

## Review Article

# Effect of Oxidative Stress on the Estrogen-NOS-NO-K<sub>Ca</sub> Channel Pathway in Uteroplacental Dysfunction: Its Implication in Pregnancy Complications

Xiang-Qun Hu , Rui Song, and Lubo Zhang 

Lawrence D. Longo, MD Center for Perinatal Biology, Department of Basic Sciences, Loma Linda University School of Medicine, Loma Linda, California 92350, USA

Correspondence should be addressed to Xiang-Qun Hu; [xhu@llu.edu](mailto:xhu@llu.edu) and Lubo Zhang; [lzhang@llu.edu](mailto:lzhang@llu.edu)

Received 8 November 2018; Revised 19 December 2018; Accepted 14 January 2019; Published 10 February 2019

Guest Editor: Ylva Hellsten

Copyright © 2019 Xiang-Qun Hu et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

During pregnancy, the adaptive changes in uterine circulation and the formation of the placenta are essential for the growth of the fetus and the well-being of the mother. The steroid hormone estrogen plays a pivotal role in this adaptive process. An insufficient blood supply to the placenta due to uteroplacental dysfunction has been associated with pregnancy complications including preeclampsia and intrauterine fetal growth restriction (IUGR). Oxidative stress is caused by an imbalance between free radical formation and antioxidant defense. Pregnancy itself presents a mild oxidative stress, which is exaggerated in pregnancy complications. Increasing evidence indicates that oxidative stress plays an important role in the maladaptation of uteroplacental circulation partly by impairing estrogen signaling pathways. This review is aimed at providing both an overview of our current understanding of regulation of the estrogen-NOS-NO-K<sub>Ca</sub> pathway by reactive oxygen species (ROS) in uteroplacental tissues and a link between oxidative stress and uteroplacental dysfunction in pregnancy complications. A better understanding of the mechanisms will facilitate the development of novel and effective therapeutic interventions.

## 1. Introduction

During pregnancy, maternal circulation undergoes significant physiological changes to meet the increased metabolic demand of the growing fetus and the well-being of the mother [1]. Throughout pregnancy, cardiac output rises by increasing heart rate and stroke volume, reaching ~50% above prepregnancy baseline in the third trimester. Systemic vascular resistance decreases by ~20% in the second trimester, leading to reduced mean arterial blood pressure. In addition, blood volume increases by 40-50%. Nevertheless, marked changes also occur at the maternal-fetal interface. The placenta formation and structural and physiological remodeling of uterine arteries lead to the establishment of the low-resistance uteroplacental circulation. In human and sheep, uterine blood flow increases from 20 to 50 ml/min in nonpregnant state to ≥1000 ml/min at near-term pregnancy. Elevated steroid hormones such as 17β-estradiol (E<sub>2</sub>) and progesterone are believed to play

an important role in the cardiovascular adaptation during pregnancy [2-4].

Aberrant uteroplacental adaptation leads to pregnancy complications such as preeclampsia and intrauterine (fetal) growth restriction (IUGR). These complications are associated with diminished uteroplacental blood flow [5, 6]. Both preeclampsia and IUGR are major causes of maternal and/or fetal morbidity and mortality. Accumulating evidence suggests that preeclampsia and IUGR also have detrimental effects on the health of both the mother beyond pregnancy and offspring. Women with a history of preeclampsia have increased risk of cardiovascular disease [7]. Moreover, offspring born from preeclamptic pregnancy also have high incidence of high blood pressure and stroke later in life [8, 9]. Similarly, IUGR is associated with increased prevalence of metabolic syndrome, diabetes, and cardiovascular disease in later life of offspring [10, 11].

Although the etiologies of preeclampsia and IUGR are not fully elucidated, placental insufficiency (or uteroplacental

vascular insufficiency), the inability to deliver an adequate supply of oxygen and nutrients to the fetus due to reduced blood flow to the placenta, is generally considered as a major contributor to the development of these disorders. Soleymanlou et al. revealed a remarkable similarity of global gene expression in hypoxia-treated placenta explants, high-altitude placentas, and preeclamptic placentas [12], implying an important causative role of hypoxia in these complications. This notion is further substantiated by observations in animal models in which gestational hypoxia imitated placental insufficiency, reduced fetal growth, and induced preeclampsia-like symptoms [13–15].

Oxidative stress is defined as an imbalance between oxidants and antioxidants in favor of the oxidants [16]. Prolonged hypoxia is shown to elicit oxidative stress [17]. Consistently, placental insufficiency also promotes oxidative stress in preeclampsia, IUGR, and high-altitude pregnancy [18, 19]. Accumulating evidence suggests a critical role of reactive oxygen species (ROS) in the pathogenesis of pregnancy complications [20, 21]. However, the mechanistic insights into ROS-induced maladaptation of uteroplacental circulation remain largely elusive. In this article, we provide a succinct review of effects of oxidative stress on  $E_2\beta$  signaling pathways in the uteroplacental circulation in pregnancy complications.

## 2. $E_2\beta$ Signaling and Uteroplacental Circulation in Physiological and Pathophysiological Conditions

**2.1. Estrogen and Estrogen Receptors (ERs) in Normal Pregnancy and Pregnancy Complications.** Both  $E_2\beta$  and its metabolites are essential for the success of pregnancy. Starting from approximately week 9 of gestation, the placenta becomes the primary site of estrogen synthesis involving enzymes such as aromatase (CYP19) and hydroxysteroid  $17\beta$ -dehydrogenases 1 (HSD17B1,  $17\beta$ -HSD1) [22]. Circulating estrogen rises progressively throughout pregnancy, and plasma  $17\beta$ -estradiol ( $E_2\beta$ ) level at term is ~100-fold higher than that in nonpregnant subjects. Similarly,  $E_2\beta$  metabolites produced by cytochrome P450s and catechol-O-methyltransferase (COMT) such as catecholestradiols also elevated during pregnancy [23]. However, estrogen biosynthesis and metabolism are apparently impaired in pregnancy complications. Maternal plasma  $E_2\beta$  levels are significantly lower in preeclamptic [24–26] and IUGR [27] pregnancies. Low circulating  $E_2\beta$  was also observed in high-altitude human and sheep pregnancy [28–30], although one study showed an increase in plasma estrogen [31]. The metabolism of  $E_2\beta$  is also impaired in preeclampsia, leading to reduced 2-methoxyestrone and 2-methoxyestradiol [25, 32]. It appears that the reduced circulating levels of  $E_2\beta$  and its metabolites in pregnancy complications are the result of dysregulation of steroidogenic enzyme expression in the placenta. Preeclamptic placenta displayed deficiency of aromatase, HSD17B1, and COMT [24, 25, 32–34]. The impaired estrogen steroidogenesis and metabolism in these disorders are evidently caused by placental insufficiency. Aromatase in cultured human

trophoblast cells and in trophoblast cell line JEG-3 was downregulated by hypoxia [24, 35], and the expression of placental aromatase was reduced in a rabbit model of placental ischemia [24]. Aberrant production of  $E_2\beta$  and its metabolites could contribute to the pathogenesis of pregnancy complications due to their key roles in regulating trophoblast invasion, angiogenesis, and uterine vascular tone, which will be discussed in later sections.

Estrogen produces its plethoric effects *via* interacting with its receptors involving both nongenomic and genomic mechanisms. To elicit genomic actions, estrogen binds to the nuclear estrogen receptor  $\alpha$  (ER $\alpha$ ) or estrogen receptor  $\beta$  (ER $\beta$ ). The receptors become dimerized and bind to the estrogen response element (ERE) located in the target gene promoter, triggering or suppressing gene expression [36]. Estrogen can also activate membrane G-protein-coupled estrogen receptor (GPER, or GPR30) and membrane-associated ER $\alpha$  and ER $\beta$ , which in turn stimulate adenylate cyclase to generate cAMP or activate kinases such as tyrosine kinase Src, phosphoinositide 3-kinase (PI3K), extracellular-signal-regulated kinase (ERK), and protein kinase B (PKB or AKT) [37]. Activation of membrane or membrane-associated estrogen receptors can lead both acute and long-term effects. The presence of ER $\alpha$ , ER $\beta$ , and GPER in uterine arteries and the placenta has been demonstrated by real-time polymerase chain reaction (PCR), Western blot, and immunohistochemistry [38–41]. The expression of all forms of estrogen receptors in uterine arteries and the placenta increases as pregnancy advances [38–40, 42]. The maintenance or upregulation of ERs in the uteroplacental tissues apparently requires continuous estrogen stimulation. Ovariectomy in sheep reduced ER $\beta$  expression in the endothelium of uterine arteries [42]. In addition, chronic treatment with  $E_2\beta$  *in vivo* and *ex vivo* significantly increased ER $\alpha$  expression in uterine arteries [40, 42]. The expression of GPER in HTR8/SVneo cells derived from first trimester extravillous trophoblast and placental extravillous explants was also upregulated by  $E_2\beta$  [43].

Information on estrogen receptor expression in pregnancy complications is scant, and conflicting observations have been reported. ER $\alpha$  expression was described as increased, decreased, or unchanged in the preeclamptic placenta [44–46]. No conclusion could be drawn currently, and more rigorous studies are needed to clarify the discrepancy. The expression of ER $\alpha$  in uteroplacental tissues was suppressed in high-altitude pregnancy [40], and hypoxia appeared to be the causative factor responsible for ER $\alpha$  downregulation [45, 47]. Defective expression of ER $\alpha$  could have profound effects on uteroplacental function including gene expression. Intriguingly, the placental expression of ER $\beta$  appears to be differently affected in preeclampsia and IUGR. Whereas ER $\beta$  expression was reduced in the IUGR placenta [44], an upregulation of ER $\beta$  was observed in preeclamptic placentas [44, 45]. These observations suggest that the etiologies of preeclampsia and IUGR may differ. It remains to be determined whether/how the distinct regulations of ER $\beta$  contribute to the pathogenesis of these two complications. The placental expression of GPER was reduced in preeclamptic pregnancy [43, 48], which may lead to dysfunction of uteroplacental vessels.

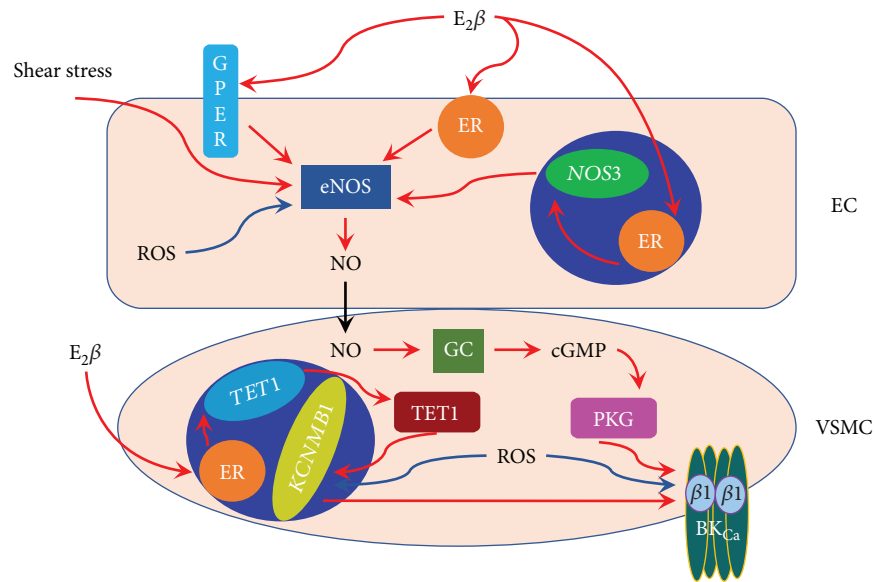


FIGURE 1: Estrogen ( $E_2\beta$ ) regulates uterine artery function partly *via* its actions on endothelial nitric oxide synthase (eNOS) in the endothelial cell (EC) and the large-conductance  $Ca^{2+}$ -activated  $K^+$  ( $BK_{Ca}$ ) channel in vascular smooth muscle cell (VSMC) during pregnancy. Shear stress stimulates eNOS activity, leading to increased NO production.  $E_2\beta$  could increase the expression of eNOS in ECs via interacting with nuclear estrogen receptors (ERs) and/or elevate eNOS activity via interacting with the G protein-coupled estrogen receptor (GPER, GPR30) or membrane-associated ER $\alpha$  and ER $\beta$ . In addition,  $E_2\beta$  increases the expression of the  $BK_{Ca}$  channel  $\beta_1$  subunit encoded by *KCNMB1* and channel activity via upregulating ten-eleven translocation methylcytosine dioxygenase 1 (TET1, encoded by *TET1*) in VSMCs. Moreover, the activity of the  $BK_{Ca}$  channel can be enhanced by NO-PKG signaling. In pregnancy complications, excessive oxygen species (ROS) impair the estrogen-NOS-NO- $BK_{Ca}$  channel pathway.

