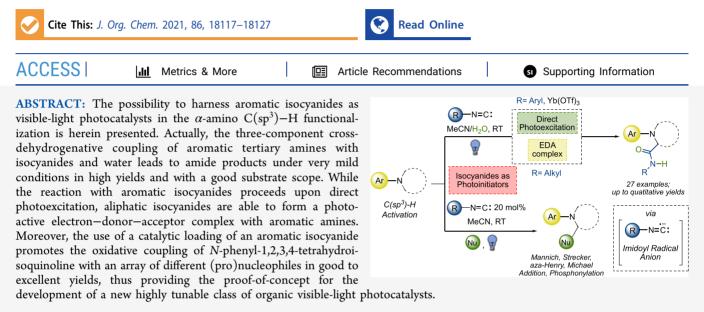
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The Dark Side of Isocyanides: Visible-Light Photocatalytic Activity in the Oxidative Functionalization of C(sp³)–H Bonds

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INTRODUCTION

Isocyanides represent a class of very complex and fascinating compounds, thanks to their chameleonic electronic properties, which have been enabling, in the last decades, the development of well-defined and stimulating research areas.¹ Accordingly, their unique reactivity features have been widely exploited in isocyanide-based multicomponent reactions² (nucleophile/carbene reactivity, Figure 1a) and in Lewis acidcatalyzed migratory insertions into nucleophiles⁵⁻⁸ (electrophile reactivity, Figure 1b). Furthermore, their ability to form complexes with π -electron-releasing transition metals has been shown as key to promote imidoylative cross couplings, with isocyanides undergoing 1,1-migratory insertions into either σ - or π -bonds.⁹⁻¹⁴ On the other hand, the presence of a lone pair on the (carbenic) isocyanide carbon atom makes them excellent geminal radical acceptors in reactions involving the formation of open-shell species (somophile reactivity, Figure 1c).^{15–18}

Recently, with the advent of the visible-light photocatalysis era,^{19–22} the somophile reactivity has been investigated in a plethora of transformations involving intramolecular cyclization of 2-isocyanobiphenyls and related analogues,^{16,17,23} as well as in two- or multicomponent reactions leading to amides, keto-amides, and other interesting molecular scaffolds.^{24–26} More in detail, these processes involve the formation of an imidoyl radical intermediate upon the addition of a radical species to isocyanide (Figure 1c). The imidoyl radical, depending on the reaction conditions and the species involved, can meet different fates such as oxidation to nitrilium ions, α -fragmentation (α -FGM) to give back

isocyanide and the radical species, β -fragmentation (β -FGM) to nitrile (Figure 2a), and intramolecular interception of a radical acceptor (e.g., phenyl ring in the *ortho*-position of 2-isocyanobiphenyl, Figure 2b).

While most of the current literature is based on these reactivity profiles, mainly triggered by either metal-based or organic visible-light photocatalysts as well as by thermal initiators (e.g., di*tert*-butyl peroxide, DTBP; *tert*-butyl hydroperoxide, TBHP; and so forth), processes involving the formation of imidoyl radical anions have been marginally reported (Figure 1d). The latter are indeed considered quite unstable,²⁷ albeit a recent involvement of their generation has been accounted upon photoinduced single-electron transfer (SET) from arylsulfinate anions following the formation of an EDA complex (EDA: electron donor–acceptor).²⁸

Following our interest in isocyanide-involving photochemical reactions, 29,30 we wondered whether the exploitation of such isocyanide radical anions could lead to developing new useful chemical transformations proceeding via EDA complex formation. $^{31-35}$ Accordingly, a screening of possible electron donors endowed with different redox potentials (i.e., carboxylic and boronic acids and dimethylani-

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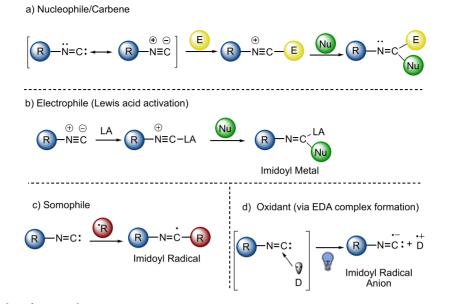


Figure 1. Reactivity modes of isocyanides.

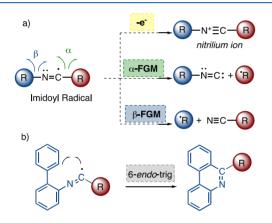


Figure 2. Possible reaction pathways of imidoyl radicals.

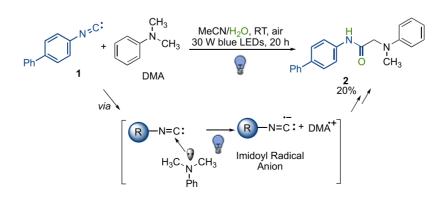
line, DMA) was performed. Interestingly, the test reaction of isocyanide 1 with DMA in MeCN at room temperature and under irradiation with 30 W blue light-emitting diodes (LEDs), open in air and in the presence of 10 equivalents of water, led to the formation of amide 2 in 20% yield (Scheme 1).³⁶

Scheme 1. Test Reaction between Isocyanide 1 and DMA

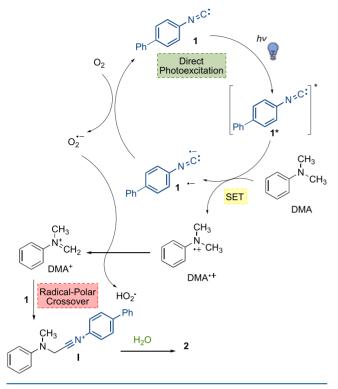
RESULTS AND DISCUSSION

Direct Photoexcitation of Aromatic Isocyanides. Intrigued by the mechanism underlying its formation and with the aim to gain information useful to optimize reaction conditions, the UV–vis absorption spectra of 1, DMA, and a mixture of both were recorded to check the presence of an EDA complex. While no bathochromic shift was observed for the mixture of 1 and DMA, we noticed that the absorption spectrum of 1 was characterized by two bands with λ_{max} at 265 and 360 nm (Figure S1, Supporting Information). This observation prompted us to propose a mechanistic hypothesis entailing a catalytic role of isocyanide 1 upon its direct photoexcitation with visible light (Scheme 2).

More in detail, 1, upon light absorption, reached an electronically excited state, thus acting as a photocatalyst: a SET from DMA to 1^* led to the formation of the imidoyl radical anion $1^{\bullet-}$ and the radical cation of dimethylaniline DMA^{$\bullet+$}. Molecular oxygen was then able to regenerate isocyanide 1, while forming a superoxide radical anion $O_2^{\bullet-}$, which abstracted a hydrogen atom from DMA^{$\bullet+$}, thus leading to the formation of the iminium ion DMA^{$\pm+$}. The latter was then intercepted by the ground-state isocyanide 1, to afford a nitrilium ion I, and eventually, after the addition of water, the amide 2. Overall, formation of amide 2 proceeded via a radical/polar crossover pathway.



Scheme 2. Mechanistic Hypothesis for the Formation of 2



Experimental data further supporting this mechanistic hypothesis were provided by Stern-Volmer quenching of 1 with increasing amounts of DMA (Figure 3) and by the formation of product $2-{}^{18}O$ in the presence of $H_2{}^{18}O$, as detected by means of high-resolution mass spectrometry (HRMS) analysis of the crude reaction mixture (Figure S2, Supporting Information). Taken together, such results provided a rational basis to identify optimum reaction conditions. Changing the ratio of 1:DMA as well as the reaction times or the light source did not improve the yield of amide 2 (Table S1, Supporting Information). Therefore, we wondered if the poor yield of 2 could be ascribed to a very short half-life of imidoyl radical anion 1^{•-}, which would lead to a back-electron transfer (BET) event to isocyanide 1 and DMA, thus preventing a quantitative oxidation of DMA to DMA⁺.

