



Antiangiogenesis therapy: an update after the first decade

Sandro De Falco

Angiogenesis LAB, Institute of Genetics and Biophysics 'Adriano Buzzati-Traverso', Napoli, Italy

Received: October 8, 2013 Accepted: November 19, 2013

Correspondence to Sandro De Falco, M.D.

Angiogenesis LAB, Institute of Genetics and Biophysics 'Adriano Buzzati-Traverso', Via Pietro Castellino 111, Napoli 80131, Italy Tel: +39-81-613-2354 Fax: +39-81-613-2706 E-mail: sandro.defalco@igb.cnr.it Angiogenesis is a complex biological phenomenon that forms new blood vessels from the pre-existing vasculature. Aberrant angiogenesis has been implicated in a variety of diseases such as cancer, atherosclerosis, arthritis, obesity, pulmonary hypertension, diabetic retinopathy, and age-related macular degeneration. These conditions collectively affect nearly 10% of the global population. Much effort has focused on identifying new therapeutic agents that inhibit pathological angiogenesis since 1971, when Judah Folkman published the hypothesis that tumor growth is angiogenesis-dependent and that its inhibition may be therapeutic. In 2004, the U.S. Food and Drug Administration approved the first antiangiogenic drug for the treatment of metastatic colon cancer, bevacizumab (Avastin, Genentech). This drug is a humanized monoclonal antibody that neutralizes the vascular endothelial growth factor. It is used in combination with chemotherapy, and its use began the era of antiangiogenesis therapy. Several new therapeutic agents have been added to the list of approved drugs, and clinical trials of new therapeutic options and antiangiogenic agents are ongoing. This review describes the progress made in the first decade of antiangiogenesis therapy, and addresses both validated and possible targets for future drug development.

Keywords: Angiogenesis; Neoplasms; Macular degeneration; Antibodies, monoclonal; Tyrosine kinase inhibitor

INTRODUCTION

From embryonic development to adulthood, blood vessels play a fundamental physiological role in supplying oxygen and nutrients, removing catabolic waste, and circulating cells for immune surveillance [1,2].

It is unsurprising that structural alterations or functional aberrations of vessels are involved in a plethora of diseases [3,4]. These diseases may be divided into two groups. The first involves inadequate vessel maintenance and growth; it includes diseases such as myocardial infarction, stroke, neurodegenerative or obesity-associated disorders, and requires proangiogenic therapy. The second involves disproportionate vascular growth and abnormal remodeling. This group includes cancer, inflammatory disorders, ophthalmic neovascular diseases, and requires antiangiogenic therapy (Table 1).

Previous therapeutic efforts that focused on stimulating angiogenesis using proangiogenic factors have failed. Drugs that block vessel growth have been successful, and have led to the approval of antiangiogenic drugs for some cancers and neovascular ophthalmic diseases [5-8].

FACTORS DRIVING PHYSIOLOGICAL AND PATHOLOGICAL ANGIOGENESIS

The development of functional vessels by angiogenesis



Table 1. Diseases that involve angiogenesis

Diseases with increased vascularization					
Cancer					
Hemangioma and vascular malformation					
Age-related macular degeneration					
Diabetic retinopathy					
Rheumatoid arthritis					
Osteoarthritis					
Atherosclerotic plaque progression					
Obesity					
Infectious diseases					
Psoriasis					
Asthma					
Endometriosis					
Diseases with decreased vascularization					
Ischemic cardiomyopathy					
Hind limb ischemia					
Stroke					
Systemic and pulmonary hypertension					
Atherosclerotic peripheral diseases					
Neurodegenerative disorders					
Alzheimer disease					
Nephropathy					

and arteriogenesis requires the cooperation of several growth factor families, their related receptors, multiple cell types, and the presence of certain conditions, such as hypoxia [9].

Understanding this process has allowed the identification of a large number of targets for the inhibition of angiogenesis. Some of these targets have been used for antiangiogenic therapy, whereas many others have the potential to become new validated targets. The following is a summary of the different activities of the molecule families that are active in angiogenesis.

The vascular endothelial growth factor (VEGF) family

VEGF (also known as VEGF-A) is the main member of the VEGF family, and plays a major role in angiogenesis. Its activity is exerted through the binding of two receptors: VEGF receptor 1 (VEGFR-1; also known as Flt-1) and VEGFR-2 (also known as KDR or Flk-1). The latter plays a main role in endothelial activation in conjunction with neuropilin (NRP) receptors 1 and 2 that act as coreceptors to enhance the activity of VEGFR-2 [10]. The soluble isoforms of VEGF stimulate vessel enlargement, whereas the isoforms that bind to the extracellular matrix promote vessel branching [11,12]. VEGF produced by endothelial cells maintains vascular homeostasis.

VEGF-C is a ligand of the VEGFR-2 and VEGFR-3 receptors. It plays an important role in stimulating endothelial cells to express the tip cell phenotype. These endothelial cells become motile, invasive, and protrude filopodia, which drives new vessel formation [13]. VEG-FR-3 plays a role in vascular formation during early embryogenesis. Later, it becomes a key regulator of lymphangiogenesis or the formation of new lymphatic vessels from pre-existing ones [14].

Placental growth factor (PlGF) is relevant only in pathological conditions [15-17]. The activation of its specific receptor, VEGFR-1, directly or indirectly stimulates angiogenesis. PlGF is able to recruit and stimulate bone marrow-derived endothelial progenitor and myeloid cells needed to sustain the angiogenic process [18]. PlGF contributes to the unequal polarization of tumor-associated macrophages (TAMs) between the M1 and M2 phenotypes [19].