**2.2. Estrogen and the Regulation of Uteroplacental Circulation.** Several lines of evidence have implicated a critical role of estrogen in the adaptation of the uteroplacental circulation. First, the high ratio of  $E_2\beta$  to progesterone in the follicular phase was associated with increased blood to the uterus [49, 50]. Second, reduced uterine vascular resistance and increased uterine blood flow concurred with progressively rising plasma  $E_2\beta$  levels during pregnancy [51–53]. Third, acute treatment with exogenous  $E_2\beta$  markedly increased uterine blood flow and/or reduced uterine vascular resistance in nonpregnant animals [54–56]. Fourth, chronic administration of  $E_2\beta$  into nonpregnant sheep also significantly increased uterine blood flow and/or reduced uterine vascular resistance [57, 58]. *Ex vivo* treatment of uterine arteries from nonpregnant sheep with  $E_2\beta$  reduced uterine arterial myogenic tone [59]. The chronic effects of  $E_2\beta$  simulated pregnancy-induced hemodynamic changes in the uterine circulation. Fifth, the nonselective ER $\alpha$ /ER $\beta$  antagonist ICI 182,780 reduced the increase in uterine blood flow induced by exogenous  $E_2\beta$  in nonpregnant sheep and by endogenous  $E_2\beta$  in the follicular phase of nonpregnant sheep by ~60% [53]. Intriguingly, the same antagonist also lowered basal uterine blood flow in pregnant sheep by 37% [53]. Importantly,  $E_2\beta$  and its metabolites also play an important role in uteroplacental adaptation.  $E_2\beta$ , 2-hydroxyestradiol, 4-hydroxyestradiol, and 4-methoxyestradiol were implicated in angiogenesis by promoting endothelial cell proliferation [60], whereas 2-methoxyestradiol promoted the differentiation of the cytotrophoblast to an invasive phenotype [61].

**2.3. NO and  $Ca^{2+}$ -Activated  $K^+$  ( $BK_{Ca}$ ) Channels in Regulating Uteroplacental Function.** Nitric oxide (NO) is a gaseous messenger-generated nitric oxide synthase (NOS). NO contributes to the maintenance of cardiovascular homeostasis by regulating vasocontractility [62]. The large-conductance  $Ca^{2+}$ -activated  $K^+$  ( $BK_{Ca}$ ) channel is primarily expressed in vascular smooth muscle cells (VSMCs) and plays a pivotal role in regulating myogenic tone [63]. In VSMCs, the  $BK_{Ca}$  channel is a heteromeric assembly of the pore-forming  $\alpha$  subunit and accessory  $\beta_1$  subunits [64]. The  $\beta_1$  subunit encoded by *KCNMB1* increases the channel's  $Ca^{2+}$ /voltage sensitivity. Importantly, the  $BK_{Ca}$  channel is one of many targets of NO in the cardiovascular system [64]. Not surprisingly, the NO-cGMP-PKG- $BK_{Ca}$  channel axis is implicated in the adaptation of uteroplacental circulation during pregnancy [65] (Figure 1).

Activation of either of ER $\alpha$ , ER $\beta$ , or GPER induced acute vasorelaxation of uterine arteries [66]. The acute estrogen effects in regulating uterine hemodynamics involved stimulation of endothelial NOS (eNOS) activity and increased NO release in endothelial cells (ECs) [38, 67] and activation of  $BK_{Ca}$  channels in VSMCs [67]. Stimulation of eNOS activity by estrogen in uterine arterial ECs required phosphorylation of the enzyme at serine 635 and serine 1177 mediated by ER $\alpha$  [68].  $E_2\beta$  could also directly activate  $BK_{Ca}$  channels in uterine arterial VSMCs [67], possibly *via* interacting with the accessory  $\beta_1$  subunit [69].

$E_2\beta$  could also exert its genomic effect to regulate the expression of both NOS and  $BK_{Ca}$  channels in uteroplacental tissues. Expression and function of eNOS [70–72] and the

BK<sub>Ca</sub> channel  $\beta$ 1 subunit [65, 73, 74] in uterine arteries were increased in the follicular phase and during pregnancy. The upregulation of eNOS and the BK<sub>Ca</sub> channel  $\beta$ 1 subunit in uteroplacental circulation during these two physiological states was apparently stimulated by estrogen as chronic treatment with exogenous E<sub>2</sub> $\beta$  in intact nonpregnant animals [58, 75, 76] and in *ex vivo* cultured uterine arteries [73] elevated their abundance and activity.

*In vivo* studies revealed distinct contributions of eNOS and the BK<sub>Ca</sub> channel to basal uterine blood flow in nonpregnant and pregnant sheep. Intrauterine arterial infusion of the NO synthase inhibitor L-nitro-arginine methyl ester (L-NAME) demonstrated minimal contribution of NO to basal uterine blood flow in both nonpregnant and pregnant sheep [77]. However, infusion of the BK<sub>Ca</sub> channel blocker tetraethylammonium into uterine arteries revealed that at least half of the basal uterine blood flow is maintained by the BK<sub>Ca</sub> channel in pregnant sheep, whereas the channel did not contribute to basal uterine blood flow in nonpregnant animals [67, 78]. These findings are reinforced by the observations that uterine arterial myogenic tone (i.e., the major constituent of vascular tone) of pregnant subjects was regulated by the BK<sub>Ca</sub> channel [73], but not by the endothelium [79, 80]. Thus, estrogen-induced eNOS expression and activity during pregnancy are probably responsible for enhanced endothelium-dependent vasorelaxation in uterine arteries in response to given vasodilators [81, 82] and uterine artery remodeling [83], but not for regulating basal uterine vascular tone. In contrast, the upregulation of the BK<sub>Ca</sub> channel is essential for the reduced uterine vascular tone during pregnancy. In addition, the upregulated BK<sub>Ca</sub> channel also contributed to blunted vasoconstrictor responses in uterine arteries during pregnancy [65, 84]. Thus, the BK<sub>Ca</sub> channel in uteroplacental circulation functions as a negative feedback control mechanism to prevent excessive vasoconstriction. Together, these findings reinforced the notion that E<sub>2</sub> $\beta$ , through its acute and chronic actions on eNOS and BK<sub>Ca</sub> channels, plays a pivotal role in uteroplacental adaptation.

Expression/activity of placental eNOS in preeclamptic and IUGR pregnancies was reported as either unaltered [85, 86], decreased [44, 87], or increased [88, 89]. Whereas eNOS in placental chorionic plate arteries was downregulated in preeclampsia [90], this enzyme in uterine arteries was upregulated in high-altitude pregnancy [91]. Regardless of uteroplacental eNOS expression status, NO bioavailability in pregnancy complications appeared to be reduced due to substrate deficiency and enzyme inhibition. Both plasma and placental L-arginine levels were reduced in preeclampsia [86, 92]. In addition, the expression of arginase-2, which consumes eNOS's substrate L-arginine, was increased in the placenta and in omental vessels of women with preeclampsia [86, 93]. The increased arginase-2 expression could be imitated by treating human umbilical vein endothelial cells (HUVECs) with preeclamptic plasma [93]. Moreover, HUVECs from IUGR pregnancy also displayed increased arginase-2 expression and activity and placental vessels exhibited impaired eNOS-dependent relaxation [89]. A deficiency of L-arginine would not only reduce eNOS-derived NO but also increase eNOS-mediated superoxide production

leading to peroxynitrite (ONOO<sup>-</sup>) formation, evidenced by increased nitrotyrosine staining in villi and maternal vasculature of preeclamptic women [86, 94]. Similarly, nitrotyrosine staining was increased in the syncytiotrophoblast and extravillous trophoblast of high-altitude placenta [95]. Intriguingly, the circulating level of asymmetric dimethylarginine (ADMA), an endogenous NOS inhibitor, also increased in preeclamptic and IUGR pregnancies [96, 97]. Not surprisingly, NOS-dependent relaxation of placental chorionic arteries from IUGR pregnancy was impaired [98]. Moreover, both chronic blockade of NOS with L-NAME or knockout of eNOS in rodents increased maternal blood pressure and reduced fetal growth [99, 100], partly due to impaired uteroplacental vessel remodeling [101].

The expression and function of the BK<sub>Ca</sub> channel in uteroplacental vessels are also impaired in pregnancy complications. It appears that the  $\beta$ 1 subunit of the channel is selectively targeted, whereas the  $\alpha$  subunit remains unaffected in these disorders. The BK<sub>Ca</sub> channel  $\beta$ 1 subunit was downregulated in placental chorionic plate arteries in preeclampsia [102] and uterine arteries in high-altitude pregnancy [103]. High-altitude pregnancy also suppressed the ability of estrogen to upregulate the expression of the BK<sub>Ca</sub> channel  $\beta$ 1 subunit in uterine arteries [103], leading to increased uterine arterial myogenic tone. BK<sub>Ca</sub> channel-mediated vasorelaxation was also reduced in both pathological conditions. The impact of high-altitude pregnancy on the BK<sub>Ca</sub> channel was simulated by *ex vivo* hypoxia [104], implicating a causative role of hypoxia in the downregulation of the BK<sub>Ca</sub> channel  $\beta$ 1 subunit. In a preeclampsia-like murine model induced by autoantibodies against angiotensin II type 1 receptor (AT1-AA), the expression of the BK<sub>Ca</sub> channel  $\beta$ 1 subunit and channel activity in mesenteric arteries was also reduced [105].

The intermediate-conductance (IKs) and small-conductance (SKs) Ca<sup>2+</sup>-activated K<sup>+</sup> channels are predominantly expressed in ECs and also mediate endothelium-dependent vasodilation [106]. The endothelium-derived hyperpolarizing factor (EDHF) causes hyperpolarization of VSMCs by activating IKs and SKs. Both IKs and SKs are expressed in uteroplacental tissues [90, 107, 108]. IKs and SKs are also expressed in VSMCs of uterine and placental chorionic plate arteries in addition to their expression in ECs [90, 107]. In the uteroplacental system, IKs and SKs participated in the regulation of contractility of uterine and placental vessels [90, 107, 109]. Moreover, SK3 was also involved in regulating uterine vascular remodeling and placental vascularization [110, 111]. Like BK<sub>Ca</sub> channels, E<sub>2</sub> $\beta$  is required to maintain and to upregulate the expression and function of SKs in vasculature. Pregnancy *via* estrogen's action upregulated the expression of SK2 and SK3 in uterine arteries [107]. Ovariectomy reduced SK3 activity in ECs and ablated the channel's role in EDHF-mediated vasorelaxation in non-uterine arteries [112].

The expression and function of IK1, SK2, or SK3 in uteroplacental vessels and umbilical vessels were downregulated in high-altitude pregnancy and preeclampsia [90, 107, 113] as well as in a rat model of preeclampsia induced by testosterone [108]. Given the important role of estrogen in the

regulation of IKs and SKs in uteroplacental circulation, it is anticipated that impaired  $E_2\beta$ -ER signaling could contribute to the downregulation of these ion channels in high-altitude and preeclamptic pregnancies.

Together, evidence presented in this section demonstrated critical roles of both estrogen synthesis and metabolism in the adaptation of uteroplacental circulation. Preeminently,  $E_2\beta$  and its metabolites contribute to this adaptive process by promoting angiogenesis, trophoblast invasion, and remodeling and by lowering uterine vascular tone through upregulating activity and/or expression of both eNOS and  $K_{Ca}$  channels. However, the  $E_2\beta$ -NOS-NO- $K_{Ca}$  channel pathway is disrupted in pregnancy complications, which could contribute to the pathogenesis of these disorders.

### 3. Oxidative Stress and Pregnancy Complications

**3.1. Cellular Sources of ROS and Antioxidant Defense.** ROS are oxidants formed during oxygen metabolism, primarily produced during oxidative phosphorylation in the mitochondria and by oxidases such as NADPH oxidases (NOXs) and xanthine oxidase (XO) as well as uncoupled NOS [114, 115]. ROS include free radicals such as superoxide ( $O_2^{\cdot-}$ ) and hydroxyl radical ( $\cdot OH$ ) and nonradical hydrogen peroxide ( $H_2O_2$ ). In order to maintain redox hemostasis, mammalian cells have developed enzymatic and nonenzymatic defense mechanisms to balance the oxidative state. The major antioxidant enzymes involved in detoxifying ROS include superoxide dismutase (SOD), catalase, glutathione peroxidase (GPx), and peroxiredoxin (Prx) [116]. Nonenzymatic antioxidants include metabolic products such as glutathione (GSH), uric acid, and melatonin [117, 118].

ROS at low levels can act as intracellular second messengers to modulate cellular responses. The very short lifetime and diffusion distance of  $O_2^{\cdot-}$  and  $\cdot OH$  make them unsuitable to function as signaling molecules. In contrast,  $H_2O_2$  mediates reversible oxidation of cysteine residues in proteins, which can alter protein activities and functions [119]. These proteins include enzymes (i.e., mitogen-activated protein kinases (MAPKs), tyrosine kinases, and protein tyrosine phosphatases) and transcription factors (i.e., activator protein-1 (AP-1), nuclear factor kappa-light-chain-enhancer of activated B cells (NF- $\kappa B$ ), and hypoxia-inducible factor 1 (HIF-1)). When ROS production overwhelms the intrinsic antioxidant defense, due to either increased ROS formation or reduced ability to neutralize ROS or both, oxidative stress arises. As a consequence, ROS attack cellular components, leading to potential cell/tissue damage.

**3.2. Normal Pregnancy Is a Mild State of Oxidative Stress.** The metabolic activity of the placenta is high in order to meet the growth of both the placenta and the fetus, leading to increased ROS production during normal pregnancy. It has long been proposed that pregnancy is a state of oxidative stress [120, 121]. This notion is supported by the following observations: (1) increased levels of superoxide  $O_2^{\cdot-}$ , 8-iso-prostaglandin  $F_2\alpha$  (8-iso-PGF $_2\alpha$ ), and malondialdehyde (MDA) in the circulation and the placenta [122–124];

(2) reduced circulating expression and activity of enzymatic antioxidants such as SOD, GPx, and catalase [123, 125]; and (3) decreased levels of nonenzymatic antioxidants including uric acid, vitamin C, vitamin E, and GSH [123, 126]. Notably, the increased ROS production in early pregnancy plays an important role in trophoblast proliferation, differentiation, invasion, and angiogenesis [127, 128]. As gestation advances, placental SODs and catalase as well as total antioxidant capacity also increase [129, 130], which counters the increased ROS generation. Thus, a relatively physiological balance between oxidants and antioxidants is maintained in normal pregnancy.

