

Research Highlight

Cultivating Yet-to-be Cultivated Microbes: The Challenge Continues

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When looking into the current situation of microbial ecology, you would realize that the (meta) omics-driven studies associated with next generation sequencing technologies make the headlines in related journals. Currently, isolation and characterization of as-yet-uncultured, but functionally important microorganisms is, at least to a certain extent, being replaced by omics-driven approach without cultivation to decipher their functions. To date, more than 100 microbes have been identified as "Candidatus", that is a provisional status for "well-characterized but as-yet uncultured organisms" (26). Together with this trend, numerous scientists still voice the importance of the cultivation and isolation of microorganisms. Indeed, the pace of proposals on novel species, genus and even higher levels has been incredibly accelerated: *i.e.*, over the past decades, more than 6,000 prokaryotic species have been isolated and characterized on the basis of biochemical, morphological, physiological, and genetic traits (5). However, most of the described organisms are readily cultivable ones, in turn, as-yetuncultured organisms still remain uncultivable. To fill the gap between the canonical isolation methods and the stateof-the-art technologies that circumvent isolation, developing new approaches to cultivate those as-yet-uncultured organisms in hand is one of the most intriguing challenges in microbial ecology (12, 24).

Significant progress can be highlighted by the description on microorganisms within the class Anaerolineae of the phylum Chloroflexi (formerly known as Chloroflexi subphylum I). Until 2003, the subphylum I within the phylum Chloroflexi had not have any cultured representatives whereas it had been well-known as cosmopolitan based on culture-independent molecular analyses. Since the first cultured microorganism was obtained and the novel class Anaerolineae was coined, it has gained increased universality. The first cultivated organism named Anaerolinea thermophila has been followed by a number of newly isolated organisms within the new genera Bellilinea, Leptolinea, Levilinea, Longilinea, Thermanaerothrix, and Ornatilinea, all of which were within the class Anaerolineae, were difficult to isolate but were eventually isolated in pure culture (7, 23, 32). Those were isolated from anaerobic wastewater treatment process, rice paddy soil, and deep terrestrial hot aquifer, and characterized as anaerobic heterotrophic bacteria

** Editor-in-Chief, Microbes and Environments; E-mail: y.kamagata@aist.go.jp (32). Moreover, the 16S rRNA gene clones associated with the class Anaerolineae were frequently observed in anaerobic wastewater treatment processes, and they may play a role in the degradation of organic compounds such as carbohydrates and amino acids, probably to a great extent, associated with methanogens via interspecies hydrogen transfer (1, 20, 32). In the current issue of Microbes and Environments, Nunoura et al. (21) report that Anaerolineae-type organisms were predominated in an *in situ* colonization system placed on the shallow submarine hydrothermal vent. They successfully isolate a bacterial strain SW7 and propose the new genus Thermomarinilinea with type species T. lacunofontalis. Among a number of novel isolates described over the last decade, these organisms are exceptionally well-coordinated on nomenclature basis. Together with genome sequencing of these organisms, we will now know the entity of those organisms that allows us to know exactly who they would be and what they would do, once close relatives are isolated, or omics data that hints at the presence of relatives are obtained.

Methanogenic archaea (methanogens) have a key role in anaerobic ecosystems, where electron accepters other than carbon dioxide (e.g., oxygen, sulfate, and ferric iron) are limited. Previously characterized methanogens have been classified into the orders Methanosarcinales, Methanocellales, Methanomicrobiales, Methanobacteriales, Methanococcales, and Methanopyrales of the phylum Eurvarchaeota (Fig. 1). Generally, we use laborious culturing techniques with specific apparatuses (e.g., roll tube and agar shake tube) for isolation of the methanogens as well as other obligate anaerobic microbes to eliminate oxygen in the culture medium (8, 21, 30, 31). Such obstacles have led to the difficulty in isolation of obligate anaerobic microorganisms. Nakamura et al. (18) developed a simple technique for cultivation of such fastidious anaerobic organisms by using six-well plate and anaerobic gas pack system. They demonstrated the usefulness of this technique to cultivate the methanogens, syntrophic substrate-oxidizing bacteria (syntrophs), and sulfate- or thiosulfate-reducing bacteria. Subsequently, a thermophilic and hydrogenotrophic methanogen, Methanothermobacter tenebrarum strain RMAS, was successfully isolated from natural gas field by using this technique (17). More recently, a methanogenic archaeon, Methanomassiliicoccus luminyensis strain B10, was isolated from human feces (3). This strain is the first cultured methanogenic representative of the class Thermoplasmata. Thereafter, Iino et al. (9) report the methanogenic enrichment culture derived from the sludge