Like PlGF, VEGF-B is not required for physiological angiogenesis and it specifically recognizes VEGFR-1. Its angiogenic activity is limited to certain tissues such as the heart [20]. Interestingly, PlGF and VEGF-B can stimulate the growth of new vessels without inducing adverse effects such as increased permeability or leakage, as observed in diverse preclinical models [21,22].

VEGFR-1 remains the most elusive in terms of angiogenic function. This is most likely due to the fact that it is expressed in different cell types and it is activated by three members of the VEGF family: VEGF-A, VEGF-B, and PlGF [23]. Based on its weak tyrosine kinase activity, VEGFR-1 has been defined as a decoy receptor for VEGF, and determines the amount of free VEGF available to activate VEGFR-2. This explains why VEGFR-1 loss results in vessel overgrowth [24]. Alternatively, VEGFR-1 activation on angiogenic endothelial, stromal, and myeloid cells stimulates pathological angiogenesis [25].

The platelet derived growth factor (PDGF) family

The maturation of new blood vessels by arteriogenesis occurs through the covering of nascent vessels by mu-



ral cells. The PDGF family contributes to this process along with angiopoietins (ANGs) and transforming growth factor β (TGF- β) [26]. PDGF-B is produced by endothelial cells to recruit pericytes from the local environment or perivascular pericyte progenitors from the bone marrow [27,28] that express PDGF receptor β . In turn, pericytes produce VEGF, which is essential for endothelial survival and protects vessels from VEGF blockade.

Other members of this family also have angiogenic functions. PDGF-CC stimulates vessel growth and maturation, but attenuates the response to anti-VEGF treatment [29]. Inhibition of PDGF-DD suppresses ocular neovascularization, whereas PDGF-DD overexpression normalizes tumor vessels and improves drug delivery.

The fibroblast growth factor (FGF) family

Basic FGF was one of the first angiogenic factors to be discovered. Other members of this family, such as FGF1 and FGF9, have angiogenic and arteriogenic properties. The activation of FGF receptors (FGFRs), on endothelial cells or other cell types, sustains angiogenesis directly or by inducing the release of angiogenic factors. This has been confirmed by data indicating that FGF signaling promotes tumor angiogenesis [30].

Low levels of FGF are needed to maintain vascular integrity because inhibition of FGFR signaling in quiescent endothelial cells causes vessel disintegration [31].

The ANG and tyrosine kinase endothelial (TIE) family

This family plays a crucial role in the maintenance of vessel quiescence but vessels must be able to respond to angiogenic stimuli [32].

The two main ligands of the ANG family, ANG1 and ANG2, have the same target receptor, TIE2. ANG2 acts as a competitive antagonist of ANG-1. A second receptor, TIE1, exists but a ligand has not yet been identified. The function of this orphan receptor remains elusive [32].

ANG1 stimulates TIE2 clustering at cell-cell junctions, which sustains vessel quiescence and promotes mural cell coverage of vessels [33]. In the presence of angiogenic stimuli, activated endothelial cells produce ANG2 that antagonizes ANG1/TIE2 signaling enhancing mural cell detachment, vascular permeability, and endothelial cell sprouting. ANG2 also promotes angiogenesis by recruiting proangiogenic TIE2-expressing monocytes (TEMs)[34].

The cadherin family

In quiescent vessels, the interconnections between endothelial cells are crucial to form a continuous monolayer. This formation has been described as a cellular phalanx. Vascular endothelial cadherin (VE-cadherin) is vital for endothelial cell-cell adhesion. VE-cadherin also promotes vessel stabilization by inhibiting VEG-FR-2 signaling and activating TGFR pathways [35]. VE-cadherin is strictly regulated by oxygen concentration in a feedback loop so that vessel perfusion may be optimized depending on the oxygen supply [36]. A reduction in VE-cadherin between adjacent cells by endocytosis occurs in response to angiogenic stimuli and supports new vessel formation. The localization of VE-cadherin in the filopodia of tip cells allows the establishment of new connections with other sprouting vessels.

The notch- and delta-like ligand 4 (DLL4) signaling

During new vessel formation, the tip cells migrate toward the angiogenic stimuli, while the neighboring endothelial cells (stalk cells) proliferate [8]. The activation of VEGFR-2 by VEGF in tip cells upregulates DLL4 expression, which determines the activation of NOTCH in stalk cells [37]. There is downregulation of VEGFR-2 and upregulation of VEGFR-1 in stalk cells, which become less responsive to VEGF as opposed to other VEGF family members able to bind VEGFR1. This demonstrates a complexity of endothelial cells by activation of a fine-tuning of VEGFR-1 and VEGFR-2 expression in response to neighboring stimuli that determine the tip or stalk functional phenotypes [38].

The ephrin (EPH) family

EPH receptors and ligands modulate cell contact dependent patterns. During vasculogenesis, arterial and venous territories are specified by EPH-B2 and EPHB4 expression, respectively. Their interactions avoid vessel repulsion while also separating the respective vessel territories. EPH-B2 also promotes the recruitment of mural cells and bone marrow-derived endothelial progenitor cells [39,40].



The semaphorin (SEMA) family

The SEMA family is composed of a large number of secreted or Gpi-anchored proteins that are able to bind plexin receptors or NRP coreceptors [41]. Many SEMAs inhibit tumor angiogenesis (SEMA3A, SEMA3B, SE-MA3D, SEMA3F, and SEMA4A), whereas SEMA3C and SEMA4D promote tumor angiogenesis. The loss of the *plexin-d1* receptor in mice induces erroneous vessel navigation, because the endothelial cells cannot recognize the repulsive SEMA3E signals.