In light of these observations, we reasoned that the use of a σ -electron acceptor, such as a Lewis acid, could promote the stabilization of such imidoyl radical anion, thus preventing unproductive BETs. Actually, the reaction performed in the

presence of 30 mol % of Yb(OTf)₃ afforded amide 2 in 95% isolated yield.³⁷ While the irradiation with black light did not change the yield, no product was obtained when the light was carefully excluded, proving the photocatalytic nature of the transformation. Other Lewis acids such as silver, lanthanum, and copper triflates as well as transition metals and different ytterbium sources showed to have either no or poorer catalytic activity (Table S1, Supporting Information). A further survey of the minimum amount of Yb(OTf)₃ required to get optimum yields revealed that a 10 mol % equivalent was effective with a reaction time of 20 h. A tentative scaling up to 0.25 and 0.8 mmol afforded 41 and 23% yields, respectively, with a recovery of about 50% of the starting isocyanide. Interestingly, the use of 30 W blue LEDs was crucial to the transformation, as when the reaction was performed under irradiation with 1 W blue LEDs, only traces of 2 could be detected.

With the optimized conditions in hand, the substrate scope of both aromatic isocyanides and amines was investigated. As shown in Figure 4, both electron-poor (3, 4) and electronrich isocyanides (5-7) gave excellent yields, with the paraand meta-substitution patterns not affecting the formation of the desired products. Interestingly, even ortho-isocyanobiphenyl, usually reacting intramolecularly when involved in the formation of imidoyl radical intermediates, gave the linear amide 10 in 94% yield, thus supporting the radical/polar crossover mechanistic hypothesis. As for the amine partners, both electron-poor (11) and electron-rich (12-14) N,Ndimethylanilines were competent starting materials, as well as cyclic tertiary aromatic amines (15 and 16). Heteroaromatic dimethylanilines, such as N,N-dimethyl aminopyridine, as well as N-methyldiphenylamine, secondary anilines, tertiary aliphatic amines, and amides, were all unsuccessful substrates. These results were in accordance with related existing reactions.38

Reaction of Aliphatic Isocyanides. With the aim of further expanding the isocyanide scope to aliphatic ones and to prove the ability of aromatic isocyanides to act as photocatalysts, a test reaction involving cyclohexylisocyanide 17, DMA, a catalytic 20 mol % loading of isocyanide 1 and 10 mol % of Yb(OTf)₃ was performed (Scheme 3). After 20 h, the desired amide 18 was obtained in 16% yield. Optimization of reaction conditions (Table S2, Supporting Information) surprisingly led to the formation of product 18 in 53% yield in the absence of both 1 and Yb(OTf)₃ (Scheme 3).

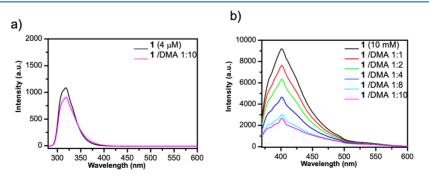


Figure 3. (a) Fluorescence spectra of 1 in the absence and presence of DMA (10 equiv) at 25 $^{\circ}$ C and an excitation wavelength of 265 nm and (b) Stern–Volmer fluorescence quenching of 1* with increasing amounts of DMA at 25 $^{\circ}$ C and an excitation wavelength of 360 nm.

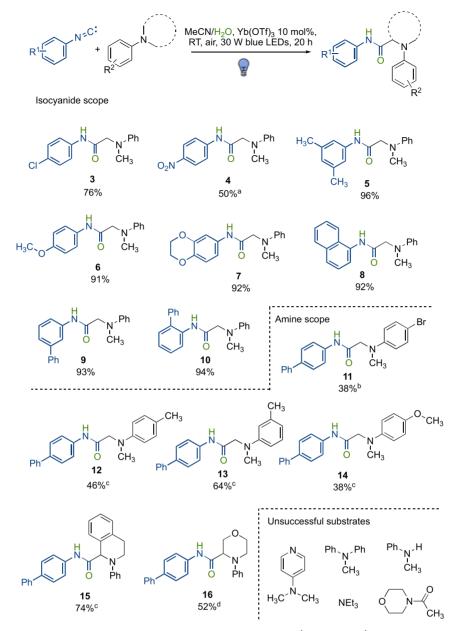
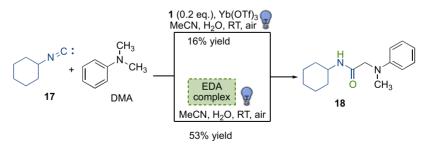


Figure 4. Substrate scope for aromatic isocyanides and amines [^awithout Yb(OTf)₃; ^b5 d; ^c48 h; ^d72 h].

Scheme 3. Reaction of Aliphatic Isocyanides



In order to shed light on the nature of this unexpected outcome, UV–Vis absorption spectra of DMA alone and in combination with an aliphatic nonvolatile isocyanide such as 1-adamantyl isocyanide **19** were recorded. As apparent from Figure 5, such experiments indicated a charge-transfer band due to the formation of an EDA complex between DMA and **19**, thus leading to propose the mechanistic hypothesis shown in Scheme 4. More in detail, visible-light excitation of the EDA complex II triggered a SET from DMA (the donor) to the isocyanide 17 (the acceptor), thus forming the radical cation $DMA^{\bullet+}$ and the isocyanide radical anion $17^{\bullet-}$.

The latter, similar to the pathway proposed for aromatic isocyanide 1 (Scheme 2), was oxidized back to 17 by molecular oxygen furnishing a superoxide radical anion $O_2^{\bullet-}$,

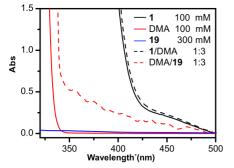
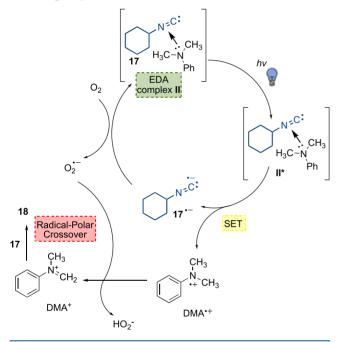


Figure 5. UV–Vis absorption spectra at 25 $^{\circ}$ C of 19, DMA alone, and in combination with 19. For comparison, spectra of isocyanide 1 alone and upon the addition of DMA were also reported.

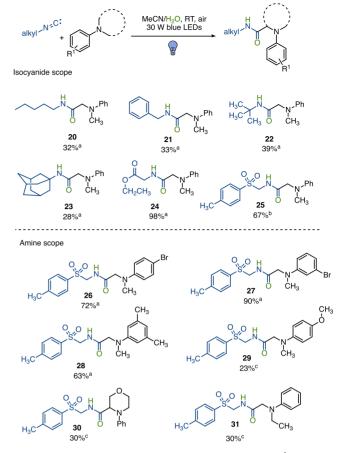
Scheme 4. Mechanistic Hypothesis for the Reaction Involving Aliphatic Isocyanides



which was responsible for hydrogen atom abstraction from DMA^{•+}, eventually affording the iminium ion DMA⁺. As stated above, overall, the formation of product **18** proceeded via a radical-polar crossover mechanism.

