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The potential predictive value of DEK expression for neoadjuvant chemoradiotherapy response in locally advanced rectal cancer

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Abstract

Background: Limited data are available regarding the ability of biomarkers to predict complete pathological response to neoadjuvant chemoradiotherapy in locally advanced rectal cancer. Complete response translates to better patient survival. DEK is a transcription factor involved not only in development and progression of different types of cancer, but is also associated with treatment response. This study aims to analyze the role of DEK in complete pathological response following chemoradiotherapy for locally advanced rectal cancer.

Methods: Pre-treated tumour samples from 74 locally advanced rectal-cancer patients who received chemoradiation therapy prior to total mesorectal excision were recruited for construction of a tissue microarray. DEK immunoreactivity from all samples was quantified by immunohistochemistry. Then, association between positive stained tumour cells and pathologic response to neoadjuvant treatment was measured to determine optimal predictive power.

Results: DEK expression was limited to tumour cells located in the rectum. Interestingly, high percentage of tumour cells with DEK positiveness was statistically associated with complete pathological response to neoadjuvant treatment based on radiotherapy and fluoropyrimidine-based chemotherapy and a marked trend toward significance between DEK positiveness and absence of treatment toxicity. Further analysis revealed an association between DEK and the pro-apoptotic factor P38 in the pre-treated rectal cancer biopsies.

Conclusions: These data suggest DEK as a potential biomarker of complete pathological response to treatment in locally advanced rectal cancer.

Keywords: DEK, Chemoradiotherapy, Neoadjuvant treatment, Rectal cancer, Predictive biomarker, Complete pathological response

Background

Colorectal cancer is one of the most common gastrointestinal malignant tumours in the world and has one of the highest rates of morbidity and mortality worldwide. It is not only the third most common malignancy in United States but also the third leading cause of cancer-related deaths [1]. Rectal cancer accounts for between 27% and

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58% of all cases of colorectal cancer, with variations attributable to the cancer registry studied and the method used to classify rectosigmoid tumours [2]. Of the 304,930 new cases of digestive-tract cancer diagnosed in 2016 in the United States, 39,220 were rectal, with higher incidence seen among males than females (23,110 vs. 16,110) [1]. Further information about the global incidence of rectal cancer can be obtained from the World Health Organization (WHO)-GLOBOCAN [3, 4].

A distinction must be made between rectal and colon carcinoma, as rectal cancer has a distinct dissemination pattern. Furthermore, surgical resection is the mainstay



© The Author(s). 2018 **Open Access** This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated. of curative treatment for rectal adenocarcinomas [5]. Colon carcinoma is located in the peritoneal cavity, an area that is highly accessible and facilitates surgical intervention with wide resection margins. In contrast, rectal cancer is located extraperitoneally, within the pelvis, thus it makes harder the surgical resection that in most of cases involve low anterior or abdominoperineal resection. Some rectal tumours are superficial (T0/T1) and small enough (< 3 cm) to be successfully resected by local excision. However, most patients have more deeply invasive tumours that are adherent or fixed to adjoining structures (e.g., sacrum, pelvic sidewalls, prostate, or bladder) that requires more extensive resection [6].

Rectal tumours tend toward local recurrence, and surgery alone only provides a high cure rate for patients with early-stage disease [7]. In fact, the five-year survival rate for patients with stage I tumours is around 80 to 90%, while this rate is below 80 for those with stage II or III disease [8].

To increase long-term survival, the Swedish Study Group has introduced neoadjuvant treatment for locally advanced tumours based on chemotherapy combined with radiation [9]. The effects of chemoradiotherapy are the results of DNA damage produced directly by ionizing radiations; or indirectly, by the action of chemical radicals generated from ionization [10]. Chemoradiotherapy improves survival rates and local recurrence by reducing tumour size and stage, and also has the ability to achieve pathologic downstaging [11, 12]. For these reasons, neoadjuvant chemotherapy is the standard of care for stage II-III rectal tumours, not only to increase the effectiveness of radiotherapy but also to attain negative surgical margins [13] and enhance the possibility for sphincter-preserving surgery [14]. As described by Ryan et al., tumour regression grade is a useful method of scoring pathologic response to chemoradiotherapy in rectal carcinomas [15]. However, complete pathological response has been reported in only 10% to 30% of patients, and around 40% show partial or no response [16].

To predict response to neoadjuvant treatment, translational research has focused on the search for potential biomarkers of response to preoperative treatment [17–19].

DEK was identified fusioned with the CAN nucleoporin due to the translocation t (6;9) in a subtype of acute myeloid leukemia [20]. DEK is overexpressed in multiple neoplasms, including bladder cancer [21], breast cancer [22], glioblastoma [23], hepatocellular carcinoma [24], melanoma [25], retinoblastoma [26, 27], and other types, such as oral, ovarian, or uterine-cervical cancer [28–31].

Functionally, DEK is involved in the DNA damage repair machinery from the interaction with PARP-1 [32], suppresses apoptosis, senescence, differentiation, and promotes cell transformation both in vitro and in vivo [33–35]. Our group has previously associated DEK expression with adjuvant-treatment response in colorectal cancer [36]. Here, we observed a significant increase in apoptotic cells after the combination of irinotecan treatment and DEK knock-down, compared to those treated with irinotecan or DEK knock-down individually. However, this effect was not observed with 5FU or oxaliplatin treatments alone or in combination with DEK knock-down [36].

DEK has also been described to have a high statistical power to predict pathological complete response for neoadjuvant chemotherapy in breast cancer [37].