**3.3. Pregnancy Complications Are Associated with Heightened Oxidative Stress.** Both acute and chronic hypoxia has been shown to elevate ROS [17, 131]. Mitochondria and NOXs are the major sources of ROS in response to oxygen deprivation [132–134]. Placental insufficiency is believed to be a critical element in the pathogenesis of preeclampsia and IUGR. Not surprisingly, these disorders display heightened oxidative stress compared to normal pregnancy. Apparently, both overproduction of ROS and reduction of antioxidant defense contribute to the heightened oxidative stress in pregnancy complications. The activity and expression of oxidant enzymes such as NOXs and XO increased in the preeclamptic placenta and/or circulation [135–137]. In contrast, levels and activity of circulating and placental antioxidant enzymes such as SOD, catalase, and GPx as well as thioredoxin (Trx) were decreased in preeclamptic and IUGR pregnancies [135, 138, 139]. Similarly, activities of SOD, GPx, and Trx reductase (TrxR) were reduced in placentas from high-altitude pregnancy [95]. Moreover, mitochondria in the placenta became dysfunctional in pregnancy complication. Mitochondria appear to be damaged as evidenced by swelling and broken cristae in the preeclamptic placenta [140]. Respiratory chain enzyme expression and activity of mitochondrial complexes were suppressed in preeclamptic and IUGR placentas as well as in placentas from high-altitude pregnancy [140–142], uncoupling respiration from oxidative phosphorylation. Furthermore, circulating and placental nonenzymatic antioxidants including GSH, vitamin C, and melatonin were lower in preeclampsia and IUGR [139, 143, 144]. Concomitantly, both complications exhibited higher ROS [124, 145, 146] and oxidative stress markers [147–149] in the circulation and the placenta, leading to lipid peroxidation and oxidative DNA damage [144, 149]. Increased nitrotyrosine immunostaining was observed in villous vessels of the placenta [94, 135] and systemic vessels [150] in preeclamptic pregnancy, suggesting that preeclampsia promotes NOS uncoupling and OONO $^-$  generation.

**3.4. Animal Models Replicate Oxidative Stress in Pregnancy Complications.** The elevated oxidative stress in preeclampsia and IUGR has been imitated in animal models. Increased placental  $O_2^{\cdot-}$  was observed in an eNOS $^{-/-}$  mouse model of fetal growth restriction [151]. The reduced uterine perfusion pressure model reduced levels of SODs and GPx, increased levels of MDA, and decreased mitochondrial complexes I and II expression [152–154] in rat placentas. Rodent models

of preeclampsia and/or IUGR also promoted eNOS uncoupling in the aorta and placenta [14, 155] and decreased placental GSH content [156]. Therefore, similar to human pregnancy complications, an imbalance between oxidant and antioxidant systems apparently accounts for the heightened oxidative stress in these animal models and hypoxia appeared to be a major cause of the heightened oxidative stress in these models. Increased uterine arterial ROS generation was detected in a sheep model of high-altitude pregnancy due to increased NOX2 expression, which could be replicated by ex vivo hypoxic treatment of uterine arteries [157]. Naïve high-altitude pregnant sheep exhibited higher circulating MDA than low-altitude pregnant sheep and native high-altitude pregnant sheep [30]. In addition, gestational hypoxia increased levels of 4-hydroxynonenal (4-HNE), a lipid peroxidation product, in rat placentas [158].

In conclusion, oxidative stress is an inherent feature of normal pregnancy and plays an important role in the development of the placenta. The uteroplacental system is particularly vulnerable to oxidative stress. When unchecked, oxidative stress becomes augmented and could give rise to pathological conditions such as preeclampsia and IUGA, harming both the mother and the fetus. Therefore, oxidative can play both physiological and pathological roles in the progression and outcome of pregnancy.

#### 4. Regulation of $E_2\beta$ Production and $E_2\beta$ Signaling Pathway by ROS in Pregnancy Complications

As aforementioned,  $E_2\beta$  is an essential element in the adaptation of the uteroplacental circulation during pregnancy. Given heightened oxidative stress in preeclampsia, IUGR and high-altitude pregnancy, and diverse effects of ROS on macromolecules, it is not surprising that excessive ROS plays a critical role in the pathogenesis of these complications by disrupting the  $E_2\beta$  signaling pathway. ROS could directly or indirectly exert their detrimental effects on the targets, and their actions could be acute or chronic. Unfortunately, there is limited information regarding the impacts of ROS on the  $E_2\beta$  signaling pathway in uteroplacental circulation under pathophysiological conditions. In this section, findings from both uteroplacental and nonuteroplacental tissues/cells will be discussed.

**4.1. ROS and Estrogen Synthesis.** Aromatase and HSD17B1, two key enzymes in estrogen biosynthesis catalyze the interconversion between testosterone and  $E_2\beta$  and between estrone and  $E_2\beta$ , respectively, using cofactors NADPH [159, 160]. In fact, NADPH is a key component against cellular oxidation. Maintaining an adequate NADPH/NADP<sup>+</sup> ratio is essential to activities of these enzymes and  $E_2\beta$  generation. In HUVECs, high glucose elevated ROS [161] but reduced the NADPH level [162]. Lowering the NADPH/NADP<sup>+</sup> ratio markedly reduced the conversion of estrone to  $E_2\beta$  in HEK293 cells [163]. Interestingly, the reduced  $E_2\beta$  level in preeclamptic placental explants was mimicked by the treatment of placental explants from normal pregnancy with H<sub>2</sub>O<sub>2</sub> [164]. Moreover, H<sub>2</sub>O<sub>2</sub> treatment

of the homogenate of the human ovary suppressed aromatase activity, which could be prevented by GPx [165]. These observations suggest that oxidative stress could impair estrogen synthesis by suppressing key enzyme activities.

**4.2. ROS and Estrogen Receptor Expression.** ER $\alpha$  expression is also subject to ROS modulation. In general, the expression of ER $\alpha$  is negatively regulated by ROS. The following observations were made in cancer cell lines. In MCF-7 cells, a brief treatment with glucose oxidase, which catalyzes the oxidation of glucose to H<sub>2</sub>O<sub>2</sub> and D-glucono- $\delta$ -lactone, resulted in marked ER $\alpha$  level reduction 24 hours after the treatment [166]. Chronic (16 hours) H<sub>2</sub>O<sub>2</sub> treatment of ZR-75-1 cells also decreased ER $\alpha$  protein level [167]. The detrimental effect of H<sub>2</sub>O<sub>2</sub> on ER $\alpha$  expression could be normalized by increasing antioxidant capacity. Overexpression of Prx-1, a H<sub>2</sub>O<sub>2</sub> scavenger, ablated H<sub>2</sub>O<sub>2</sub>-induced downregulation of ER $\alpha$ , whereas inhibition of Prx-1/2 activity with adenanthin promoted ER $\alpha$  downregulation [167].

**4.3. ROS and NO Production.** NOS are also regulated by ROS. ROS affect NO production apparently through altering eNOS expression/activity and eNOS cofactors. In HUVECs, H<sub>2</sub>O<sub>2</sub> treatment for 2 hours increased eNOS phosphorylation of serine 1177 and enzyme activity, whereas catalase did the opposite [168]. However, H<sub>2</sub>O<sub>2</sub> was found to decrease NO bioavailability in porcine aortic ECs by inactivation of eNOS cofactors without altering enzyme activity [169]. Long-term treatment with H<sub>2</sub>O<sub>2</sub> or superoxide treatment resulted in downregulation of eNOS in HUVECs [170, 171]. NOXs appeared to be major sources of ROS responsible for eNOS downregulation. HUVECs from women with preeclampsia exhibited NOX2 upregulation and eNOS downregulation [113]. In addition, the upregulation of NOX4 by angiotensin II and high glucose promoted eNOS uncoupling, leading to increased generation of O<sub>2</sub><sup>-</sup> and OONO<sup>-</sup> in glomerular mesangial cells [172, 173]. Thus, it is expected that inhibiting oxidant generation or enhancing antioxidant defense could potentially normalize the adverse effect of ROS on eNOS. As expected, eNOS expression was partially rescued or restored by NOX inhibitor apocynin or overexpression of SOD2 [113, 174]. Administration of the GSH synthase inhibitor buthionine sulfoximine into rats decreased total GSH level in the liver, reduced urinary excretion of NOx, and increased nitrotyrosine staining in the kidney without altering renal eNOS level [175].

**4.4. ROS and K<sub>Ca</sub> Channels.** ROS display complex actions toward the BK<sub>Ca</sub> channel. H<sub>2</sub>O<sub>2</sub> could be stimulatory or inhibitory on BK<sub>Ca</sub> channel activity depending on experimental conditions. H<sub>2</sub>O<sub>2</sub> increased BK<sub>Ca</sub> channel activity in human and porcine artery VSMCs and HUVECs [176–178], whereas it decreased BK<sub>Ca</sub> channel-mediated currents in porcine renal artery ECs and vascular smooth muscle-type BK<sub>Ca</sub> channel reconstituted in HEK293 cells [179]. A study by Tang et al. revealed that both cysteine and methionine residues of the BK<sub>Ca</sub> channel were subject to redox modulation [180]. Interestingly, oxidation of cysteine and methionine produced opposite regulations of BK<sub>Ca</sub> channel activity.

Whereas cysteine oxidation decreased BK<sub>Ca</sub> channel currents, methionine oxidation increased channel activity. Moreover, oxidation of a cysteine residue near the Ca<sup>2+</sup> bowl of the BK<sub>Ca</sub> channel  $\alpha$  subunit by H<sub>2</sub>O<sub>2</sub> almost abolished physiological activation of the channel [181]. It is likely that distinct actions of H<sub>2</sub>O<sub>2</sub> on the BK<sub>Ca</sub> channel resulted from selectively targeting cysteine and methionine residues. Whereas O<sub>2</sub><sup>-</sup> did not alter currents mediated by the BK<sub>Ca</sub> channel, ONOO<sup>-</sup> exhibited an inhibitory effect on BK<sub>Ca</sub> channel activity [182, 183]. It appears that the BK<sub>Ca</sub> channel in uterine artery VSMCs of high-altitude pregnant sheep is under tonic inhibition by ROS. An acute application of antioxidants such as N-acetylcysteine (NAC), the NOX inhibitor apocynin, and the synthetic SOD/catalase mimetic EUK-134 partially reversed gestational hypoxia-induced suppression of BK<sub>Ca</sub> channel-mediated currents and vasorelaxation [104, 184]. As NOX2 was upregulated in gestational hypoxia, the superoxide generated by this enzyme and its dismutation product H<sub>2</sub>O<sub>2</sub> probably contributed to the gestational hypoxia-induced suppression of BK<sub>Ca</sub> channel activity/function in uterine arteries [157]. IK channel-mediated currents in HUVECs were also inhibited by the superoxide donors, xanthine/xanthine oxidase (X/XO) mixture [185].

In addition to direct modulation of K<sub>Ca</sub> channel activity, ROS also exert a significant impact on the expression of K<sub>Ca</sub> channels. High-altitude pregnancy increased uterine vascular tone owing to NOX2 overexpression and *KCNMB1* downregulation as well as decreased BK<sub>Ca</sub> channel activity [103, 157]. These detrimental effects could be simulated by ex vivo hypoxic treatment of uterine arteries of low-altitude pregnancy [104]. A cause-and-effect relationship was established by the observation that antioxidants apocynin and NAC largely eliminated gestational hypoxia-induced reduction of *KCNMB1* expression and channel activity [104, 157]. In addition, estrogen-induced upregulation of the BK<sub>Ca</sub> channel  $\beta$ 1 subunit and channel activity in uterine arteries was eradicated by gestational hypoxia, which was restored by NAC in ex vivo experiments [104, 184]. Similarly, preeclampsia reduced the expression of *KCNMB1* along with upregulation of NOX2 and superoxide in HUVECs [113]. Importantly, the *KCNMB1* downregulation was partially rescued by treating cultured HUVECs with apocynin [113]. The *KCNMB1* downregulation appeared to be directly induced by ROS. Exposure of the cultured human coronary artery VSMCs to H<sub>2</sub>O<sub>2</sub> for 12 hours led to reduced *KCNMB1* expression [186]. These observations signal a contributing role of ROS in the dysfunction of the BK<sub>Ca</sub> channel in uteroplacental circulation. Targeting *KCNMB1* expression by ROS is also observed in diabetes. The BK<sub>Ca</sub> channel  $\beta$ 1 subunit protein level was downregulated in diabetic mouse aorta, which was accompanied by increased expression of NOX1 and NOX4, decreased expression of SOD and catalase, and elevated O<sub>2</sub><sup>-</sup> generation [186].

The expression of SK and IK channels is also regulated by ROS in pregnancy complications. Pregnancy/estrogen-induced upregulation of SK2 (K<sub>Ca</sub>2.2) and SK3 (K<sub>Ca</sub>2.3) channel expression/activity in ovine uterine arteries was diminished at high altitude [107], and a causative role of ROS was evidenced by the reversal of gestational

hypoxia-induced detrimental effects with NAC [184]. Treatment of human uterine microvascular ECs with serum from preeclamptic women also reduced SK3 and IK1 expression, which was reversed by silencing NOX4 with siRNA or treatment with a membrane-permeable SOD [187]. The reduced expression of SK3 and IK1 (K<sub>Ca</sub>3.1) in the placenta, umbilical vessels, and HUVECs was also associated with the upregulation of NOX2 or NOX4 and heightened oxidative stress in preeclamptic pregnancy [113, 187, 188]. The contributing role of ROS to the downregulation of SK<sub>Ca</sub> and IK<sub>Ca</sub> channels was substantiated based on the following findings: (1) restoration of channel expression by antioxidants such as apocynin, tempol, and tirion and (2) simulation of the downregulation by oxidants such as superoxide generated by exogenous X/XO mixture and H<sub>2</sub>O<sub>2</sub> [113, 188, 189].

