





Complete Genome Sequences of Bacteriophages Kaya, Guyu, Kopi, and TehO, Which Target Clinical Strains of *Pseudomonas aeruginosa*

[®]Belinda Loh,^a Xiaoqing Wang,^b [®]Xiaoting Hua,^c Junhan Luo,^b Tanye Wen,^b Liwei Zhang,^b Long Ma,^b Prasanth Manohar,^b Ramesh Nachimuthu,^d Ian Grainge,^e Yunsong Yu,^c [®]Sebastian Leptihn^{b,c,f}

^aDepartment of Biological Sciences, Xi'an Jiaotong-Liverpool University, Suzhou, PR China

^bZhejiang University-University of Edinburgh (ZJU-UoE) Institute, Zhejiang University, International Campus, Haining, Zhejiang, PR China

cDepartment of Infectious Diseases, Sir Run Run Shaw Hospital, Zhejiang University School of Medicine, Hangzhou, Zhejiang, PR China

^dAntibiotic Resistance and Phage Therapy Laboratory, School of Biosciences and Technology, Vellore Institute of Technology (VIT), Vellore, Tamil Nadu, India

eSchool of Environmental and Life Sciences, The University of Newcastle, Callaghan, New South Wales, Australia College of Medicine & Veterinary Medicine, University of Edinburgh Medical School, Edinburgh, United Kingdom

ABSTRACT *Pseudomonas aeruginosa* is a major public health concern, as drug-resistant strains increase mortality in hospital-acquired infections. We report the isolation and complete genome sequences of four lytic bacteriophages that target clinical multidrug-resistant *P. aeruginosa* strains.

P seudomonas aeruginosa is an important nosocomial opportunistic pathogen that is able to live in a wide range of environments (1). The motile rod-shaped bacterium can cause lethal infections, such as sepsis in immunocompromised hosts and hospitalized patients (e.g., burn wounds), and infects a wide range of organs, including the lungs, urinary tract, and kidneys. Some strains of *P. aeruginosa* exhibit extensive drug resistance to available antibiotics, and the species has hence been listed as a priority 1 pathogen by the WHO (2, 3). Therefore, novel antibiotics or clinical therapeutic options are needed. One strategy is the use of therapeutic bacteriophages (4, 5). Here, we report the complete genome sequences of four lytic bacteriophages (Kaya, Guyu, Kopi, and TehO) that have been isolated using clinical multidrug-resistant strains of *P. aeruginosa*.

Water was collected in January 2020 from a river in Haining, China (120.605111°E, 30.481146°N). The water was filtered (pore size, 0.45 μ m) before phage enrichment using cultures of P. aeruginosa. P. aeruginosa host strains were grown in lysogeny broth (LB) at 37°C overnight with agitation; the strains used to isolate each phage are provided in Table 1. Phages were obtained from clear single plaques and grown in the presence of the bacterial host in LB overnight. Bacterial cells were removed by centrifugation, and the supernatant was filtered through a $0.22-\mu$ m membrane (6). Nucleic acids were extracted using the Biomed virus rapid DNA/RNA kit (Beijing, China) according to the manufacturer's instructions. Sequencing libraries were prepared using the NEBNext Ultra II DNA library prep kit for Illumina, and the genomes were sequenced using the Illumina HiSeg platform. The average read length obtained was 150 bp. The assembly pipeline Unicycler v0.4.8 (7) was used to conduct quality control of raw reads, assemble the genomes, and determine the completion of the assembled genomes. Genome annotation was completed using the CPT Galaxy and Web Apollo interfaces (8). tRNAs were predicted using ARAGORN v2.36 (9) and tRNA-scan-SE v2.0 (10). Open reading frames (ORFs) were predicted using GeneMarkS v4.28 (11), Glimmer v3.0 (12), and MetaGeneAnnotator v1.0 (13) and were then manually validated using BLAST v2.9.0 searches (14) against the

Editor Simon Roux, DOE Joint Genome Institute

Copyright © 2021 Loh et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Belinda Loh, belinda.loh@gmail.com, or Sebastian Leptihn, sebastian.leptihn@ed.ac.uk.