Therefore, our hypothesis to link DEK with neoadjuvant therapy in rectal cancer has been based on the above-mentioned reports that associated DEK with treatment response.

This study aimed to explore the precise role of DEK as a novel biomarker of pathologic response in rectal adenocarcinoma. To achieve this, 74 biopsies obtained from pre-treated locally advanced rectal-adenocarcinoma patients were immunostained with DEK. Association with neoadjuvant chemoradiotherapy response was assessed in light of these findings.

Methods

Patient samples

The follow-up of 91 consecutive patients with stage II or stage III rectal adenocarcinoma according to American Joint Committee on Cancer [38] who underwent standardized neoadjuvant chemoradiotherapy followed by total mesorectal excision, from December 2006 to January 2014, were reviewed for the study. However, only those patients with available endoscopic biopsies for immunohistochemical analysis were selected for this study. A total of 74 patients with locally advanced rectal adenocarcinoma, from General and Digestive-Tract Surgery Department of University Hospital Fundación Jiménez Díaz were assessed for eligibility.

Sixty-three percent of the rectal tumours included in the study were determined to be of a high grade based on the recommendations of the College of American Pathologists [39]. Magnetic resonance imaging (MRI), computed tomography, endorectal ultrasound, and/or endoscopy revealed a high prevalence of stage III tumours (93%). The criteria published by Ryan et al. were applied to classify patients according to response to neoadjuvant treatment [15]. According to this classification system, complete pathological response was indicated by an absence of tumour cells; partial pathologic response by fibrosis with presence of isolated tumour cells; and minimum pathologic response by tumour nests outgrown by fibrosis or no tumour kill. Tand N-downstaging were also assessed. Radiotherapy administered as neoadjuvant treatment was dosed over 28 sessions (45 Gy to the pelvic area and 50.4 Gy to the tumour area).

Tissue microarray

Samples from 74 patients were used to construct a paraffin block containing 148 cores (2 cores per patient) to allow for immunohistochemistry analysis. A hollow needle (MTA-1 tissue arrayer, Beecher Instruments, Sun Prairie, USA) was used to perform a punch biopsy from pre-selected tumour areas in paraffin-embedded (FFPE) tissues. These tissue cores were then inserted in a recipient paraffin block. Sections from this FFPE block were cut using a microtome and mounted on a microscope slide to be analyzed by immunohistochemistry.

Immunohistochemistry and quantification

Staining was conducted in 2-µm sections. Slides were deparaffinized by incubation at 60 °C for 10 min and then incubated with PT-Link (Dako, Denmark) for 20 min at 95 °C in a high pH-buffered solution. To block endogenous peroxidase, holders were incubated with peroxidase blocking reagent (Dako, Denmark). Biopsies were stained for 20 min with a 1:50 dilution of DEK antibody (610,948, BD Biosciences) and with 1:150 of phospho-P38 (ab38238, Abcam) followed by incubation with anti-Ig horseradish peroxidase-conjugated polymer (EnVision, Dako, Denmark) to detect antigen-antibody reaction. A single human normal rectum tissue was used as a positive control for immunohistochemical staining. Sections were then visualized with 3,3'-diaminobenzidine as the chromogen for 5 min and counterstained with hematoxylin. Photographs were taken with a stereo microscope (Leica DMi1, Wetzlar, Germany). Immunoreactivity was quantified by two independent pathologists as the percentage of positive stained cells over the total number of tumour cells. Positiveness was defined as medium to high DEK expression levels according to The Human Protein Atlas (http://www.proteinatlas.org) and quantification of each biopsy was calculated using the average of both cores.

Statistical analysis

The association between DEK expression (categorized as low or high percentage of positive stained cells) and clinicopathologic variables, including pathologic response, was evaluated by *Fisher's exact* or Chi-square (χ^2) test. χ^2 test was used to analyze the relationship between DEK expression and clinicopathologic parameters. *Fisher's exact* test was used when one or more variable had a frequency of five or less. Association between phospho-P38

Characteristics	Patients ($N = 74$)
Median age-years (range)	72 (46–89)
> 60 years	60 (81%)
< 60 years	14 (19%)
Sex	
Male	45 (61%)
Female	29 (39%)
ECOG	
0	41 (55%)
1	31 (42%)
2	2 (3%)
Status	
Death	7 (10%)
Alive without disease	59 (78%)
Alive with disease	7 (10%)
N/A	1 (1%)
T Downstaging	
0	28 (38%)
1	39 (53%)
N/A	7 (9%)
N Downstaging	
0	20 (27%)
1	47 (64%)
N/A	7 (9%)
Grade	
Low	19 (26%)
High	47 (63%)
N/A	8 (11%)
Stage	
II	4 (6%)
III	69 (93%)
N/A	1 (1%)
Neoadjuvant Treatment	
RT + Fluoropyrimidines based	73 (99%)
Other	1 (1%)
Treatment toxicity	
Yes	30 (41%)
No	44 (59%)
Pathological Response	
Complete	9 (12%)
Partial	27 (37%)
Minimun	38 (51%)
DEK	
Low	26 (35%)
High	48 (65%)

N/A not available, RT Radiotherapy

expression (categorized as low or high percentage of positive stained cells) with pathologic response was assessed by *Fisher's exact* test. Association between DEK and phospho-P38 expression was analysed by χ^2 test. *P* values ≤ 0.05 were considered significant. Analysis was performed with the IBM SPSS program, version 20.0.

Results

Patient characteristics

The clinical features of the resected rectal-cancer patients are summarized in Table 1. The median age of

the patients was 72 years (range 46–89 years), and male population has higher incidence (n = 45; 61%) with good performance status (ECOG 0) (n = 41; 55%).