Overwhelming evidence suggests that the E<sub>2</sub> $\beta$ -NOS--NO-K<sub>Ca</sub> channel pathway in uteroplacental tissue is a target of oxidative stress in pregnancy complications. Overall, excessive ROS inhibited E<sub>2</sub> $\beta$  synthesis and estrogen receptor expression. In addition, NOS and K<sub>Ca</sub> channel expression/activity could also be suppressed by oxidative stress, leading to reduced NO bioavailability and impaired K<sub>Ca</sub> functions.

## 5. The Interplay among Hypoxia, ROS, and Epigenetic Modifications in Pregnancy Complications

Although it is now well-recognized that placental insufficiency and oxidative stress are important contributors to the pathogenesis of preeclampsia and IUGR, the mechanisms underlying their actions in these complications are not fully resolved. Recent studies have identified epigenetic modifications as important mechanisms underlying various human diseases [190]. In this section, we will try to establish a link among hypoxia, ROS, and epigenome in preeclampsia and IUGR.

**5.1. ROS in O<sub>2</sub> Sensing.** HIFs are transcription factors and function as master regulators of cellular responses to hypoxia. HIFs are heterodimers composed of a HIF- $\alpha$  subunit (HIF-1 $\alpha$  and HIF-2 $\alpha$ ) and a constitutively expressed HIF-1 $\beta$  subunit. Under normoxia, HIF- $\alpha$  subunits are hydroxylated on proline residues by the O<sub>2</sub>-dependent prolyl hydroxylases (PHDs), resulting in ubiquitination and successive proteasomal degradation by the von Hippel-Lindau protein (pVHL) E3-ubiquitin ligase. In hypoxia, PHD activity is suppressed. Subsequently, HIF- $\alpha$  is accumulated, translocated into the nucleus, and dimerized with HIF-1 $\beta$ , leading to gene expression by binding to hypoxia-responsive element (HRE) in the promoter of the target gene. Interestingly, ROS appear to participate in cellular oxygen sensing and hypoxic activation of HIFs. ROS generated by mitochondrial complex III in response to hypoxia were found to stabilize HIF-1 $\alpha$  [132, 191]. The stabilization of HIF-1 $\alpha$  was mimicked by exogenous H<sub>2</sub>O<sub>2</sub> and by genetic suppression of SOD2 under normoxia [191, 192]. However, HIF-1 $\alpha$  stabilization was attenuated by silencing Rieske iron-sulfur protein of complex III and by enzymatic and nonenzymatic antioxidants

[193–195]. ROS produced by NOXs could also lead to accumulation of HIF-1 $\alpha$  [196, 197] and HIF-2 $\alpha$  [198, 199]. ROS stabilized HIF- $\alpha$  apparently through suppressing the ability of the PHDs to hydroxylate HIF- $\alpha$  protein [200]. ROS-mediated stabilization of HIFs thus constitutes an important mechanism for hypoxia to stimulate gene expression.

**5.2. Crosstalk between ROS and Epigenome.** Whereas genome confers genetic information for making and maintaining an organism, the epigenome describes all the chemical modifications to DNA and histone proteins. Epigenetic modifications of the genome determine how the information in genes is expressed by switching genes on and off without altering the DNA sequence. The major mechanisms of the epigenetic modification include DNA methylation, histone modifications, and noncoding-RNA-based silencing [201]. Several lines of evidence suggest existence of a crosstalk between ROS and epigenetic modifications. ROS are found to promote DNA hypermethylation by altering DNA methylation/demethylation machineries and enzyme recruitment. In vitro studies demonstrated that H<sub>2</sub>O<sub>2</sub> treatment increased expression/activity of DNA methyltransferases (DNMTs) [202–204], although many of these studies were conducted in cancer cell lines. In addition, H<sub>2</sub>O<sub>2</sub> could facilitate DNA methylation by recruiting DNMT1 to the CpG sites in gene promoters [203, 205]. The linking of ROS induced by hypoxia and other stimuli to DNA hypermethylation was further confirmed by findings that antioxidants such as NAC and apocynin were able to prevent both ROS-induced global methylation or specific gene methylation [202, 206, 207] and upregulation of DNMTs [202]. ROS could also impair DNA demethylation. In a cell-free system, H<sub>2</sub>O<sub>2</sub> suppressed enzymatic activity of ten-eleven translocation (TET) dioxygenase [208]. The catalytic activity of TETs requires vitamin C and Fe<sup>2+</sup> as cofactors [209, 210]. To maintain an active dioxygenase enzyme, vitamin C is required to reduce Fe<sup>3+</sup> to Fe<sup>2+</sup>. Thus, vitamin C depletion in pregnancy complications [144, 211] would reduce TET activity. Histone modifications are also subject to ROS regulation. It is found that increasing oxidative stress by H<sub>2</sub>O<sub>2</sub> upregulated histone deacetylase 1 (HDAC1) in cancer cell lines [204]. Prolonged treatment with H<sub>2</sub>O<sub>2</sub> also increased global histone methylation marks H3K4me3 and H3K27me3 in human bronchial epithelial cells [208]. It appears that ROS produced from both mitochondria and NOX promotes microRNA-210 (miR-210) generation. Whereas Nox4 siRNA partially decreased hypoxia-induced miR-210 expression, mitochondrial complexes I and III inhibitors rotenone and antimycin increased miR-210 biogenesis in adipose-derived stem cells [212].

Conversely, ROS production could be altered by epigenetic modifications of genes for enzymatic oxidants and antioxidants. It appears that hypermethylation promotes oxidative stress, whereas demethylation boosts antioxidation. In human pulmonary arterial hypertension, a CpG island in an enhancer region of intron 2 and another in the promoter of SOD2 were hypermethylated in pulmonary arterial smooth muscle cells (PASMCs) owing to upregulation of DNMT1

and DNMT3b, leading to downregulation of the antioxidant enzyme [213]. Similarly, hypoxia also reduced SOD2 expression in the rat carotid body via hypermethylation of a single CpG dinucleotide close to the transcription start site [214]. H<sub>2</sub>O<sub>2</sub> promoted methylation of a CpG island in the catalase promoter and downregulated catalase [215]. TET1 deficiency produced by TET1 siRNA enhanced H<sub>2</sub>O<sub>2</sub>-induced increase apoptosis of cerebellar granule cells [216], suggesting that TET1-mediated demethylation may upregulate antioxidant mechanisms to counter oxidative stress. Histone modification also contributes to the hemostasis of the oxidant-antioxidant system. The expression/activity of SOD3 in the lung from human idiopathic pulmonary arterial hypertension was reduced, and this downregulation could be reversed by the treatment of PASMCs with class I HDAC inhibitors or HDAC3 siRNA [217], suggesting that histone deacetylation negatively regulates SOD3 expression. In contrast, histone deacetylation mediated by HDAC3 upregulated NOX4 in HUVECs as HDAC3 siRNA and pan-HDAC inhibitor scriptaid reduced NOX4 expression [218]. Furthermore, miRs also participate in the regulation of mitochondrial metabolism and function. The downregulation of iron-sulfur cluster assembly enzyme (ISCU) in mitochondria by miR-210 in hypoxia would block electron exit from complex I, promoting its leakage to generation of ROS [219]. Overall, it appears that there exists a positive feed-forward loop between ROS generation and epigenetic modifications.

**5.3. Epigenetic Mechanisms in Regulating Uteroplacental Circulation during Normal Pregnancy.** In sheep, the upregulation of ER $\alpha$  in uterine arteries was conferred by an epigenetic mechanism [220]. The specificity protein 1- (Sp1-) binding site (Sp1<sub>-520</sub>) at the promoter of the ER $\alpha$  encoding gene *ESR1*, to which Sp1 or Sp1-ER $\alpha$  binds, was essential for E2 $\beta$ -stimulated promoter activity. The CpG dinucleotide of this site was hypermethylated in nonpregnant animals, and the gene is thus kept quiescent. However, the Sp1 site became less methylated in pregnant animals and enabled the expression of the gene, leading to increased ER $\alpha$  mRNA and protein abundance in uterine arteries and subsequent attenuation of uterine vascular tone.

E2 $\beta$  also epigenetically upregulates *KCNMB1* expression in uterine arteries [221, 222]. Similar to ER $\alpha$ , the CpG dinucleotide in the Sp1-binding site (-380) at the promoter of *KCNMB1* was highly methylated in uterine arteries of nonpregnant sheep, resulting in gene silence. During pregnancy, E2 $\beta$  through ER $\alpha$  stimulated *TET1* (TET1 encoding gene) promoter activity and gene expression. The upregulation of *TET1* in turn promoted Sp1<sub>-380</sub> demethylation of the *KCNMB1* promoter. Consequently, the expression of *KCNMB1* and the activity of the BK<sub>Ca</sub> channel increased in uterine arteries, leading to reduced myogenic tone.

**5.4. Aberrant Epigenetic Modifications in Pregnancy Complications.** Epigenetic mechanisms play an important role in the pathophysiological processes of pregnancy complications. Global hypermethylation was observed in pre-eclamptic placenta [223, 224]. In addition, various genes including *ESR1* and *KCNMB1* in the uterine arteries of



high-altitude pregnant sheep [52, 220, 221, 225] and *IGF1*, *HSD11B2*, *H19*, and *HLA-G* in the placenta from preeclamptic and IUGR pregnancies [224, 226, 227] were hypermethylated. The increased methylation in the uteroplacental tissues was accompanied by upregulation of DNMT1 and DNMT3b expression/activity [224, 225, 227, 228] and downregulation of TET1, TET2, and TET3 expression [52, 227, 229, 230]. Pregnancy complications also alter histone modification in the placenta. JMJD6 histone demethylase activity was suppressed in preeclamptic placenta [231]. Moreover, miR-210 was also upregulated in both uterine arteries and placenta of high-altitude pregnancy [52, 142]. Increased miR-210 level was also observed in preeclamptic and IUGR placenta [140, 230, 232]. These changes undoubtedly would contribute to the aberrant expression of key elements in the  $E_2\beta$ -NOS-NO- $K_{Ca}$  pathway in uteroplacental circulation.

The aforementioned changes in epigenetic modifications of the uteroplacental system in pregnancy complications are apparently caused by hypoxia/ischemia. HIF-1 $\alpha$  overexpression in uteroplacental tissues is a characterized feature in pregnancy complications and high-altitude pregnancy [157, 233, 234]. Both ex vivo hypoxia treatment of tissues or pharmacologically induced hypoxia in intact animal models induced the expression of DNMTs and miR-210 [142, 225, 235] and repressed both histone demethylase activity [231] and TETs expression/activity [235, 236]. Although not investigated in the uteroplacental tissues, studies conducted in other tissues/cells suggest that hypoxia-induced alterations in epigenetic machineries is HIF-1 $\alpha$ -dependent. *DNMT1*, *DNMT3b*, and *miR210* all contain hypoxia-responsive element (HRE) in their promoters, and the binding of HIF-1 $\alpha$  to HRE stimulates the expression of these genes [237]. Hypoxia *via* HIF-1 $\alpha$  also induced the expression of histone demethylases JHDM1B/KDM2B and JARID1B/KDM5B, which demethylate the activating mark H3K4me2/3, leading to gene repression [238]. The  $E_2\beta$  metabolite 2-methoxyestradiol is an endogenous HIF inhibitor [239]. The reduced 2-methoxyestradiol level in preeclampsia probably contributes to aberrant epigenetic modifications in uteroplacental tissues due to the relief of HIF inhibition.