APPROVED ANTIANGIOGENIC DRUGS FOR CANCER THERAPY AND AGE-RELATED MACU-LAR DEGENERATION

Despite the large number of putative targets for antiangiogenesis therapy and multiple reported compounds with antiangiogenic activity [5,9], the vast majority have failed despite encouraging preclinical results [7,42]. The exceptions are the drugs that target VEGF ligands or receptors [10]. Most of the antiangiogenic agents that entered the drug development pipeline in the past decade target VEGF signaling pathways. However, efforts using other targets such as ANGs and integrins are ongoing. The approved antiangiogenic drugs treat various tumors or age-related macular degeneration (AMD) (Table 2).

Bevacizumab (Avastin, Genentech, San Francisco, CA, USA), a humanized anti-VEGF-A monoclonal antibody [43], was the first antiangiogenic drug approved by the U.S. Food and Drug Administration. The clinical trial was a randomized double-blind phase-III study in which bevacizumab was administered in combination with a bolus of irinotecan, 5-fluorouracil, leucovorin chemotherapy as a first-line therapy for previously untreated metastatic colorectal cancer (mCRC). The median survival and progression-free survival times were increased with the addition of bevacizumab [44].

Currently, bevacizumab is approved for first- and second-line treatment of mCRC in conjunction with 5-fluorouracil-irinotecan or 5-fluorouracil-oxaliplatin-based chemotherapy. It is also approved for first-line treatment of advanced nonsquamous nonsmall cell lung cancer in combination with carboplatin and paclitaxel in patients who have not yet received chemotherapy. More recently, it has been approved as a single agent in adult patients with recurrent glioblastoma multiforme who had tumor progression after initial treatment, and in conjunction with interferon α to treat metastatic renal cell carcinoma (RCC) [45].

Although bevacizumab is generally well tolerated, some serious and unusual toxicities were observed, including: gastrointestinal perforation, hemorrhage, proteinuria, venous or arterial thromboembolic events, and hypertension. The most common adverse events were asthenia, abdominal pain, headache, diarrhea, nausea, and vomiting [6].

Aflibercept (Zaltrap, Regeneron and Sanofi Aventis, Bridgewater, NJ, USA), previously known as VEGF-trap, is a recombinant fusion protein that consists of VEGF-binding portions from the extracellular domains of human VEGFR 1 and 2 fused to the Fc portion of human immunoglobulin G1. It functions as a decoy receptor for proangiogenic members of the VEGF family: VEGF-A, VEGF-B, and PIGF. It has been approved for second-line treatment of mCRC that is resistant or has progressed following treatment with an oxaliplatin-containing regimen. It is used in combination with 5-fluorouracil, leucovorin, and irinotecan (FOLFIRI) [46].

The adverse effects were similar to those observed with bevacizumab, but common adverse reactions such as neutropenia, diarrhea, hypertension, leukopenia, stomatitis, fatigue, proteinuria, and asthenia, were reported more frequently with those treated with aflibercept and FOLFIRI.

Several multitargeted tyrosine kinase inhibitors (TKIS), which block the signaling of pathways such as VEGF, have been approved for cancer therapy. Sorafenib (Nexavar, Bayer HealthCare Pharmaceuticals, West Haven, CT, USA) targets VEGFRs, PDGFRs, FGFR1, Kit, and Raf kinases (mainly C-Raf as opposed to B-Raf). It was approved for first-line treatment of metastatic RCC in 2005 [47] and for the treatment of unresectable hepatocellular carcinoma in 2007 [48].

Sunitinib (Sutent, Pfizer Inc., New York, NY, USA) inhibits the tyrosine phosphorylation of VEGFRs, PDG-FRs, c-kit, and Flt-3, and was approved for first line treatment of metastatic RCC in 2006 [49]. It was also approved for the treatment of gastrointestinal stromal tumors (GIST) following progression during treatment with imatinib (Gleevec, Novartis, Basel, Switzerland) or



Table 2. Approved antiangiogenic drugs

Drug/Company	Type of inhibitor	Targets	Use	Indication
Bevacizumab	Monoclonal Antibody	VEGF-A		Metastatic colorectal cancer
(Avastin)			First line	Nonsmall cell lung cancer
Genentech/Roche			First line	Recurrent glioblastoma
			First line	Metastatic renal cell carcinoma
			Off-label	Wet age-related macular degeneration
A (1:1	C1 '		C	Macular edema following CRVO
Aflibercept	Chimeric soluble	VEGF-A, VEGF-B,	Second line	Metastatic colorectal cancer
(Zaltrap)	receptor	PlGF		
Regeneron/Sanofi Pegaptanib	Descripted entermon	VECE A.G.	First line	Wet and related meaning decomposition
(Macugen)	Pegylated aptamer	VEGF-A165	First line	Wet age-related macular degeneration
Eyetech Pharms				
Ranibizumab	Fab fragment of	VEGF-A	First line	Wet age-related macular degeneration
(Lucentis)	antibody	VLGI-M	1 list line	Macular edema following CRVO
Genentech/Roche	antibody			Macular cucina following City o
Aflibercept	Chimeric soluble	VEGF-A, VEGF-B,	First line	Wet age-related macular degeneration
(Eleya)	receptor	PlGF	1110011110	Macular edema following CRVO
Regeneron/Bayer				
Sorafenib	Tyrosine kinase	VEGFR, PDGFRs,	First line	Metastatic renal cell carcinoma
(Nexavar)	inhibitor	FGFR1, KIT, RAF	First line	Unresectable hepatocellular
Bayer/Onyx		, ,		carcinoma
Sunitinib	Tyrosine kinase	VEGFRs, PDGFRs,	First line	Metastatic renal cell carcinoma
(Sutent)	inhibitor	KIT, FLT-3	Second line	Gastrointestinal stromal tumor
Pfizer			First line	Unresectable pancreatic
				neuroendocrine tumors
Pazopanib	Tyrosine kinase	VEGFRs, PDGFRs,	First line	Metastatic renal cell carcinoma
(Votrient)	inhibitor	KIT	Second line	Advanced soft tissue sarcoma
GlaxoSmithKline				
Axitinib	Tyrosine kinase	VEGFRs, PDGFRs,	First line	Metastatic renal cell carcinoma
(Inlyta)	inhibitor	KIT		
Pfizer				
Vandetanib	Tyrosine kinase	VEGFRs, EGFR,	First line	Late-stage medullary thyroid cancer
(Caprelsa)	inhibitor	RET		
Astra Zeneca				
Cabozantinib	Tyrosine kinase	VEGFR2, RET,	First line	Progressive medullary thyroid cancer
(Cometriq)	inhibitor	MET		
Exelixis	m · 1·		c 11:	
Regorafenib	Tyrosine kinase	VEGFRs, TIE ₂ ,	Second line	Metastatic colorectal cancer
(Stivarga)	inhibitor	PDGFRs, RET, KIT,	Second line	Gastrointestinal stromal tumor
Bayer/Onyx		FGFRs		