As for the generality of this transformation, as shown in Figure 6, it proved to be efficient with electron-rich aliphatic primary, secondary, and bulky tertiary isocyanides (20-23), while excellent yields were obtained with isocyanides bearing electron-withdrawing groups (24 and 25). When *p*-toluenesulfonylmethyl isocyanide (TosMIC) was reacted with different *N*,*N*-dimethylanilines, excellent yields were observed in the presence of electron-withdrawing substituents regardless of the *para-* or *meta-*substituents (28 and 29, Figure 6) as well as cyclic tertiary aromatic amines (30, Figure 6) led to yields from good to moderate. When nonsymmetrical *N*-ethyl-*N*-methylaniline was reacted as the starting amine, the selective formation of regioisomer 31 was obtained in 30% yield (Figure 6).

Catalytic Role of Aromatic Isocyanides: Proof-of-Concept. If the unearthing of EDA complex formation between aliphatic isocyanides and tertiary aliphatic amines



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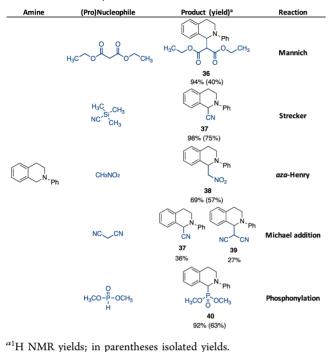
Figure 6. Substrate scope of aliphatic isocyanides ($^{a}48$ h; $^{b}20$ h; and $^{c}72$ h).

contributed to explain the mechanism underlying the generation of amide derivatives 18, 20-31 (Figure 6), on the other hand, the need for further proving the catalytic role of aromatic isocyanides, for example 1, was still disappointed.

Accordingly, in order to investigate whether this unexplored catalytic reactivity of isocyanides could be harnessed at a more general level, a range of both carbon and hetero-atom (pro)nucleophiles, such as diethyl malonate (Mannich-type reaction),³⁹ cyanotrimethylsilane (Strecker-type reaction),⁴⁰ nitromethane (*aza*-Henry reaction),⁴¹ malononitrile,³⁹ and dimethyl phosphite,⁴² were made to react with *N*-phenyl-1,2,3,4-tetrahydroisoquinoline in the presence of a 20 mol % loading of isocyanide 1 (Scheme 5).

Here, the resonance-stabilized carbanions (Michael donors) were formed upon deprotonation by the in situ formed hydroperoxide anion HO₂⁻, without requiring the addition of any further base. As shown in Scheme 5, all the (pro)nucleophiles gave the cross dehydrogenative coupling adducts in good to excellent yields; note that the reaction between Nphenyl-1,2,3,4-tetrahydroisoquinoline and diethyl malonate performed in the absence of 1 afforded a 4% yield of product 32. Cyanotrimethylsilane and nitromethane led to the Strecker and aza-Henry products 33 and 34 in 98 and 69% yields, respectively. Malononitrile showed to be effective as a Michael donor under standard reaction conditions leading to a mixture of α -aminonitrile 33 and adduct 35. These results were consistent with previous reports in the literature.³⁸ Similarly, dimethyl phosphite afforded phosphonylated product 36 in 92% yield.

Scheme 5. Michael-Type Addition Promoted by Isocyanide 1 as a Photocatalyst



CONCLUSIONS

In conclusion, the possibility to trigger completely new reactivities by enlightening a dark side of isocyanide was herein disclosed. Indeed, aromatic isocyanides proved able to act as photocatalysts triggering the oxidation of α -amino $C(sp^3)$ -H bonds. When stoichiometric amounts of aromatic isocyanides were used, the self-catalyzed reaction led to the formation of amide derivatives upon the radical/polar crossover reaction in good to excellent yields and with a wide substrate scope. Aliphatic isocyanides proved to form photoactive EDA complexes with aromatic amines and when combined with water, they were able to provide amide adducts under very mild and, to our knowledge, unprecedented, metal-free oxidative conditions. Importantly, it was also shown that a catalytic loading of an aromatic isocyanide can promote the cross dehydrogenative coupling of a tertiary aromatic amine, such as N-phenyl-1,2,3,4-tetrahydroisoquinoline, with a range of (pro)nucleophiles affording Mannich, Strecker, aza-Henry, Michael-addition, and phosphonylated adducts in good to excellent yields. As a whole, the experimental data provided evidence of the potential exploitation of aromatic isocyanides as a new class of highly tunable organic photocatalysts. Worth of note, isocyanides are nowadays considered a class of widely accessible compounds, with thousands of derivatives endowed with no or limited toxicity reported in the literature and readily synthesizable from the corresponding amines.^{43,44} Actually, the addition of electron-withdrawing or electron-donor groups on their aromatic moiety, as well as the ring expansion to polycyclic scaffolds, could lead to easily accessible and fine modulation of their redox and optical properties, which could expand the substrate scope to a broader range of $C(sp^3)$ -H bonds. Such investigations are currently in progress in our laboratories.

EXPERIMENTAL SECTION

General Methods. Commercially available reagents and solvents were used without further purification. Photochemical reactions were carried out using a PhotoRedOx Box (EvoluChem) with a 30 W blue LED (EvoluChem, model: HCK1012-01-008, wavelength 450 nm, LED: CREE XPE; for the emission spectrum of the light source see: https://www.hepatochem.com/photoreactors-leds-accessories/ led-evoluchem/). A holder suitable for 4 mL scintillation vials (45 × 14.7 mm) has been fitted within the box: this allows a fixed sample placement distance from the light source. All NMR spectra were obtained using a Bruker AVANCE NEO 400 or 700 MHz instrument. Experiments for structure elucidation were performed in CDCl₃ at 25 °C with a RT-DR-BF/1H-5 mm-OZ SmartProbe. High-resolution electrospray ionization mass spectrometry (ESI-MS) spectra were recorded on a Thermo LTQ Orbitrap XL mass spectrometer. The spectra were recorded by infusion into the ESI source using MeOH as the solvent. Chemical shifts (δ) are reported in parts per million (ppm) relative to the residual solvent peak. Column chromatography was performed on silica gel (70-230 mesh ASTM) using the reported eluents. Thin-layer chromatography (TLC) was carried out on 5×20 cm plates with a layer thickness of 0.25 mm (Silica gel 60 F_{254}) to monitor the reaction by using UV as the revelation method.

General Procedure for the Preparation of Compounds 2– 16. To a 4 mL colorless screw-cap glass vial equipped with a magnetic stir bar were added isocyanide (0.08 mmol) and Yb(OTf)₃ (0.008 mmol, 0.1 equiv). Then, 800 μ L of dry MeCN (0.1 M), 14.4 μ L of microfiltered water (0.8 mmol, 10 equiv), and the aniline derivative (0.16 mmol, 2 equiv) were added into the reaction vial via a syringe. The resulting mixture was stirred open flask in a Photoredox box (EvoluChem), under 30 W blue LED irradiation, at room temperature, until the completion of the reaction, as monitored by TLC (specific reaction times are available for each compound). Then, the solvent was removed under vacuum and the crude mixture was purified by silica gel chromatography.

N-([1,1'-Biphenyl]-4-yl)-2-(methyl(phenyl)amino)acetamide (2). The crude material (reaction time: 20 h) was purified by column chromatography (*n*-hexane/ethyl acetate 97:3) to give the product as a light pink solid [23.0 mg, 91% yield; 62.6 mg, 23% when the reaction was performed on a 0.8 mmol scale, with a recovery of 51% (76 mg) of the starting isocyanide (reaction time: 5dd); 41% NMR yield when the reaction was performed on a 0.25 mmol scale, with 50% of unreacted 4-isocyanobiphenyl (reaction time: 5dd)]. ¹H NMR (700 MHz, CDCl₃): δ 8.50 (br s, -NH), 7.61 (d, J = 8.5 Hz, 2H), 7.57–7.56 (m, 4H), 7.43 (t, J = 7.6 Hz, 2H), 7.34–7.31 (m, 3H), 6.92 (t, J = 7.4 Hz, 1H), 6.86 (d, J = 7.9 Hz, 2H), 3.99 (s, 2H), 3.10 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 168.8, 149.5, 140.5, 137.5, 136.5, 129.6, 128.8, 127.6, 127.2, 126.9, 120.2, 119.5, 113.8, 60.1, 40.1; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₂₁H₂₁N₂O⁺, 317.1648; found, [M + H]⁺ 317.1650.