Neoadjuvant treatment was based on fluoropyrimidines (5FU or FOLFOX) and combined with radiotherapy was administered in 73 patients (99%). The majority of patients did not present treatment toxicity (n = 44; 59%). Concerning pathological response, complete response was achieved in 9 patients (12%) and partial and minimum response in 27 patients (37%), and 38 patients (51%) respectively.



High DEK expression associated with complete response to neoadjuvant chemoradiotherapy

Based on our previous reports [36], we hypothesized that DEK could be related to neoadjuvant response and serve as a predictive biomarker in patients with rectal adenocarcinoma prior to surgery. For this purpose, a tissue microarray was constructed and stained to quantify the percentage of DEK positive cells over the total number of tumour cells. All samples were obtained before the patients received neoadjuvant treatment. After immunohistochemical staining, the biopsies were observed to have nuclear localization and DEK stained only tumour cells (Fig. 1a to d). Distribution of samples according to the percentage of positive tumour cells staining showed a uniform cumulative distribution (Fig. 1e). The biopsies were then stratified into low or high DEK expression using the mean percentage of positive stained tumor cells as a cut-off point. The results showed that 9 (19%) patients out of the 45 patients with high DEK expression achieved a complete response to neoadjuvant treatment; while none of those with low DEK expression obtained a complete response. In fact, all patients who showed complete response (n = 9) had high DEK expression. Moreover, 82% of patients (n = 39) with high expression achieved partial or minimal response, while all patients (n= 26; 100%) with low DEK expression achieved partial or none response (Table 2). Statistical analysis showed significant differences between both groups of response to neoadjuvant chemoradiotherapy (complete vs. partial or minimal) and the low or high DEK expression (Chi*squared:* P = 0,018; *Fisher's exact:* P = 0,023) (Table 2).

Further analysis revealed no statistical association between DEK expression and the rest of the clinicopathologic variables studied, including gender (P = 0.553), age (P = 0.758), T-downstaging (P = 0.840), N-downstaging (P = 0.626), grade (P = 0.312), ECOG (P = 0.843), status (P = 0.544), tumour size (P = 0.703), and stage (P =0.613). Concerning treatment toxicity, a considerable trend was observed between high DEK expression and the absence of treatment toxicity (P = 0.086) (Table 3).

Table 2 Statistical association between neoadjuvant treatmentresponse and low- or high-percentage of DEK positive tumor cells

	Treatment Response	3		
DEK	No. Complete (% of DEK subpopulation)	No. Partial or minimum (% of DEK subpopulation)	P (chi-square)	P (Fisher)
High	9	39		
n = 48	(19%)	(82%)		
			0,018	0,023
Low	0	26		
n = 26	(0%)	(100%)		

No Number of patients

DEK expression associated with phospho-P38 expression in pre-treated rectal cancer biopsies

P38 is an important component of the mitogen-activated protein kinases (MAPK) [40] and plays a central role in cell proliferation and apoptosis in multiple neoplasias [41]. Furthermore, P38 has been recently associated to chemotherapy response in colorectal cancer [42]. Therefore, we quantified the immunoreactivity of the active form of P38 (phospho-P38) in all rectal cancers biopsies by immunohistochemistry. Phospho-P38 expression was then categorized as low or high according to median percentage of positive stained tumor cells as cut-off point. Although we did not find statistically significant

Table 3 Statistical association between low- or high-percentage of

 DEK positive stained tumor cells and clinico-pathological parameters

	DEK		
Parameter	Low	High	Р
Gender			0.553
Male	17	28	
Female	9	20	
Age			0.758
< 60 years	4	10	
> 60 years	22	38	
T_Downstaging			0.840
No	10	18	
Yes	13	26	
N_Downstaging			0.626
No	6	14	
Yes	17	30	
Grade			0.312
Low	9	10	
High	16	31	
Treatment toxicity			0.086
Yes	14	16	
No	12	32	
ECOG			0.843
0	14	27	
1–2	12	21	
Status			0.544
Alive with disease or death	6	8	
Alive without disease	20	40	
Tumor size			0.703
< 3 cm	2	6	
> 3 cm	24	41	
Stage			0.613
II	2	2	
	24	45	

association between phospho-P38 expression and pathological response to neoadjuvant treatment (P = 0.296; data not shown), a direct association was found between phospho-P38 and DEK expression (P = 0.027; Table 4). In fact, seven patients of whom showed not only complete response but also high DEK expression (n = 9) revealed high expression of phospho-P38, while two patients presented low phospho-P38 expression.

These results suggest that high DEK expression in tumour biopsies could be used as a potential biomarker of pathological response that follows neoadjuvant therapy in rectal cancer. Moreover, the association between DEK and phospho-P38 expression supports and provides a highly robust predictive model of cell-death revealed by the complete response to neoadjuvant treatment.