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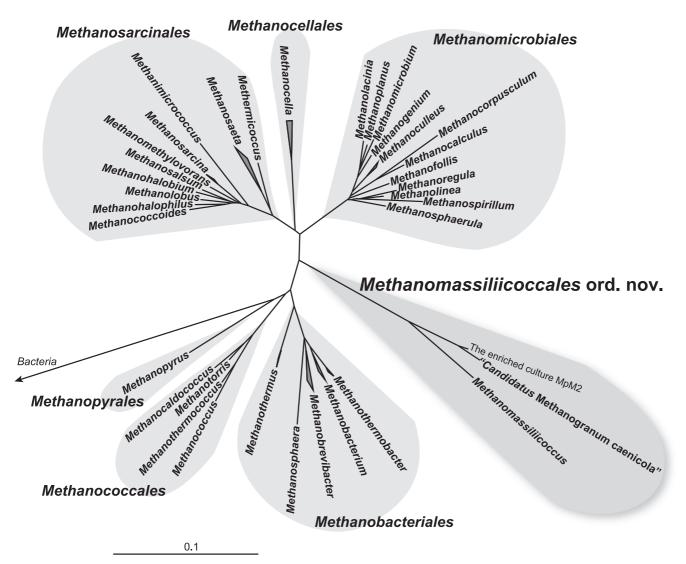


Fig. 1. Phylogeny of methanogens. The neighbor-joining tree was constructed on the basis of 16S rRNA gene sequences of previously known methanogens (19) and *Methanomassiliicoccales*-related strains (3, 9, 22) using the ARB software (15). The 16S rRNA gene sequences of the *Thermodesulfobacterium* (AF418169, AF334601, NR_025146, NR_029311) were used as outgroups (not shown). The bar indicates 10% base substitution.

of an anaerobic digestion process, that contains a novel methanogenic archaeon Kjm51a as a sole archaeal population. Phylogenetic analysis based on the 16S rRNA gene sequences indicates that archaeon Kjm51a is a relative of the *Methanomassiliicoccus luminyensis* but the identity between them is relatively low. According to the phylogenetic and physiological traits of archaeon Kjm51a, they propose "*Candidatus* Methanogranum caenicola" as the provisional taxonomic assignment. Together with *Methanomassiliicoccus luminyensis*, they also propose novel taxa, the family *Methanomassiliicoccaeae* and the order *Methanomassiliicoccus luminyensis*, for a methanogenic linage of the class *Thermoplasmata* (Fig. 1).

Besides the anaerobic microorganisms, remarkable efforts have been made to cultivate aerobic microorganisms. Fujitani *et al.* (6) develop a bioreactor-based selective culturing technique for the enrichment of *Nitrospira*-type nitriteoxidizing bacteria (NOB). The affinity to nitrite strikingly affects the growth of dominant NOB, thus nitrite concentration is maintained at a low level to facilitate the specific growth of *Nitrospira*-type NOB and to inhibit the growth of *Nitrobacter*-type NOB. Because the bioreactor-based culturing strategy has the advantage of setting up the stable culture conditions suitable for targeted uncultured microbes, this approach was applied to enrich the yet-to-be cultured organisms: for example, anaerobic ammonium oxidation (anammox) bacteria in coastal sediment (13) and phylogenetically diverse anaerobic microorganisms in subseafloor sediment (10). Tanaka *et al.* (29) successfully isolated a novel aerobic bacterial strain YO-36 from the rhizoplane of an aquatic plant in freshwater environment by using low-nutrient agar medium. Bacterium YO-36 was assigned to the candidate phylum OP10, and proposed as *Armatimonas rosea* of the novel phylum *Armatimonadetes* (28).

In addition, functionally important microorganisms have recently been isolated or enriched: ammonia-oxidizing archaeon (16), aromatic-hydrocarbon-degrading bacteria (11), cellulolytic bacteria (4), chitinolytic bacteria (25), denitrifying bacteria (27), methane-oxidizing bacteria (2), sulfate-reducing bacteria (8), uranium-tolerant bacteria (14), and uric acid-degrading bacteria (30). Clearly, cultivation and (meta) omics approaches should be complimentary. Omics information will give us a clue to the way of isolation of yet-to-be cultivated organisms, and conversely, characterization and genome information of isolates will provide convincing information that would make omics data far more robust. The challenge still continues.

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