



Draft Genome Sequence of the Plant Pathogen *Streptomyces* sp. Strain 11-1-2

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ABSTRACT *Streptomyces* sp. strain 11-1-2 is a Gram-positive filamentous bacterium that was isolated from a common scab lesion on a potato tuber. The strain is highly pathogenic to plants but does not produce the virulence-associated *Streptomyces* phytotoxin thaxtomin A. Here, we report the draft genome sequence of *Streptomyces* sp. 11-1-2.

Members of the *Streptomyces* genus are Gram-positive filamentous spore-forming bacteria that are best known for their ability to produce a wide variety of secondary metabolites with useful biological activities (1). A small number of *Streptomyces* spp. can additionally form parasitic relationships with plants and cause economically important crop diseases, such as potato common scab (2). Most scab-causing streptomycetes produce thaxtomin A, a phytotoxic secondary metabolite that is essential for scab disease development (3). Recently, we isolated *Streptomyces* sp. strain 11-1-2 from a common scab lesion on a potato tuber harvested in Newfoundland, Canada (4). This strain was shown to be highly pathogenic to plants, and it produces at least one organic-soluble phytotoxin, but it does not produce thaxtomin A (4).

Genomic DNA was extracted from *Streptomyces* sp. 11-1-2 using the salting-out protocol (5) and was sequenced using the Pacific Biosciences RSII platform (Menlo Park, CA, USA) at the McGill University and Génome Québec Innovation Centre (Montréal, Québec, Canada). This produced a total of 198,563 raw subreads, with an average length of 11,877 bp, using two single-molecule real-time (SMRT) cells. The reads were assembled using the hierarchical genome assembly process (HGAP) (6) into two contigs totaling 11,655,206 bp, with an average coverage of 230× and an average G+C content of 70.85%. The analysis indicated that the genome consists of a single linear chromosome (11,603,877 bp) with a centrally located origin of replication, as well as a single plasmid (51,329 bp).

Gene prediction and annotation were carried out using the NCBI Prokaryotic Genome Annotation Pipeline version 4.2 (7). A total of 9,036 protein-coding genes, 64 tRNAs, and 18 rRNA genes (six 5S, six 16S, six 23S) were identified. Secondary metabolite biosynthetic gene clusters (BGCs) were predicted using antiSMASH version 4.0 (8), which identified 47 putative BGCs in the genome. This includes nine type I polyketide synthase (PKS) gene clusters, one type II PKS gene cluster, five nonribosomal peptide synthetase (NRPS) gene clusters, three hybrid PKS-NRPS gene clusters, and other BGCs for producing siderophores, lantipeptides, and terpenes. One BGC was found to be identical to the gene cluster known for producing the polyether ionophore nigericin (9). Also identified was a type I PKS BGC that is highly similar to the gene cluster that produces the phytotoxic ansamycin herbimycin in *Streptomyces hygrosopicus* (10).

Nucleotide BLAST analysis indicated that the *Streptomyces* sp. 11-1-2 chromosome sequence is most similar to that of *Streptomyces violaceusniger* Tü 4113 (96.8% nucleotide identity). It is currently unclear if the sequence differences between *Streptomyces* sp. 11-1-2 and *S. violaceusniger* exist at the strain level or the species level.

Received 4 August 2017 Accepted 8 August 2017 Published 14 September 2017

Citation Bown L, Bignell DRD. 2017. Draft genome sequence of the plant pathogen *Streptomyces* sp. strain 11-1-2. *Genome Announc* 5:e00968-17. <https://doi.org/10.1128/genomeA.00968-17>.

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The *Streptomyces* sp. 11-1-2 genome sequence will be a valuable resource for characterizing the phytotoxic secondary metabolite(s) produced by this strain, as well as other virulence factors that contribute to the plant-pathogenic phenotype of the organism.

Accession number(s). The draft genome sequence of *Streptomyces* sp. 11-1-2 has been deposited in DDBJ/ENA/GenBank under the accession numbers [CP022545](#) and [CP022546](#). The versions described in this paper are the first versions, CPS022545.1 and CP022546.1, respectively.

ACKNOWLEDGMENTS

This work was supported by Natural Sciences and Engineering Research Council of Canada Discovery Grant 386696-2010 to D.R.D.B. L.B. was supported in part through a fellowship from the School of Graduate Studies at Memorial University of Newfoundland.

REFERENCES

1. Bérđy J. 2005. Bioactive microbial metabolites. *J Antibiot* (Tokyo) 58: 1–26. <https://doi.org/10.1038/ja.2005.1>.
2. Loria R, Kers J, Joshi M. 2006. Evolution of plant pathogenicity in *Streptomyces*. *Annu Rev Phytopathol* 44:469–487. <https://doi.org/10.1146/annurev.phyto.44.032905.091147>.
3. Bignell DRD, Fyans JK, Cheng Z. 2014. Phytotoxins produced by plant pathogenic *Streptomyces* species. *J Appl Microbiol* 116:223–235. <https://doi.org/10.1111/jam.12369>.
4. Fyans JK, Bown L, Bignell DRD. 2016. Isolation and characterization of plant-pathogenic *Streptomyces* species associated with common scab-infected potato tubers in Newfoundland. *Phytopathology* 106:123–131. <https://doi.org/10.1094/PHYTO-05-15-0125-R>.
5. Kieser T, Bibb MJ, Buttner MJ, Chater KF, Hopwood DA. 2000. Practical *Streptomyces* genetics. John Innes Foundation, Norwich, United Kingdom.
6. Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Non-hybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <https://doi.org/10.1038/nmeth.2474>.
7. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
8. Blin K, Wolf T, Chevrette MG, Lu X, Schwalen CJ, Kautsar SA, Suarez Duran HG, de los Santos ELC, Kim HU, Nave M, Dickschat JS, Mitchell DA, Shelest E, Breitling R, Takano E, Lee SY, Weber T, Medema MH. 2017. antiSMASH 4.0—improvements in chemistry prediction and gene cluster boundary identification. *Nucleic Acids Res* 45:W36–W41. <https://doi.org/10.1093/nar/gkx319>.
9. Harvey BM, Mironenko T, Sun Y, Hong H, Deng Z, Leadlay PF, Weissman KJ, Haydock SF. 2007. Insights into polyether biosynthesis from analysis of the nigericin biosynthetic gene cluster in *Streptomyces* sp. DSM4137. *Chem Biol* 14:703–714. <https://doi.org/10.1016/j.chembiol.2007.05.011>.
10. Rascher A, Hu Z, Buchanan GO, Reid R, Hutchinson CR. 2005. Insights into the biosynthesis of the benzoquinone ansamycins geldanamycin and herbimycin, obtained by gene sequencing and disruption. *Appl Environ Microbiol* 71:4862–4871. <https://doi.org/10.1128/AEM.71.8.4862-4871.2005>.