Discussion

Neoadjuvant chemoradiotherapy is the standard care approach for stage II and III rectal-cancer patients. The aim of this treatment is to achieve pathologic downstaging and complete response. Therefore, extensive investigation is currently being devoted to biomarkers that predict response to neoadjuvant treatment. Genetic profiling platforms have become a useful tool for analyzing DNA, RNA, and other factors that may or may not be translated into protein, such as miRNA. In the era of genomics, transcriptomics, and proteomics, these methodologies have helped elucidate potential biomarkers of treatment response in rectal cancer [17, 43–47]. DNA microarrays have been used to differentiate rectal-cancer patients into responders and non-responders. A study using DNA microarrays to assess 17 rectal-cancer samples discovered 17 genes differentially expressed between responders and non-responders [44]. Some of these genes included MMP, NFKB2, TGFB1, TOP1, and ITGB1 [44]. The most highly overexpressed gene, MMP7, was validated by immunohistochemistry, and it was found that none of the non-responders (n = 7) overexpressed the gene. However, only four of the responders (n = 10)overexpressed MMP7 [44]. Palma et al. analyzed the gene-expression profiles of 26 pre-treatment biopsies by expression microarray and demonstrated that high levels

Table 4 Statistical association between phospho-P38 and DEKpositiveness in rectal cancer patients treated with neoadjuvantchemoradiotherapy

DEK \ phospho-P38	Low	High	Total	P (chi-square)
Low	15	15	30	
(%)	(20%)	(20%)	(40%)	
High	11	33	44	
(%)	(15%)	(45%)	(60%)	
Total	26	48	74	0,027
(%)	(35%)	(65%)	(100%)	

of Gng4, c-Myc, Pola1, and Rrm1 expression were significant factors when predicting neoadjuvant response in rectal cancer [45]. Others studies with 23 patient samples [17] and with 43 patient samples [43] revealed 54 and 43 differentially expressed genes, respectively, though no concordance was found between both studies. Some studies based on miRNA microarrays revealed higher miR-223 levels in responders compared to nonresponders, one in a cohort of 43 rectal-cancer patients [46], and a more recent in a cohort of 59 patients [47].

Post-translational modifications may affect the concordance between gene-expression profile and proteinexpression pattern, which could lead to controversial results. Proteins are the main agents in biologic pathways, and thus the results of protein-expression analysis may be the key to treatment decision-making. Regarding the prediction of response to chemoradiotherapy in rectal cancer by immunohistochemistry, Kuremsky et al. reported that the most commonly biomarkers evaluated were p53, EGFR, TYMS, Ki-67, p21, BCL-2, and BAX [48].

High DEK expression has been described previously by our group as a crucial event for aggressive tumour phenotype and as a biomarker for poor response to irinotecan in metastatic colorectal cancer [36]. In the present study, high DEK expression was related to pathological response in 74 locally advanced rectal adenocarcinomas. This enabled us to establish a new model based on DEK expression that was statistically associated with complete pathological response. Here, it is supported that rectal cancer patients with high DEK expression have a 19% probability to achieve complete response. Otherwise, low DEK expression predicts lack of complete response to neoadjuvant treatment. Moreover, the fact that DEK expression associated with the proapoptotic factor P38 supports the role of DEK as a predictive biomarker for pathological complete response to chemoradiotherapy prior to surgery in rectal cancer patients.

The findings showed in the present study seem to disagree with those obtained in our previous work with colorectal cancer [36]. However, our previous research was performed with stage IV colorectal cancer samples, while the present work only focused on stage II–III rectal tumours that only represent a part of colorectal tumors. Moreover, the potential effect of DEK in our previous study to predict irinotecan response was not observed with 5FU or oxaliplatin, drugs used in the present study to evaluate pathological response. Indeed, DEK has also been related to neoadjuvant treatment response in breast cancer, independently of estrogenreceptor status [49]. Consequently, our study agree with Witkiewicz et al., who reported a strong association between high DEK expression and a low residual cancer