Intriguingly, hypoxia-induced *TET1* repression in uterine arteries was mediated by miR-210 and the binding of miR-210 to the 3'-untranslated region (3'UTR) of *TET1* mRNA resulted in degradation of the transcript [52]. The overall effects of upregulation of DNMT3b and downregulation of *TET1* in uterine arteries promoted *ESR1* and *KCNMB1* hypermethylation and gene repression [52, 220, 221, 225, 235].  $ER\alpha$  and the  $BK_{Ca}$  channel are two key elements contributing to reduced uterine vascular tone in pregnancy [59, 73]. Consequently, the downregulation of both  $ER\alpha$  and the  $BK_{Ca}$  channel impaired pregnancy-induced attenuation of uterine vascular tone, leading to maladaptation of uteroplacental circulation [40, 47, 225] (Figure 2). Increased DNA methylation may also contribute to impaired spiral artery remodeling. The downregulation of *TET2* reduced *in vitro* trophoblast migration and invasion [230]. The overexpression of miR-210 in the preeclamptic placenta suppressed ISCU and impaired mitochondrial respiration [140, 142, 232]. It is

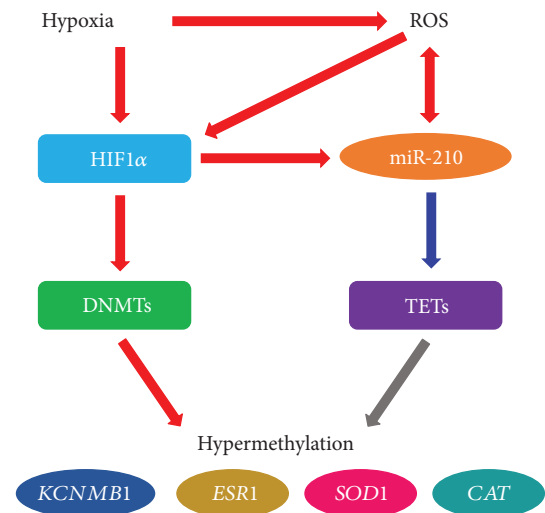


FIGURE 2: Crosstalk among hypoxia, ROS, and DNA methylation. The cellular responses to hypoxia are primarily mediated by hypoxia-inducible factor-1 (HIF-1). Hypoxia could induce HIF-1 either directly or indirectly through the stabilization of HIF-1 $\alpha$  by ROS. HIF-1 could upregulate DNA methyltransferases (DNMTs) and miR-210. ROS may be able directly to induce miR-210. *TET1* mRNA is a target of miR-210 and is degraded upon the binding of miR-210 to the 3'UTR of the transcript. The upregulation of DNMTs and downregulation of *TET1* result in hypermethylation of *ESR*, *KCNMB1*, *SOD1*, and *CAT* (catalase encoding gene) and subsequent gene repression. The repression of *ESR* and *KCNMB1* ultimately increases uterine vascular tone, whereas the repression of *SOD1* and *CAT* elevates ROS. Moreover, miR-210 could also target iron-sulfur cluster scaffold (ISCU) in the mitochondria leading to increased ROS formation. Red arrow: stimulatory effect, blue arrow: inhibitory effect, and grey arrow: indirect action due to *TET1* downregulation.

probably that both the miR-210-mediated mitochondrial dysfunction and DNA hypermethylation (indirectly *via* downregulating TETs) disrupt trophoblast invasion and impair spiral artery remodeling in high-altitude pregnancy and pregnancy complications. In addition, miR-210 also targeted potassium channel modulatory factor 1 (KCMF1) and thrombospondin type I domain-containing 7A (THSD7A), which could also contribute to the impaired trophoblast invasion [240, 241]. The expression of *CYP19A1* and *HSD17B1* is also regulated by DNA methylation. Methylation of CpG islands in the promoters of both genes suppressed their expression [242, 243]. Although not examined in the placenta, it is probably DNA methylation-mediated downregulation of aromatase and *HSD17B1* also occurs in preeclampsia, IUGR, and high-altitude pregnancy. Furthermore, the expression of *HSD17B1* was downregulated by miR-210 in preeclamptic placenta [33]. The epigenetic modifications of key enzymes in estrogen biosynthesis could then reduce circulating  $E_2\beta$  level in pregnancy complications.

## 6. Concluding Remarks

Preeclampsia and IUGR are leading causes of maternal and perinatal mortality and morbidity and have great impacts

on maternal and offspring health. Unfortunately, there is currently no cure for them. Preeclampsia, IUGR, and high-altitude pregnancy all exhibit uteroplacental hypoxia/ischemia and oxidative stress concurrently. Moreover, these pregnancy complications are associated with altered epigenome. There exist interplays among ROS, HIFs, and epigenome. The ROS-HIF pathway appears to be a potential cause in the changes of epigenetic modifications in these complications. In uterine arteries, HIF-1 $\alpha$  apparently functions as an important link between ROS and aberrant epigenetic modifications, leading to disrupted E<sub>2</sub> $\beta$ -BK<sub>Ca</sub> axis and increased uterine vascular tone. In the placenta, the ROS-HIF-epigenome interplay impairs estrogen synthesis, trophoblast invasion, and spiral artery transformation. Both preeclampsia and IUGR are multifactorial disorders. What we know about these complications is only the tip of the iceberg. Further studies are needed to advance our understanding on the pathogenesis of them in order to develop effective therapeutics.

## Abbreviations

3'UTR:	3'-Untranslated region	H <sub>2</sub> O <sub>2</sub> :	Hydrogen peroxide
4-HNE:	4-Hydroxynonenal	HRE:	Hypoxia-responsive element
8-iso-PGF <sub>2</sub> $\alpha$ :	8-Iso-prostaglandin F <sub>2</sub> $\alpha$	<i>HSD11B2</i> :	The gene encoding hydroxysteroid 11 $\beta$ -dehydrogenase 2
ADMA:	Asymmetric dimethylarginine	HSD17B1 (17 $\beta$ -HSD1):	Hydroxysteroid 17 $\beta$ -dehydrogenases 1
AP-1:	Activator protein-1	HUVEC:	Human umbilical vein endothelial cell
AT1-AA:	Autoantibodies against angiotensin II type I receptor	<i>IGF1</i> :	The gene encoding insulin-like growth factor 1
BK <sub>Ca</sub> :	Large-conductance Ca <sup>2+</sup> -activated K <sup>+</sup> channel	IK:	Intermediate-conductance Ca <sup>2+</sup> -activated K <sup>+</sup> channel
cAMP:	Cyclic adenosine monophosphate	IUGR:	Intrauterine growth restriction
cGMP:	Cyclic guanosine monophosphate	ISCU:	Iron-sulfur cluster scaffold
COMT:	Catechol-O-methyltransferase	K <sub>Ca</sub> :	Ca <sup>2+</sup> -activated K <sup>+</sup> channel
CpG:	Cytosine-guanine dinucleotide	KCMF1:	Potassium channel modulatory factor 1
CYP19:	Aromatase	<i>KCNMB1</i> :	The gene encoding BK <sub>Ca</sub> channel $\beta$ subunit 1
<i>CYP19A1</i> :	The gene encoding aromatase	L-NAME:	L-Nitro-arginine methyl ester or N <sup>ω</sup> -nitro-L-arginine methyl ester
DNMT:	DNA methyltransferase	MDA:	Malondialdehyde
E2 $\beta$ :	17 $\beta$ -Estradiol	MAPKs:	Mitogen-activated protein kinases
ECs:	Endothelial cells	miR:	MicroRNA
EDHF:	Endothelium-derived hyperpolarizing factor	NAC:	N-Acetylcysteine
eNOS:	Endothelial nitric oxide synthase	NADP:	Nicotinamide adenine dinucleotide phosphate
ER $\alpha$ :	Estrogen receptor $\alpha$	NADPH:	Reduced form of NADP <sup>+</sup>
ER $\beta$ :	Estrogen receptor $\beta$	NF- $\kappa$ B:	Nuclear factor kappa-light-chain-enhancer of activated B cells
ERK:	Extracellular signal-regulated kinase	NO:	Nitric oxide
ERE:	Estrogen response element	NOS:	Nitric oxide synthases
<i>ESR1</i> :	The gene encoding ER $\alpha$	NOX:	NADPH oxidase
GPER (GPR30):	G-protein-coupled estrogen receptor	O <sub>2</sub> :	Oxygen
GPx:	Glutathione peroxidase	O <sub>2</sub> <sup>-•</sup> :	Superoxide
GSH:	Glutathione	•OH:	Hydroxyl radical
<i>H19</i> :	The gene encoding imprinted maternally expressed transcript	ONOO <sup>-</sup> :	Peroxynitrite
HRE:	Hypoxia-responsive element	PASMC:	Pulmonary arterial smooth muscle cell
HDAC:	Histone deacetylase	PHD:	Prolyl hydroxylases
HIF:	Hypoxia-inducible factor	PCR:	Polymerase chain reaction
<i>HLA-G</i> :	The gene encoding major histocompatibility complex, class I, G	PI3K:	Phosphoinositide 3-kinase
		PKB (AKT):	Protein kinase B
		PKG:	Protein kinase G
		Prx:	Peroxiredoxin
		pVHL:	von Hippel-Lindau protein
		ROS:	Reactive oxygen species
		siRNA:	Small interfering RNA
		SK:	Small-conductance Ca <sup>2+</sup> -activated K <sup>+</sup> channel
		SOD:	Superoxide dismutase
		Sp1:	Specificity protein 1
		TET:	Ten-eleven translocation dioxygenase
		THSD7A:	Thrombospondin type I domain containing 7A
		Trx:	Thioredoxin
		TrxR:	Thioredoxin reductase
		VSMCs:	Vascular smooth muscle cells

X: Xanthine  
 XO: Xanthine oxidase.

## Conflicts of Interest

None of the authors has any conflict of interests to disclose.

## Acknowledgments

This work was supported by National Institutes of Health Grants HD083132 (L. Zhang), HL128209 (L. Zhang), and HL137649 (L. Zhang).

## References

- [1] C. A. Ducusay, R. Goyal, W. J. Pearce, S. Wilson, X. Q. Hu, and L. Zhang, "Gestational hypoxia and developmental plasticity," *Physiological Reviews*, vol. 98, no. 3, pp. 1241–1334, 2018.
- [2] T. Napso, H. E. J. Yong, J. Lopez-Tello, and A. N. Sferruzzi-Perri, "The role of placental hormones in mediating maternal adaptations to support pregnancy and lactation," *Frontiers in Physiology*, vol. 9, p. 1091, 2018.
- [3] K. Chang and L. Zhang, "Review article: steroid hormones and uterine vascular adaptation to pregnancy," *Reproductive Sciences*, vol. 15, no. 4, pp. 336–348, 2008.
- [4] M. B. Pastore, S. O. Jobe, J. Ramadoss, and R. R. Magness, "Estrogen receptor- $\alpha$  and estrogen receptor- $\beta$  in the uterine vascular endothelium during pregnancy: functional implications for regulating uterine blood flow," *Seminars in Reproductive Medicine*, vol. 30, no. 1, pp. 46–61, 2012.
- [5] N. O. Lunell, L. E. Nylund, R. Lewander, B. Sarby, and S. Thornström, "Uteroplacental blood flow in pre-eclampsia measurements with indium-113m and a computer-linked gamma camera," *Clinical and Experimental Hypertension Part B: Hypertension in Pregnancy*, vol. 1, no. 1, pp. 105–117, 1982.
- [6] J. C. Konje, E. S. Howarth, P. Kaufmann, and D. J. Taylor, "Longitudinal quantification of uterine artery blood volume flow changes during gestation in pregnancies complicated by intrauterine growth restriction," *BJOG: An International Journal of Obstetrics & Gynaecology*, vol. 110, no. 3, pp. 301–305, 2003.
- [7] L. Brouwers, A. J. van der Meiden-van Roest, C. Savelkoul et al., "Recurrence of pre-eclampsia and the risk of future hypertension and cardiovascular disease: a systematic review and meta-analysis," *BJOG: An International Journal of Obstetrics & Gynaecology*, vol. 125, no. 13, pp. 1642–1654, 2018.
- [8] E. Kajantie, J. G. Eriksson, C. Osmond, K. Thornburg, and D. J. P. Barker, "Pre-eclampsia is associated with increased risk of stroke in the adult offspring: the Helsinki birth cohort study," *Stroke*, vol. 40, no. 4, pp. 1176–1180, 2009.
- [9] J. J. M. Geelhoed, A. Fraser, K. Tilling et al., "Preeclampsia and gestational hypertension are associated with childhood blood pressure independently of family adiposity measures: the Avon longitudinal study of parents and children," *Circulation*, vol. 122, no. 12, pp. 1192–1199, 2010.
- [10] B. T. Alexander, J. H. Dasinger, and S. Intapad, "Fetal programming and cardiovascular pathology," *Comprehensive Physiology*, vol. 5, no. 2, pp. 997–1025, 2015.
- [11] U. Neitzke, T. Harder, and A. Plagemann, "Intrauterine growth restriction and developmental programming of the metabolic syndrome: a critical appraisal," *Microcirculation*, vol. 18, no. 4, pp. 304–311, 2011.
- [12] N. Soleymanlou, I. Jurisica, O. Nevo et al., "Molecular evidence of placental hypoxia in preeclampsia," *The Journal of Clinical Endocrinology and Metabolism*, vol. 90, no. 7, pp. 4299–4308, 2005.
- [13] J. Zhou, D. Xiao, Y. Hu et al., "Gestational hypoxia induces preeclampsia-like symptoms via heightened endothelin-1 signaling in pregnant rats," *Hypertension*, vol. 62, no. 3, pp. 599–607, 2013.
- [14] C. F. Rueda-Clausen, J. L. Stanley, D. F. Thambiraj, R. Poudel, S. T. Davidge, and P. N. Baker, "Effect of prenatal hypoxia in transgenic mouse models of preeclampsia and fetal growth restriction," *Reproductive Sciences*, vol. 21, no. 4, pp. 492–502, 2014.
- [15] K. L. Brain, B. J. Allison, Y. Niu et al., "Induction of controlled hypoxic pregnancy in large mammalian species," *Physiological Reports*, vol. 3, no. 12, article e12614, 2015.
- [16] H. Sies, C. Berndt, and D. P. Jones, "Oxidative stress," *Annual Review of Biochemistry*, vol. 86, no. 1, pp. 715–748, 2017.
- [17] T. Miyata, S. Takizawa, and C. van Ypersele de Strihou, "Hypoxia. 1. Intracellular sensors for oxygen and oxidative stress: novel therapeutic targets," *American Journal of Physiology-Cell Physiology*, vol. 300, no. 2, pp. C226–C231, 2011.
- [18] G. J. Burton, H. W. Yung, T. Cindrova-Davies, and D. S. Charnock-Jones, "Placental endoplasmic reticulum stress and oxidative stress in the pathophysiology of unexplained intrauterine growth restriction and early onset preeclampsia," *Placenta*, vol. 30, Supplement, pp. 43–48, 2009.
- [19] J. M. Roberts, "Pathophysiology of ischemic placental disease," *Seminars in Perinatology*, vol. 38, no. 3, pp. 139–145, 2014.
- [20] A. Agarwal, S. Gupta, and R. K. Sharma, "Role of oxidative stress in female reproduction," *Reproductive Biology and Endocrinology*, vol. 3, no. 1, p. 28, 2005.
- [21] R. Aouache, L. Biward, D. Vaiman, and F. Miralles, "Oxidative stress in preeclampsia and placental diseases," *International Journal of Molecular Sciences*, vol. 19, no. 5, p. 1496, 2018.
- [22] N. Berkane, P. Liere, J. P. Oudinet et al., "From pregnancy to preeclampsia: a key role for estrogens," *Endocrine Reviews*, vol. 38, no. 2, pp. 123–144, 2017.
- [23] P. Ball and R. Knuppen, "Catecholestrogens (2- and 4-hydroxyoestrogens): chemistry, biogenesis, metabolism, occurrence and physiological significance," *Acta Endocrinologica Supplementum*, vol. 232, pp. 1–127, 1980.
- [24] A. Perez-Sepulveda, L. J. Monteiro, A. Dobierzewska et al., "Placental aromatase is deficient in placental ischemia and preeclampsia," *PLoS One*, vol. 10, no. 10, article e0139682, 2015.
- [25] S. O. Jobe, C. T. Tyler, and R. R. Magness, "Aberrant synthesis, metabolism, and plasma accumulation of circulating estrogens and estrogen metabolites in preeclampsia implications for vascular dysfunction," *Hypertension*, vol. 61, no. 2, pp. 480–487, 2013.
- [26] A. Hertig, P. Liere, N. Chabbert-Buffet et al., "Steroid profiling in preeclamptic women: evidence for aromatase deficiency," *American Journal of Obstetrics and Gynecology*, vol. 203, no. 5, pp. 477.e1–477.e9, 2010.