VEGF-A, vascular endothelial growth factor A; CRVO, central retinal vein occlusion; VEGF-B, vascular endothelial growth factor B; PlGF, placental growth factor; VEGFR, vascular endothelial growth factor receptor; PDGFR, platelet derived growth factor receptor; FGFR, fibroblast growth factor receptor; KIT, v-kit feline sarcoma viral oncogene homolog; RAF, v-raf-1 murine leukemia viral oncogene homolog 1; FLT-3, fms-related tyrosine kinase 3; EGFR, epidermal growth factor receptor; RET, ret proto-oncogene; MET, met proto-oncogene; TIE, TEK tyrosine kinase endothelial.

with intolerance to the drug [50]. More recently (May 2011), sunitinib has been approved for the treatment of pancreatic neuroendocrine tumors in patients with un-

resectable, locally advanced, or metastatic disease [51]. Pazopanib (Votrient, GlaxoSmithKline, Stevenage, UK) targets VEGFRs, PDGFRs, and c-kit, and was approved in

кјім≁

2009 for first-line treatment of metastatic RCC [52]. In April 2012, it was also approved for the treatment of advanced soft tissue sarcoma in patients who had received prior chemotherapy [53]. Axitinib (Inlyta, Pfizer Inc.) has the same targets as Pazopanib and was approved in January 2012 for the treatment of advanced RCC that progressed despite first-line therapy [54].

Vandetanib (Caprelsa, AstraZeneca, Wilmington, DE, USA) inhibits VEGFRs, EGFR, and RET-tyrosine kinase. It was approved in April 2011 for the treatment of symptomatic or progressive medullary thyroid cancer (MTC) in patients with unresectable, locally advanced, or metastatic disease [55].

Cabozantinib (Cometriq, Exselixis Inc., San Francisco, CA, USA) inhibits the activity of multiple tyrosine kinases, including RET, MET, and VEGFR2. It was approved in November 2012 for the first-line treatment of patients with progressive metastatic MTC [56].

Regorafenib (Stivarga, Bayer HealthCare Pharmaceuticals) was approved in September 2012 for second-line treatment of patients with mCRC who have been previously been treated with 5-fluorouracil, oxaliplatin, and irinotecan based chemotherapy, with an anti-VEGF therapy, and, if KRAS wild type, with an anti-EGFR therapy [57]. In February 2013, it was also approved for the second-line treatment of patients with advanced GIST that cannot be surgically resected and no longer responds to imatinib and sunitinib [58]. Regorafenib and its active metabolites inhibit multiple membrane-bound and intracellular kinases that are involved in normal cellular functions and pathologic processes, including: RET, VEGFRs, KIT, PDGFRs, FGFRs, TIE2, and Raf.

The listed TKIs have many adverse effects; the most common include: fatigue, diarrhea, nausea, and vomiting. Hypertension, as with bevacizumab and aflibercept, also occurs with TKIs, and often requires medical intervention with standard antihypertensive therapy. Hypertension developed in 11% of patients receiving bevacizumab and is now recognized as a standard side effect of VEGF blockers [6].

Three anti-VEGF drugs are used for the treatment of the wet (neovascular) form of AMD. AMD causes blindness due to the formation of leaky neovessels [59].

The first, Pegaptanib (Macugen, Eyetech, New York, NY, USA), is a pegylated, 28-base ribonucleic aptamer that specifically targets the VEGF165 isoform that was ap-

proved in 2004. Macugen slowed the progression of vision loss, but did not yield clinically significant improvements in visual acuity [60].

The second drug, ranibizumab (Lucentis, Genentech), is a recombinant, humanized monoclonal antibody Fab fragment derived from bevacizumab that was approved in 2006. Phase 3 clinical studies demonstrated that monthly intravitreal injections of ranibizumab prevented the loss of visual acuity in ~95% of patients and improved visual acuity in 25% to 33% of patients during a 24-month treatment period [60,61]. Ocular adverse events, such as endophthalmitis or uveitis, attributable to either the injection procedure (intravitreal) or Ranibizumab were observed in 1% to 2% of patients during the 2-year treatment period. With respect to the potential systemic anti-VEGF side effects, the rates of hypertension were not imbalanced, and adverse events associated with proteinuria were not reported. Ranibizumab has been also approved for the treatment of macular edema following central retinal vein occlusion (CRVO)[62].

