Empirical or susceptibility-guided treatment for *Helicobacter pylori* infection? A comprehensive review

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Abstract: Although susceptibility-quided therapy is frequently recommended for *Helicobacter* pylori infection, the evidence available to date supporting this strategy is limited. The aim of the present article is to review the advantages and limitations of the susceptibility-guided and the empirical strategies to treat this infection. We performed a bibliographic search to identify studies investigating *H. pylori* susceptibility-quided therapy. Culture is not the only way to assess antibiotic resistance, as different polymerase chain reaction-based approaches have been developed as alternative methods. For detecting *H. pylori* antimicrobial resistance, a molecular approach based on a stool sample might enable more convenient, time-saving methods. Unfortunately, the antimicrobial susceptibility cannot be obtained in all cases. Furthermore, antibiotic susceptibility testing in clinical practice yields useful information only for a few antibiotics: clarithromycin, metronidazole, and quinolones. In addition, susceptibility towards clarithromycin and metronidazole in vitro does not necessarily lead to eradication in vivo. In the case of H. pylori therapy failure, we should not re-administer any of the antibiotics against which H. pylori has probably become resistant. Our updated meta-analysis showed that susceptibility-guided treatment is not better than empirical treatment of H. pylori infection in first-line therapy if the most updated guadruple regimens are empirically prescribed. and similar efficacy results were also demonstrated with the two strategies for secondline therapy. Cumulative H. pylori eradication rate with several successive rescue therapies empirically prescribed reaches almost 100%. Finally, the studies that have evaluated the cost-effectiveness of the susceptibility-guided treatment have achieved contradictory results. In summary, we can conclude that the evidence is too limited to support the generalized use of susceptibility-quided therapy for *H. pylori* treatment in routine clinical practice, either as firstline or as rescue treatment. Nevertheless, it would be recommended that susceptibility tests are performed routinely, even before prescribing first-line treatment, in specialized centers with an interest in *H. pylori* management.

Keywords: culture, empirical, Helicobacter pylori, susceptibility, tailored

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Introduction

Helicobacter pylori (*H. pylori*) infection affects billions of people worldwide, being the main cause of gastritis, peptic ulcer disease, and gastric cancer.¹ Antibiotic resistance is the major factor affecting our ability to cure *H. pylori* infection, and the rate of resistance to several antibiotics, mainly clarithromycin, is steadily increasing in

many geographic areas.^{2–5} A recent systematic review and meta-analysis assessed the distribution of *H. pylori* resistance to commonly used antibiotics in 65 countries, and found that primary resistance rates to clarithromycin, metronidazole, and levofloxacin were $\geq 15\%$ in most regions. Furthermore, increasing antibiotic resistance was observed in most regions.⁶ Accordingly,

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the World Health Organization has designated clarithromycin-resistant *H. pylori* a high priority for antibiotic research and development.

As antibiotic resistance is an evolving process, it seems mandatory to carry out point-prevalence surveys on a regular basis.³ A strategy that has been suggested to increase the eradication rate is to provide individualized (personalized) treatment according to antibiotic susceptibility testing. However, the true utility of performing antibiotic susceptibility testing and the moment when it must be performed (before the first treatment or only after eradication failure) are both controversial. Of note, H. pylori culture is time consuming, not always available on a routine basis, offering quite low sensitivity, and obviously implying the performance of an endoscopic exploration.^{7,8} Furthermore, culture is relatively expensive, not because of the cost of the procedure per se, but mainly because of the costs of the associated endoscopy required to obtain biopsy specimens.

Although susceptibility-guided therapy has been recommended in many *H. pylori* consensus conferences,^{9–11} the studies evaluating this strategy are, however, quite limited, and the evidence available to date about when and in whom culture should be performed is surprisingly scant. In fact, currently, most physicians treat *H. pylori* infection without relying on antimicrobial susceptibility testing to choose the best regimen.

The aim of the present article was to review the advantages and limitations of susceptibilityguided and empirical strategies to treat *H. pylori* infection.

Search strategy

Bibliographical searches were performed in the MEDLINE and EMBASE electronic database up to April 2020 based on the following words (all fields): pylori AND [(culture OR culture-based OR culture-guided OR tailored OR susceptibility OR susceptibility-guided OR "antimicrobial susceptibility" OR "susceptibility testing") OR (empiric OR empirical)]. Articles published in any language were included. Reference lists from the articles selected by electronic searching were examined in detail to further identify relevant studies. Abstracts of the articles selected in each of these multiple searches were reviewed, and those dealing with susceptibility-guided treatment of *H. pylori* infection were recorded. The number of articles identified with PubMed was 6355, and with EMBASE 9607; after excluding duplicates, 12,736 articles were finally identified and reviewed.

What are the main arguments for assessing the antibiotic susceptibility of *H. pylori*?

The proportion of patients who achieve *H. pylori* eradication depends mainly, on the prevalence of antimicrobial resistance in the particular population being treated. Therefore, if the proportion of resistant infections is unknown, the results in any population cannot be generalized to another population with a different proportion of antimicrobial resistance.¹² Empirical therapy that takes into consideration the local resistance patterns may be superior to predict the efficacy of any *H. pylori* eradication regimen. Therefore, the local resistance patterns and the efficacy rates in the context of a specific environment are essential for establishing a correct treatment of the infection in real-world settings.¹³

Furthermore, it has been suggested that it appears unjustified to prescribe an antibiotic that will lack efficacy, generate higher cost, and will induce adverse events. A disadvantage of empirical treatment is that it often contains three, partly unnecessary, antibiotics, implicating the misuse of antibiotics. Susceptibility testing has been proposed to reduce unnecessary antibiotic prescription.¹⁴ For example, performing antimicrobial susceptibility testing before first-line therapy might still allow the administration of the standard clarithromycin-based triple therapy to patients with an H. pylori clarithromycin-susceptible strain in areas with high overall clarithromycin resistance.15

Another obvious benefit of this strategy is that through the application of susceptibility testing prior to treatment, the development of antimicrobial resistance can be minimized,¹⁵ as antibiotic use in the outpatient community is positively correlated with antibiotic resistance.³ Emergence of antibiotic resistance after widespread use of antibiotics is an important concern especially for mass screening and eradication of *H. pylori* in asymptomatic subjects in the community. Nevertheless, the transient increase of antibiotic resistance to certain bacteria observed immediately after antibiotic treatment may be restored to basal state shortly (2 months) after *H. pylori* eradication treatment has been administered.¹⁶ Finally, it has been reported that there is a significant shortterm perturbation of gut microbiota after *H. pylori* eradication.^{17,18} However, the diversity of gut microbiota may be fully restored several months or years after prescribing eradication therapy.^{19,20}

Is culture the only way to assess antibiotic resistance to *H. pylori*?

In routine clinical practice, the detection of *H. pylori* antimicrobial resistance is mainly based on culture, including gradient diffusion susceptibility testing (E-test) and the agar dilution method.¹⁵ These phenotypic assays offer the opportunity to determine the minimal inhibitory concentrations of the antibiotics. Antibiogram is the only available way to test the susceptibility to all antibiotics.

Recently, different polymerase chain reaction (PCR)-based approaches have been developed as alternative methods to culture. These techniques allow assessing point mutations responsible for antibiotic resistance.21 PCR-based tests are, at present, available mainly for the detection of clarithromycin and levofloxacin resistance. A recent systematic review evaluated the feasibility of genotypic detection methods compared with phenotypic detection methods, and concluded that the genotypic detection methods were reliable for the diagnosis of clarithromycin and quinolone resistance in the strain and biopsy specimens; the A2142G/C and/or A2143G combination had the best sensitivity and specificity for the detection of clarithromycin resistance.²² Although genotypic methods to evaluate antibiotic resistance were initially developed for macrolides, more recently they have also been evaluated for other antibiotics: the rdxA and frxA genes were found to contribute to metronidazole resistance, a relationship was found between 16S ribosomal DNA mutation and tetracycline resistance, and the role of the gyrA gene in fluoroquinolone-resistant strains was also determined.²² However, the use of genotypic methods for the detection of metronidazole resistance is not vet established.