N-(4-Chlorophenyl)-2-(methyl(phenyl)amino)acetamide (3).³⁶ The crude material (reaction time: 20 h) was purified by column chromatography (*n*-hexane/ethyl acetate 97:3) to give the product as a pink solid (16.7 mg, 76% yield). ¹H NMR (400 MHz, CDCl₃): δ 8.43 (br s, -NH), 7.49-7.47 (m, 2H), 7.33-7.27 (m, 4H), 6.91 (t, *J* = 7.3 Hz, 1H), 6.83 (d, *J* = 8.0 Hz, 2H), 3.96 (s, 2H), 3.08 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 168.8, 149.4, 135.8, 129.6, 129.6, 129.0, 121.2, 119.6, 113.8, 60.1, 40.2; HRMS (ESI) *m*/*z*: calcd [M + H]⁺ for C₁₅H₁₆ClN₂O⁺, 275.0946; found, [M + H]⁺ 275.0943.

2-(Methyl(phenyl)amino)-N-(4-nitrophenyl)acetamide (4).⁴⁴ The crude material [reaction performed without Yb(OTf)₃; reaction time: 5 days] was purified by column chromatography (*n*-hexane/ ethyl acetate 94:6) to give the product as a yellow amorphous solid (11.5 mg, 50% yield). ¹H NMR (700 MHz, CDCl₃): δ 8.78 (br s, -NH), 8.21–8.20 (m, 2H), 7.73–7.72 (m, 2H), 7.32 (dd, *J* = 8.7, 7.4 Hz, 2H), 6.93 (t, *J* = 7.3 Hz, 1H), 6.84 (d, *J* = 8.1 Hz, 2H), 3.99 (s, 2H), 3.10 (s, 3H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 169.5, 149.3, 143.8, 142.9, 129.7, 125.1, 120.0, 119.3, 114.0, 60.3,

40.4; HRMS (ESI) m/z: calcd $[M + H]^+$ for $C_{15}H_{16}N_3O_3^+$, 286.1186; found, $[M + H]^+$ 286.1185.

N-(3,5-*Dimethylphenyl*)-2-(*methyl(phenyl)amino)acetamide* (5). The crude material (reaction time: 20 h) was purified by column chromatography (*n*-hexane/ethyl acetate 97:3) to give the product as a brownish solid (20.7 mg, 96% yield). ¹H NMR (700 MHz, CDCl₃): δ 8.32 (br s, −NH), 7.32−7.29 (m, 2H), 7.16 (s, 2H), 6.90 (t, *J* = 7.3 Hz, 1H), 6.83 (d, *J* = 8.0 Hz, 2H), 6.77 (s, 1H), 3.94 (s, 2H), 3.07 (s, 3H), 2.29 (s, 6H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 168.5, 149.5, 138.8, 137.1, 129.5, 126.3, 119.4, 117.6, 113.7, 60.1, 40.0, 21.3; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₁₇H₂₁N₂O⁺, 269.1648; found, [M + H]⁺ 269.1647.

N-(4-*Methoxyphenyl*)-2-(*methyl*(*phenyl*)*amino*)*acetamide* (6).⁴⁵ The crude material (reaction time: 20 h) was purified by column chromatography (*n*-hexane/ethyl acetate 95:5) to give the product as a brown solid (19.6 mg, 91% yield). ¹H NMR (700 MHz, CDCl₃): δ 8.32 (br s, -NH), 7.43-7.41 (m, 2H), 7.32-7.29 (m, 2H), 6.89 (t, *J* = 7.4 Hz, 1H), 6.86-6.83 (m, 4H), 3.95 (s, 2H), 3.78 (s, 3H), 3.08 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 168.5, 156.7, 149.5, 130.4, 129.5, 121.8, 119.3, 114.2, 113.7, 59.9, 55.5, 40.1; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₁₆H₁₉N₂O₂⁺, 271.1441; found, [M + H]⁺ 271.1439.

N-(2,3-*Dihydrobenzo*[*b*][1,4]*dioxin*-6-*y*])-2-(*methyl*(*phenyl*)*amino*)-*acetamide* (7). The crude material (reaction time: 20 h) was purified by column chromatography (*n*-hexane/ethyl acetate 85:15) to give the product as a brownish solid (22.0 mg, 92% yield). ¹H NMR (400 MHz, CDCl₃): δ 8.25 (br s, −NH), 7.32– 7.28 (m, 2H), 7.17 (d, *J* = 2.5 Hz, 1H), 6.90–6.86 (m, 2H), 6.83– 6.77 (m, 3H), 4.24–4.21 (m, 4H), 3.93 (s, 2H), 3.06 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 168.4, 149.5, 143.5, 140.7, 130.9, 129.5, 119.3, 117.2, 113.7, 113.6, 109.8, 64.4, 64.3, 59.9, 40.1; HRMS (ESI) *m*/*z*: calcd [M + H]⁺ for C₁₇H₁₉N₂O₃⁺, 299.1390; found, [M + H]⁺ 299.1390.

2-(Methyl(phenyl)amino)-N-(naphthalen-1-yl)acetamide (8).⁴⁴ The crude material (reaction time: 20 h) was purified by column chromatography (*n*-hexane/ethyl acetate 96:4) to give the product as a brown solid (21.3 mg, 92% yield). ¹H NMR (400 MHz, CDCl₃): δ 8.96 (br s, -NH), 8.08 (d, J = 7.5 Hz, 1H), 7.84 (d, J =8.1 Hz, 1H), 7.68 (d, J = 8.2 Hz, 1H), 7.49 (t, J = 7.9 Hz, 1H), 7.45–7.42 (m, 2H), 7.38–7.34 (m, 3H), 6.97–6.92 (m, 3H), 4.12 (s, 2H), 3.21 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 168.9, 149.3, 134.1, 131.7, 129.7, 128.8, 126.6, 126.4, 126.0, 125.8, 125.6, 120.0, 120.0, 119.6, 113.9, 59.9, 40.4; HRMS (ESI) *m*/*z*: calcd [M + H]⁺ for C₁₉H₁₉N₂O⁺, 291.1492; found, [M + H]⁺ 291.1489.

N-([1,1'-Biphenyl]-3-yl)-2-(methyl(phenyl)amino)acetamide (9). The crude material (reaction time: 20 h) was purified by column chromatography (*n*-hexane/ethyl acetate 98:2) to give the product as a reddish solid (23.5 mg, 93% yield). ¹H NMR (400 MHz, CDCl₃): δ 8.50 (br s, -NH), 7.73-7.72 (m, 1H), 7.59-7.55 (m, 3H), 7.44-7.40 (m, 3H), 7.38-7.29 (m, 5H), 6.91 (t, *J* = 7.3 Hz, 1H), 6.86 (d, *J* = 8.0 Hz, 2H), 3.98 (s, 2H), 3.10 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 168.8, 149.5, 142.2, 140.6, 137.7, 129.6, 129.4, 128.7, 127.5, 127.2, 123.4, 119.5, 118.8, 118.7, 113.8, 60.2, 40.1; HRMS (ESI) *m*/*z*: calcd [M + H]⁺ for C₂₁H₂₁N₂O⁺, 317.1648; found, [M + H]⁺ 317.1648.

