jensen, A.C.; Karsdal, M.A.; Vassiliadis, E.; Wichuk, S.; Marcher-Mikkelsen, K.; Lories, R.; Christiansen, C.; Maksymowych, W.P. Circulating citrullinated vimentin fragments reflect disease burden in ankylosing spondylitis and have prognostic capacity for radiographic progression. *Arthritis Rheum.* 2013, 65, 972–980. [CrossRef]
- Klingberg, E.; Nurkkala, M.; Carlsten, H.; Forsblad-d'Elia, H. Biomarkers of bone metabolism in ankylosing spondylitis in relation to osteoproliferation and osteoporosis. J. Rheumatol. 2014, 41, 1349–1356. [CrossRef]
- 110. Yilmaz, Y.; Kurt, R.; Eren, F.; Imeryuz, N. Serum osteocalcin levels in patients with nonalcoholic fatty liver disease: Association with ballooning degeneration. *Scand. J. Clin. Lab. Investig.* **2011**, *71*, 631–636. [CrossRef]
- 111. Kwon, S.R.; Lim, M.J.; Suh, C.H.; Park, S.G.; Hong, Y.S.; Yoon, B.Y.; Kim, H.A.; Choi, H.J.; Park, W. Dickkopf-1 level is lower in patients with ankylosing spondylitis than in healthy people and is not influenced by anti-tumor necrosis factor therapy. *Rheumatol. Int.* 2012, *32*, 2523–2527. [CrossRef]
- 112. Pavliuk, O.; Shevchuk, S. Levels of osteocalcin and procollagen i n-terminal propeptide (pinp) in men suffering from ankylosing spondylitis. *Wiad. Lek.* **2021**, *74*, 2384–2391. [CrossRef]
- 113. Wu, M.; Chen, M.; Ma, Y.; Yang, J.; Han, R.; Yuan, Y.; Hu, X.; Wang, M.; Zhang, X.; Xu, S.; et al. Dickkopf-1 in ankylosing spondylitis: Review and meta-analysis. *Clin. Chim. Acta* 2018, 481, 177–183. [CrossRef] [PubMed]
- 114. Garcia, S. Role of Semaphorins in Immunopathologies and Rheumatic Diseases. Int. J. Mol. Sci. 2019, 20, 374. [CrossRef] [PubMed]
- McGovern, K.E.; Wilson, E.H. Role of Chemokines and Trafficking of Immune Cells in Parasitic Infections. *Curr. Immunol. Rev.* 2013, 9, 157–168. [CrossRef] [PubMed]
- 116. Ivanovska, M.; Abdi, Z.; Murdjeva, M.; Macedo, D.; Maes, A.; Maes, M. CCL-11 or Eotaxin-1: An Immune Marker for Ageing and Accelerated Ageing in Neuro-Psychiatric Disorders. *Pharmaceuticals* **2020**, *13*, 230. [CrossRef] [PubMed]
- 117. Kindstedt, E.; Holm, C.K.; Sulniute, R.; Martinez-Carrasco, I.; Lundmark, R.; Lundberg, P. CCL11, a novel mediator of inflammatory bone resorption. *Sci. Rep.* 2017, *7*, 5334. [CrossRef]
- 118. Kalayci, M.; Gul, E. Eotaxin-1 Levels in Patients with Myocardial Infarction. Clin. Lab. 2022, 68, 3. [CrossRef]
- Komsalova, L.Y.; Martínez Salinas, M.P.; Jiménez, J.F.G. Predictive values of inflammatory back pain, positive HLA B27 antigen and acute and chronic magnetic resonance changes in early diagnosis of Spondyloarthritis. A study of 133 patients. *PLoS ONE* 2020, 15, e0244184. [CrossRef]
- 120. Rudwaleit, M.; Claudepierre, P.; Wordsworth, P.; Cortina, E.L.; Sieper, J.; Kron, M.; Carcereri-De-Prati, R.; Kupper, H.; Kary, S. Effectiveness, safety, and predictors of good clinical response in 1250 patients treated with adalimumab for active ankylosing spondylitis. *J. Rheumatol.* **2009**, *36*, 801–808. [CrossRef]