Ophthalmologists have begun to use off label Bevacizumab instead of Ranibizumab due to the enormous difference in terms of cost and reports of similar beneficial effects [63].

In November 2011, Aflibercept (Eylea, Regeneron, Tarrytown, NY, USA) has also been approved for the treatment of patients with AMD and macular edema following CRVO [64].

Since VEGF plays an important role in the physiological homeostasis of the retina [65], important side effects have been reported with respect to retinal function [66,67]. However, several clinical trials with Ranibizumab, Bevacizumab, and Aflibercept are ongoing to explore the treatment of neovascular ocular pathologies, such as diabetic retinopathy and retinopathy of prematurity [68,69].

ANTIANGIOGENIC DRUGS IN CLINICAL DE-VELOPMENT

Tens of clinical trials are ongoing for new antiangiogenic agents, many of which are receptor TKIs. Please see http://www.cancer.gov/clinicaltrials or http://clinicaltrials.gov/ct2/home for more information. For other

клм≁

agents, it is interesting to note drugs directed towards specific targets that are not related to the mechanism of action of currently approved drugs.

Trebanabib (Amgen, Thousand Oaks, CA, USA), a peptide fused to the Fc portion of an antibody (peptibody), potently and selectively inhibits ANG1 and ANG2 binding to the TIE2 tyrosine kinase receptor [70]. It is currently in phase 3 clinical trials for the treatment of multiple tumor types.

Velociximab (PDL BioPharma, Fremont, CA, USA; and Biogen Idec, Cambridge, MA, USA), a chimeric monoclonal antibody that inhibits a5b1 integrin functional activity has passed phase 2 trials [71].

The first agent able to target integrins that has reached phase 3 of clinical development, Cilengitide (Merck KGaA, Darmstadt, Germany) [72], is a cyclic peptide that blocks av integrins. Unfortunately, this drug did not meet the primary endpoint in patients with newly diagnosed glioblastoma.

In addition, two promising human antibodies that also interfere with VEGF pathways, specifically targeting VEGFR-1 or VEGFR-2, are in clinical development. Ramucirumab (ImClone/Lilly, ImClone Systems, Branchburg, NJ, USA) blocks VEGFR-2 signaling and has recently met the primary endpoint of improved overall survival. It also prolonged the progression-free survival time in patients with metastatic gastric cancer [73]. Ramucirumab and Icrucumab (ImClone/Lilly), which blocks VEGFR-1 [74] are currently undergoing diverse phase 3 studies for multiple tumor types.

CONCLUSIONS

Despite great success in the development of several antiangiogenic drugs, the clinical outcomes in cancer patients treated with antiangiogenic therapy have indicated this strategy to be more challenging than anticipated [9,75-77].

Clinical benefit is reflected by the lengthening of progression-free survival time in advanced colorectal, lung, renal, pancreatic, neuroendocrine, and ovarian cancer and by the longer overall survival times in patients with metastatic colorectal and renal cancer. Antiangiogenic treatment generally prolongs the survival of responsive patients in the order of months, but the clinical benefit is not usually sustained or is decreased to absent with some types of cancer. Many cancer therapies have modest effects on overall survival times. Increases in overall survival were obtained in only 12% of 73 randomized phase 3 trials for bevacizumab, trastuzumab, and other targeted therapies in addition to a range of chemotherapeutic agents for metastatic breast cancer over the past 30 years [78].

Another limitation of antiangiogenic cancer treatment is evidenced by the results of both preclinical and clinical research into angiogenic mechanisms. Tumors can adapt to the presence of angiogenesis inhibitors so as to functionally evade the therapeutic blockade of angiogenesis [75]. In contrast to the traditional concept of drug resistance acquisition based on target gene mutation or alteration in drug uptake and efflux [79], evasive resistance for angiogenesis is essentially indirect [75]. VEGF blockade can aggravate hypoxia resulting in upregulation of angiogenic factors, such as PIGF, FGFs, chemokines, and EPHs, and also lead to recruitment of proangiogenic bone marrow-derived cells, including TEMs and TAMs. These responses may rescue tumor vascularization.

The observation that cancer cells can adapt by aggressively migrating into normal tissue after VEGF blockade [80,81] has opened an important debate on whether antiangiogenic treatment may lead to more invasive and metastatic tumors [82]. This raises new concerns that must be considered in future angiogenic therapy development.

Currently, the main concern is how antiangiogenic cancer therapy can be improved. An important focus will be the ability of drugs to recognize more angiogenic targets. The importance of this quality is evidenced by the fact that TKIs are effective as monotherapy since they act on several targets, while anti-VEGF antibodies or recombinant proteins require a combination with chemotherapy. Another consideration regards the dose and duration optimization for antiangiogenic drug delivery since these have not been clinically optimized [83]. Moreover, anti-VEGF agents could be combined with agents that target the escape pathways identified in clinical studies of ANG2, PIGF, stromal cell-derived growth factor 1 α , and chemokine receptor type 4 [84].

The ability to prevent inflammatory cell recruitment [85] and tumor vessel normalization [86] should be ex-

кјім≁

plored in the future to improve anticancer immune therapy and prevent metastatic spread.

In contrast to cancer therapy outcomes, those obtained for AMD or CRVO have been more impressive. This may be due to the fact that the neovascularization associated with AMD is the primary cause for irreversible blindness in advanced countries. Anti-VEGF agents used as monotherapy prevented the loss of visual acuity in ~95% of patients, and improved visual acuity in 25% to 33%, which was never obtained with previous treatments [61,63]. Also for ocular therapy, the ability to block more targets involved in neovascular formation, particularly those associated with the immunological component of the disease [87], as well as preventing the numerous side effects, could represent a further advancement in therapeutic success.