N-([1,1'-Biphenyl]-2-yĪ)-2-(methyl(phenyl)amino)acetamide (10). The crude material (reaction time: 20 h) was purified by column chromatography (*n*-hexane/ethyl acetate 98:2) to give the product as a reddish sticky solid (23.8 mg, 94% yield). ¹H NMR (400 MHz, CDCl₃): δ 8.65 (br s, −NH), 8.46 (d, *J* = 8.1 Hz, 1H), 7.33–7.29 (m, 1H), 7.18–7.04 (m, 7H), 7.00–6.98 (m, 2H), 6.77 (t, *J* = 7.3 Hz, 1H), 6.48 (d, *J* = 8.0 Hz, 2H), 3.74 (s, 2H), 2.58 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 168.6, 148.7, 137.6, 134.6, 131.9, 129.8, 129.2, 129.0, 128.8, 128.5, 127.6, 124.1, 119.9, 118.8, 113.2, 59.6, 39.4; HRMS (ESI) *m*/*z*: calcd [M + H]⁺ for C₂₁H₂₁N₂O⁺, 317.1648; found, [M + H]⁺ 317.1646.

N-([1,1'-Biphenyl]-4-yl)-2-((4-bromophenyl)(methyl)amino)-acetamide (11). The crude material (reaction time: 5 days) was purified by column chromatography (*n*-hexane/ethyl acetate 95:5) to give the product as an off-white solid (12.1 mg, 38% yield). ¹H

NMR (700 MHz, CDCl_3): δ 8.31 (br s, -NH), 7.59–7.55 (m, 6H), 7.43 (t, J = 7.7 Hz, 2H), 7.40–7.38 (m, 2H), 7.33 (t, J = 7.4 Hz, 1H), 6.71 (d, J = 9.0 Hz, 2H), 3.96 (s, 2H), 3.09 (s, 3H); $^{13}\text{C}\{^{1}\text{H}\}$ NMR (176 MHz, CDCl_3): δ 168.2, 148.4, 140.4, 137.7, 136.3, 132.3, 128.8, 127.7, 127.2, 126.9, 120.2, 115.3, 111.8, 59.9, 40.3; HRMS (ESI) m/z: calcd $[M + H]^+$ for $C_{21}H_{20}\text{BrN}_2\text{O}^+$, 395.0754; found, $[M + H]^+$ 395.0753.

N-([1,1'-Biphenyl]-4-yl)-2-(methyl(p-tolyl)amino)acetamide (12). The crude material (reaction time: 48 h) was purified by column chromatography (*n*-hexane/ethyl acetate 97:3) and preparative TLC (dichloromethane) to give the product as a light pink solid (12.1 mg, 46% yield). ¹H NMR (700 MHz, CDCl₃): δ 8.57 (br s, −NH), 7.61 (d, *J* = 8.6 Hz, 2H), 7.57−7.55 (m, 4H), 7.42 (t, *J* = 7.7 Hz, 2H), 7.33 (t, *J* = 7.4 Hz, 1H), 7.12 (d, *J* = 8.4 Hz, 2H), 6.77 (d, *J* = 8.5 Hz, 2H), 3.93 (s, 2H), 3.06 (s, 3H), 2.29 (s, 3H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 169.0, 147.4, 140.5, 137.4, 136.6, 130.0, 129.0, 128.8, 127.6, 127.2, 126.9, 120.2, 114.1, 60.5, 40.4, 20.3; HRMS (ESI) *m*/*z*: calcd [M + H]⁺ for C₂₂H₂₃N₂O⁺, 331.1805; found, [M + H]⁺ 331.1806. *N*-([1,1'-Biphenyl]-4-yl)-2-(methyl(m-tolyl)amino)acetamide

N-([1,1'-Biphenyl]-4-yl)-2-(methyl(m-tolyl)amino)acetamide (13). The crude material (reaction time: 48 h) was purified by column chromatography (*n*-hexane/ethyl acetate 98:2) to give the product as a light pink solid (16.9 mg, 64% yield). ¹H NMR (700 MHz, CDCl₃): δ 8.51 (br s, −NH), 7.62–7.61 (d, 2H), 7.57–7.56 (m, 4H), 7.43 (t, *J* = 7.7 Hz, 2H), 7.33 (t, *J* = 7.4 Hz, 1H), 7.22– 7.19 (m, 1H), 6.74 (d, *J* = 7.5 Hz, 1H), 6.67–6.66 (m, 2H), 3.97 (s, 2H), 3.08 (s, 3H), 2.35 (s, 3H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 168.9, 149.6, 140.5, 139.5, 137.5, 136.5, 129.4, 128.8, 127.7, 127.2, 126.9, 120.4, 120.2, 114.6, 111.0, 60.2, 40.1, 21.8; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₂₂H₂₃N₂O⁺, 331.1805; found, [M + H]⁺ 331.1806.

N-([1,1'-Biphenyl]-4-yl)-2-((4-methoxyphenyl)(methyl)-amino)acetamide (**14**). The crude material (reaction performed with 3 equiv of 4-methoxy-*N*,*N*-dimethylaniline; reaction time: 48 h) was purified by column chromatography (*n*-hexane/ethyl acetate 95:S) to give the product as a dark green solid (10.5 mg, 38% yield). ¹H NMR (700 MHz, CDCl₃): δ 8.71 (br s, -NH), 7.63-7.62 (m, 2H), 7.57-7.56 (m, 4H), 7.43 (t, *J* = 7.7 Hz, 2H), 7.33 (t, *J* = 7.4 Hz, 1H), 6.89-6.88 (m, 2H), 6.84-6.82 (m, 2H), 3.88 (s, 2H), 3.78 (s, 3H), 3.02 (s, 3H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 169.0, 153.6, 144.0, 140.5, 137.4, 136.6, 128.8, 127.6, 127.1, 126.9, 120.1, 115.9, 114.9, 61.0, 55.7, 41.0; HRMS (ESI) *m*/*z*: calcd [M + H]⁺ for C₂₂H₂₃N₂O₂⁺, 347.1754; found, [M + H]⁺ 347.1750.

N-([1,1⁻-*B*iphenyl]-4-yl)-2-phenyl-1,2,3,4-tetrahydroisoquinoline-1-carboxamide (**15**). The crude material (reaction time: 48 h) was purified by column chromatography (*n*-hexane/ethyl acetate 98:2) to give the product as an off-white solid (24.0 mg, 74% yield). ¹H NMR (400 MHz, CDCl₃): δ 8.89 (br s, -NH), 7.69 (d, J = 7.2 Hz, 1H), 7.60–7.52 (m, 6H), 7.43 (t, J = 7.6 Hz, 2H), 7.39–7.27 (m, 5H), 7.21 (d, J = 7.1 Hz, 1H), 7.06 (d, J = 8.2 Hz, 2H), 6.98 (t, J =7.3 Hz, 1H), 5.13 (s, 1H), 4.00–3.95 (m, 1H), 3.47–3.41 (m, 1H), 3.19–3.03 (m, 2H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 170.6, 149.4, 140.5, 137.3, 136.8, 134.5, 132.1, 129.6, 129.1, 128.7, 127.8, 127.7, 127.5, 127.1, 126.8 (3 C), 120.4, 120.1, 115.3, 66.4, 45.4, 28.8; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₂₈H₂₅N₂O⁺, 405.1961; found, [M + H]⁺ 405.1957.