Molecular tests are faster than conventional culturebased assays. Culture is time consuming (it takes approximately 10 days to culture *H. pylori* and measure the minimal inhibitory concentration), especially when a low bacterial load is present, as generally occurs after eradication failure.⁷ In this respect, PCR-based, culture-free techniques are accurate in genotypically finding even minimal traces of certain resistant strains. Finally, PCR is technically feasible for clinical application in small- and medium-sized hospitals in developing countries.²³

The correlation between antimicrobial susceptibility testing performed by culture and antibiogram versus a molecular test, essentially real-time PCR, is not perfect. This relatively low concordance may be due to different factors: the relative low sensitivity of phenotypic investigation, the possibility that an E-test may identify resistant strains with point mutations different from those tested by PCR, or its inability to detect heteroresistance (defined as the coexistence of strains susceptible and resistant to the same antibiotic in the same patient).^{21,24} In this respect, molecular tests are able to detect more cases of hetero-resistance than culture, and an isolate could be mistakenly considered susceptible if a single biopsy is used for antimicrobial tests.13,25

Are gastric biopsies the only useful samples to study antibiotic susceptibility?

For detecting H. pylori antimicrobial resistance, a molecular approach based on a stool sample might enable more convenient, time-saving methods that facilitate the applicability of susceptibility-guided treatment. Some studies have shown an overall high sensitivity and specificity when comparing fecal DNA samples with culture or PCR on gastric biopsies to evaluate clarithromycin susceptibility.²¹ However, noninvasive molecular tests are currently at a very early phase of development.²¹ In fact, the aim of a recent meta-analysis was to provide pooled diagnostic accuracy measures for stool PCR test in the diagnosis of *H. pylori* infection.²⁶ Overall, 26 studies identified met the eligibility criteria, and it was found that stool PCR test had a performance of only 71% sensitivity (the main limitation of these stool tests seems to be the presence of PCR inhibitors of the feces that may cause false-negative results), although with a 96% specificity. The authors concluded that, in descending order of significance, the most diagnostic candidate genes using PCR detection were 23S rRNA, 16S rRNA, and glmM, and that PCR for the 23S rRNA gene, which has the highest performance, could be applicable to detect H. pylori infection.²⁶

Susceptibility-based strategy and test-and-treat strategy: a contradiction?

Endoscopy has several obvious disadvantages: firstly, it is expensive and uncomfortable. In addition, it frequently involves prolonged waiting times. Furthermore, as the majority of endoscopy findings are normal they do not contribute to management. In summary, although performing an endoscopic evaluation of the upper gastrointestinal tract in all dyspeptic patients is a theoretical option, it is not realistic in clinical practice.

Several diagnostic strategies have been proposed for selecting patients with dyspeptic symptoms who are expected to benefit most from endoscopy. The "test-and-treat" strategy is based on the investigation of the presence of *H. pylori* and its subsequent eradication when detected. Several decision analyses and prospective studies support the use of the test-and-treat strategy for dyspeptic patients,^{27,28} therefore it has been recommended by all international consensus conferences.^{9–11} To avoid the theoretical risk of delaying the diagnosis of a malignant neoplasm, this strategy has been recommended only in young patients with no alarm symptoms; otherwise, endoscopy should be performed.

Taking into account that dyspepsia is the main indication for *H. pylori* eradication, it seems that a contradiction exists in recommending the application of a susceptibility-based strategy and the test-and-treat strategy, as culture obviously implies the performance of an endoscopic exploration to obtain biopsy specimens. However, this apparent contradiction could disappear in the near future if we had noninvasive methods to evaluate antibiotic susceptibility, for example from stool samples. Unfortunately, as previously discussed, stool molecular tests are currently at a very early phase of development.²¹

Finally, the feasibility of the susceptibility-based (endoscopic) strategy has not been properly evaluated. Most studies using susceptibility-guided therapy only include patients with a positive culture. Therefore, the number of susceptibilityguided therapy failures due to patients' refusal of endoscopy has not been estimated or included.²⁹ In fact, when the applicability and effectiveness of this strategy was reviewed,²⁹ the rate of acceptance of endoscopy for biopsy and culture was described only in one article with only 60 patients, and was reported to be as low as 60%.³⁰

What is the success rate of culture to provide information about antimicrobial susceptibility?

Unfortunately, the antimicrobial susceptibility cannot be obtained in all cases (i.e. the sensitivity of bacterial culture is not 100%).³¹ Even in the optimal conditions usually encountered in clinical trials, when both gastroenterologists and microbiologists are highly motivated, a culture sensitivity of no more than 90% is achieved in treatmentnaïve patients. Furthermore, the bacterium was isolated in <80% of cases in several studies, including patients who had failed at least one eradication treatment.⁷

Growth of H. pylori can be affected by many environmental factors, such as the number of obtained gastric specimens, the duration and temperature of the transport period, the microaerophilic conditions and the selectivity of the culture medium.³² Moreover, certain drugs such as bismuth salts, antibiotics and proton pump inhibitors (PPIs) may negatively influence H. pylori detection.³² Therefore, in routine clinical practice an even lower (60-80%) probability of isolating the bacterium is to be expected.^{7,29,32} Taking into account that a rate of acceptance of endoscopy of only 50-60% has been estimated, resistance would be finally determined in only approximately 50% of the patients to whom endoscopy and culture were offered.29

What useful information, from which antibiotics, can be obtained from culture?

The standard method (culture and antibiogram) is the only way to test the susceptibility to all antibiotics. However, antibiotic susceptibility testing in clinical practice yields useful information only for a few antibiotics. Antibiotics effective and generally used against H. pylori are mainly the following five: amoxicillin, clarithromycin, metronidazole, tetracycline and quinolones. Resistance to amoxicillin has been estimated to be as low as 0-1%; similarly, resistance to tetracycline has been reported to range from 0% to 5%. Hence, their role in clinical practice may even be marginalized. Finally, the relevance of in vitro metronidazole resistance for the in vivo treatment is quite limited (see next section). On the other hand, resistance to clarithromycin and quinolones is rapidly increasing and has reached alarming levels worldwide, and its clinical relevance is doubtless. Therefore, it may even be

assumed that antibiotic susceptibility testing in clinical practice yields useful information only regarding clarithromycin and quinolones.

What is the correlation between *in vitro* (culture) and *in vivo* (eradication rate) results?

On one hand, susceptibility towards clarithromycin and metronidazole in vitro does not necessarily lead to eradication in vivo. Thus, even knowing the susceptibility of H. pylori, eradication rates do not achieve 100%.33,34 In this respect, retreatment of H. pylori using a therapy regimen including metronidazole achieves only a 70–90% eradication rate in those patients harboring metronidazole-susceptible strains. Likewise, only a 70-80% success rate is obtained in those patients infected with clarithromycin-sensitive strains when using a clarithromycin-based regimen.⁷ A recent systematic review assessed infection cure rates in patients harboring strains found to be susceptible to the antibiotics administered in clinical trials in which the efficacy of second-line treatments was evaluated. This review reported a cure rate of only 72% in the patients harboring a clarithromycin-susceptible strain after previous clarithromycin treatment.²⁹ Therefore, the authors concluded that susceptibility-guided treatment alone did not achieve adequate cure rates for rescue therapies.29

On the other hand, the contrary is also possible: H. pylori eradication may be achieved in the presence of H. pylori metronidazole- or clarithromycin-resistant strains even with a drug combination including these antibiotics. This translates that in vitro resistance to either clarithromycin or metronidazole could sometimes be overcome in vivo by prescribing these antibiotics.⁷ As an example, the European Registry on H. pylori Management (Hp-EuReg)³⁵ showed that the standard triple therapy with a PPI, clarithromycin and amoxicillin was effective in almost 80% of patients with clarithromycin resistance.³⁶ Furthermore, probably due to the synergistic effect of bismuth, the addition of this drug to a triple therapy with clarithromycin may allow achieving a cure rate of approximately 90% even in patients with resistance against this antibiotic.37,38 Finally, it has been shown that vonoprazan, a novel gastric acid suppressant, is superior to conventional PPIbased therapy for the eradication of clarithromycin-resistant H. pylori strains.³⁹

The lack of concordance between *in vitro* and *in vivo* results may be due to the fact that the *in vitro* test might not reflect the actual levels of active antibiotics in the gastric lumen, where pH may exert an influence on antimicrobial activity. In addition, some discrepancies between antibiotic susceptibility and *H. pylori* eradication may occur, due, for example, to the possibility of coinfection with different *H. pylori* strains.^{13,40}

Should antibiotics be repeated, especially after *H. pylori* eradication failure?