In conclusion, the clinical data from the first decade of antiangiogenesis therapy along with the knowledge that pathological angiogenesis plays a role in a large number of complex diseases should result in further advances in angiogenic therapy.

Conflict of interest

No potential conflict of interest relevant to this article is reported.

Acknowledgments

The author would like to thank Anna Maria Aliperti for editing the manuscript. S.D.F. was supported by the Associazione Italiana Ricerca sul Cancro (AIRC grant no. IG 11420).

REFERENCES

- 1. Folkman J, Shing Y. Angiogenesis. J Biol Chem 1992; 267:10931-10934.
- 2. Carmeliet P. Angiogenesis in health and disease. Nat Med 2003;9:653-660.
- 3. Ferrara N, Kerbel RS. Angiogenesis as a therapeutic target. Nature 2005;438:967-974.
- 4. Carmeliet P. Angiogenesis in life, disease and medicine. Nature 2005;438:932-936.
- 5. Folkman J. Angiogenesis: an organizing principle for drug discovery? Nat Rev Drug Discov 2007;6:273-286.
- 6. Ferrara N, Mass RD, Campa C, Kim R. Targeting VEGF-A

to treat cancer and age-related macular degeneration. Annu Rev Med 2007;58:491-504.

- 7. Crawford Y, Ferrara N. VEGF inhibition: insights from preclinical and clinical studies. Cell Tissue Res 2009;335:261-269.
- 8. Potente M, Gerhardt H, Carmeliet P. Basic and therapeutic aspects of angiogenesis. Cell 2011;146:873-887.
- Carmeliet P, Jain RK. Molecular mechanisms and clinical applications of angiogenesis. Nature 2011;473:298-307.
- Tugues S, Koch S, Gualandi L, Li X, Claesson-Welsh L. Vascular endothelial growth factors and receptors: anti-angiogenic therapy in the treatment of cancer. Mol Aspects Med 2011;32:88-111.
- Lee S, Chen T'T, Barber CL, et al. Autocrine VEGF signaling is required for vascular homeostasis. Cell 2007;130:691-703.
- 12. Stockmann C, Doedens A, Weidemann A, et al. Deletion of vascular endothelial growth factor in myeloid cells accelerates tumorigenesis. Nature 2008;456:814-818.
- 13. Tvorogov D, Anisimov A, Zheng W, et al. Effective suppression of vascular network formation by combination of antibodies blocking VEGFR ligand binding and receptor dimerization. Cancer Cell 2010;18:630-640.
- 14. Tammela T, Alitalo K. Lymphangiogenesis: molecular mechanisms and future promise. Cell 2010;140:460-476.
- 15. Carmeliet P, Moons L, Luttun A, et al. Synergism between vascular endothelial growth factor and placental growth factor contributes to angiogenesis and plasma extravasation in pathological conditions. Nat Med 2001;7:575-583.
- 16. Gigante B, Morlino G, Gentile MT, Persico MG, De Falco S. Plgf-/-eNos-/- mice show defective angiogenesis associated with increased oxidative stress in response to tissue ischemia. FASEB J 2006;20:970-972.
- 17. De Falco S. The discovery of placenta growth factor and its biological activity. Exp Mol Med 2012;44:1-9.
- 18. Tarallo V, Vesci L, Capasso O, et al. A placental growth factor variant unable to recognize vascular endothelial growth factor (VEGF) receptor-1 inhibits VEGF-dependent tumor angiogenesis via heterodimerization. Cancer Res 2010;70:1804-1813.
- 19. Rolny C, Mazzone M, Tugues S, et al. HRG inhibits tumor growth and metastasis by inducing macrophage polarization and vessel normalization through down-



regulation of PlGF. Cancer Cell 2011;19:31-44.

- 20. Hagberg CE, Falkevall A, Wang X, et al. Vascular endothelial growth factor B controls endothelial fatty acid uptake. Nature 2010;464:917-921.
- Luttun A, Tjwa M, Moons L, et al. Revascularization of ischemic tissues by PlGF treatment, and inhibition of tumor angiogenesis, arthritis and atherosclerosis by anti-Flt1. Nat Med 2002;8:831-840.
- 22. Bry M, Kivela R, Holopainen T, et al. Vascular endothelial growth factor-B acts as a coronary growth factor in transgenic rats without inducing angiogenesis, vascular leak, or inflammation. Circulation 2010;122:1725-1733.
- 23. De Falco S, Gigante B, Persico MG. Structure and function of placental growth factor. Trends Cardiovasc Med 2002;12:241-246.
- 24. Fischer C, Mazzone M, Jonckx B, Carmeliet P. FLT1 and its ligands VEGFB and PlGF: drug targets for anti-angiogenic therapy? Nat Rev Cancer 2008;8:942-956.
- 25. Schwartz JD, Rowinsky EK, Youssoufian H, Pytowski B, Wu Y. Vascular endothelial growth factor receptor-1 in human cancer: concise review and rationale for development of IMC-18F1 (human antibody targeting vascular endothelial growth factor receptor-1). Cancer 2010;116(4 Suppl):1027-1032.
- 26. Jain RK. Molecular regulation of vessel maturation. Nat Med 2003;9:685-693.
- 27. Hellberg C, Ostman A, Heldin CH. PDGF and vessel maturation. Recent Results Cancer Res 2010;180:103-114.
- Song S, Ewald AJ, Stallcup W, Werb Z, Bergers G. PDG-FRbeta+ perivascular progenitor cells in tumours regulate pericyte differentiation and vascular survival. Nat Cell Biol 2005;7:870-879.
- 29. di Tomaso E, London N, Fuja D, et al. PDGF-C induces maturation of blood vessels in a model of glioblastoma and attenuates the response to anti-VEGF treatment. PLoS One 2009;4:e5123.
- Beenken A, Mohammadi M. The FGF family: biology, pathophysiology and therapy. Nat Rev Drug Discov 2009;8:235-253.
- Murakami M, Nguyen LT, Zhuang ZW, et al. The FGF system has a key role in regulating vascular integrity. J Clin Invest 2008;118:3355-3366.
- 32. Augustin HG, Koh GY, Thurston G, Alitalo K. Control of vascular morphogenesis and homeostasis through the angiopoietin-Tie system. Nat Rev Mol Cell Biol 2009;10:165-177.