N-([1,1'-Biphenyl]-4-yl)-4-phenylmorpholine-3-carboxamide (**16**). The crude material (reaction time: 72 h) was purified by column chromatography (*n*-hexane/ethyl acetate 95:5) to give the product as a light purple solid (15.0 mg, 52% yield). ¹H NMR (700 MHz, CDCl₃): δ 8.17 (br s, −NH), 7.53 (d, *J* = 7.3 Hz, 2H), 7.50 (d, *J* = 8.5 Hz, 2H), 7.44 (d, *J* = 8.6 Hz, 2H), 7.41 (t, *J* = 7.7 Hz, 2H), 7.36–7.30 (m, 3H), 7.07 (d, *J* = 8.1 Hz, 2H), 7.00 (t, *J* = 7.3 Hz, 1H), 4.20–4.19 (m, 1H), 4.16–4.15 (m, 1H), 4.10–4.09 (m, 1H), 3.96–3.89 (m, 2H), 3.53–3.50 (m, 1H), 3.32–3.29 (m, 1H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 168.5, 149.6, 140.4, 137.5, 136.5, 129.9, 128.8, 127.6, 127.2, 126.9, 122.2, 120.2, 117.7, 67.7, 66.5, 62.0, 49.0; HRMS (ESI) *m*/*z*: calcd [M + H]⁺ for C₂₃H₂₃N₂O₂⁺, 359.1754; found, [M + H]⁺ 319.1754. General Procedure for the Preparation of Compounds 18, 20–31. To a 4 mL colorless screw-cap glass vial equipped with a magnetic stir bar were added isocyanide (0.08 mmol), 800 μ L of dry MeCN (0.1 M), 14.4 μ L of microfiltered water (0.8 mmol, 10 equiv), and the aniline derivative (0.16 mmol, 2 equiv). The resulting mixture was stirred open flask in a Photoredox box (EvoluChem), under 30 W blue LED irradiation, at room temperature, until the completion of the reaction, as monitored by TLC (specific reaction times are available for each compound). Then, the solvent was removed under vacuum and the crude mixture was purified by silica gel chromatography.

N-Cyclohexyl-2-(methyl(phenyl)amino)acetamide (**18**).³⁶ The crude material (reaction time: 72 h) was purified by column chromatography (*n*-hexane/ethyl acetate 9:1) to give the product as an off-white solid (10.4 mg, 53% yield). ¹H NMR (400 MHz, CDCl₃): δ 7.29–7.25 (m, 2H), 6.84 (t, *J* = 7.3 Hz, 1H), 6.74 (d, *J* = 8.1 Hz, 2H), 6.44 (br d, -NH), 3.87–3.78 (m, 1H), 3.83 (s, 3H), 2.99 (s, 3H), 1.86–1.82 (m, 2H), 1.67–1.56 (m, 3H), 1.40–1.28 (m, 2H), 1.16–1.03 (m, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 169.3, 149.5, 129.3, 118.7, 113.3, 59.2, 47.8, 39.7, 33.0, 25.4, 24.7; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₁₅H₂₃N₂O⁺, 247.1805; found, [M + H]⁺ 247.1804.

2-(Methyl(phenyl)amino)-N-pentylacetamide (20).⁴⁵ The crude material (reaction time: 48 h) was purified by column chromatography (*n*-hexane/ethyl acetate 9:1) to give the product as an off-white amorphous solid (6.0 mg, 32% yield). ¹H NMR (700 MHz, CDCl₃): δ 7.29–7.26 (m, 2H), 6.84 (t, *J* = 7.3 Hz, 1H), 6.74 (d, *J* = 8.3 Hz, 2H), 6.57 (br t, -NH), 3.85 (s, 2H), 3.27 (q, *J* = 9.4 Hz, 2H), 3.01 (s, 3H), 1.46 (m, 2H), 1.30–1.22 (m, 4H), 0.85 (t, *J* = 7.3 Hz, 3H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 170.2, 149.3, 129.4, 118.6, 113.1, 59.0, 39.8, 39.2, 29.3, 29.0, 22.3, 14.0; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₁₄H₂₃N₂O⁺, 235.1805; found, [M + H]⁺ 235.1808.

N-*Benzyl-2-(methyl(phenyl)amino)acetamide* (21).³⁶ The crude material (reaction time: 48 h) was purified by column chromatography (*n*-hexane/ethyl acetate 9:1) to give the product as a brownish solid (6.7 mg, 33% yield). ¹H NMR (400 MHz, CDCl₃): δ 7.31–7.24 (m, 5H), 7.21–7.19 (m, 2H), 6.90 (br t, -NH), 6.84 (t, *J* = 7.3 Hz, 1H), 6.74 (d, *J* = 8.2 Hz, 2H), 4.49 (d, *J* = 6.0 Hz, 2H), 3.92 (s, 2H), 3.00 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 170.4, 149.3, 138.0, 129.4, 128.7, 127.5, 127.5, 118.8, 113.3, 59.0, 43.1, 39.9; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₁₆H₁₉N₂O⁺, 255.1492; found, [M + H]⁺ 255.1496.

N-((*Methyl(phenyl)amino)methyl)pivalamide* (**22**).³⁶ The crude material (reaction time: 48 h) was purified by column chromatography (*n*-hexane/ethyl acetate 96:4) to give the product as an off-white amorphous solid (6.9 mg, 39% yield). ¹H NMR (400 MHz, CDCl₃): δ 7.29–7.25 (m, 2H), 6.85 (t, *J* = 7.3 Hz, 1H), 6.74 (d, *J* = 8.0 Hz, 2H), 6.38 (br s, −NH), 3.73 (s, 2H), 2.98 (s, 3H), 1.33 (s, 9H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 169.6, 149.5, 129.3, 118.7, 113.4, 59.9, 50.9, 39.8, 28.7; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₁₃H₂₁N₂O⁺, 221.1648; found, [M + H]⁺ 221.1654.

N-((3*s*, 5*s*, 7*s*)-Adamantan-1-*y*])-2-(methyl(phenyl)amino)acetamide (**23**). The crude material (reaction time: 48 h) was purified by column chromatography (*n*-hexane/ethyl acetate 9:1) to give the product as an off-white solid (6.7 mg, 28% yield). ¹H NMR (400 MHz, CDCl₃): δ 7.29–7.25 (m, 3H), 6.84 (t, *J* = 7.3 Hz, 1H), 6.74 (d, *J* = 8.0 Hz, 2H), 6.25 (br s, −NH), 3.71 (s, 2H), 2.98 (s, 3H), 2.06 (s, 3H), 1.96 (d, *J* = 2.8 Hz, 6H), 1.67–1.65 (m, 6H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 169.2, 149.5, 129.3, 118.7, 113.4, 59.9, 51.6, 41.6, 39.7, 36.3, 29.4; HRMS (ESI) *m*/*z*: calcd [M + H]⁺ for C₁₉H₂₇N₂O⁺, 299.2118; found, [M + H]⁺ 299.2117.

Ethyl N-Methyl-N-phenylglycylglycinate (24).³⁶ The crude material (reaction time: 48 h) was purified by column chromatography (*n*-hexane/ethyl acetate 85:15) to give the product as a yellow solid (19.6 mg, 98% yield). ¹H NMR (400 MHz, CDCl₃): δ 7.30–7.26 (m, 2H), 7.05 (br t, -NH), 6.85 (t, J = 7.3 Hz, 1H), 6.78 (d, J = 8.1 Hz, 2H), 4.18 (q, J = 7.1 Hz, 2H), 4.06 (d, J = 5.7 Hz, 2H), 3.91 (s, 2H), 3.04 (s, 3H), 1.26 (t, J = 7.1 Hz, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 171.0, 169.5, 149.4, 129.4, 118.8, 113.4, 61.5,

58.8, 41.0, 39.7, 14.1; HRMS (ESI) m/z: calcd $[M + H]^+$ for $C_{13}H_{19}N_2O_3^+$, 251.1390; found, $[M + H]^+$ 251.1391.