When a regimen has to be selected to treat H. pylori infection, we have several data that will aid us in suspecting resistance to a particular antibiotic, without the necessity of a culture. In naïve patients, a suggested strategy is to choose the best regimen for a population according to the prevalence of antibiotic resistance. The effectiveness of a treatment for *H. pylori* can be predicted as long as its efficacy in resistant and susceptible strains and the prevalence of antibiotic resistance are known in the specific population.⁴¹ For this to be possible, epidemiological surveys evaluating resistances in each country or region should be conducted on a regular basis. Moreover, it is very important to ask patients about previous exposure to antibiotics, particularly macrolides and fluoroquinolones, for any reason, as this provides a proxy for underlying antibiotic resistance to H. pylori.^{3,42} Thus, in areas where H. pylori clarithromycin resistance is known to be low (<15%) and in patients with no previous history of macrolide exposure (for any reason), clarithromycin triple therapy may still be a valid first-line treatment option.10

Regarding rescue treatment, after failure of a firstline eradication regimen, the remaining H. pylori will show very high resistance to some (though not all) of the prescribed antibiotics.43,44 Resistance to amoxicillin and tetracycline is extremely rare, even after failure of treatment including these antibiotics. The same applies to bismuth: no in vitro resistance to this drug has been described.⁴⁵ By contrast, after treatment failure, resistance to clarithromycin, quinolones and metronidazole reach virtually 100%. As the efficacy of clarithromycin- and quinolone-containing regimens is strongly affected by clarithromycin and quinolone resistance, repeating these drugs in rescue treatments is discouraged.^{33,46,47} Even if resistance to these antibiotics does not appear, it remains uncertain whether their re-administration is adequate, as they were not efficacious (for unknown reasons) for the first time. In fact, a major finding of a recent systematic review was that, even if the culture shows a clarithromycin-susceptible strain, repeating clarithromycin after a first treatment failure with this drug should be discouraged.²⁹

With regard to metronidazole, some studies suggest that in vitro metronidazole resistance has a limited impact on the efficacy of H. pylori treatments when sufficiently long treatments and high metronidazole doses are used. However, a recent multicenter study showed that cure rates of a 14-day, high-dose, rescue triple metronidazoleamoxicillin-PPI therapy were as low as 37% in patients with previous metronidazole administration.⁴⁸ Therefore, it is suggested that repeating this antibiotic is only recommended when it is indispensable and in the setting of bismuth-based quadruple therapies.^{37,49,50} In this respect, an advantage of prescribing a bismuth-based quadruple therapy is that we do not need to worry about previous antibiotic use as the risk of having a tetracycline resistant strain is extremely low and metronidazole resistance has limited impact on effectiveness of this regimen.

Regarding the optimal dose of the antibiotics, both in first-line but especially in rescue therapies, to obtain maximal pharmacodynamic effect, although amoxicillin is generally dosed twice daily, prescribing this antibiotic in more frequent doses (e.g. three or four times daily) has been suggested to improve efficacy.37,38 On the other hand, the recommended metronidazole dosage for the majority of infectious diseases is 7.5 mg/kg three times a day with a plasma half-life of the molecule between 7 and 10h. Several studies have shown good eradication rates using high doses and three-times-a-day schedules of both amoxicillin and metronidazole.37,38 Regarding the optimal duration of H. pylori eradication treatment, it should be 14 days, unless a shorter scheme has been shown locally to be equally effective.^{37,38} Finally, high-dose PPI therapy is recommended for triple therapy, and may probably increase the efficacy of a nonbismuth concomitant regimen as well. Nevertheless, more pharmacokinetic and pharmacodynamic studies are necessary to clarify these issues.

It has been suggested that endoscopy with culture may be appropriate after failure of two eradication therapies. For example, Cammarota et al. assessed the efficacy of a third-line, culture-guided treatment approach.⁵¹ After the first two eradication attempts, 95% of patients were resistant to clarithromycin, and all were resistant to metronidazole. Consequently, most patients received a quadruple therapy consisting of PPI, bismuth, tetracycline and amoxicillin, and H. pylori eradication was achieved in 90% of the cases. The authors concluded that a third-line culture-guided therapeutic approach is effective; however, it would seem more appropriate to conclude instead that the bismuth-, tetracycline- and amoxicillin-based quadruple regimen would be an appropriate empirical third-line rescue treatment option (as it would not be necessary to know antibiotic susceptibilities to choose a regimen that simply implies not readministering clarithromycin or metronidazole).

In summary, the position in the case of *H. pylori* therapy failure would be clear: not to re-administer any of the antibiotics against which *H. pylori* has probably become resistant. Although it may seem illogical, some studies have demonstrated that the repetition, even of exactly the same antibiotic regimen after *H. pylori* eradication failure, is not exceptional in clinical practice.^{52,53}

As a representative example of this empirical strategy of not repeating key antibiotics, rifabutin-based rescue therapy constitutes an encouraging fourthline strategy after multiple previous eradication failures with key antibiotics such as clarithromycin, metronidazole and levofloxacin.⁵⁴ The use of furazolidone, an antimicrobial drug that is active against a broad spectrum of bacteria and protozoa, may also be a good alternative for empirical treatment after several eradication failures.⁵⁵

What is the comparative effectiveness of susceptibility-guided *versus* empirical strategy for first-line treatment?

The Maastricht V consensus report stated that "it is recommended to perform clarithromycin susceptibility testing when a standard clarithromycin-based treatment is considered as the firstline therapy, except in populations or regions with well documented low clarithromycin resistance (<15%)".⁹ However, the scientific evidence supporting this statement is limited. Several meta-analyses have compared cure rates of susceptibility-guided *versus* empirical therapy for *H. pylori* first-line treatment. The first meta-analysis was published by Wenzhen *et al.* in 2012 and was focused specifically on first-line treatment.⁵⁶ Only five randomized controlled trials (RCTs) were included, and it was concluded that culture-guided triple therapy was more effective than standard triple therapy for first-line treatment.

The second meta-analysis was published by Lopez-Gongora *et al.*⁵⁷ RCTs were selected and analyzed separately for first- and second-line treatments. In first-line treatment (nine studies), susceptibility-guided therapy was more efficacious than empirical 7–10 day triple therapy (which was the regimen prescribed in most studies).

The third meta-analysis was published by Chen *et al.* in 2016, and included both randomized and nonrandomized controlled clinical trials (nine studies in total).⁵⁸ First-line tailored therapy achieved higher eradication rates than empirical regimens.

We have just performed (in 2020) an updated meta-analysis comparing empirical versus susceptibility-guided treatment of H. pylori,59 including 40 studies.⁶⁰⁻¹⁰⁰ When only assessing first-line treatments, better efficacy results were obtained, overall, with the susceptibility-guided strategy (although the results were borderline statistically significant). However, when considering only empirical up-to-date first-line quadruple regimens (that is excluding the suboptimal triple therapies) no differences in efficacy were found versus the susceptibility-guided group. This lack of difference was confirmed when only RCTs were included and studies based on CYP2C19 gene polymorphism were excluded. Therefore, we concluded that susceptibility-guided treatment is not better than empirical treatment of H. pylori infection in first-line if the most updated quadruple regimens are empirically prescribed.

What is the comparative effectiveness of susceptibility-guided *versus* empirical strategy for second-line treatment?

Some meta-analyses have compared *H. pylori* cure rates of susceptibility-guided therapies *versus* those of empirical therapy specifically for second-line treatment. In the previously mentioned meta-analysis by Lopez-Gongora *et al.* only four RCTs that included *H. pylori* second-line rescue therapies were found.⁵⁷ Results were highly heterogeneous and no significant differences were found between susceptibility-guided and empirical strategies in terms of efficacy. The other meta-analysis previously mentioned, performed by Chen *et al.* also found no differences between tailored and empirical rescue regimens, although only three studies were included.⁵⁸ Finally, in the updated metaanalysis we have just performed, when including only second-line regimens, similar efficacy results were demonstrated with the two strategies (tailored and empirical) both when all the comparative studies were included and when only the RCTs were considered.⁵⁹

What is the effectiveness of susceptibilityguided strategy for third-line treatment?