- 33. Saharinen P, Eklund L, Miettinen J, et al. Angiopoietins assemble distinct Tie2 signalling complexes in endothelial cell-cell and cell-matrix contacts. Nat Cell Biol 2008;10:527-537.
- 34. De Palma M, Venneri MA, Galli R, et al. Tie2 identifies a hematopoietic lineage of proangiogenic monocytes required for tumor vessel formation and a mesenchymal population of pericyte progenitors. Cancer Cell 2005;8:211-226.
- 35. Dejana E, Tournier-Lasserve E, Weinstein BM. The control of vascular integrity by endothelial cell junctions: molecular basis and pathological implications. Dev Cell 2009;16:209-221.
- 36. Mazzone M, Dettori D, Leite de Oliveira R, et al. Heterozygous deficiency of PHD2 restores tumor oxygenation and inhibits metastasis via endothelial normalization. Cell 2009;136:839-851.
- Thurston G, Noguera-Troise I, Yancopoulos GD. The Delta paradox: DLL4 blockade leads to more tumour vessels but less tumour growth. Nat Rev Cancer 2007;7:327-331.
- Jakobsson L, Franco CA, Bentley K, et al. Endothelial cells dynamically compete for the tip cell position during angiogenic sprouting. Nat Cell Biol 2010;12:943-953.
- 39. Yancopoulos GD, Davis S, Gale NW, Rudge JS, Wiegand SJ, Holash J. Vascular-specific growth factors and blood vessel formation. Nature 2000;407:242-248.
- 40. Mosch B, Reissenweber B, Neuber C, Pietzsch J. Eph receptors and ephrin ligands: important players in angiogenesis and tumor angiogenesis. J Oncol 2010; 2010:135285.
- Gu C, Giraudo E. The role of semaphorins and their receptors in vascular development and cancer. Exp Cell Res 2013;319:1306-1316.
- 42. Kerbel R, Folkman J. Clinical translation of angiogenesis inhibitors. Nat Rev Cancer 2002;2:727-739.
- 43. Presta LG, Chen H, O'Connor SJ, et al. Humanization of an anti-vascular endothelial growth factor monoclonal antibody for the therapy of solid tumors and other disorders. Cancer Res 1997;57:4593-4599.
- 44. Hurwitz H, Fehrenbacher L, Novotny W, et al. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. N Engl J Med 2004;350:2335-2342.
- 45. Scott AM, Wolchok JD, Old LJ. Antibody therapy of can-



cer. Nat Rev Cancer 2012;12:278-287.

- 46. Ciombor KK, Berlin J, Chan E. Aflibercept. Clin Cancer Res 2013;19:1920-1925.
- 47. Escudier B, Eisen T, Stadler WM, et al. Sorafenib in advanced clear-cell renal-cell carcinoma. N Engl J Med 2007;356:125-134.
- 48. Llovet JM, Ricci S, Mazzaferro V, et al. Sorafenib in advanced hepatocellular carcinoma. N Engl J Med 2008;359:378-390.
- 49. Motzer RJ, Michaelson MD, Redman BG, et al. Activity of SU11248, a multitargeted inhibitor of vascular endothelial growth factor receptor and platelet-derived growth factor receptor, in patients with metastatic renal cell carcinoma. J Clin Oncol 2006;24:16-24.
- 50. Demetri GD, van Oosterom AT, Garrett CR, et al. Efficacy and safety of sunitinib in patients with advanced gastrointestinal stromal tumour after failure of imatinib: a randomised controlled trial. Lancet 2006;368:1329-1338.
- Motzer RJ, Hutson TE, Tomczak P, et al. Sunitinib versus interferon alfa in metastatic renal-cell carcinoma. N Engl J Med 2007;356:115-124.
- 52. Ward JE, Stadler WM. Pazopanib in renal cell carcinoma. Clin Cancer Res 2010;16:5923-5927.
- 53. Schoffski P. Pazopanib in the treatment of soft tissue sarcoma. Expert Rev Anticancer Ther 2012;12:711-723.
- 54. Ho TH, Jonasch E. Axitinib in the treatment of metastatic renal cell carcinoma. Future Oncol 2011;7:1247-1253.
- 55. Wells SA Jr, Robinson BG, Gagel RF, et al. Vandetanib in patients with locally advanced or metastatic medullary thyroid cancer: a randomized, double-blind phase III trial. J Clin Oncol 2012;30:134-141.
- 56. Elisei R, Schlumberger MJ, Muller SP, et al. Cabozantinib in progressive medullary thyroid cancer. J Clin Oncol 2013;31:3639-3646.
- 57. Andre T, Dumont SN. Regorafenib approved in metastatic colorectal cancer. Bull Cancer 2013;100:1027-1029.
- Lyseng-Williamson KA. Regorafenib: a guide to its use in advanced gastrointestinal stromal tumor (GIST) after failure of imatinib and sunitinib. BioDrugs 2013;27:525-531.
- 59. Ambati J, Ambati BK, Yoo SH, Ianchulev S, Adamis AP. Age-related macular degeneration: etiology, pathogenesis, and therapeutic strategies. Surv Ophthalmol 2003;48:257-293.
- 60. Brown DM, Kaiser PK, Michels M, et al. Ranibizumab versus verteporfin for neovascular age-related macular

degeneration. N Engl J Med 2006;355:1432-1444.