- 121. Robinson, P.C.; Claushuis, T.A.; Cortes, A.; Martin, T.M.; Evans, D.M.; Leo, P.; Mukhopadhyay, P.; Bradbury, L.A.; Cremin, K.; Harris, J.; et al. Genetic dissection of acute anterior uveitis reveals similarities and differences in associations observed with ankylosing spondylitis. *Arthritis Rheumatol.* 2015, 67, 140–151. [CrossRef]
- Lin, H.; Gong, Y.Z. Reply to "Association of HLA-B27 and its subtypes with ankylosing spondylitis and clinical manifestations of ankylosing spondylitis in different HLA-B27 subtypes: Comment on the article by Lin et al.". *Rheumatol. Int.* 2017, 37, 1685. [CrossRef]
- 123. Li, M.Y.; Yao, Z.Q.; Liu, X.Y. Advance of research on HLA-B27 and the immunological mechanism of ankylosing spondylitis. *Sheng Li Ke Xue Jin Zhan* **2011**, *42*, 16–20. [PubMed]
- 124. Caba, L.; Florea, L.; Gug, C.; Dimitriu, D.C.; Gorduza, E.V. Circular RNA-Is the Circle Perfect? *Biomolecules* 2021, *11*, 1755. [CrossRef] [PubMed]

- 125. Kumar, L.; Shamsuzzama; Jadiya, P.; Haque, R.; Shukla, S.; Nazir, A. Functional Characterization of Novel Circular RNA Molecule, circzip-2 and Its Synthesizing Gene zip-2 in C. elegans Model of Parkinson's Disease. *Mol. Neurobiol.* 2018, 55, 6914–6926. [CrossRef] [PubMed]
- 126. Qu, S.; Yang, X.; Li, X.; Wang, J.; Gao, Y.; Shang, R.; Sun, W.; Dou, K.; Li, H. Circular RNA: A new star of noncoding RNAs. *Cancer Lett.* **2015**, *365*, 141–148. [CrossRef]
- 127. Wang, L.; Long, H.; Zheng, Q.; Bo, X.; Xiao, X.; Li, B. Circular RNA circRHOT1 promotes hepatocellular carcinoma progression by initiation of NR2F6 expression. *Mol. Cancer* **2019**, *18*, 119. [CrossRef]
- Ouyang, Q.; Wu, J.; Jiang, Z.; Zhao, J.; Wang, R.; Lou, A.; Zhu, D.; Shi, G.P.; Yang, M. Microarray Expression Profile of Circular RNAs in Peripheral Blood Mononuclear Cells from Rheumatoid Arthritis Patients. *Cell. Physiol. Biochem.* 2017, 42, 651–659. [CrossRef]
- Luo, Q.; Zhang, L.; Li, X.; Fu, B.; Deng, Z.; Qing, C.; Su, R.; Xu, J.; Guo, Y.; Huang, Z.; et al. Identification of circular RNAs hsa\_circ\_0044235 in peripheral blood as novel biomarkers for rheumatoid arthritis. *Clin. Exp. Immunol.* 2018, 194, 118–124. [CrossRef]
- 130. Luo, Q.; Li, X.; Fu, B.; Zhang, L.; Fang, L.; Qing, C.; Guo, Y.; Huang, Z.; Li, J. Expression profile and diagnostic value of circRNAs in peripheral blood from patients with systemic lupus erythematosus. *Mol. Med. Rep.* **2021**, *23*, 1. [CrossRef]
- Zhang, Y.; Li, Q.; Liu, C.; Gao, S.; Ping, H.; Wang, J.; Wang, P. MiR-214-3p attenuates cognition defects via the inhibition of autophagy in SAMP8 mouse model of sporadic Alzheimer's disease. *Neurotoxicology* 2016, 56, 139–149. [CrossRef]
- 132. Li, D.; Liu, J.; Guo, B.; Liang, C.; Dang, L.; Lu, C.; He, X.; Cheung, H.Y.; Xu, L.; Lu, C. Osteoclast-derived exosomal miR-214-3p inhibits osteoblastic bone formation. *Nat. Commun.* **2016**, *7*, 10872. [CrossRef]