Table 5	5 Dataset of pa	itient biopsies recruited	in the study									
Biopsy	Age ECOG_PS	5 Status	T-Downstaging	N-Downstaging	Grade	Stage	Neoadjuvant treatment	Treatment toxicity	Pathological Response	Tumor size	DEK (% positive tumor cells)	Phospho-P38 (% positive tumor cells)
	> 70 1	alive without disease	0	1	High	≡	RDT + FOLFOX	No	Partial	> 3 cm		35
2	> 60 1	alive without disease	0	1	High	≡	RDT + 5FU	No	Minimum	> 3 cm	S	35
e	> 70 0	Death	-	0	High	≡	RDT + FOLFOX	Yes	Minimum	< 3 cm	7	65
4	> 70 0	alive without disease	0	1	Low	≡	RDT + 5FU	No	Minimum	> 3 cm	10	80
5	> 60 0	alive with desease	0	1	Low	≡	RDT + FOLFOX	Yes	Minimum	> 3 cm	15	45
9	> 70 0	alive without disease	-	1	High	≡	RDT + FOLFOX	Yes	Partial	> 3 cm	15	80
7	> 40 0	alive with desease	0	0	High	≡	RDT + 5FU	Yes	Partial	> 3 cm	20	25
00	> 40 1	alive with desease	-	0	High	=	RDT + 5FU	Yes	Minimum	> 3 cm	20	25
6	> 70 0	alive without disease	-	1	Low	≡	RDT + 5FU	No	Partial	> 3 cm	30	45
10	> 70 0	alive without disease	-	1	High	≡	RDT + FOLFOX	Yes	Minimum	> 3 cm	35	55
11	> 70 0	alive without disease	N/A	N/A	High	≡	RDT + FOLFOX	No	Minimum	> 3 cm	35	10
12	> 50 0	alive without disease	0	1	High	≡	RDT + 5FU	Yes	Minimum	> 3 cm	35	70
13	> 70 0	alive without disease	1	1	High	≡	RDT + 5FU	No	Partial	> 3 cm	35	25
14	> 50 0	alive without disease	-	1	Low	≡	RDT + 5FU	No	Minimum	> 3 cm	35	25
15	> 70 1	alive without disease	-	1	High	≡	RDT + 5FU	Yes	Minimum	< 3 cm	35	06
16	> 60 1	alive without disease	1	1	Low	≡	RDT + 5FU	No	Partial	> 3 cm	35	06
17	> 60 0	N/A	0	0	High	≡	RDT + FOLFOX	Yes	Minimum	> 3 cm	40	5
18	> 80 0	Death	-	0	High	≡	RDT + 5FU	Yes	Partial	> 3 cm	40	06
19	> 70 1	alive without disease	0	1	Low	≡	RDT + 5FU	Yes	Minimum	> 3 cm	40	85
20	> 60 0	alive without disease	0	1	High	≡	RDT + 5FU	No	Minimum	> 3 cm	40	65
21	> 70 2	alive without disease	-	0	N/A	=	RDT + 5FU	Yes	Minimum	> 3 cm	40	100
22	> 70 1	Death	0	1	High	≡	RDT + 5FU	Yes	Partial	> 3 cm	45	65
23	> 80 1	alive without disease	N/A	N/A	High	≡	RDT + 5FU	No	Partial	> 3 cm	45	40
24	> 70 1	alive without disease	N/A	N/A	Low	≡	RDT + 5FU	Yes	Partial	> 3 cm	45	80
25	> 80 1	alive without disease	-	1	Low	≡	RDT + 5FU	No	Partial	> 3 cm	50	80
26	> 70 1	alive without disease	-	1	Low	≡	RDT + 5FU	No	Partial	> 3 cm	55	80
27	> 50 0	alive without disease	-	1	High	≡	RDT + FOLFOX	Yes	Minimum	> 3 cm	60	80
28	> 80 1	alive without disease	N/A	N/A	High	N/A	RDT + 5FU	No	Complete	> 3 cm	60	60
29	> 80 1	alive without disease	-	1	High	≡	RDT + FOLFOX	Yes	Minimum	> 3 cm	60	75
30	> 60 0	alive without disease	0	0	High	≡	RDT + 5FU	No	Minimum	> 3 cm	60	40
31	> 50 1	alive with desease	-	1	High	≡	RDT + 5FU	No	Complete	< 3 cm	60	100
32	> 60 0	alive without disease	N/A	N/A	High	≡	RDT + 5FU	No	Complete	> 3 cm	60	45

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Biopsy	Age	ECOG_P5	S Status T-D	ownstaging N	J-Downstaging	Grade	Stage	Neoadjuvant treatment	Treatment toxicity	Pathological Response	Tumor size	DEK (% positive tumor cells)	Phospho-P38 (% positive tumor cells)
33	> 40	0	alive without disease 0	0		High	=	RDT + FOLFOX	Yes	Minimum	> 3 cm	65	100
34	> 80	0	alive without disease 1	-		High	≡	RDT + 5FU	Yes	Partial	> 3 cm	65	75
35	> 80	0	alive without disease 1	0		High	≡	RDT + 5FU	No	Partial	> 3 cm	65	75
36	> 80	-	alive without disease 1			N/A	≡	RDT + 5FU	Yes	Partial	< 3 cm	65	06
37	> 70	0	alive without disease 1			N/A	≡	RDT + 5FU	No	Complete	> 3 cm	65	06
38	> 70		alive without disease 1	<i>~</i>		Low	≡	RDT + 5FU	No	Minimum	> 3 cm	65	15
39	> 60	0	alive without disease 1	<i>—</i>		High	≡	RDT + 5FU	No	Partial	> 3 cm	65	80
40	> 60		alive without disease 1	0		High	≡	RDT + 5FU	No	Partial	> 3 cm	65	50
41	> 40	0	alive without disease 1	-		Low	≡	RDT + 5FU	Yes	Partial	> 3 cm	70	60
42	> 80	-	alive without disease 1	,		High	≡	RDT + 5FU	No	Complete	> 3 cm	75	80
43	> 70	0	alive with desease	0		Low	≡	RDT + FOLFOX	No	Minimum	< 3 cm	75	06
44	> 70	-	alive without disease N/A	Z	1/A	High	≡	RDT + 5FU	No	Minimum	> 3 cm	75	75
45	> 40	0	alive without disease 1	,		N/A	≡	RDT + 5FU	No	Complete	> 3 cm	75	95
46	> 60	0	alive without disease 1			Low	≡	RDT + 5FU	Yes	Complete	< 3 cm	75	06
47	> 80	-	alive without disease 0	<i>~</i>		High	≡	RDT + 5FU	Yes	Minimum	> 3 cm	75	95
48	> 60		alive with desease 0	<i>—</i>		High	≡	RDT + 5FU	Yes	Complete	> 3 cm	75	70
49	> 70	0	Death 1	<i>~</i>		Low	≡	others	Yes	Minimum	< 3 cm	80	ſ
50	> 80	-	Death 0	0		High	≡	RDT + FOLFOX	Yes	Minimum	> 3 cm	80	35
51	> 70	0	alive without disease 0	0		High	≡	RDT + 5FU	No	Partial	> 3 cm	80	100
52	> 60	0	alive without disease 0			High	≡	RDT + 5FU	Yes	Partial	< 3 cm	80	95
53	> 70		alive without disease 1	0		N/A	≡	RDT + 5FU	No	Partial	> 3 cm	80	30
54	> 60	0	alive without disease 0	0		N/A	=	RDT + 5FU	No	Partial	> 3 cm	80	10
55	> 70		alive without disease 1	<i>—</i>		N/A	≡	RDT + FOLFOX	No	Partial	> 3 cm	85	06
56	> 80	-	alive without disease 0	<i>—</i>		High	≡	RDT + 5FU	Yes	Minimum	> 3 cm	85	45
57	> 50	0	alive without disease 1	<i>—</i>		Low	≡	RDT + 5FU	No	Partial	> 3 cm	85	75
58	> 50	-	alive without disease 0	0		High	≡	RDT + 5FU	No	Minimum	> 3 cm	85	85
59	> 60	0	alive without disease 1	-		Low	≡	RDT + 5FU	No	Partial	> 3 cm	85	30
60	> 80		alive without disease 1			High	≡	RDT + 5FU	No	Minimum	> 3 cm	85	80
61	> 60	0	alive without disease 1	<i>—</i>		Low	≡	RDT + 5FU	No	Partial	> 3 cm	85	0
62	> 50	0	alive without disease N/A	Z	1/A	Low	≡	RDT + FOLFOX	No	Minimum	> 3 cm	85	100
63	> 70		alive without disease 1			High	≡	RDT + 5FU	No	Minimum	N/A	06	95
64	> 70	0	alive without disease 0	-		High	≡	RDT + 5FU	No	Minimum	> 3 cm	06	85