It has been frequently recommended that performing culture at first-line treatment or after a first eradication failure may not be necessary and therefore assessing *H. pylori* sensitivity to antibiotics in clinical practice may be suggested only after failure of the second treatment.¹⁰¹ However, previous meta-analyses could not find any RCT comparing cure rates of susceptibility-guided therapies *versus* those of empirical third-line therapy.⁵⁷ In our updated meta-analysis, only two studies including at least third-line rescue regimens were identified, and similar efficacy results were found between both tailored and empirical strategies.^{86,88}

The aim of a recent systematic review was to evaluate the effectiveness of susceptibility-guided therapy as third-line therapy (without comparing it with empirical treatment).¹⁰² Four observational studies were included (no comparative studies were found), and the pooled mean eradication rate with susceptibility-guided therapy was only 72%. Therefore, the authors concluded that susceptibility-guided therapy may be an acceptable option as rescue treatment, but cure rates are, at best, moderate; therefore, the evidence in favor of susceptibility-guided therapy as rescue therapy is currently insufficient to recommend its use.¹⁰²

Are the results from studies comparing empirical *versus* susceptibility-guided strategies reliable?

There are some relevant limitations affecting comparative studies, and consequently also the reliability of the meta-analyses including these studies, which are summarized as follows. A major limitation of the current evidence regarding susceptibility-guided therapy is that comparative studies of susceptibility-guided therapy randomized patients after diagnostic endoscopy or even after successful culture has been performed.⁵⁷ Therefore, the comparative effectiveness of susceptibility-guided therapy versus the noninvasive diagnosis and empirical treatment strategy in patients where H. pylori infection is suspected (but not yet proven) has not been evaluated in randomized trials. Thus, a study adequately evaluating the effectiveness of susceptibility-guided therapy as a first-line treatment should randomize patients with uninvestigated dyspepsia into noninvasive testing versus endoscopy plus culture groups. In this same line, most of the studies evaluating the effectiveness of susceptibility-guided therapy as rescue therapy included the patients when culture had been already obtained. Therefore, the effectiveness of susceptibility-guided therapy (with endoscopy) and empirical rescue therapy (without endoscopy) has never been properly compared.¹⁰² In this respect, the Maastricht V consensus report stated that "after a first failure, if an endoscopy is carried out, culture and standard antimicrobial susceptibility testing are recommended to tailor the treatment".9

A second limitation is that previous meta-analyses showed that susceptibility-guided therapy was more effective than empirical 7-10 day triple therapy, which was the standard treatment when most of the studies were conducted. However, clarithromycin-containing triple therapies are currently known to generally achieve poor cure rates and, therefore, are suboptimal comparators in most settings (mainly in regions with high clarithromycin resistance). However, there is limited evidence comparing this empirical approach with the highly effective bismuth or nonbismuth quadruple therapies currently recommended.9 In fact, as previously noted in our updated meta-analysis, when only empirical first-line quadruple regimens were included, no differences in efficacy were found versus the susceptibility-guided group.⁵⁹ This results are in agreement with the well-known high effectiveness of bismuth quadruple therapy, even in patients with clarithromycin or metronidazole resistance; and also with the encouraging results of nonbismuth quadruple concomitant therapy, even when single clarithromycin or metronidazole resistance is present (only dual clarithromycin and metronidazole resistance

seems to jeopardize effectiveness with this regimen). $^{103}\,$

A final limitation is that many of the comparative studies evaluating susceptibility-guided *versus* empirical treatment included susceptibility testing for only one antibiotic (clarithromycin); metronidazole susceptibility was assessed in only some cases; and quinolone resistance was only exceptionally evaluated.

What is the cumulative *H. pylori* eradication rate with successive empirical treatments?

In designing a treatment strategy we should not focus on the results of primary therapy alone (although this should obviously be our primary goal); an adequate strategy for treating this infection should use several therapies which, if used consecutively, come as close to the 100% cure rate as possible.¹⁰¹ In this sense, some studies have evaluated different empirical regimens after failure of two or more eradication treatments and have achieved a final (overall) eradication rate of almost 100%.^{7,104–118}

In our previous experience, we aimed to evaluate the efficacy of different rescue therapies empirically prescribed during 10 years to 500 patients in whom at least one eradication regimen had failed.¹⁰⁴ Antibiotic susceptibility was unknown, and therefore rescue regimens were chosen empirically. Overall, *H. pylori* cure rates with the second-, third-, and fourth-line rescue regimens were 70%, 74%, and 76%, respectively. Thus, although the effectiveness of rescue regimens could not be considered ideal, cumulative *H. pylori* eradication rate with four successive treatments was 99.5%.

These results have been recently updated, including 1200 patients and 18 years of follow-up, and the cumulative effectiveness after five consecutive therapies was 99.8%, demonstrating that eradication can be achieved virtually in all cases by the administration of several consecutive empirical therapies, based just on the previously prescribed regimens.¹¹⁹ Empirical strategy should be based on the avoidance of repeating similar eradicating schemes in the same patients during the course of different eradicating regimens.⁴⁶ In this last study, the most effective second-line strategy was the administration of a bismuth-containing quadruple therapy (either classical or with amoxicillin and levofloxacin); the most effective third-line strategy was the administration of a not previously used bismuth-containing quadruple therapy; finally, a good alternative as a fourth-line therapy was the administration of rifabutin (PPI, rifabutin, amoxicillin and bismuth).¹¹⁹

Is the susceptibility-guided approach cost-effective?

Several studies have evaluated the cost-effectiveness of the susceptibility-guided strategy, generally using decision models. These studies are chronologically reviewed below.

The first cost-effectiveness study was performed by Breuer and Graham in 1999, by using a decision model, and showed that an eradication strategy driven by antimicrobial susceptibility testing would be able to save US \$37 per patient treated in a US population.¹²⁰ However, it should be noted that, in this study, the empirical group also included the performance of an endoscopy plus biopsy (which may be avoided by the use of noninvasive *H. pylori* diagnostic tests).

In 2003, Romano *et al.* achieved, in a setting of treatment-naïve Italian patients, savings of approximately US \$5 per patient in the susceptibility testing group.⁶⁶ However, some authors calculated the eradication costs from these data, and pointed out that the mere 5% eradication benefit achieved by an invasive approach may not be justifiable because of the extra cost of US \$148/ patient for pretreatment susceptibility testing among young dyspeptic patients.^{121,122}

In 2013, Cosme *et al.* reported that, in Spain, the culture-based approach was more cost-effective than standard first-line therapy given empirically (\notin 571 *versus* \notin 666 per patient).⁷⁷

In 2007, Furuta *et al.* calculated a similar cost per successful eradication with both strategies in Japan: US \$669 per patient for the tailored and US \$657 for the standard regimen group.⁷⁰

More recently, in 2018, Liou *et al.* found that, in Taiwan, US \$6920 would be required to additionally cure one patient with refractory *H. pylori* infection using the genotypic resistance-guided therapy, compared to empirical therapy, which was obviously not cost-effective.⁸⁶

Also in 2018, Gweon *et al.* showed that the cost of a successful eradication using PCR (US \$120)

would be similar or superior to the expected cost of a successful eradication with empirical treatment (US \$92).⁸⁷

Finally, in 2019, Cho *et al.* reported that, compared with empirical triple therapy, the incremental cost-effectiveness ratios of tailored therapy (using PCR as diagnostic test) were US \$3.96 per patient, concluding that, in Korea, tailored eradication may be cost-effective.⁹³

In summary, the different studies that have evaluated the cost-effectiveness of the H. pylori susceptibility-guided treatment have achieved contradictory results. As H. pylori eradication rates depend on several factors (i.e. treatment regimen, compliance, number of prior eradication attempts, among others), the cost-effectiveness of a strategy will also depend on them.³² In addition, an eradication strategy based on culture consists of several parts, each of which has a precise cost, such as endoscopic procedures and drug regimens.³² Also, H. pylori antibiotic resistance varies among different geographical areas, which may limit the applicability of the results of the cost-effectives analysis to other populations. Furthermore, savings of a strategy are linked with the characteristics of the specific setting; for example, performing pretreatment susceptibility testing in patients with previous, independent indication of upper endoscopy would be obviously more cost-effective.32 Finally, the costeffectiveness may vary according to the cost of care in a given country, and therefore the same conclusion may not be applied to other settings.