2-(Methyl(phenyl)amino)-N-(tosylmethyl)acetamide (25).³⁶ The crude material (reaction time: 20 h) was purified by column chromatography (*n*-hexane/ethyl acetate 85:15) to give the product as a white solid (17.8 mg, 67% yield). ¹H NMR (400 MHz, CDCl₃): δ 7.72 (d, J = 8.2 Hz, 2H), 7.34–7.29 (m, 4H), 6.89 (t, J = 7.3 Hz, 1H), 6.67 (d, J = 8.2 Hz, 2H), 4.68 (d, J = 6.9 Hz, 2H), 3.75 (s, 2H), 2.98 (s, 3H), 2.46 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 170.3, 149.0, 145.5, 133.8, 129.9, 129.5, 128.8, 119.3, 113.5, 59.9, 58.5, 40.1, 21.8; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₁₇H₂₁N₂O₃S⁺, 333.1267; found, [M + H]⁺ 333.1267.

2-((4-Bromophenyl)(methyl)amino)-N-(tosylmethyl)acetamide (26).⁴⁶ The crude material (reaction time: 48 h) was purified by column chromatography (*n*-hexane/ethyl acetate 7:3) to give the product as an off-white solid (23.7 mg, 72% yield). ¹H NMR (700 MHz, CDCl₃): δ 7.69 (d, J = 8.2 Hz, 2H), 7.34–7.33 (m, 4H), 7.24 (br t, -NH), 6.52–6.50 (m, 2H), 4.68 (d, J = 6.9 Hz, 2H), 3.73 (s, 2H), 2.97 (s, 3H), 2.46 (s, 3H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 169.8, 148.0, 145.7, 133.7, 132.2, 130.0, 128.8, 115.0, 111.4, 59.8, 58.3, 40.2, 21.8; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₁₇H₂₀BrN₂O₃S⁺, 411.0373; found, [M + H]⁺ 411.0373.

2-((3-Bromophenyl)(methyl)amino)-N-(tosylmethyl)acetamide (27).³⁶ The crude material (reaction time: 48 h) was purified by column chromatography (*n*-hexane/ethyl acetate 75:25) to give the product as a beige solid (29.5 mg, 90% yield). ¹H NMR (700 MHz, CDCl₃): δ 7.71 (d, *J* = 8.2 Hz, 2H), 7.35 (d, *J* = 8.2 Hz, 2H), 7.24 (br t, -NH), 6.99–6.97 (m, 1H), 6.98 (d, *J* = 7.9 Hz, 1H), 6.81– 6.80 (m, 1H), 6.56 (dd, *J_a* = 8.4, *J_b* = 2.5 Hz, 1H), 4.67 (d, *J* = 6.9 Hz, 2H), 3.75 (s, 2H), 2.96 (s, 3H), 2.45 (s, 3H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 169.7, 150.2, 145.7, 133.7, 130.7, 130.1, 128.8, 123.6, 121.9, 116.1, 111.8, 59.9, 57.9, 39.9, 21.8; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₁₇H₂₀BrN₂O₃S⁺, 411.0373; found, [M + H]⁺ 411.0377.

2-((3,5-Dimethylphenyl)(methyl)amino)-N-(tosylmethyl)-acetamide (28).³⁶ The crude material (reaction time: 48 h) was purified by column chromatography (*n*-hexane/ethyl acetate 8:2) to give the product as a beige sticky solid (18.1 mg, 63% yield). ¹H NMR (700 MHz, CDCl₃): δ 7.72 (d, J = 8.2 Hz, 2H), 7.35–7.32 (m, 3H), 6.55 (s, 1H), 6.33 (s, 2H), 4.67 (d, J = 6.9 Hz, 2H), 3.73 (s, 2H), 2.95 (s, 3H), 2.45 (s, 3H), 2.30 (s, 6H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 170.6, 149.2, 145.5, 139.2, 133.9, 130.0, 128.8, 121.2, 111.4, 59.9, 58.6, 40.1, 21.8, 21.7. HRMS (ESI) *m*/*z*: calcd [M + H]⁺ for C₁₉H₂₅N₂O₃S⁺, 361.1580; found, [M + H]⁺ 361.1579.

2-((4-Methoxyphenyl)(methyl)amino)-N-(tosylmethyl)-acetamide (29).⁴⁵ The crude material (reaction time: 72 h) was purified by column chromatography (*n*-hexane/ethyl acetate 75:25) to give the product as a brownish sticky solid (6.8 mg, 23% yield). ¹H NMR (700 MHz, CDCl₃): δ 7.72 (d, *J* = 8.1 Hz, 2H), 7.53 (br t, -NH), 7.33 (d, *J* = 8.1 Hz, 2H), 6.85 (d, *J* = 9.0 Hz, 2H), 6.67 (d, *J* = 9.0 Hz, 2H), 4.70 (d, *J* = 6.9 Hz, 2H), 3.78 (s, 3H), 3.65 (s, 2H), 2.91 (s, 3H), 2.45 (s, 3H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 170.5, 153.6, 145.6, 143.7, 133.8, 130.0, 128.8, 115.7, 114.8, 59.9, 59.6, 55.7, 41.1, 21.8; HRMS (ESI) *m*/*z*: calcd [M + H]⁺ for C₁₈H₂₃N₂O₄S⁺, 363.1373; found, [M + H]⁺ 363.1375.

4-Phenyl-N-(tosylmethyl)morpholine-3-carboxamide (**30**). The crude material (reaction time: 72 h) was purified by column chromatography (*n*-hexane/ethyl acetate 75:25) to give the product as a white solid (9.0 mg, 30% yield). ¹H NMR (700 MHz, CDCl₃): δ 7.59 (d, *J* = 8.1 Hz, 2H), 7.33 (t, *J* = 7.8 Hz, 2H), 7.27 (d, *J* = 8.0 Hz, 2H), 7.02–6.99 (m, 2H), 6.91 (d, *J* = 8.3 Hz, 2H), 4.73–4.69 (m, 1H), 4.47–4.44 (m, 1H), 3.96–3.94 (m, 1H), 3.90–3.88 (m, 2H), 3.82–3.78 (m, 2H), 3.43–3.40 (m, 1H), 3.24–3. 21 (m, 1H), 2.43 (s, 3H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 170.1, 149.2, 145.4, 133.9, 129.9, 129.8, 128.7, 121.9, 117.1, 67.7, 66.5, 60.6, 59.9, 48.1, 21.8; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₁₉H₂₃N₂O₄S⁺, 375.1373; found, [M + H]⁺ 375.1375.

2-(Ethyl(phenyl)amino)-N-(tosylmethyl)acetamide (31).³⁶ The crude material (reaction time: 72 h) was purified by column chromatography (*n*-hexane/ethyl acetate 85:15) to give the product

as a beige amorphous solid (8.3 mg, 30% yield). ¹H NMR (700 MHz, CDCl₃): δ 7.68–7.67 (m, 2H), 7.32–7.26 (m, 4 H), 6.87–6.85 (m, 1H), 6.65 (d, *J* = 6.3 Hz, 2H), 4.66–4.65 (m, 2H), 3.73 (s, 2H), 3.43–3.39 (m, 2H), 2.45 (s, 3H), 1.18–1.16 (m, 3H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 170.4, 147.3, 145.5, 133.8, 129.9, 129.6, 128.8, 119.0, 113.7, 59.8, 55.4, 46.5, 21.8, 11.5; HRMS (ESI) *m*/*z*: calcd [M + H]⁺ for C₁₈H₂₃N₂O₃S⁺, 347.1424; found, [M + H]⁺ 347.1426.