Biopsy	Age	ECOG_PS	Status	T-Downstaging	N-Downstaging	Grade	Stage	Neoadjuvant treatment	Treatment toxicity	Pathological Response	Tumor size	DEK (% positive tumor cells)	Phospho-P38 (% positive tumor cells
65	> 70		Death	0	0	High	≡	RDT + 5FU	Yes	Minimum	< 3 cm	06	45
66	> 50	0	alive without disease	-	1	Low	≡	RDT + 5FU	No	Minimum	< 3 cm	06	55
67	> 60	2	Death	0	0	High	≡	RDT + 5FU	Yes	Minimum	< 3 cm	95	100
68	> 70	0	alive without disease	-	0	N/A	=	RDT + 5FU	No	Minimum	< 3 cm	95	85
69	> 60	0	alive without disease	0	1	High	≡	RDT + 5FU	No	Partial	< 3 cm	95	80
70	> 60	0	alive without disease	0	-	High	≡	RDT + 5FU	Yes	Minimum	< 3 cm	95	06
71	> 70	0	alive with desease	0	-	High	≡	RDT + 5FU	No	Minimum	< 3 cm	95	75
72	> 70	0	alive without disease	-	0	High	≡	RDT + 5FU	No	Complete	< 3 cm	95	75
73	> 80	,	alive without disease	0	-	High	≡	RDT + 5FU	No	Minimum	< 3 cm	100	75
74	> 50	.	alive without disease	0	1	High	≡	RDT + 5FU	No	Minimum	> 3 cm	100	80

burden, indicative of preferred response to neoadjuvant chemotherapy [49].

Conclusions

This retrospective study supports DEK as a potential predictive biomarker for neoadjuvant treatment response in rectal cancer. Moreover, the methodology performed here is easy and reproducible enough to be implemented in the routine clinical practise.

Although further research is needed, this preliminary study could be used to prospectively validate the predictive value of DEK expression in rectal and other types of tumours prior neoadjuvant treatment.

Abbreviations

5FU: 5-Fluorouracil; BAX: BCL2-associated X protein; BCL-2: B-cell lymphoma 2; c-MYC: c-myelocytomatosis viral oncogene; DEK: DEK proto-oncogen; ECOG: Eastern cooperative oncology group; EGFR: Epidermal growth factor receptor; FFPE: Formalin-fixed paraffin-embedded; FOLFOX: Folinic acid + 5-Fluorouracil + Oxaliplatin; GNG4: G protein subunit gamma 4; Gy: Gray; ITGB1: Integrin subunit beta 1; Ki-67: Marker of proliferation Ki-67; MAPK: Mitogen-activated protein kinases; MMP: Matrix metallopeptidases; MRI: Magnetic resonance imaging; N/A: Not available; NFKB2: Nuclear factor of kappa light polypeptide gene enhancer in B-cells 2; POLA1: Polymerase (DNA) alpha 1; RRM1: Ribonucleotide reductase M1; RT: Radiotherapy; TGFB1: Transforming growth factor beta 1; TOP1: Topoisomerase (DNA) I; TYMS: Thymidylate synthetase

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Availability of data and materials

All data supporting the findings of the present manuscript can be found in the additional supporting file (Table 5. Dataset of patient biopsies recruited in the study).

Authors' contributions

JM-U and JG-F designed research; JM-U, IM, MR-R, AB-P, AP-O, NP, and L dP-N performed research; JM-U, AC, TG delP, MSS, MJF-A and JG-F contributed to analytic tools; JM-U, W.L., and JG-F analysed data; and JM-U wrote the paper. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The clinical samples used in the study were kindly supplied by the BioBank of the Fundacion Jimenez Diaz – Universidad Autonoma de Madrid (PT13/0010/0012). All patients gave written informed consent for the use of their biological samples for research purposes. The institutional review board (IRB) of the Fundacion Jimenez Diaz Hospital evaluated the study, granting approval on December 9, 2014 under approval number 17/14. The FJD-IRB also certified that this study belongs to the RNA-Reg Consolider-Ingenio Network (CSD2009–0080) and Spanish Health Research Project Funds (P116/01468) from *Instituto de Salud Carlos III* (ISCIII)-Fondos FEDER.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interest.