Conclusion

Resistance of *H. pylori* to antibiotics has reached alarming levels worldwide. Local surveillance networks are required to select appropriate eradication regimens for each region. Tailored treatment of H. pylori infection according to systematic antimicrobial susceptibility testing may be useful to limit the emergence of antibiotic resistance worldwide. However, whether patients should systematically undergo an upper endoscopy for bacterial culture (or PCR) before administering H. pylori eradication treatment in clinical practice remains a debatable matter. In the present article we have reviewed the advantages and limitations of the susceptibility-guided and the empirical strategies to treat H. pylori infection, which are summarized in Table 1.

 Table 1. Main advantages and limitations of susceptibility-guided and empirical treatment of Helicobacter pylori infection.

Susceptibility-guided treatment	Empirical treatment
Advantages	
Allows performing resistance surveys	"Test-and-treat" strategy for dyspepsia is recommended by all consensus conferences
Provide personalized treatment	Resistance to amoxicillin and tetracycline is extremely rare, so they can be empirically prescribed
Reduce unnecessary antibiotic prescription	No <i>in vitro</i> resistance to bismuth has been described, so it can be also empirically prescribed
May limit the emergence of antibiotic resistance worldwide	<i>In vitro</i> metronidazole resistance has a limited impact on the efficacy of treatments when sufficiently long treatments and high metronidazole doses are used
Might allow the administration of the standard clarithromycin-based triple therapy to patients with clarithromycin-susceptible strains in areas with high overall clarithromycin resistance	The position in the case of failure is clear: not to re- administer any of the antibiotics against which <i>H. pylori</i> has probably become resistant
Molecular tests (PCR) based on a stool sample might enable more convenient methods	Rifabutin and furazolidone are good alternatives for empirical treatment after several eradication failures
It would be recommendable that susceptibility tests are routinely performed in specialized centers, with the aim to evaluate the prevalence of antibiotic resistance in the treatment of naïve patients and the influence of such resistances on the efficacy of treatments	Cumulative <i>H. pylori</i> eradication rate with several successive rescue therapies empirically prescribed reaches almost 100%
Limitations	
The moment when it must be performed (before the first treatment or after failure) is controversial	Resistance of <i>H. pylori</i> to antibiotics has reached alarming levels worldwide
Implies the performance of endoscopy, which is expensive and uncomfortable	Empirical treatment may increase the emergence of antibiotic resistance worldwide
Low rate of acceptance of endoscopy by patients	In some cases, it will imply prescribing an antibiotic that will lack efficacy
Since the majority of endoscopy findings are normal they do not contribute to management	Increase unnecessary antibiotic prescription
Culture is time consuming	Do not allow performing resistance surveys
Culture is not always available on a routine basis	Do not provide personalized treatment
Culture has low sensitivity (<80%)	May induce transient increase of antibiotic resistance to certain bacteria
Imperfect correlation between susceptibility testing performed by culture and PCR	May induce short-term perturbation of gut microbiota after <i>H. pylori</i> eradication
Culture provides useful information only for clarithromycin, metronidazole and quinolones	
Imperfect correlation between <i>in vitro</i> and <i>in vivo</i> results (mainly for metronidazole)	
Expensive (mainly because of endoscopy)	
PCR, polymerase chain reaction.	

It would be recommendable that susceptibility tests (culture or PCR) are routinely performed, even before prescribing first-line treatment, in specialized centers with interest in *H. pylori* management, with the intention to evaluate the prevalence of antibiotic resistance in the treatment of naïve patients and the influence of such resistances on the efficacy of up-to-date first-line eradication treatments. Furthermore, it would also seem recommendable that susceptibility tests are routinely performed in some specialized centers after *H. pylori* eradication failures, to evaluate the development of resistances in this setting and to assess how they may reduce the effectiveness of rescue regimens.

However, the evidence is too limited to support the generalized use of susceptibility-guided therapy for H. pylori treatment in routine clinical practice, either as first-line or as rescue treatment. In particular, it seems that despite the use of susceptibility-guided combinations of drugs, rescue treatments are frequently unsuccessful, indicating that other factors different from in vitro antibiotic susceptibility influence eradication rates Practical, economical, and logistical issues should be evaluated and addressed according to the target population and the clinical situation prior to the application of susceptibility-guided H. pylori therapy. In the future, stool sample-based molecular approach for detecting H. pylori antimicrobial resistance might enable more convenient, less invasive methods that facilitate the applicability of susceptibility-guided treatment.

What is undoubted is that we always must prescribe the most effective first-line *H. pylori* eradication treatments (that is those regimens that have demonstrated to achieve cure rates \geq 90% in our setting) and that the rescue treatment should be carefully chosen depending on which treatment was used initially. The results (*H. pylori* cure rates) of our clinical practice should be continuously audited to confirm that we always maintain a high success rate.

Conflict of interest statement

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References

- 1. Hooi JKY, Lai WY, Ng WK, *et al.* Global prevalence of Helicobacter pylori infection: systematic review and meta-analysis. *Gastroenterology* 2017; 153: 420–429.
- Dore MP, Leandro G, Realdi G, et al. Effect of pretreatment antibiotic resistance to metronidazole and clarithromycin on outcome of *Helicobacter pylori* therapy: a meta-analytical approach. *Dig Dis Sci* 2000; 45: 68–76.
- 3. Megraud F, Coenen S, Versporten A, *et al. Helicobacter pylori* resistance to antibiotics in Europe and its relationship to antibiotic consumption. *Gut* 2013; 62: 34–42.
- Camargo MC, Garcia A, Riquelme A, et al. The problem of *Helicobacter pylori* resistance to antibiotics: a systematic review in Latin America. *Am J Gastroenterol* 2014; 109: 485–495.
- 5. Thung I, Aramin H, Vavinskaya V, *et al.* Review article: the global emergence of *Helicobacter pylori* antibiotic resistance. *Aliment Pharmacol Ther* 2016; 43: 514–533.
- Savoldi A, Carrara E, Graham DY, et al. Prevalence of antibiotic resistance in *Helicobacter* pylori: a systematic review and meta-analysis in World Health Organization regions. *Gastroenterology* 2018; 155: 1372–1382.e17.
- Zullo A, Hassan C, Lorenzetti R, et al. A clinical practice viewpoint: to culture or not to culture *Helicobacter pylori? Dig Liver Dis* 2003; 35: 357–361.
- Gisbert JP. Is culture necessary before first-line treatment for *Helicobacter pylori* infection? *Intern Med* 2011; 50: 2717; author reply 9–20.
- Malfertheiner P, Megraud F, O'Morain CA, et al. Management of *Helicobacter pylori* infection-the Maastricht V/Florence Consensus Report. *Gut* 2017; 66: 6–30.
- Chey WD, Leontiadis GI, Howden CW, et al. ACG clinical guideline: treatment of *Helicobacter* pylori infection. Am J Gastroenterol 2017; 112: 212–239.
- Fallone CA, Chiba N, van Zanten SV, et al. The Toronto consensus for the treatment of *Helicobacter pylori* infection in adults. *Gastroenterology* 2016; 151: 51–69.e14.