- Rosenfeld PJ, Brown DM, Heier JS, et al. Ranibizumab for neovascular age-related macular degeneration. N Engl J Med 2006;355:1419-1431.
- 62. Garnock-Jones KP. Ranibizumab: in macular oedema following retinal vein occlusion. Drugs 2011;71:455-463.
- CAT'T Research Group, Martin DF, Maguire MG, et al. Ranibizumab and bevacizumab for neovascular age-related macular degeneration. N Engl J Med 2011;364:1897-1908.
- 64. Schmidt-Erfurth U, Kaiser PK, Korobelnik JF, et al. Intravitreal aflibercept injection for neovascular age-related macular degeneration: ninety-six-week eesults of the VIEW studies. Ophthalmology 2013.
- 65. Famiglietti EV, Stopa EG, McGookin ED, Song P, LeBlanc V, Streeten BW. Immunocytochemical localization of vascular endothelial growth factor in neurons and glial cells of human retina. Brain Res 2003;969:195-204.
- 66. Sayanagi K, Sharma S, Kaiser PK. Photoreceptor status after antivascular endothelial growth factor therapy in exudative age-related macular degeneration. Br J Ophthalmol 2009;93:622-626.
- 67. Yodoi Y, Tsujikawa A, Nakanishi H, et al. Central retinal sensitivity after intravitreal injection of bevacizumab for myopic choroidal neovascularization. Am J Ophthalmol 2009;147:816-824.
- Mintz-Hittner HA, Kennedy KA, Chuang AZ; BEAT-ROP Cooperative Group. Efficacy of intravitreal bevacizumab for stage 3+ retinopathy of prematurity. N Engl J Med 2011;364:603-615.
- 69. Kimoto K, Kubota T. Anti-VEGF agents for ocular angiogenesis and vascular permeability. J Ophthalmol 2011;2012:852183.
- 70. Peeters M, Strickland AH, Lichinitser M, et al. A randomised, double-blind, placebo-controlled phase 2 study of trebananib (AMG 386) in combination with FOLFIRI in patients with previously treated metastatic colorectal carcinoma. Br J Cancer 2013;108:503-511.
- 71. Besse B, Tsao LC, Chao DT, et al. Phase Ib safety and pharmacokinetic study of volociximab, an anti-alpha-5beta1 integrin antibody, in combination with carboplatin and paclitaxel in advanced non-small-cell lung cancer. Ann Oncol 2013;24:90-96.
- 72. Macdonald TJ, Vezina G, Stewart CF, et al. Phase II study of cilengitide in the treatment of refractory or relapsed high-grade gliomas in children: a report from the Chil-



dren's Oncology Group. Neuro Oncol 2013;15:1438-1444.

- 73. Clarke JM, Hurwitz HI. Targeted inhibition of VEGF receptor 2: an update on ramucirumab. Expert Opin Biol Ther 2013;13:1187-1196.
- 74. Wu Y, Zhong Z, Huber J, et al. Anti-vascular endothelial growth factor receptor-1 antagonist antibody as a therapeutic agent for cancer. Clin Cancer Res 2006;12:6573-6584.
- 75. Bergers G, Hanahan D. Modes of resistance to anti-angiogenic therapy. Nat Rev Cancer 2008;8:592-603.
- 76. Ribatti D. The inefficacy of antiangiogenic therapies. J Angiogenes Res 2010;2:27.
- 77. Sennino B, McDonald DM. Controlling escape from angiogenesis inhibitors. Nat Rev Cancer 2012;12:699-709.
- 78. Verma S, McLeod D, Batist G, Robidoux A, Martins IR, Mackey JR. In the end what matters most? A review of clinical endpoints in advanced breast cancer. Oncologist 2011;16:25-35.
- 79. O'Connor R, Clynes M, Dowling P, O'Donovan N, O'Driscoll L. Drug resistance in cancer: searching for mechanisms, markers and therapeutic agents. Expert Opin Drug Metab Toxicol 2007;3:805-817.
- 80. Rubenstein JL, Kim J, Ozawa T, et al. Anti-VEGF antibody

treatment of glioblastoma prolongs survival but results in increased vascular cooption. Neoplasia 2000;2:306-314.

- Du R, Lu KV, Petritsch C, et al. HIF1alpha induces the recruitment of bone marrow-derived vascular modulatory cells to regulate tumor angiogenesis and invasion. Cancer Cell 2008;13:206-220.
- Ebos JM, Kerbel RS. Antiangiogenic therapy: impact on invasion, disease progression, and metastasis. Nat Rev Clin Oncol 2011;8:210-221.
- Kerbel RS, Kamen BA. The anti-angiogenic basis of metronomic chemotherapy. Nat Rev Cancer 2004;4:423-436.
- Jain RK, Duda DG, Willett CG, et al. Biomarkers of response and resistance to antiangiogenic therapy. Nat Rev Clin Oncol 2009;6:327-338.
- 85. Grunewald M, Avraham I, Dor Y, et al. VEGF-induced adult neovascularization: recruitment, retention, and role of accessory cells. Cell 2006;124:175-189.
- Jain RK. Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. Science 2005;307:58-62.
- Ambati J, Atkinson JP, Gelfand BD. Immunology of age-related macular degeneration. Nat Rev Immunol 2013;13:438-451.