- [27] U. Pecks, W. Rath, N. Kleine-Eggebrecht et al., "Maternal serum lipid, estradiol, and progesterone levels in pregnancy, and the impact of placental and hepatic pathologies," *Geburtshilfe und Frauenheilkunde*, vol. 76, no. 7, pp. 799–808, 2016.
- [28] L. A. Sobrevilla, I. Romero, F. Kruger, and J. Whittembury, "Low estrogen excretion during pregnancy at high altitude," *American Journal of Obstetrics and Gynecology*, vol. 102, no. 6, pp. 828–833, 1968.
- [29] S. Zamudio, K. K. Leslie, M. White, D. D. Hagerman, and L. G. Moore, "Low serum estradiol and high serum progesterone concentrations characterize hypertensive pregnancies at high altitude," *Journal of the Society for Gynecologic Investigation*, vol. 1, no. 3, pp. 197–205, 1994.
- [30] V. H. Parraguez, S. Mamani, E. Cofre et al., "Disturbances in maternal steroidogenesis and appearance of intrauterine growth retardation at high-altitude environments are established from early pregnancy. Effects of treatment with antioxidant vitamins," *PLoS One*, vol. 10, no. 11, article e0140902, 2015.
- [31] S. M. Charles, C. G. Julian, E. Vargas, and L. G. Moore, "Higher estrogen levels during pregnancy in Andean than European residents of high altitude suggest differences in aromatase activity," *The Journal of Clinical Endocrinology and Metabolism*, vol. 99, no. 8, pp. 2908–2916, 2014.
- [32] K. Kanasaki, K. Palmsten, H. Sugimoto et al., "Deficiency in catechol-O-methyltransferase and 2-methoxyoestradiol is associated with pre-eclampsia," *Nature*, vol. 453, no. 7198, pp. 1117–1121, 2008.
- [33] O. Ishibashi, A. Ohkuchi, M. M. Ali et al., "Hydroxysteroid (17- $\beta$ ) dehydrogenase 1 is dysregulated by miR-210 and miR-518c that are aberrantly expressed in preeclamptic placentas: a novel marker for predicting preeclampsia," *Hypertension*, vol. 59, no. 2, pp. 265–273, 2012.
- [34] N. Berkane, P. Liere, G. Lefevre et al., "Abnormal steroidogenesis and aromatase activity in preeclampsia," *Placenta*, vol. 69, pp. 40–49, 2018.
- [35] B. Jiang, A. Kamat, and C. R. Mendelson, "Hypoxia prevents induction of aromatase expression in human trophoblast cells in culture: potential inhibitory role of the hypoxia-inducible transcription factor Mash-2 (mammalian achaete-scute homologous protein-2)," *Molecular Endocrinology*, vol. 14, no. 10, pp. 1661–1673, 2000.
- [36] M. Marino, P. Galluzzo, and P. Ascenzi, "Estrogen signaling multiple pathways to impact gene transcription," *Current Genomics*, vol. 7, no. 8, pp. 497–508, 2006.
- [37] E. R. Levin, "Plasma membrane estrogen receptors," *Trends in Endocrinology and Metabolism*, vol. 20, no. 10, pp. 477–482, 2009.
- [38] T. Tropea, E. M. de Francesco, D. Rigracciolo et al., "Pregnancy augments G protein estrogen receptor (GPER) induced vasodilation in rat uterine arteries via the nitric oxide - cGMP signaling pathway," *PLoS One*, vol. 10, no. 11, article e0141997, 2015.
- [39] J. Fujimoto, Y. Nakagawa, H. Toyoki, H. Sakaguchi, E. Sato, and T. Tamaya, "Estrogen-related receptor expression in placenta throughout gestation," *The Journal of Steroid Biochemistry and Molecular Biology*, vol. 94, no. 1-3, pp. 67–69, 2005.
- [40] K. Chang, D. Xiao, X. Huang et al., "Chronic hypoxia inhibits sex steroid hormone-mediated attenuation of ovine uterine arterial myogenic tone in pregnancy," *Hypertension*, vol. 56, no. 4, pp. 750–757, 2010.
- [41] W. X. Liao, R. R. Magness, and D. B. Chen, "Expression of estrogen receptors- $\alpha$  and - $\beta$  in the pregnant ovine uterine artery endothelial cells in vivo and in vitro," *Biology of Reproduction*, vol. 72, no. 3, pp. 530–537, 2005.
- [42] M. J. Byers, A. Zangl, T. M. Phernetton, G. Lopez, D. B. Chen, and R. R. Magness, "Endothelial vasodilator production by ovine uterine and systemic arteries: ovarian steroid and pregnancy control of ER $\alpha$  and ER $\beta$  levels," *The Journal of Physiology*, vol. 565, no. 1, pp. 85–99, 2005.
- [43] C. Tong, X. Feng, J. Chen et al., "G protein-coupled receptor 30 regulates trophoblast invasion and its deficiency is associated with preeclampsia," *Journal of Hypertension*, vol. 34, no. 4, pp. 710–718, 2016.
- [44] B. Schiessl, I. Mylonas, P. Hantschmann et al., "Expression of endothelial NO synthase, inducible NO synthase, and estrogen receptors alpha and beta in placental tissue of normal, preeclamptic, and intrauterine growth-restricted pregnancies," *The Journal of Histochemistry and Cytochemistry*, vol. 53, no. 12, pp. 1441–1449, 2005.
- [45] M. N. Park, K. H. Park, J. E. Lee et al., "The expression and activation of sex steroid receptors in the preeclamptic placenta," *International Journal of Molecular Medicine*, vol. 41, no. 5, pp. 2943–2951, 2018.
- [46] G. Yin, X. Zhu, C. Guo et al., "Differential expression of estradiol and estrogen receptor  $\alpha$  in severe preeclamptic pregnancies compared with normal pregnancies," *Molecular Medicine Reports*, vol. 7, no. 3, pp. 981–985, 2013.
- [47] M. Chen, D. Xiao, X. Q. Hu, C. Dasgupta, S. Yang, and L. Zhang, "Hypoxia represses ER- $\alpha$  expression and inhibits estrogen-induced regulation of Ca<sup>2+</sup>-activated K<sup>+</sup> channel activity and myogenic tone in ovine uterine arteries: causal role of DNA methylation," *Hypertension*, vol. 66, no. 1, pp. 44–51, 2015.
- [48] J. Li, Z. Chen, X. Zhou et al., "Imbalance between proliferation and apoptosis-related impaired GPR30 expression is involved in preeclampsia," *Cell and Tissue Research*, vol. 366, no. 2, pp. 499–508, 2016.
- [49] S. P. Ford, "Control of uterine and ovarian blood flow throughout the estrous cycle and pregnancy of ewes, sows and cows," *Journal of Animal Science*, vol. 55, Supplement 2, pp. 32–42, 1982.
- [50] T. C. Gibson, T. M. Phernetton, M. C. Wiltbank, and R. R. Magness, "Development and use of an ovarian synchronization model to study the effects of endogenous estrogen and nitric oxide on uterine blood flow during ovarian cycles in sheep," *Biology of Reproduction*, vol. 70, no. 6, pp. 1886–1894, 2004.
- [51] B. J. Sprague, T. M. Phernetton, R. R. Magness, and N. C. Chesler, "The effects of the ovarian cycle and pregnancy on uterine vascular impedance and uterine artery mechanics," *European Journal of Obstetrics, Gynecology, and Reproductive Biology*, vol. 144, Supplement 1, pp. S170–S178, 2009.
- [52] X. Q. Hu, C. Dasgupta, D. Xiao, X. Huang, S. Yang, and L. Zhang, "MicroRNA-210 targets ten-eleven translocation methylcytosine dioxygenase 1 and suppresses pregnancy-mediated adaptation of large conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channel expression and function in ovine uterine arteries," *Hypertension*, vol. 70, no. 3, pp. 601–612, 2017.
- [53] R. R. Magness, T. M. Phernetton, T. C. Gibson, and D. B. Chen, "Uterine blood flow responses to ICI 182 780 in ovariectomized oestradiol-17 $\beta$ -treated, intact follicular and

- pregnant sheep,” *The Journal of Physiology*, vol. 565, no. 1, pp. 71–83, 2005.
- [54] L. L. Penney, R. J. Frederick, and G. W. Parker, “ $17\beta$ -Estradiol stimulation of uterine blood flow in oophorectomized rabbits with complete inhibition of uterine ribonucleic acid synthesis,” *Endocrinology*, vol. 109, no. 5, pp. 1672–1676, 1981.
- [55] E. Majid and J. Senior, “Anti-oestrogen modification of uterine responses to oestrogen in the rat,” *Journal of Reproduction and Fertility*, vol. 66, no. 1, pp. 79–85, 1982.
- [56] R. R. Magness and C. R. Rosenfeld, “Local and systemic estradiol-17 beta: effects on uterine and systemic vasodilation,” *The American Journal of Physiology*, vol. 256, 4 Part 1, pp. E536–E542, 1989.
- [57] R. R. Magness, T. M. Phernetton, and J. Zheng, “Systemic and uterine blood flow distribution during prolonged infusion of  $17\beta$ -estradiol,” *The American Journal of Physiology*, vol. 275, 3 Part 2, pp. H731–H743, 1998.
- [58] W. A. Salhab, P. W. Shaul, B. E. Cox, and C. R. Rosenfeld, “Regulation of types I and III NOS in ovine uterine arteries by daily and acute estrogen exposure,” *American Journal of Physiology. Heart and Circulatory Physiology*, vol. 278, no. 6, pp. H2134–H2142, 2000.
- [59] D. Xiao, X. Huang, S. Yang, and L. Zhang, “Direct chronic effect of steroid hormones in attenuating uterine arterial myogenic tone: role of protein kinase C/extracellular signal-regulated kinase 1/2,” *Hypertension*, vol. 54, no. 2, pp. 352–358, 2009.
- [60] S. O. Jobe, J. Ramadoss, J. M. Koch, Y. Jiang, J. Zheng, and R. R. Magness, “Estradiol- $17\beta$  and its cytochrome P450- and catechol- O -methyltransferase-derived metabolites stimulate proliferation in uterine artery endothelial cells: role of estrogen receptor- $\alpha$  versus estrogen receptor- $\beta$ ,” *Hypertension*, vol. 55, no. 4, pp. 1005–1011, 2010.
- [61] A. Perez-Sepulveda, P. P. Espana-Perrot, E. R. Norwitz, and S. E. Illanes, “Metabolic pathways involved in 2-methoxyestradiol synthesis and their role in preeclampsia,” *Reproductive Sciences*, vol. 20, no. 9, pp. 1020–1029, 2013.
- [62] C. Farah, L. Y. M. Michel, and J. L. Balligand, “Nitric oxide signalling in cardiovascular health and disease,” *Nature Reviews Cardiology*, vol. 15, no. 5, pp. 292–316, 2018.
- [63] M. A. Hill, Y. Yang, S. R. Ella, M. J. Davis, and A. P. Braun, “Large conductance,  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channels ( $\text{BK}_{\text{Ca}}$ ) and arteriolar myogenic signaling,” *FEBS Letters*, vol. 584, no. 10, pp. 2033–2042, 2010.
- [64] X. Q. Hu and L. Zhang, “Function and regulation of large conductance  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channel in vascular smooth muscle cells,” *Drug Discovery Today*, vol. 17, no. 17-18, pp. 974–987, 2012.
- [65] C. R. Rosenfeld, X. T. Liu, and K. DeSpain, “Pregnancy modifies the large conductance  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channel and cGMP-dependent signaling pathway in uterine vascular smooth muscle,” *American Journal of Physiology. Heart and Circulatory Physiology*, vol. 296, no. 6, pp. H1878–H1887, 2009.
- [66] J. J. Corcoran, C. Nicholson, M. Sweeney et al., “Human uterine and placental arteries exhibit tissue-specific acute responses to  $17\beta$ -estradiol and estrogen-receptor-specific agonists,” *Molecular Human Reproduction*, vol. 20, no. 5, pp. 433–441, 2014.
- [67] C. R. Rosenfeld, R. E. White, T. Roy, and B. E. Cox, “Calcium-activated potassium channels and nitric oxide coregulate estrogen-induced vasodilation,” *American Journal of Physiology. Heart and Circulatory Physiology*, vol. 279, no. 1, pp. H319–H328, 2000.
- [68] M. B. Pastore, S. Talwar, M. R. Conley, and R. R. Magness, “Identification of differential ER-alpha versus ER-beta mediated activation of eNOS in ovine uterine artery endothelial cells,” *Biology of Reproduction*, vol. 94, no. 6, p. 139, 2016.
- [69] M. A. Valverde, P. Rojas, J. Amigo et al., “Acute activation of maxi-K channels (*hSlo*) by estradiol binding to the  $\beta$  subunit,” *Science*, vol. 285, no. 5435, pp. 1929–1931, 1999.
- [70] S. H. Nelson, O. S. Steinsland, Y. Wang, C. Yallampalli, Y. L. Dong, and J. M. Sanchez, “Increased nitric oxide synthase activity and expression in the human uterine artery during pregnancy,” *Circulation Research*, vol. 87, no. 5, pp. 406–411, 2000.
- [71] R. R. Magness, J. A. Sullivan, Y. Li, T. M. Phernetton, and I. M. Bird, “Endothelial vasodilator production by uterine and systemic arteries. VI. Ovarian and pregnancy effects on eNOS and NOx,” *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 280, no. 4, pp. H1692–H1698, 2001.
- [72] K. E. Vagnoni, C. E. Shaw, T. M. Phernetton, B. M. Meglin, I. M. Bird, and R. R. Magness, “Endothelial vasodilator production by uterine and systemic arteries. III. Ovarian and estrogen effects on NO synthase,” *The American Journal of Physiology*, vol. 275, 5 Part 2, pp. H1845–H1856, 1998.
- [73] X. Q. Hu, D. Xiao, R. Zhu et al., “Pregnancy upregulates large-conductance  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channel activity and attenuates myogenic tone in uterine arteries,” *Hypertension*, vol. 58, no. 6, pp. 1132–1139, 2011.
- [74] L. H. Khan, C. R. Rosenfeld, X. T. Liu, and R. R. Magness, “Regulation of the cGMP-cPKG pathway and large-conductance  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channels in uterine arteries during the ovine ovarian cycle,” *American Journal of Physiology. Endocrinology and Metabolism*, vol. 298, no. 2, pp. E222–E228, 2010.
- [75] D. Nagar, X. T. Liu, and C. R. Rosenfeld, “Estrogen regulates  $\beta_1$ -subunit expression in  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channels in arteries from reproductive tissues,” *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 289, no. 4, pp. H1417–H1427, 2005.
- [76] H. L. Rupnow, T. M. Phernetton, C. E. Shaw, M. L. Modrick, I. M. Bird, and R. R. Magness, “Endothelial vasodilator production by uterine and systemic arteries. VII. Estrogen and progesterone effects on eNOS,” *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 280, no. 4, pp. H1699–H1705, 2001.
- [77] C. R. Rosenfeld, B. E. Cox, T. Roy, and R. R. Magness, “Nitric oxide contributes to estrogen-induced vasodilation of the ovine uterine circulation,” *The Journal of Clinical Investigation*, vol. 98, no. 9, pp. 2158–2166, 1996.
- [78] C. R. Rosenfeld, T. Roy, K. DeSpain, and B. E. Cox, “Large-conductance  $\text{Ca}^{2+}$ -dependent  $\text{K}^+$  channels regulate basal uteroplacental blood flow in ovine pregnancy,” *Journal of the Society for Gynecologic Investigation*, vol. 12, no. 6, pp. 402–408, 2005.
- [79] K. R. Kublickiene, M. Kublickas, B. Lindblom, N. O. Lunell, and H. Nisell, “A comparison of myogenic and endothelial properties of myometrial and omental resistance vessels in