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References

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. CA Cancer J Clin. 2016; 66(1):7–30.
- Ferlay J, Steliarova-Foucher E, Lortet-Tieulent J, Rosso S, Coebergh JW, Comber H, et al. Cancer incidence and mortality patterns in Europe: estimates for 40 countries in 2012. Eur J Cancer. 2013;49(6):1374–403.
- Ferlay J SI, Ervik M, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F Cancer Incidence and Mortality Worldwide: IARC. GLOBOCAN 2012 v10 2013, No. 11.
- Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer. 2014;136(5):E359–86.
- 5. McCourt M, Armitage J, Monson JR. Rectal cancer. Surgeon. 2009;7(3):162-9.
- 6. Fazeli MS, Keramati MR. Rectal cancer: a review. Med J Islam Repub Iran.
- 2015;29:171–2015. 7. Maeda K, Koide Y, Katsuno H. When is local excision appropriate for early
- rectal cancer? Surg Today 2014;44(11):2000-2014 Epub 2013 Nov 21 doi: 101007/s00595-013-0766-3.
- Minsky BD, Mies C, Recht A, Rich TA, Chaffey JT. Resectable adenocarcinoma of the rectosigmoid and rectum. I. Patterns of failure and survival. Cancer. 1988;61(7):1408–16.
- Dahlberg M, Glimelius B, Pahlman L. Improved survival and reduction in local failure rates after preoperative radiotherapy: evidence for the generalizability of the results of Swedish rectal cancer trial. Ann Surg. 1999; 229(4):493–7.
- Katz D, Ito E, Liu FF. On the path to seeking novel radiosensitizers. Int J Radiat Oncol Biol Phys. 2009;73(4):988–96.
- van Gijn W, Marijnen CA, Nagtegaal ID, Kranenbarg EM, Putter H, Wiggers T, et al. Preoperative radiotherapy combined with total mesorectal excision for resectable rectal cancer: 12-year follow-up of the multicentre, randomised controlled TME trial. Lancet Oncol. 2011;12(6):575–82.
- Yoon WH, Kim HJ, Kim CH, Joo JK, Kim YJ, Kim HR. Oncologic impact of pathologic response on clinical outcome after preoperative chemoradiotherapy in locally advanced rectal cancer. Ann Surg Treat Res. 2015;88(1):15–20.
- Schrag D. Evolving role of neoadjuvant therapy in rectal cancer. Curr Treat Options in Oncol. 2013;14(3):350–64.
- Dimitriou N, Michail O, Moris D, Griniatsos J. Low rectal cancer: sphincter preserving techniques-selection of patients, techniques and outcomes. World J Gastrointest Oncol. 2015;7(7):55–70.
- Ryan R, Gibbons D, Hyland JM, Treanor D, White A, Mulcahy HE, et al. Pathological response following long-course neoadjuvant chemoradiotherapy for locally advanced rectal cancer. Histopathology. 2005; 47(2):141–6.
- Wheeler JM, Dodds E, Warren BF, Cunningham C, George BD, Jones AC, et al. Preoperative chemoradiotherapy and total mesorectal excision surgery for locally advanced rectal cancer: correlation with rectal cancer regression grade. Dis Colon rectum. 2004;47(12):2025–31.
- Ghadimi BM, Grade M, Difilippantonio MJ, Varma S, Simon R, Montagna C, et al. Effectiveness of gene expression profiling for response prediction of rectal adenocarcinomas to preoperative chemoradiotherapy. J Clin Oncol. 2005;23(9):1826–38.