Received 26 October 2021 Accepted 6 November 2021 Published 2 December 2021

	Pseudomonas	Total no. of reads	Genome	Genome	GC	No. of	Accession no.	
Isolate	host strain	(forward/reverse)	coverage (×)	length (bp)	content (%)	ORFs	GenBank	SRA
Kaya	2081	11,290,334	2.56	43,067	54	60	MZ927745.1	SRR16248205
Guyu	2072	13,776,770	89.19	43,141	55	56	MZ927746	SRR16248204
Корі	2072	7,620,976	142.96	42,820	53	55	OK330455.1	SRR16248203
TehO	2081	8,705,734	86.39	43,015	54	56	OK330456.1	SRR16248202

TABLE 1 Characteristics of *Pseudomonas* phage genomes

NCBI nonredundant and Swiss-Prot databases (15). Pairwise nucleotide alignments between the phages were evaluated using NCBI blastn. Default parameters were used unless stated otherwise.

The characteristics of all four phage genomes are listed in Table 1. The phages are novel but are close relatives of each other, with their genes showing the mosaicism typical of bacterial viruses (Fig. 1). No genes were found to encode toxins or antibiotic resistance factors according to blastn searches against the Bacterial Virulence Factor Database (VFDB) (16). The phages were categorized as lytic using PhageAI (17). The most closely related phages of Kaya and Guyu are *Xanthomonas* phage Samson (GenBank accession number MN062187) and *Pseudomonas* phage PaMx42 (JQ067092), with genome coverage between 90% and 92% at sequence identities between 85% and 97%. Kopi and TehO are most closely related to *Stenotrophomonas* phage vB_SmaS-DLP2 (KR537871) and *Pseudomonas* phage vB_Pae-Kakheti25 (JQ307387), with sequence coverage between 91% and 95% at 95.48% to 97.89% sequence identity. With these sequence similarities, Kaya, Guyu, Kopi, and TehO are predicted to be *Siphoviridae* of the order *Caudovirales*.

Data availability. The sequencing data for bacteriophages Kaya, Guyu, Kopi, and TehO are available in GenBank under BioProject accession number PRJNA751744. The accession numbers for the genomes and sequencing reads are listed in Table 1.

	Kaya	Guyu	Корі	TehO	Samson	PaMx42	SmaS- DLP2	Kakheti25
Kaya	100%	99%	83%	77%	92%	90%	81%	80%
	100%	88.44%	97.32%	97.10%	85.30%	85.02%	96.01%	96.27%
Guyu	99%	100%	75%	75%	90%	90%	73%	70%
	88.44%	100%	81.54%	81.88%	97.26%	97.41%	81.70%	81.51%
Корі	83%	75%	100%	89%	71%	64%	95%	93%
	97.32%	81.54%	100%	98.49%	81.45%	81.53%	97.89%	96.70%
TehO	77%	75%	89%	100%	73%	76%	91%	95%
	97.10%	81.88%	98.00%	100%	81.50%	81.56%	95.48%	96.22%
Samson	92%	90%	71%	73%	100%	99%	71%	72%
	85.30%	97.26%	81.45%	81.50%	100%	96.05%	81.48%	81.35%
PaMx42	90%	90%	64%	76%	99%	100%	64%	75%
	85.02%	97.41%	81.53%	81.56%	96.05%	100%	81.49%	81.39%
SmaS-	81%	73%	95%	91%	71%	64%	100%	95%
DLP2	96.01%	81.70%	97.89%	95.48%	81.48%	81.49%	100%	97.08%
Kakheti25	80%	70%	93%	95%	72%	75%	95%	100%
	96.27%	81.51%	96.70%	96.22%	81.35%	81.39%	97.08%	100%

FIG 1 Genome sequence coverage (top number in each cell) and nucleotide identity (bottom number) of *Pseudomonas* phages with their closest relatives. The green and brown boxes indicate phages from this study. The gray boxes indicate phages from other studies: Samson (*Xanthomonas* phage; GenBank accession number MN062187), PaMx42 (*Pseudomonas* phage; JQ067092), SmaS-DLP2 (*Stenotrophomonas* phage; KR537871), and Kakheti25 (vB_Pae-Kakheti25) (*Pseudomonas* phage; JQ307387).