General Procedure for the Preparation of Compounds 32, 33, and 36. To a 4 mL colorless screw-cap glass vial equipped with a magnetic stir bar were added the pro-nucleophile (0.08 mmol), *N*phenyl-1,2,3,4-tetrahydroisoquinoline (33.4 mg, 0.16 mmol, 2 equiv.), and 800 μ L of dry MeCN (0.1 M). Then, 4isocyanobiphenyl 1 (2.9 mg, 0.016 mmol, 20% mol) was added to the resulting mixture, which was stirred open flask in a Photoredox box (EvoluChem), under 30 W blue LED irradiation, at room temperature, for 20 h. After the completion of the reaction, as monitored by TLC, the solvent was removed under vacuum and the crude mixture was purified by silica gel chromatography.

Diethyl 2-(2-Phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)malonate (**32**).³⁹ The crude material was purified by column chromatography (*n*-hexane + triethylamine 0.1% v/v) to give the product as a yellowish sticky solid (11.7 mg, 40% isolated yield; NMR yield: 94%). ¹H NMR (400 MHz, CDCl₃): δ 7.30–7.08 (m, 6H), 6.98 (d, J = 8.1 Hz, 2H), 6.75 (t, J = 7.3 Hz, 1H), 5.72 (d, J = 9.1 Hz, 1H), 4.16–3.95 (m, 4H), 3.89 (d, J = 9.2 Hz, 1H), 3.71–3.62 (m, 2H), 3.11–3.03 (m, 1H), 2.91–2.85 (m, 1H), 1.17 (t, J = 7.1 Hz, 3H); 1.09 (t, J = 7.1 Hz, 3H); 1³C{¹H} NMR (176 MHz, CDCl₃): δ 168.0, 167.2, 148.9, 136.0, 134.83, 129.1, 128.9, 127.5, 127.2, 126.0, 118.5, 115.1, 61.6, 59.6, 57.9, 42.3, 26.1, 13.9, 13.9; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₂₂H₂₆NO₄⁺, 368.1856; found, [M + H]⁺ 368.1856.

2-Phenyl-1,2,3,4-tetrahydroisoquinoline-1-carbonitrile (33).⁴⁰ The crude material was purified by column chromatography (*n*-hexane + triethylamine 0.05% v/v) to give the product as a yellowish solid (14.0 mg, 75% isolated yield; NMR yield: 98%). ¹H NMR (700 MHz, CDCl₃): δ 7.39–7.36 (m, 2H), 7.34–7.27 (m, 3H), 7.25 (d, *J* = 7.5 Hz, 1H), 7.10 (d, *J* = 8.0 Hz, 2H), 7.03 (t, *J* = 7.3 Hz, 1H), 5.53 (s, 1H), 3.80–3.77 (m, 1H), 3.52–3.48 (m, 1H), 3.20–3.15 (m, 1H), 3.00–2.97 (m, 1H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 148.4, 134.6, 129.6, 129.4, 128.8, 127.1, 126.9, 121.9, 117.8, 117.6, 53.3, 44.2, 28.6. HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₁₆H₁₅N₂⁺, 235.1230; found, [M + H]⁺ 235.1232.

Dimethyl¹ (2-Phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphonate (**36**).⁴² The crude material was purified by column chromatography (*n*-hexane/ethyl acetate 85:15) to give the product as a yellow sticky solid (16.1 mg, 63% yield). ¹H NMR (700 MHz, CDCl₃): δ 7.39–7.37 (m, 2H), 7.30–7.28 (m, 2H), 7.24–7.18 (m, 2H), 7.00 (d, J = 7.7 Hz, 2H), 6.84 (t, J = 7.0 Hz, 1H), 5.23 (d, J = 20.3 Hz, 1H), 4.06–4.02 (m, 1H), 3.69 (d, J = 10.5 Hz, 3H), 3.67 (d, J = 10.5 Hz, 3H), 3.67–3.65 (m, 1H), 3.11–3.01 (m, 2H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 149.2 (d, $J_{C-P} = 6.3$ Hz), 136.4 (d, $J_{C-P} = 6.0$ Hz), 130.4, 129.3, 128.8 (d, $J_{C-P} = 2.5$ Hz), 127.9 (d, $J_{C-P} = 4.9$ Hz), 127.5 (d, $J_{C-P} = 3.7$ Hz), 126.0 (d, $J_{C-P} =$ 2.5 Hz), 118.7, 114.8, 58.8 (d, $J_{C-P} = 160.0$ Hz), 53.9 (d, $J_{C-P} = 6.9$ Hz), 52.9 (d, $J_{C-P} = 7.6$ Hz), 43.6, 26.7; HRMS (ESI) *m*/*z*: calcd [M + K]⁺ for C₁₇H₂₀KNO₃P⁺, 356.0812; found, [M + K]⁺ 356.0813.

1-(Nitromethyl)-2-phenyl-1,2,3,4-tetrahydroisoquinoline (34).⁴¹ To a 4 mL colorless screw-cap glass vial equipped with a magnetic stir bar were added N-phenyl-1,2,3,4-tetrahydroisoquinoline (16.7 mg, 0.08 mmol), nitromethane (130 μ L, 2.4 mmol, 30 equiv.), and 800 μ L of dry MeCN (0.1 M). Then, 4-isocyanobiphenyl 1 (2.9 mg, 0.016 mmol, 20% mol) was added to the resulting mixture, which was stirred open flask in a Photoredox box (EvoluChem), under 30 W blue LED irradiation, at room temperature, for 20 h. After the completion of the reaction, as monitored by TLC, the solvent and the excess of nitromethane were removed under vacuum and the crude mixture was purified by preparative TLC (*n*-hexane/ tetrahydrofuran 8:2) to give the product as a yellow amorphous

solid (12,2 mg, 57% isolated yield; NMR yield: 69%). ¹H NMR (700 MHz, CDCl₃): δ 7.29–7.25 (m, 3H), 7.22–7.19 (m, 2H), 7.14 (d, *J* = 7.5 Hz, 1H), 6.98 (d, *J* = 8.3 Hz, 2H), 6.85 (t, *J* = 7.3 Hz, 1H), 5.55 (t, *J* = 7.2 Hz, 1H), 4.88 (dd, *J_a* = 11.9, *J_b* = 7.8 Hz, 1H), 4.57 (dd, *J_a* = 11.9, *J_b* = 6.7 Hz, 1H), 3.69–3.61 (m, 2H), 3.11–3.07 (m, 1H), 2.82–2.78 (m, 1H); ¹³C{¹H} NMR (176 MHz, CDCl₃): δ 148.4, 135.3, 132.9, 129.5, 129.2, 128.1, 127.0, 126.7, 119.4, 115.1, 78.8, 58.2, 42.1, 26.5; HRMS (ESI) *m/z*: calcd [M + H]⁺ for C₁₆H₁₇N₂O₂⁺, 269.1285; found, [M + H]⁺ 269.1285.

2-(2-Phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)malononitrile (**35**).³⁹ To a 4 mL colorless screw-cap glass vial equipped with a magnetic stir bar were added N-phenyl-1,2,3,4-tetrahydroisoquinoline (16.7 mg, 0.08 mmol), malononitrile (26.6 μ L, 0.48 mmol, 6 equiv.), and 800 μ L of dry MeCN (0.1 M). Then, 4isocyanobiphenyl 1 (2.9 mg, 0.016 mmol, 20% mol) was added to the resulting mixture, which was stirred open flask in a Photoredox box (EvoluChem), under 30 W blue LED irradiation, at room temperature, for 20 h. After the completion of the reaction, as monitored by TLC, the solvent and the excess of malononitrile were removed under vacuum and the crude ¹H NMR spectrum in the presence of 1,3,5-trimethoxybenzene as the internal standard was registered (NMR yield: 27%).

ASSOCIATED CONTENT

G Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.joc.1c02378.

Absorption/fluorescence spectra, complete experimental procedures, characterization data, ¹H- and ¹³C NMR, HRMS, and absorption/fluorescence spectra (PDF)

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Notes

The authors declare no competing financial interest.

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