- Smith FM, Reynolds JV, Miller N, Stephens RB, Kennedy MJ. Pathological and molecular predictors of the response of rectal cancer to neoadjuvant radiochemotherapy. Eur J Surg Oncol. 2006;32(1):55–64.
- Grade M, Wolff HA, Gaedcke J, Ghadimi BM. The molecular basis of chemoradiosensitivity in rectal cancer: implications for personalized therapies. Langenbeck's Arch Surg. 2012;397(4):543–55.
- von Lindern M, Breems D, van Baal S, Adriaansen H, Grosveld G. Characterization of the translocation breakpoint sequences of two DEK-CAN fusion genes present in t(6;9) acute myeloid leukemia and a SET-CAN fusion gene found in a case of acute undifferentiated leukemia. Genes Chromosomes Cancer. 1992;5(3):227–34.
- Datta A, Adelson ME, Mogilevkin Y, Mordechai E, Sidi AA, Trama JP. Oncoprotein DEK as a tissue and urinary biomarker for bladder cancer. BMC Cancer. 2011;11:234.
- Privette Vinnedge LM, McClaine R, Wagh PK, Wikenheiser-Brokamp KA, Waltz SE, Wells SI. The human DEK oncogene stimulates beta-catenin signaling, invasion and mammosphere formation in breast cancer. Oncogene. 2011;30(24):2741–52.
- Kroes RA, Jastrow A, McLone MG, Yamamoto H, Colley P, Kersey DS, et al. The identification of novel therapeutic targets for the treatment of malignant brain tumors. Cancer Lett. 2000;156(2):191–8.
- 24. Kondoh N, Wakatsuki T, Ryo A, Hada A, Aihara T, Horiuchi S, et al. Identification and characterization of genes associated with human hepatocellular carcinogenesis. Cancer Res. 1999;59(19):4990–6.
- Khodadoust MS, Verhaegen M, Kappes F, Riveiro-Falkenbach E, Cigudosa JC, Kim DS, et al. Melanoma proliferation and chemoresistance controlled by the DEK oncogene. Cancer Res. 2009;69(16):6405–13.
- Grasemann C, Gratias S, Stephan H, Schuler A, Schramm A, Klein-Hitpass L, et al. Gains and overexpression identify DEK and E2F3 as targets of chromosome 6p gains in retinoblastoma. Oncogene. 2005;24(42):6441–9.
- 27. Paderova J, Orlic-Milacic M, Yoshimoto M, da Cunha Santos G, Gallie B, Squire JA. Novel 6p rearrangements and recurrent translocation breakpoints in retinoblastoma cell lines identified by spectral karyotyping and mBAND analyses. Cancer Genet Cytogenet. 2007;179(2):102–11.
- Carro MS, Spiga FM, Quarto M, Di Ninni V, Volorio S, Alcalay M, et al. DEK expression is controlled by E2F and deregulated in diverse tumor types. Cell Cycle. 2006;5(11):1202–7.
- Han S, Xuan Y, Liu S, Zhang M, Jin D, Jin R, et al. Clinicopathological significance of DEK overexpression in serous ovarian tumors. Pathol Int. 2009;59(7):443–7.
- Wu Q, Li Z, Lin H, Han L, Liu S, Lin Z. DEK overexpression in uterine cervical cancers. Pathol Int. 2008;58(6):378–82.
- Nagpal JK, Das BR. Identification of differentially expressed genes in tobacco chewing-mediated oral cancer by differential display-polymerase chain reaction. Eur J Clin Investig. 2007;37(8):658–64.
- Gamble MJ, Fisher RP. SET and PARP1 remove DEK from chromatin to permit access by the transcription machinery. Nat Struct Mol Biol. 2007; 14(6):548–55.
- Wise-Draper TM, Allen HV, Thobe MN, Jones EE, Habash KB, Munger K, et al. The human DEK proto-oncogene is a senescence inhibitor and an upregulated target of high-risk human papillomavirus E7. J Virol. 2005; 79(22):14309–17.
- Wise-Draper TM, Allen HV, Jones EE, Habash KB, Matsuo H, Wells SI. Apoptosis inhibition by the human DEK oncoprotein involves interference with p53 functions. Mol Cell Biol. 2006;26(20):7506–19.
- Kim D, Kim J, Kang SS, Jin EJ. Transforming growth factor-beta3-induced Smad signaling regulates actin reorganization during chondrogenesis of chick leg bud mesenchymal cells. J Cell Biochem. 2009;107(4):622–9.
- Martinez-Useros J, Rodriguez-Remirez M, Borrero-Palacios A, Moreno I, Cebrian A, Gomez del Pulgar T, et al. DEK is a potential marker for aggressive phenotype and irinotecan-based therapy response in metastatic colorectal cancer. BMC Cancer. 2014;14:965.
- Witkiewicz AK, Balaji U, Knudsen E. Systematically defining single gene determinants of response to neoadjuvant chemotherapy reveals specific biomarkers. Clin Cancer Res.
- Edge SB, Compton CC. The American joint committee on cancer: the 7th edition of the AJCC cancer staging manual and the future of TNM. Ann Surg Oncol. 2010;17(6):1471–4.
- Adsay NV, Basturk O, Bonnett M, Kilinc N, Andea AA, Feng J, et al. A proposal for a new and more practical grading scheme for pancreatic ductal adenocarcinoma. Am J Surg Pathol. 2005;29(6):724–33.

- 40. Zarubin T, Han J. Activation and signaling of the p38 MAP kinase pathway. Cell Res. 2005;15(1):11–8.
- 41. Cuenda A, Rousseau S. p38 MAP-kinases pathway regulation, function and role in human diseases. Biochim Biophys Acta. 2007;1773(8):1358–75.
- 42. Marzi L, Combes E, Vie N, Ayrolles-Torro A, Tosi D, Desigaud D, et al. FOXO3a and the MAPK p38 are activated by cetuximab to induce cell death and inhibit cell proliferation and their expression predicts cetuximab efficacy in colorectal cancer. Br J Cancer. 2016;115(10):1223–33.
- Rimkus C, Friederichs J, Boulesteix AL, Theisen J, Mages J, Becker K, et al. Microarray-based prediction of tumor response to neoadjuvant radiochemotherapy of patients with locally advanced rectal cancer. Clin Gastroenterol Hepatol. 2008;6(1):53–61.
- Nishioka M, Shimada M, Kurita N, Iwata T, Morimoto S, Yoshikawa K, et al. Gene expression profile can predict pathological response to preoperative chemoradiotherapy in rectal cancer. Cancer Genomics Proteomics. 2011;8(2):87–92.
- Palma P, Cano C, Conde-Muino R, Comino A, Bueno P, Ferron JA, et al. Expression profiling of rectal tumors defines response to neoadjuvant treatment related genes. PLoS One. 2014;9(11):2014.
- Hotchi M, Shimada M, Kurita N, Iwata T, Sato H, Morimoto S, et al. microRNA expression is able to predict response to chemoradiotherapy in rectal cancer. Mol Clin Oncol. 2013;1(1):137–42.
- Nakao T, Iwata T, Hotchi M, Yoshikawa K, Higashijima J, Nishi M, et al. Prediction of response to preoperative chemoradiotherapy and establishment of individualized therapy in advanced rectal cancer. Oncol Rep. 2015;34(4):1961–7.
- Kuremsky JG, Tepper JE, McLeod HL. Biomarkers for response to neoadjuvant chemoradiation for rectal cancer. Int J Radiat Oncol Biol Phys. 2009;74(3):673–88.
- 49. Witkiewicz AK, Balaji U, Knudsen ES. Systematically defining single-gene determinants of response to neoadjuvant chemotherapy reveals specific biomarkers. Clin Cancer Res. 2014;20(18):4837–48.

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