- late pregnancy," *American Journal of Obstetrics and Gynecology*, vol. 176, no. 3, pp. 560–566, 1997.
- [80] N. I. Gokina, O. Y. Kuzina, R. Fuller, and G. Osol, "Local uteroplacental influences are responsible for the induction of uterine artery myogenic tone during rat pregnancy," *Reproductive Sciences*, vol. 16, no. 11, pp. 1072–1081, 2009.
- [81] D. Xiao, W. J. Pearce, and L. Zhang, "Pregnancy enhances endothelium-dependent relaxation of ovine uterine artery: role of NO and intracellular  $Ca^{2+}$ ," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 281, no. 1, pp. H183–H190, 2001.
- [82] C. L. M. Cooke and S. T. Davidge, "Pregnancy-induced alterations of vascular function in mouse mesenteric and uterine arteries," *Biology of Reproduction*, vol. 68, no. 3, pp. 1072–1077, 2003.
- [83] S. A. Hale, L. Weger, M. Mandala, and G. Osol, "Reduced NO signaling during pregnancy attenuates outward uterine artery remodeling by altering MMP expression and collagen and elastin deposition," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 301, no. 4, pp. H1266–H1275, 2011.
- [84] C. R. Rosenfeld, L. S. Hyman, X. T. Liu, and T. Roy, "Large conductance  $Ca^{2+}$ -activated  $K^+$  channels modulate uterine  $\alpha_1$ -adrenergic sensitivity in ovine pregnancy," *Reproductive Sciences*, vol. 21, no. 4, pp. 456–464, 2014.
- [85] K. P. Conrad and A. K. Davis, "Nitric oxide synthase activity in placentae from women with pre-eclampsia," *Placenta*, vol. 16, no. 8, pp. 691–699, 1995.
- [86] M. Noris, M. Todeschini, P. Cassis et al., "L-arginine depletion in preeclampsia orients nitric oxide synthase toward oxidant species," *Hypertension*, vol. 43, no. 3, pp. 614–622, 2004.
- [87] S. P. Brennecke, N. M. Gude, J. L. Di Iulio, and R. G. King, "Reduction of placental nitric oxide synthase activity in pre-eclampsia," *Clinical Science*, vol. 93, no. 1, pp. 51–55, 1997.
- [88] L. Myatt, A. L. Eis, D. E. Brockman, I. A. Greer, and F. Lyall, "Endothelial nitric oxide synthase in placental villous tissue from normal, pre-eclamptic and intrauterine growth restricted pregnancies," *Human Reproduction*, vol. 12, no. 1, pp. 167–172, 1997.
- [89] B. J. Krause, I. Carrasco-Wong, A. Caniuguir, J. Carvajal, M. Faras, and P. Casanello, "Endothelial eNOS/arginase imbalance contributes to vascular dysfunction in IUGR umbilical and placental vessels," *Placenta*, vol. 34, no. 1, pp. 20–28, 2013.
- [90] F. F. Li, M. Z. He, Y. Xie et al., "Involvement of dysregulated  $IK_{Ca}$  and  $SK_{Ca}$  channels in preeclampsia," *Placenta*, vol. 58, pp. 9–16, 2017.
- [91] D. Xiao, I. M. Bird, R. R. Magness, L. D. Longo, and L. Zhang, "Upregulation of eNOS in pregnant ovine uterine arteries by chronic hypoxia," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 280, no. 2, pp. H812–H820, 2001.
- [92] Y. J. Kim, H. S. Park, H. Y. Lee et al., "Reduced L-arginine level and decreased placental eNOS activity in preeclampsia," *Placenta*, vol. 27, no. 4-5, pp. 438–444, 2006.
- [93] S. Sankaralingam, H. Xu, and S. T. Davidge, "Arginase contributes to endothelial cell oxidative stress in response to plasma from women with preeclampsia," *Cardiovascular Research*, vol. 85, no. 1, pp. 194–203, 2010.
- [94] L. Myatt, R. B. Rosenfield, A. L. W. Eis, D. E. Brockman, I. Greer, and F. Lyall, "Nitrotyrosine residues in placenta: evidence of peroxynitrite formation and action," *Hypertension*, vol. 28, no. 3, pp. 488–493, 1996.
- [95] S. Zamudio, O. Kovalenko, J. Vanderlelie et al., "Chronic hypoxia in vivo reduces placental oxidative stress," *Placenta*, vol. 28, no. 8-9, pp. 846–853, 2007.
- [96] M. D. Savvidou, A. D. Hingorani, D. Tsikas, J. C. Frolich, P. Vallance, and K. H. Nicolaidis, "Endothelial dysfunction and raised plasma concentrations of asymmetric dimethylarginine in pregnant women who subsequently develop pre-eclampsia," *Lancet*, vol. 361, no. 9368, pp. 1511–1517, 2003.
- [97] M. Laskowska, K. Laskowska, B. Leszczyńska-Gorzela, and J. Oleszczuk, "Asymmetric dimethylarginine in normotensive pregnant women with isolated fetal intrauterine growth restriction: a comparison with preeclamptic women with and without intrauterine growth restriction," *The Journal of Maternal-Fetal & Neonatal Medicine*, vol. 24, no. 7, pp. 936–942, 2011.
- [98] D. Schneider, C. Hernandez, M. Farias, R. Uauy, B. J. Krause, and P. Casanello, "Oxidative stress as common trait of endothelial dysfunction in chorionic arteries from fetuses with IUGR and LGA," *Placenta*, vol. 36, no. 5, pp. 552–558, 2015.
- [99] A. L. Diket, M. R. Pierce, U. K. Munshi et al., "Nitric oxide inhibition causes intrauterine growth retardation and hind-limb disruptions in rats," *American Journal of Obstetrics and Gynecology*, vol. 171, no. 5, pp. 1243–1250, 1994.
- [100] L. C. Kusinski, J. L. Stanley, M. R. Dilworth et al., "eNOS knockout mouse as a model of fetal growth restriction with an impaired uterine artery function and placental transport phenotype," *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, vol. 303, no. 1, pp. R86–R93, 2012.
- [101] S. Kulandavelu, K. J. Whiteley, D. Qu, J. Mu, S. A. Bainbridge, and S. L. Adamson, "Endothelial nitric oxide synthase deficiency reduces uterine blood flow, spiral artery elongation, and placental oxygenation in pregnant mice," *Hypertension*, vol. 60, no. 1, pp. 231–238, 2012.
- [102] M. He, F. Li, M. Yang et al., "Impairment of  $BK_{Ca}$  channels in human placental chorionic plate arteries is potentially relevant to the development of preeclampsia," *Hypertension Research*, vol. 41, no. 2, pp. 126–134, 2018.
- [103] X. Q. Hu, D. Xiao, R. Zhu et al., "Chronic hypoxia suppresses pregnancy-induced upregulation of large-conductance  $Ca^{2+}$ -activated  $K^+$  channel activity in uterine arteries," *Hypertension*, vol. 60, no. 1, pp. 214–222, 2012.
- [104] X. Q. Hu, X. Huang, D. Xiao, and L. Zhang, "Direct effect of chronic hypoxia in suppressing large conductance  $Ca^{2+}$ -activated  $K^+$  channel activity in ovine uterine arteries via increasing oxidative stress," *The Journal of Physiology*, vol. 594, no. 2, pp. 343–356, 2016.
- [105] P. Wang, S. Zhang, J. Ren et al., "The inhibitory effect of  $BK_{Ca}$  channels induced by autoantibodies against angiotensin II type 1 receptor is independent of AT1R," *Acta Biochimica et Biophysica Sinica*, vol. 50, no. 6, pp. 560–566, 2018.
- [106] M. Feletou and P. M. Vanhoutte, "EDHF: an update," *Clinical Science*, vol. 117, no. 4, pp. 139–155, 2009.
- [107] R. Zhu, X. Q. Hu, D. Xiao et al., "Chronic hypoxia inhibits pregnancy-induced upregulation of  $SK_{Ca}$  channel expression and function in uterine arteries," *Hypertension*, vol. 62, no. 2, pp. 367–374, 2013.