- Graham DY. Editorial—avoiding unethical Helicobacter pylori clinical trials: susceptibilitybased studies and probiotics as adjuvants. Helicobacter 2015; 20: 321–325.
- 13. Mascellino MT, Porowska B, De Angelis M, et al. Antibiotic susceptibility, heteroresistance, and updated treatment strategies in *Helicobacter* pylori infection. Drug Des Dev Ther 2017; 11: 2209–2220.
- Dang BN and Graham DY. *Helicobacter pylori* infection and antibiotic resistance: a WHO high priority? *Nat Rev Gastroenterol Hepatol* 2017; 14: 383–384.
- Arslan N, Yilmaz O and Demiray-Gurbuz

 E. Importance of antimicrobial susceptibility
 testing for the management of eradication in
 Helicobacter pylori infection. World J Gastroenterol
 2017; 23: 2854–2869.
- Liou JM, Chen CC, Chang CM, et al. Longterm changes of gut microbiota, antibiotic resistance, and metabolic parameters after *Helicobacter pylori* eradication: a multicentre, open-label, randomised trial. *Lancet Infect Dis* 2019; 19: 1109–1120.
- Rajilic-Stojanovic M, Figueiredo C, Smet A, et al. Systematic review: gastric microbiota in health and disease. *Aliment Pharmacol Ther* 2020; 51: 582–602.
- Prast-Nielsen S, McNicholl AG, O'Morain C, et al. Comparison of the impact of *Helicobacter* pylori eradication treatment with bismuth and non-bismuth quadruple regimens on the gut microbiota. *Microb Health Dis* 2019; 1: e145.
- Liou JM, Lee YC and Wu MS. Treatment of *Helicobacter pylori* infection and its long-term impacts on gut microbiota. J Gastroenterol *Hepatol* 2020; 35: 1107–1116.
- O'Connor A, Liou JM, Gisbert JP, et al. Review: treatment of *Helicobacter pylori* infection 2019. *Helicobacter* 2019; 24(Suppl. 1): e12640.
- Ierardi E, Giorgio F, Iannone A, et al. Noninvasive molecular analysis of *Helicobacter* pylori: is it time for tailored first-line therapy? World J Gastroenterol 2017; 23: 2453–2458.
- 22. Wang YH, Li Z, Wang L, *et al.* A systematic review and meta-analysis of genotypic methods for detecting antibiotic resistance in *Helicobacter pylori. Helicobacter* 2018; 23: e12467.
- Xuan SH, Wu LP, Zhou YG, et al. Detection of clarithromycin-resistant *Helicobacter pylori* in clinical specimens by molecular methods: a review. J Glob Antimicrob Resist 2016; 4: 35-41.

- 24. Jung DH, Kim JH, Jeong SJ, *et al.* Peptide nucleic acid probe-based analysis as a new detection method for clarithromycin resistance in *Helicobacter pylori. Gut Liver* 2018; 12: 641–647.
- 25. Papastergiou V, Georgopoulos SD and Karatapanis S. Treatment of *Helicobacter pylori* infection: meeting the challenge of antimicrobial resistance. *World J Gastroenterol* 2014; 20: 9898–9911.
- 26. Khadangi F, Yassi M and Kerachian MA. Review: diagnostic accuracy of PCR-based detection tests for *Helicobacter pylori* in stool samples. *Helicobacter* 2017; 22: e12444.
- 27. Gisbert JP and Calvet X. Helicobacter pylori "test-and-treat" strategy for management of dyspepsia: a comprehensive review. *Clin Transl Gastroenterol* 2013; 4: e32.
- 28. Beresniak A, Malfertheiner P, Franceschi F, et al. Helicobacter pylori "Test-and-Treat" strategy with urea breath test: a cost-effective strategy for the management of dyspepsia and the prevention of ulcer and gastric cancer in Spain—results of the Hp-Breath initiative. Helicobacter 2020: e12693.
- 29. Baylina M, Munoz N, Sanchez-Delgado J, et al. Systematic review: would susceptibilityguided treatment achieve acceptable cure rates for second-line *Helicobacter pylori* therapy as currently practiced? *Helicobacter* 2019; 24: e12584.
- Matsumoto Y, Miki I, Aoyama N, et al. Levofloxacin-versus metronidazole-based rescue therapy for H. pylori infection in Japan. *Dig Liver Dis* 2005; 37: 821–825.
- Working Party of the European Helicobacter pylori Study Group. Technical annex: tests used to assess Helicobacter pylori infection. Gut 1997; 41(Suppl. 2): S10–S18.
- Cammarota G, Ianiro G, Bibbo S, et al. Cultureguided treatment approach for *Helicobacter* pylori infection: review of the literature. World J Gastroenterol 2014; 20: 5205–5211.
- Gisbert JP and Pajares JM. Review article: *Helicobacter pylori* "rescue" regimen when proton pump inhibitor-based triple therapies fail. *Aliment Pharmacol Ther* 2002; 16: 1047–1057.
- Guslandi M. Review article: alternative antibacterial agents for *Helicobacter pylori* eradication. *Aliment Pharmacol Ther* 2001; 15: 1543–1547.
- 35. McNicholl AG, O'Morain CA, Megraud F, *et al.*; As Scientific Committee of the Hp-Eureg

on Behalf of the National Coordinators. Protocol of the European Registry on the management of *Helicobacter pylori* infection (Hp-EuReg). *Helicobacter* 2019; 24: e12630.

- Bujanda L, Nyssen OP, Cosme A, , et al.Impact of *Helicobacter pylori* clarithromycin resistance on the treatment effectiveness: data of the European Registry on H. pylori management (Hp-EuReg). *Helicobacter* 2020; 25(Suppl. 1): 30.
- Gisbert JP and McNicholl AG. Optimization strategies aimed to increase the efficacy of H. pylori eradication therapies. *Helicobacter* 2017; 22: e12392.
- Gisbert JP and Nyssen OP. Ten common errors in the treatment of *Helicobacter pylori* infection. In: Probert C (ed.) *Recent advances in* gastroenterology; in press.
- 39. Li M, Oshima T, Horikawa T, *et al.* Systematic review with meta-analysis: vonoprazan, a potent acid blocker, is superior to proton-pump inhibitors for eradication of clarithromycin-resistant strains of *Helicobacter pylori. Helicobacter* 2018; 23: e12495.
- Kim JJ, Kim JG and Kwon DH. Mixedinfection of antibiotic susceptible and resistant *Helicobacter pylori* isolates in a single patient and underestimation of antimicrobial susceptibility testing. *Helicobacter* 2003; 8: 202–206.
- Liou JM, Chen PY, Kuo YT, et al. Toward population specific and personalized treatment of *Helicobacter pylori* infection. *J Biomed Sci* 2018; 25: 70.
- 42. McMahon BJ, Hennessy TW, Bensler JM, et al. The relationship among previous antimicrobial use, antimicrobial resistance, and treatment outcomes for *Helicobacter pylori* infections. *Ann Intern Med* 2003; 139: 463–469.
- Lee JW, Kim N, Kim JM, et al. Prevalence of primary and secondary antimicrobial resistance of *Helicobacter pylori* in Korea from 2003 through 2012. *Helicobacter* 2013; 18: 206–214.
- 44. Munoz N, Sanchez-Delgado J, Baylina M, et al. Prevalence of *Helicobacter pylori* resistance after failure of first-line therapy. A systematic review. *Gastroenterol Hepatol* 2018; 41: 654–662.
- 45. Dore MP, Lu H and Graham DY. Role of bismuth in improving *Helicobacter pylori* eradication with triple therapy. *Gut* 2016; 65: 870–878.
- 46. Roccarina D, Franceschi F, Zocco MA, *et al.* Different Antibiotic No Culture Eradicating (DANCE) strategy: an easy way to manage

H. pylori eradication. *Dig Liver Dis* 2012; 44: 889–892.

- Calvet X. Dealing with uncertainty in the treatment of *Helicobacter pylori*. Ther Adv Chronic Dis 2018; 9: 93–102.
- Puig I, Gonzalez-Santiago JM, Molina-Infante J, et al. Fourteen-day high-dose esomeprazole, amoxicillin and metronidazole as third-line treatment for *Helicobacter pylori* infection. Int J Clin Pract 2017; 71: e13004.
- 49. Muller N, Amiot A, Le Thuaut A, *et al.* Rescue therapy with bismuth-containing quadruple therapy in patients infected with metronidazole-resistant *Helicobacter pylori* strains. *Clin Res Hepatol Gastroenterol* 2016; 40: 517–524.
- Nyssen OP, McNicholl AG and Gisbert JP. Meta-analysis of three-in-one single capsule bismuth-containing quadruple therapy for the eradication of *Helicobacter pylori*. *Helicobacter* 2019; 24: e12570.
- Cammarota G, Martino A, Pirozzi G, et al. High efficacy of 1-week doxycycline- and amoxicillinbased quadruple regimen in a culture-guided, third-line treatment approach for *Helicobacter* pylori infection. Aliment Pharmacol Ther 2004; 19: 789–795.
- Ribaldone DG, Astegiano M and Pellicano R. Helicobacter pylori eradication: poor medical compliance from East to West of the world. Scand J Gastroenterol 2018; 53: 265.
- Li H, Liang X, Chen Q, et al. Inappropriate treatment in *Helicobacter pylori* eradication failure: a retrospective study. *Scand J Gastroenterol* 2018; 53: 130–133.
- 54. Gisbert JP and Calvet X. Review article: rifabutin in the treatment of refractory *Helicobacter pylori* infection. *Aliment Pharmacol Ther* 2012; 35: 209–221.
- 55. Song C, Qian X, Zhu Y, et al. Effectiveness and safety of furazolidone-containing quadruple regimens in patients with *Helicobacter pylori* infection in real-world practice. *Helicobacter* 2019; 24: e12591.
- 56. Wenzhen Y, Yumin L, Quanlin G, et al. Is antimicrobial susceptibility testing necessary before first-line treatment for *Helicobacter pylori* infection? Meta-analysis of randomized controlled trials. *Intern Med* 2010; 49: 1103–1109.
- 57. Lopez-Gongora S, Puig I, Calvet X, *et al.* Systematic review and meta-analysis: susceptibility-guided versus empirical antibiotic

treatment for *Helicobacter pylori* infection. J Antimicrob Chemother 2015; 70: 2447–2455.

