



Commentary

The thymoma tale

Thymic epithelial tumours form the most important differential diagnosis for anterior mediastinal masses, among which thymomas constitute nearly 50 per cent of these lesions. These are infrequent and possess certain unique characteristics¹. The thymus plays a central role in the development of self-immunological tolerance by appropriate conditioning of T-cell progenitors that enter the gland. Consequently, thymic pathology (including thymomas) leads to the generation of a variety of autoantibodies. Hence, a significant proportion of patients with thymomas present with features of paraneoplastic syndromes (PNSs) to the extent of exclusion of chest-related symptoms. The most common manifestation is myasthenia gravis, induced by anti-acetylcholine esterase antibodies, identified in close to 95 per cent of patients with available clinical data in a large series of Indian patients published in this issue². It must be borne in mind that there are also other PNSs (including multiorgan autoimmunity), secondary cancers as well as opportunistic infections, which are explained in part by acquired T-cell immunodeficiency³. The presence of PNSs, paradoxically, was associated with favourable features⁴.

On histology, the thymomas, in general, are seen to be composed of epithelial cells and lymphocytes in varying proportions. However, this seemingly innocuous admixture has led to several classification systems over decades⁵. The classification used currently and in this article by Guleria *et al*⁶ is the WHO 2015⁶, which identifies five subtypes (A, AB, B₁, B₂ and B₃) depending on the morphology of the epithelial cells (spindloid cells, epithelioid cells or their combinations) and proportion of lymphocytes. The authors concluded that the thymomas of B₂ and AB subtypes were the most common in the Indian population, in contrast to the studies from the Western world. However, in a recent, multi-institutional study of 1470 patients with thymomas from 11 countries, a similar distribution was found⁷. Furthermore,

the Indian studies quoted² are either clinical (without a thorough histological review) or have been studies of ‘mixed bag’ of mediastinal tumours. Hence, to have an idea of the actual distribution, there is a need to have multi-institutional collaborations with thymomas in sufficient numbers. It would also be important to follow a protocol⁸ for clinicoradiological, gross and histological assessment of such tumours, as there are some studies indicating that even tumour sizes are useful in predicting prognosis⁹.

Another feature of thymomas is that the histotypes do not necessarily correlate with the clinical behaviour, prognosis and overall survival, which explains the use of the modified Masaoka-Koga staging system¹⁰. With time, there would presumably be more robust criteria for prognosis and therapeutic management such as utility of the angiogenesis patterns¹¹, immunoexpression of programmed death ligand¹², molecular profiling¹³ and even detection of proteomic signatures¹⁴.

Conflicts of Interest: None.

Pradeep Vaideeswar
Department of Pathology
(Cardiovascular & Thoracic Division),
Seth GS Medical College, Mumbai 400 012,
Maharashtra, India
shreeprajai@yahoo.co.in

Received January 28, 2019

References

1. den Bakker MA, Roden AC, Marx A, Marino M. Histologic classification of thymoma: a practical guide for routine cases. *J Thorac Oncol* 2014; 9 (9 Suppl 2) : S125-30.
2. Guleria P, Parshad R, Malik PS, Ray R, Pandey RM, Jain D. Histotyping of Indian thymomas: A clinicopathologic study from north India. *Indian J Med Res* 2019; 150 : 153-60.
3. Christopoulos P, Fisch P. Acquired T-cell immunodeficiency in thymoma patients. *Crit Rev Immunol* 2016; 36 : 315-27.

4. Padda SK, Yao X, Antonicelli A, Riess JW, Shang Y, Shrager JB, *et al.* Paraneoplastic syndromes and thymic malignancies: An examination of the International Thymic Malignancy Interest Group retrospective database. *J Thorac Oncol* 2018; 13 : 436-46.
5. Roden AC. Evolution of classification of thymic epithelial tumors in the era of Dr. Thomas V. Colby. *Arch Pathol Lab Med* 2017; 141 : 232-46.
6. Travis WD, Brambilla E, Burke AP, Marx A, Nicholson AG, editors. *WHO classification of tumours of lung, pleura, thymus and heart*. 4th ed. Geneva: WHO Press; 2015.
7. Weissferdt A, Kalhor N, Bishop JA, Jang SJ, Ro J, Petersson F, *et al.* Thymoma: A clinicopathological correlation of 1470 cases. *Hum Pathol* 2018; 73 : 7-15.
8. Nicholson AG, Detterbeck F, Marx A, Roden AC, Marchevsky AM, Mukai K, *et al.* Dataset for reporting of thymic epithelial tumours: Recommendations from the International Collaboration on Cancer Reporting (ICCR). *Histopathology* 2017; 70 : 522-38.
9. Bian D, Zhou F, Yang W, Zhang K, Chen L, Jiang G, *et al.* Thymoma size significantly affects the survival, metastasis and effectiveness of adjuvant therapies: A population based study. *Oncotarget* 2018; 9 : 12273-83.
10. Detterbeck FC, Nicholson AG, Kondo K, Van Schil P, Moran C. The Masaoka–Koga stage classification for thymic malignancies: Clarification and definition of terms. *J Thorac Oncol* 2011; 6 : S1710-6.
11. Pfister F, Hussain H, Belharazem D, Busch S, Simon-Keller K, Becker D, *et al.* Vascular architecture as a diagnostic marker for differentiation of World Health Organization thymoma subtypes and thymic carcinoma. *Histopathology* 2017; 70 : 693-703.
12. Duan J, Liu X, Chen H, Sun Y, Liu Y, Bai H, *et al.* Impact of PD-L1, transforming growth factor- β expression and tumor-infiltrating CD8+ T cells on clinical outcome of patients with advanced thymic epithelial tumors. *Thorac Cancer* 2018; 9 : 1341-53.
13. Enkner F, Pichlhöfer B, Zaharie AT, Kronic M, Holper TM, Janik S, *et al.* Molecular profiling of thymoma and thymic carcinoma: Genetic differences and potential novel therapeutic targets. *Pathol Oncol Res* 2017; 23 : 551-64.
14. Wang L, Branson OE, Shilo K, Hitchcock CL, Freitas MA. Proteomic signatures of thymomas. *PLoS One* 2016; 11 : e0166494.