- 133. Yan, J.; Parekh, V.V.; Mendez-Fernandez, Y.; Olivares-Villagómez, D.; Dragovic, S.; Hill, T.; Roopenian, D.C.; Joyce, S.; Van Kaer, L. In vivo role of ER-associated peptidase activity in tailoring peptides for presentation by MHC class Ia and class Ib molecules. J. Exp. Med. 2006, 203, 647–659. [CrossRef] [PubMed]
- Haroon, N.; Tsui, F.W.; Chiu, B.; Tsui, H.W.; Inman, R.D. Serum cytokine receptors in ankylosing spondylitis: Relationship to inflammatory markers and endoplasmic reticulum aminopeptidase polymorphisms. J. Rheumatol. 2010, 37, 1907–1910. [CrossRef] [PubMed]
- Li, L.; Batliwala, M.; Bouvier, M. ERAP1 enzyme-mediated trimming and structural analyses of MHC I-bound precursor peptides yield novel insights into antigen processing and presentation. J. Biol. Chem. 2019, 294, 18534–18544. [CrossRef] [PubMed]
- Martín-Esteban, A.; Guasp, P.; Barnea, E.; Admon, A.; López de Castro, J.A. Functional Interaction of the Ankylosing Spondylitis-Associated Endoplasmic Reticulum Aminopeptidase 2 With the HLA-B\*27 Peptidome in Human Cells. *Arthritis Rheumatol.* 2016, 68, 2466–2475. [CrossRef] [PubMed]
- 137. Lalevée, S.; Feil, R. Long noncoding RNAs in human disease: Emerging mechanisms and therapeutic strategies. *Epigenomics* **2015**, 7, 877–879. [CrossRef] [PubMed]
- Wang, Y.; Luo, T.B.; Liu, L.; Cui, Z.Q. LncRNA LINC00311 Promotes the Proliferation and Differentiation of Osteoclasts in Osteoporotic Rats Through the Notch Signaling Pathway by Targeting DLL3. *Cell. Physiol. Biochem.* 2018, 47, 2291–2306. [CrossRef] [PubMed]
- 139. Hassan, M.K.; Kumar, D.; Naik, M.; Dixit, M. The expression profile and prognostic significance of eukaryotic translation elongation factors in different cancers. *PLoS ONE* **2018**, *13*, e0191377. [CrossRef] [PubMed]
- 140. Yu, X.; Zheng, H.; Chan, M.T.; Wu, W.K. HULC: An oncogenic long non-coding RNA in human cancer. J. Cell. Mol. Med. 2017, 21, 410–417. [CrossRef]
- Ozgocmen, S.; Godekmerdan, A.; Ozkurt-Zengin, F. Acute-phase response, clinical measures and disease activity in ankylosing spondylitis. *Jt. Bone Spine* 2007, 74, 249–253. [CrossRef]
- Aimo, A.; Vergaro, G.; Passino, C.; Ripoli, A.; Ky, B.; Miller, W.L.; Bayes-Genis, A.; Anand, I.; Januzzi, J.L.; Emdin, M. Prognostic Value of Soluble Suppression of Tumorigenicity-2 in Chronic Heart Failure: A Meta-Analysis. *JACC Heart Fail.* 2017, *5*, 280–286. [CrossRef]
- 143. Demyanets, S.; Kaun, C.; Kaider, A.; Speidl, W.; Prager, M.; Oravec, S.; Hohensinner, P.; Wojta, J.; Rega-Kaun, G. The proinflammatory marker soluble suppression of tumorigenicity-2 (ST2) is reduced especially in diabetic morbidly obese patients undergoing bariatric surgery. *Cardiovasc. Diabetol.* **2020**, *19*, 26. [CrossRef] [PubMed]
- 144. De Boer, R.A.; Daniels, L.B.; Maisel, A.S.; Januzzi, J.L., Jr. State of the Art: Newer biomarkers in heart failure. *Eur. J. Heart Fail.* 2015, 17, 559–569. [CrossRef] [PubMed]