- Chen H, Dang Y, Zhou X, et al. Tailored therapy versus empiric chosen treatment for *Helicobacter pylori* eradication: a meta-analysis. *Medicine (Baltimore)* 2016; 95: e2750.
- Espada M, Nyssen OP and Gisbert JP. Empirical versus susceptibility-guided treatment of *Helicobacter pylori* infection: a meta-analysis. *Helicobacter* 2020; 25(Suppl. 1): 26.
- 60. Toracchio S, Cellini L, Di Campli E, *et al.* Role of antimicrobial susceptibility testing on efficacy of triple therapy in *Helicobacter pylori* eradication. *Aliment Pharmacol Ther* 2000; 14: 1639–1643.
- Romano M, Iovene MR, Montella F, et al. Pretreatment antimicrobial-susceptibility testing in the eradication of H. pylori infection. Am J Gastroenterol 2000; 95: 3317–3318.
- 62. Street ME, Caruana P, Caffarelli C, *et al.* Antibiotic resistance and antibiotic sensitivity based treatment in *Helicobacter pylori* infection: advantages and outcome. *Arch Dis Child* 2001; 84: 419–422.
- 63. Avidan B, Melzer E, Keller N, et al. The effect of culture results for *Helicobacter pylori* on the choice of treatment following failure of initial eradication. Isr Med Assoc J 2001; 3: 163–165.
- 64. Miwa H, Nagahara A, Kurosawa A, *et al.* Is antimicrobial susceptibility testing necessary before second-line treatment for *Helicobacter pylori* infection? *Aliment Pharmacol Ther* 2003; 17: 1545–1551.
- 65. Neri M, Milano A, Laterza F, et al. Role of antibiotic sensitivity testing before first-line *Helicobacter pylori* eradication treatments. *Aliment Pharmacol Ther* 2003; 18: 821–827.
- Romano M, Marmo R, Cuomo A, et al. Pretreatment antimicrobial susceptibility testing is cost saving in the eradication of *Helicobacter* pylori. Clin Gastroenterol Hepatol 2003; 1: 273–278.
- Lamouliatte H, Megraud F, Delchier JC, et al. Second-line treatment for failure to eradicate *Helicobacter pylori*: a randomized trial comparing four treatment strategies. *Aliment Pharmacol Ther* 2003; 18: 791–797.
- Yahav J, Samra Z, Niv Y, , *et al*.Susceptibilityguided vs. empiric retreatment of *Helicobacter pylori* infection after treatment failure. *Dig Dis Sci* 2006; 51: 2316–2321.
- 69. Marzio L, Coraggio D, Capodicasa S, *et al.* Role of the preliminary susceptibility testing for initial

and after failed therapy of *Helicobacter pylori* infection with levofloxacin, amoxicillin, and esomeprazole. *Helicobacter* 2006; 11: 237–242.

- Furuta T, Shirai N, Kodaira M, et al. Pharmacogenomics-based tailored versus standard therapeutic regimen for eradication of H. pylori. *Clin Pharmacol Ther* 2007; 81: 521–528.
- 71. Kawai T, Yamagishi T, Yagi K, et al. Tailored eradication therapy based on fecal *Helicobacter* pylori clarithromycin sensitivities. J Gastroenterol Hepatol 2008; 23(Suppl. 2): S171–S174.
- Wang G, Zhao Q and Li S. Study of drug sensitivity test in *Helicobacter pylori* eradication therapy. *J Clin Intern Med* 2008; 25: 474–477.
- Zhou J, Wu M and Jiang X. Role of drug sensitivity test in the triple therapy for eradication of *Helicobacter pylori*. *Chin J Gastroenterol* 2010; 15: 358–360.
- 74. Bontems P, Kalach N, Oderda G, et al. Sequential therapy versus tailored triple therapies for *Helicobacter pylori* infection in children. *J Pediatr Gastroenterol Nutr* 2011; 53: 646–650.
- 75. Molina-Infante J, Pazos-Pacheco C, Vinagre-Rodriguez G, et al. Nonbismuth quadruple (concomitant) therapy: empirical and tailored efficacy versus standard triple therapy for clarithromycin-susceptible *Helicobacter pylori* and versus sequential therapy for clarithromycinresistant strains. *Helicobacter* 2012; 17: 269–276.
- Lee HJ, Kim JI, Cheung DY, et al. Eradication of *Helicobacter pylori* according to 23S ribosomal RNA point mutations associated with clarithromycin resistance. J Infect Dis 2013; 208: 1123–1130.
- 77. Cosme A, Montes M, Martos M, et al. Usefulness of antimicrobial susceptibility in the eradication of *Helicobacter pylori*. Clin Microbiol Infect 2013; 19: 379–383.
- 78. Park CS, Lee SM, Park CH, et al. Pretreatment antimicrobial susceptibility-guided vs. clarithromycin-based triple therapy for *Helicobacter pylori* eradication in a region with high rates of multiple drug resistance. Am J Gastroenterol 2014; 109: 1595–1602.
- 79. Martos M, Bujanda L, Salicio Y, et al. Clarithromycin for first-line treatment of *Helicobacter pylori* infection after culture in highresistance regions. Eur J Gastroenterol Hepatol 2014; 26: 1380–1384.
- 80. Zhuo RP, Chen XP, Wu SZ, *et al.* Clinical effects of quadruple therapy based on

antimicrobial susceptibility testing in treatment of *Helicobacter pylori* associated upper digestive tract diseases. *World Chin J Digestol* 2015; 23: 196–201.

- 81. Dong F, Ji D, Huang R, *et al.* Multiple genetic analysis system-based antibiotic susceptibility testing in *Helicobacter pylori* and high eradication rate with phenotypic resistance-guided quadruple therapy. *Medicine* 2015; 94: e2056.
- Zhou L, Zhang J, Song Z, et al. Tailored versus triple plus bismuth or concomitant therapy as initial *Helicobacter pylori* treatment: a randomized trial. *Helicobacter* 2016; 21: 91–99.
- 83. Kwon YH, Kim N, Lee JY, et al. Comparison of the efficacy of culture-based tailored therapy for *Helicobacter pylori* eradication with that of the traditional second-line rescue therapy in Korean patients: a prospective single tertiary center study. *Scand J Gastroenterol* 2016; 51: 270–276.
- 84. Cosme A, Lizasoan J, Montes M, et al. Antimicrobial susceptibility-guided therapy versus empirical concomitant therapy for eradication of *Helicobacter pylori* in a region with high rate of clarithromycin resistance. *Helicobacter* 2016; 21: 29–34.
- 85. Ferenc S, Gnus J, Koscielna M, *et al.* High antibiotic resistance of *Helicobacter pylori* and its effect on tailored and empiric eradication of the organism in Lower Silesia, Poland. *Helicobacter* 2017; 22: e12365.
- Liou JM, Chen PY, Luo JC, et al. Efficacies of genotypic resistance-guided vs empirical therapy for refractory *Helicobacter pylori* infection. *Gastroenterology* 2018; 155: 1109–1119.
- Gweon TG, Kim JS and Kim BW. An economic modeling study of *Helicobacter pylori* eradication: comparison of dual priming oligonucleotidebased multiplex polymerase chain reaction and empirical treatment. *Gut Liver* 2018; 12: 648–654.
- Huang HT, Wang HM, Yang SC, et al. Efficacy of a 14-day quadruple-therapy regimen for thirdline *Helicobacter pylori* eradication. *Infect Drug Resist* 2018; 11: 2073–2080.
- Mascellino MT, Oliva A, De Angelis M, et al. Helicobacter pylori infection: antibiotic resistance and eradication rate in patients with gastritis showing previous treatment failures. New Microbiol 2018; 41: 306–309.
- 90. Tanabe H, Yoshino K, Ando K, *et al.* Vonoprazan-based triple therapy is non-inferior to susceptibility-guided proton pump

inhibitor-based triple therapy for *Helicobacter pylori* eradication. *Ann Clin Microbiol Antimicrob* 2018; 17: 29.