This work was supported by the National Natural Science Foundation of China (32011530116).

REFERENCES

- 1. Moradali MF, Ghods S, Rehm BH. 2017. Pseudomonas aeruginosa lifestyle: a paradigm for adaptation, survival, and persistence. Front Cell Infect Microbiol 7:39. https://doi.org/10.3389/fcimb.2017.00039.
- Tacconelli E, Carrara E, Savoldi A, Harbarth S, Mendelson M, Monnet DL, Pulcini C, Kahlmeter G, Kluytmans J, Carmeli Y, Ouellette M, Outterson K, Patel J, Cavaleri M, Cox EM, Houchens CR, Grayson ML, Hansen P, Singh N, Theuretzbacher U, Magrini N, WHO Pathogens Priority List Working Group. 2018. Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. Lancet Infect Dis 18: 318–327. https://doi.org/10.1016/S1473-3099(17)30753-3.
- Horcajada JP, Montero M, Oliver A, Sorlí L, Luque S, Gómez-Zorrilla S, Benito N, Grau S. 2019. Epidemiology and treatment of multidrug-resistant and extensively drug-resistant Pseudomonas aeruginosa infections. Clin Microbiol Rev 32:e00031-19. https://doi.org/10.1128/CMR.00031-19.
- Rohde C, Wittmann J, Kutter E. 2018. Bacteriophages: a therapy concept against multi-drug-resistant bacteria. Surg Infect (Larchmt) 19:737–744. https://doi.org/10.1089/sur.2018.184.
- Leptihn S. 2019. Welcome back to the pre-penicillin era: why we desperately need new strategies in the battle against bacterial pathogens. Infect Microbes Dis 1:33. https://doi.org/10.1097/IM9.000000000000009.
- Loh B, Wang X, Hua X, Chook HW, Ma L, Zhang L, Manohar P, Jin Y, Leptihn S. 2021. Complete genome sequence of the lytic bacteriophage Phab24, which infects clinical strains of the nosocomial pathogen Acinetobacter baumannii. Microbiol Resour Announc 10:e00669-21. https://doi.org/10.1128/MRA.00669-21.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Ramsey J, Rasche H, Maughmer C, Criscione A, Mijalis E, Liu M, Hu JC, Young R, Gill JJ. 2020. Galaxy and Apollo as a biologist-friendly interface

for high-quality cooperative phage genome annotation. PLoS Comput Biol 16:e1008214. https://doi.org/10.1371/journal.pcbi.1008214.

- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. https://doi.org/10.1093/nar/gkh152.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res 25: 955–964. https://doi.org/10.1093/nar/25.5.955.
- Besemer J, Lomsadze A, Borodovsky M. 2001. GeneMarkS: a self-training method for prediction of gene starts in microbial genomes. Implications for finding sequence motifs in regulatory regions. Nucleic Acids Res 29: 2607–2618. https://doi.org/10.1093/nar/29.12.2607.
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. Nucleic Acids Res 27:4636–4641. https:// doi.org/10.1093/nar/27.23.4636.
- Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting speciesspecific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. DNA Res 15:387–396. https://doi.org/ 10.1093/dnares/dsn027.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/ S0022-2836(05)80360-2.
- 15. Bairoch A, Boeckmann B. 1994. The SWISS-PROT protein sequence data bank: current status. Nucleic Acids Res 22:3578–3580.
- Liu B, Zheng D, Jin Q, Chen L, Yang J. 2019. VFDB 2019: a comparative pathogenomic platform with an interactive Web interface. Nucleic Acids Res 47:D687–D692. https://doi.org/10.1093/nar/gky1080.
- Tynecki P, Guziński A, Kazimierczak J, Jadczuk M, Dastych J, Onisko A. 2020. PhageAl—bacteriophage life cycle recognition with machine learning and natural language processing. bioRxiv https://doi.org/10.1101/2020.07.11.198606.