- 145. Roldan, C.A. Valvular and coronary heart disease in systemic inflammatory diseases: Systemic Disorders in heart disease. *Heart* 2008, 94, 1089–1101. [CrossRef] [PubMed]
- 146. Noer, H.R. An "experimental" epidemic of Reiter's syndrome. JAMA 1966, 198, 693–698. [CrossRef]
- 147. Gill, T.; Asquith, M.; Rosenbaum, J.T.; Colbert, R.A. The intestinal microbiome in spondyloarthritis. *Curr. Opin. Rheumatol.* 2015, 27, 319–325. [CrossRef]
- 148. Tiffany, C.R.; Bäumler, A.J. Dysbiosis: From fiction to function. Am. J. Physiol. Gastrointest. Liver Physiol. 2019, 317, 602–608. [CrossRef]
- Ciccia, F.; Ferrante, A.; Triolo, G. Intestinal dysbiosis and innate immune responses in axial spondyloarthritis. *Curr. Opin. Rheumatol.* 2016, 28, 352–358. [CrossRef]
- Sagard, J.; Olofsson, T.; Mogard, E.; Marsal, J.; Andréasson, K.; Geijer, M.; Kristensen, L.E.; Lindqvist, E.; Wallman, J.K. Gut dysbiosis associated with worse disease activity and physical function in axial spondyloarthritis. *Arthritis Res. Ther.* 2022, 24, 42. [CrossRef]

- Gill, T.; Brooks, S.R.; Rosenbaum, J.T.; Asquith, M.; Colbert, R.A. Novel Inter-omic Analysis Reveals Relationships Between Diverse Gut Microbiota and Host Immune Dysregulation in HLA-B27-Induced Experimental Spondyloarthritis. *Arthritis Rheumatol.* 2019, 71, 1849–1857. [CrossRef]
- 152. Oikonomopoulou, K.; Ricklin, D.; Ward, P.A.; Lambris, J.D. Interactions between coagulation and complement–their role in inflammation. *Semin. Immunopathol.* **2012**, *34*, 151–165. [CrossRef]
- 153. Phillips, C.M.; Kesse-Guyot, E.; Ahluwalia, N.; McManus, R.; Hercberg, S.; Lairon, D.; Planells, R.; Roche, H.M. Dietary fat, abdominal obesity and smoking modulate the relationship between plasma complement component 3 concentrations and metabolic syndrome risk. *Atherosclerosis* **2012**, *220*, 513–519. [CrossRef] [PubMed]
- Barbu, A.; Hamad, O.A.; Lind, L.; Ekdahl, K.N.; Nilsson, B. The role of complement factor C3 in lipid metabolism. *Mol. Immunol.* 2015, 67, 101–107. [PubMed]
- 155. Midwood, K.S.; Orend, G. The role of tenascin-C in tissue injury and tumorigenesis. *J. Cell Commun. Signal.* **2009**, *3*, 287–310. [CrossRef] [PubMed]
- Petrini, I. Biology of MET: A double life between normal tissue repair and tumor progression. *Ann. Transl. Med.* 2015, *3*, 82. [PubMed]
   Nakamura, T.; Sakai, K.; Nakamura, T.; Matsumoto, K. Hepatocyte growth factor twenty years on: Much more than a growth factor. *J. Gastroenterol. Hepatol.* 2011, *26*, 188–202. [CrossRef]
- 158. Grano, M.; Galimi, F.; Zambonin, G.; Colucci, S.; Cottone, E.; Zallone, A.Z.; Comoglio, P.M. Hepatocyte growth factor is a coupling factor for osteoclasts and osteoblasts in vitro. *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 7644–7648. [CrossRef] [PubMed]
- 159. Kuraya, M.; Ming, Z.; Liu, X.; Matsushita, M.; Fujita, T. Specific binding of L-ficolin and H-ficolin to apoptotic cells leads to complement activation. *Immunobiology* **2005**, 209, 689–697. [CrossRef] [PubMed]