- 91. Ong S, Kim SE, Kim JH, et al. Helicobacter pylori eradication rates with concomitant and tailored therapy based on 23S rRNA point mutation: a multicenter randomized controlled trial. *Helicobacter* 2019; 24: e12654.
- 92. Chen Q, Long X, Ji Y, *et al.* Randomised controlled trial: susceptibility-guided therapy versus empiric bismuth quadruple therapy for first-line *Helicobacter pylori* treatment. *Aliment Pharmacol Ther* 2019; 49: 1385–1394.
- 93. Cho JH, Jeon SR, Kim HG, et al. Costeffectiveness of a tailored *Helicobacter pylori* eradication strategy based on the presence of a 23S ribosomal RNA point mutation that causes clarithromycin resistance in Korean patients. *β Gastroenterol Hepatol* 2019; 34: 700–706.
- 94. Choi YI, Chung JW, Park DK, et al. Tailored eradication vs empirical bismuth-containing quadruple therapy for first-line *Helicobacter pylori* eradication: a comparative, open trial. World J Gastroenterol 2019; 25: 6743–6751.
- 95. Byambajav TO, Bira N, Choijamts G, et al. Initial trials with susceptibility-based and empiric anti-H. pylori therapies in Mongolia. Front Pharmacol 2019; 10: 394.
- 96. Delchier JC, Bastuji-Garin S, Raymond J, et al. Efficacy of a tailored PCR-guided triple therapy in the treatment of *Helicobacter pylori* infection. *Med Mal Infect*. Epub ahead of print 27 June 2019. DOI: 10.1016/j.medmal.2019.06.001.
- Zhang YD, Dong QW, Zhang SH, et al. [Effectiveness of eradication regimen based on the bacterial susceptibility and CYP2C19 genotype in children with refractory *Helicobacter pylori* infection]. *Zhonghua Er Ke Za Zhi* 2020; 58: 41–45.
- Saracino IM, Pavoni M, Zullo A, et al. Antibiotic resistance and therapy outcome in *H. pylori* eradication failure patients. *Antibiotics* (*Basel*) 2020; 9: 121.
- Miyaki A, Yamaguchi K, Ida A, et al. An assessment of the efficacy of first-line *Helicobacter pylori*-eradication therapy based on clarithromycin susceptibility. *Minerva Gastroenterol Dietol* 2016; 62: 234–239.
- 100. Pan J, Shi Z, Lin D, *et al.* Is tailored therapy based on antibiotic susceptibility effective? A multicenter, open-label, randomized trial. *Front Med* 2020; 14: 43–50.

- de Boer WA and Tytgat GN. Regular review: treatment of *Helicobacter pylori* infection. *BMJ* 2000; 320: 31–34.
- 102. Puig I, Lopez-Gongora S, Calvet X, et al. Systematic review: third-line susceptibilityguided treatment for *Helicobacter pylori* infection. *Ther Adv Gastroenterol* 2016; 9: 437–448.
- 103. Gisbert JP and Calvet X. Review article: nonbismuth quadruple (concomitant) therapy for eradication of Helicobater pylori. *Aliment Pharmacol Ther* 2011; 34: 604–617.
- 104. Gisbert JP, Gisbert JL, Marcos S, et al. Empirical rescue therapy after *Helicobacter pylori* treatment failure: a 10-year single-centre study of 500 patients. *Aliment Pharmacol Ther* 2008; 27: 346–354.
- 105. Gomollon F, Sicilia B, Ducons JA, et al. Third line treatment for *Helicobacter pylori*: a prospective, culture-guided study in peptic ulcer patients. *Aliment Pharmacol Ther* 2000; 14: 1335–1338.
- 106. Gasbarrini A, Ojetti V, Armuzzi A, et al. Efficacy of a multistep strategy for *Helicobacter pylori* eradication. *Aliment Pharmacol Ther* 2000; 14: 79–83.
- 107. Seppala K, Kosunen TU, Nuutinen H, et al. Cure of *Helicobacter pylori* infection after failed primary treatment: one-center results from 120 patients. *Scand J Gastroenterol* 2000; 35: 929–934.
- 108. Chan FK, Sung JJ, Suen R, *et al.* Salvage therapies after failure of *Helicobacter pylori* eradication with ranitidine bismuth citrate-based therapies. *Aliment Pharmacol Ther* 2000; 14: 91–95.
- 109. Perri F, Festa V, Clemente R, et al. Rifabutinbased 'rescue therapy' for Helicobacter pylori infected patients after failure of standard regimens. Aliment Pharmacol Ther 2000; 14: 311–316.
- Bock H, Koop H, Lehn N, et al. Rifabutin-based triple therapy after failure of *Helicobacter pylori* eradication treatment: preliminary experience. *J Clin Gastroenterol* 2000; 31: 222–225.
- Beales IL. Efficacy of *Helicobacter pylori* eradication therapies: a single centre observational study. *BMC Gastroenterol* 2001; 1: 7.

- 112. Zullo A, Hassan C, Campo SM, *et al.* A triple therapy regimen after failed *Helicobacter pylori* treatments. *Aliment Pharmacol Ther* 2001; 15: 1193–1197.
- 113. Canducci F, Ojetti V, Pola P, et al. Rifabutinbased *Helicobacter pylori* eradication 'rescue therapy'. *Aliment Pharmacol Ther* 2001; 15: 143.
- 114. Treiber G, Ammon S, Malfertheiner P, et al. Impact of furazolidone-based quadruple therapy for eradication of *Helicobacter pylori* after previous treatment failures. *Helicobacter* 2002; 7: 225–231.
- 115. Gisbert JP, Calvet X, Bujanda L, *et al.* 'Rescue' therapy with rifabutin after multiple *Helicobacter pylori* treatment failures. *Helicobacter* 2003; 8: 90–94.
- 116. Dore MP, Marras L, Maragkoudakis E, et al. Salvage therapy after two or more prior *Helicobacter pylori* treatment failures: the super salvage regimen. *Helicobacter* 2003; 8: 307–309.
- 117. Gisbert JP, Gisbert JL, Marcos S, et al. Empirical Helicobacter pylori "rescue" therapy after failure of two eradication treatments. Dig Liver Dis 2004; 36: 7–12.
- 118. Rokkas T, Sechopoulos P, Robotis I, , et al.Cumulative H. pylori eradication rates in clinical practice by adopting first and secondline regimens proposed by the Maastricht III consensus and a third-line empirical regimen. Am J Gastroenterol 2009; 104: 21–25.
- 119. Burgos-Santamaria D, McNicholl AG and Gisbert JP. Empirical *Helicobacter pylori* rescue therapy: an 18-year single-centre study of 1200 patients. *GastroHep* 2019; 1: 311–324.
- 120. Breuer T and Graham DY. Costs of diagnosis and treatment of *Helicobacter pylori* infection: when does choosing the treatment regimen based on susceptibility testing become cost effective? *Am J Gastroenterol* 1999; 94: 725–729.
- 121. Qasim A, Sebastian S, Buckley M, et al. Pretreatment antimicrobial susceptibility testing is not cost saving in the standard eradication of *Helicobacter pylori*. Clin Gastroenterol Hepatol 2004; 2: 85; discussion
- 122. Faber J, Bar-Meir M, Rudensky B, et al. Treatment regimens for *Helicobacter pylori* infection in children: is in vitro susceptibility testing helpful? *J Pediatr Gastroenterol Nutr* 2005; 40: 571–574.

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