- [108] V. Chinnathambi, C. S. Blesson, K. L. Vincent et al., "Elevated testosterone levels during rat pregnancy cause hypersensitivity to angiotensin II and attenuation of endothelium-dependent vasodilation in uterine arteries," *Hypertension*, vol. 64, no. 2, pp. 405–414, 2014.
- [109] R. H. P. Hilgers, S. Oparil, W. Wouters, and H. J. T. Coelingh Bennink, "Vasorelaxing effects of estetrol in rat arteries," *Journal of Endocrinology*, vol. 215, no. 1, pp. 97–106, 2012.
- [110] C. C. Rada, G. Murray, and S. K. England, "The SK3 channel promotes placental vascularization by enhancing secretion of angiogenic factors," *American Journal of Physiology. Endocrinology and Metabolism*, vol. 307, no. 10, pp. E935–E943, 2014.
- [111] C. C. Rada, S. L. Pierce, D. W. Nuno et al., "Overexpression of the SK3 channel alters vascular remodeling during pregnancy, leading to fetal demise," *American Journal of Physiology. Endocrinology and Metabolism*, vol. 303, no. 7, pp. E825–E831, 2012.
- [112] F. C. Yap, M. S. Taylor, and M. T. Lin, "Ovariectomy-induced reductions in endothelial SK3 channel activity and endothelium-dependent vasorelaxation in murine mesenteric arteries," *PLoS One*, vol. 9, no. 8, article e104686, 2014.
- [113] J. Chen, Q. Gao, L. Jiang et al., "The NOX2-derived reactive oxygen species damaged endothelial nitric oxide system via suppressed BK<sub>Ca</sub>/SK<sub>Ca</sub> in preeclampsia," *Hypertension Research*, vol. 40, no. 5, pp. 457–464, 2017.
- [114] D. I. Brown and K. K. Griendling, "Regulation of signal transduction by reactive oxygen species in the cardiovascular system," *Circulation Research*, vol. 116, no. 3, pp. 531–549, 2015.
- [115] A. C. Montezano and R. M. Touyz, "Reactive oxygen species and endothelial function—role of nitric oxide synthase uncoupling and Nox family nicotinamide adenine dinucleotide phosphate oxidases," *Basic & Clinical Pharmacology & Toxicology*, vol. 110, no. 1, pp. 87–94, 2012.
- [116] L. He, T. He, S. Farrar, L. Ji, T. Liu, and X. Ma, "Antioxidants maintain cellular redox homeostasis by elimination of reactive oxygen species," *Cellular Physiology and Biochemistry*, vol. 44, no. 2, pp. 532–553, 2017.
- [117] R. J. Reiter, J. C. Mayo, D. X. Tan, R. M. Sainz, M. Alatorre-Jimenez, and L. Qin, "Melatonin as an antioxidant: under promises but over delivers," *Journal of Pineal Research*, vol. 61, no. 3, pp. 253–278, 2016.
- [118] V. I. Lushchak, "Glutathione homeostasis and functions: potential targets for medical interventions," *Journal of Amino Acids*, vol. 2012, Article ID 736837, 26 pages, 2012.
- [119] A. B. Fisher, "Redox signaling across cell membranes," *Antioxidants & Redox Signaling*, vol. 11, no. 6, pp. 1349–1356, 2009.
- [120] L. Myatt and X. Cui, "Oxidative stress in the placenta," *Histochemistry and Cell Biology*, vol. 122, no. 4, pp. 369–382, 2004.
- [121] R. D. Pereira, N. E. de Long, R. C. Wang, F. T. Yazdi, A. C. Holloway, and S. Raha, "Angiogenesis in the placenta: the role of reactive oxygen species signaling," *BioMed Research International*, vol. 2015, Article ID 814543, 12 pages, 2015.
- [122] O. Ishihara, M. Hayashi, H. Osawa et al., "Isoprostanes, prostaglandins and tocopherols in pre-eclampsia, normal pregnancy and non-pregnancy," *Free Radical Research*, vol. 38, no. 9, pp. 913–918, 2004.
- [123] G. Kaur, S. Mishra, A. Sehgal, and R. Prasad, "Alterations in lipid peroxidation and antioxidant status in pregnancy with preeclampsia," *Molecular and Cellular Biochemistry*, vol. 313, no. 1–2, pp. 37–44, 2008.
- [124] D. Mannaerts, E. Faes, P. Cos et al., "Oxidative stress in healthy pregnancy and preeclampsia is linked to chronic inflammation, iron status and vascular function," *PLoS One*, vol. 13, no. 9, article e0202919, 2018.
- [125] M. Mihailovič, M. Cvetkovič, A. Ljubič et al., "Selenium and malondialdehyde content and glutathione peroxidase activity in maternal and umbilical cord blood and amniotic fluid," *Biological Trace Element Research*, vol. 73, no. 1, pp. 47–54, 2000.
- [126] M. T. Raijmakers, P. L. Zusterzeel, E. A. Steegers, M. P. Hectors, P. N. Demacker, and W. H. Peters, "Plasma thiol status in preeclampsia," *Obstetrics and Gynecology*, vol. 95, no. 2, pp. 180–184, 2000.
- [127] M. M. Aljunaidy, J. S. Morton, C. L. M. Cooke, and S. T. Davidge, "Prenatal hypoxia and placental oxidative stress: linkages to developmental origins of cardiovascular disease," *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, vol. 313, no. 4, pp. R395–R399, 2017.
- [128] F. Wu, F. J. Tian, and Y. Lin, "Oxidative stress in placenta: health and diseases," *BioMed Research International*, vol. 2015, Article ID 293271, 15 pages, 2015.
- [129] Y. Takehara, T. Yoshioka, and J. Sasaki, "Changes in the levels of lipoperoxide and antioxidant factors in human placenta during gestation," *Acta Medica Okayama*, vol. 44, no. 2, pp. 103–111, 1990.
- [130] J. Basu, B. Bendek, E. Agamasu et al., "Placental oxidative status throughout normal gestation in women with uncomplicated pregnancies," *Obstetrics and Gynecology International*, vol. 2015, Article ID 276095, 6 pages, 2015.
- [131] T. McGarry, M. Biniecka, D. J. Veale, and U. Fearon, "Hypoxia, oxidative stress and inflammation," *Free Radical Biology & Medicine*, vol. 125, pp. 15–24, 2018.
- [132] R. D. Guzy, B. Hoyos, E. Robin et al., "Mitochondrial complex III is required for hypoxia-induced ROS production and cellular oxygen sensing," *Cell Metabolism*, vol. 1, no. 6, pp. 401–408, 2005.
- [133] G. B. Waypa, R. Guzy, P. T. Mungai et al., "Increases in mitochondrial reactive oxygen species trigger hypoxia-induced calcium responses in pulmonary artery smooth muscle cells," *Circulation Research*, vol. 99, no. 9, pp. 970–978, 2006.
- [134] J. Q. Liu, I. N. Zelko, E. M. Erbynn, J. S. K. Sham, and R. J. Folz, "Hypoxic pulmonary hypertension: role of superoxide and NADPH oxidase (gp91<sup>phox</sup>)," *American Journal of Physiology-Lung Cellular and Molecular Physiology*, vol. 290, no. 1, pp. L2–10, 2006.
- [135] A. Many, C. A. Hubel, S. J. Fisher, J. M. Roberts, and Y. Zhou, "Invasive cytotrophoblasts manifest evidence of oxidative stress in preeclampsia," *The American Journal of Pathology*, vol. 156, no. 1, pp. 321–331, 2000.
- [136] X. L. Cui, D. Brockman, B. Campos, and L. Myatt, "Expression of NADPH oxidase isoform 1 (Nox1) in human placenta: involvement in preeclampsia," *Placenta*, vol. 27, no. 4–5, pp. 422–431, 2006.
- [137] A. Biri, N. Bozkurt, A. Turp, M. Kavutcu, O. Himmetoglu, and I. Durak, "Role of oxidative stress in intrauterine growth restriction," *Gynecologic and Obstetric Investigation*, vol. 64, no. 4, pp. 187–192, 2007.
- [138] Y. Wang and S. W. Walsh, "Antioxidant activities and mRNA expression of superoxide dismutase, catalase, and glutathione

- peroxidase in normal and preeclamptic placentas," *Journal of the Society for Gynecologic Investigation*, vol. 3, no. 4, pp. 179–184, 2016.
- [139] U. Mutlu-Turkoglu, E. Ademoglu, L. Ibrahimoglu, G. Aykac-Toker, and M. Uysal, "Imbalance between lipid peroxidation and antioxidant status in preeclampsia," *Gynecologic and Obstetric Investigation*, vol. 46, no. 1, pp. 37–40, 1998.
- [140] S. Muralimanoharan, A. Maloyan, J. Mele, C. Guo, L. G. Myatt, and L. Myatt, "MIR-210 modulates mitochondrial respiration in placenta with preeclampsia," *Placenta*, vol. 33, no. 10, pp. 816–823, 2012.
- [141] S. Matsubara, H. Minakami, I. Sato, and T. Saito, "Decrease in cytochrome c oxidase activity detected cytochemically in the placental trophoblast of patients with pre-eclampsia," *Placenta*, vol. 18, no. 4, pp. 255–259, 1997.
- [142] F. Colleon, N. Padmanabhan, H. W. Yung et al., "Suppression of mitochondrial electron transport chain function in the hypoxic human placenta: a role for miRNA-210 and protein synthesis inhibition," *PLoS One*, vol. 8, no. 1, article e55194, 2013.
- [143] D. Akyol, T. Mungan, H. Gorkemli, and G. Nuhoglu, "Maternal levels of vitamin E in normal and preeclamptic pregnancy," *Archives of Gynecology and Obstetrics*, vol. 263, no. 4, pp. 151–155, 2000.
- [144] E. Llubra, E. Gratacos, P. Martin-Gallan, L. Cabero, and C. Dominguez, "A comprehensive study of oxidative stress and antioxidant status in preeclampsia and normal pregnancy," *Free Radical Biology & Medicine*, vol. 37, no. 4, pp. 557–570, 2004.
- [145] Y. Wang and S. W. Walsh, "Increased superoxide generation is associated with decreased superoxide dismutase activity and mRNA expression in placental trophoblast cells in preeclampsia," *Placenta*, vol. 22, no. 2-3, pp. 206–212, 2001.
- [146] A. Aris, S. Benali, A. Ouellet, J. M. Moutquin, and S. Leblanc, "Potential biomarkers of preeclampsia: inverse correlation between hydrogen peroxide and nitric oxide early in maternal circulation and at term in placenta of women with preeclampsia," *Placenta*, vol. 30, no. 4, pp. 342–347, 2009.
- [147] S. W. Walsh, J. E. Vaughan, Y. Wang, and L. J. Roberts II, "Placental isoprostane is significantly increased in preeclampsia," *The FASEB Journal*, vol. 14, no. 10, pp. 1289–1296, 2000.
- [148] Y. Takagi, T. Nikaido, T. Toki et al., "Levels of oxidative stress and redox-related molecules in the placenta in preeclampsia and fetal growth restriction," *Virchows Archiv*, vol. 444, no. 1, pp. 49–55, 2004.
- [149] C. Kimura, K. Watanabe, A. Iwasaki et al., "The severity of hypoxic changes and oxidative DNA damage in the placenta of early-onset preeclamptic women and fetal growth restriction," *The Journal of Maternal-Fetal & Neonatal Medicine*, vol. 26, no. 5, pp. 491–496, 2013.
- [150] A. M. Roggensack, Y. Zhang, and S. T. Davidge, "Evidence for peroxynitrite formation in the vasculature of women with preeclampsia," *Hypertension*, vol. 33, no. 1, pp. 83–89, 1999.
- [151] J. L. Stanley, I. J. Andersson, C. J. Hirt et al., "Effect of the anti-oxidant tempol on fetal growth in a mouse model of fetal growth restriction1," *Biology of Reproduction*, vol. 87, no. 1, pp. 25, 1–25, 8, 2012.
- [152] M. Sedeek, J. S. Gilbert, B. B. LaMarca et al., "Role of reactive oxygen species in hypertension produced by reduced uterine perfusion in pregnant rats," *American Journal of Hypertension*, vol. 21, no. 10, pp. 1152–1156, 2008.
- [153] O. Balta, A. Boztosun, K. Deveci et al., "Reduced uterine perfusion pressure model is not successful to mimic severe preeclampsia," *Placenta*, vol. 32, no. 9, pp. 675–680, 2011.
- [154] V. R. Vaka, K. M. McMaster, M. W. Cunningham Jr. et al., "Role of mitochondrial dysfunction and reactive oxygen species in mediating hypertension in the reduced uterine perfusion pressure rat model of preeclampsia," *Hypertension*, vol. 72, no. 3, pp. 703–711, 2018.
- [155] B. M. Mitchell, L. G. Cook, S. Danchuk, and J. B. Puschett, "Uncoupled endothelial nitric oxide synthase and oxidative stress in a rat model of pregnancy-induced hypertension," *American Journal of Hypertension*, vol. 20, no. 12, pp. 1297–1304, 2007.
- [156] A. Beausejour, K. Bibeau, J. C. Lavoie, J. St-Louis, and M. Brochu, "Placental oxidative stress in a rat model of preeclampsia," *Placenta*, vol. 28, no. 1, pp. 52–58, 2007.
- [157] D. Xiao, X. Q. Hu, X. Huang et al., "Chronic hypoxia during gestation enhances uterine arterial myogenic tone via heightened oxidative stress," *PLoS One*, vol. 8, no. 9, article e73731, 2013.
- [158] H. G. Richter, E. J. Camm, B. N. Modi et al., "Ascorbate prevents placental oxidative stress and enhances birth weight in hypoxic pregnancy in rats," *The Journal of Physiology*, vol. 590, no. 6, pp. 1377–1387, 2012.
- [159] A. K. Agarwal and R. J. Auchus, "Minireview: cellular redox state regulates hydroxysteroid dehydrogenase activity and intracellular hormone potency," *Endocrinology*, vol. 146, no. 6, pp. 2531–2538, 2005.
- [160] Y. Hong, H. Li, Y. C. Yuan, and S. Chen, "Molecular characterization of aromatase," *Annals of the New York Academy of Sciences*, vol. 1155, no. 1, pp. 112–120, 2009.
- [161] Q. Hou, M. Lei, K. Hu, and M. Wang, "The effects of high glucose levels on reactive oxygen species-induced apoptosis and involved signaling in human vascular endothelial cells," *Cardiovascular Toxicology*, vol. 15, no. 2, pp. 140–146, 2015.
- [162] Y. Peristiwati, I. Indasah, and R. Ratnawati, "The effects of catechin isolated from green tea GMB-4 on NADPH and nitric oxide levels in endothelial cells exposed to high glucose," *Journal of Intercultural Ethnopharmacology*, vol. 4, no. 2, pp. 114–117, 2015.
- [163] D. P. Sherbet, O. L. Guryev, M. Papari-Zareei et al., "Biochemical factors governing the steady-state estrone/estradiol ratios catalyzed by human 17beta-hydroxysteroid dehydrogenases types 1 and 2 in HEK-293 cells," *Endocrinology*, vol. 150, no. 9, pp. 4154–4162, 2009.
- [164] J. Wan, Z. Hu, K. Zeng et al., "The reduction in circulating levels of estrogen and progesterone in women with preeclampsia," *Pregnancy Hypertension*, vol. 11, pp. 18–25, 2018.
- [165] Y. Okatani, N. Morioka, A. Wakatsuki, Y. Nakano, and Y. Sagara, "Role of the free radical-scavenger system in aromatase activity of the human ovary," *Hormone Research*, vol. 39, no. 1, pp. 22–27, 1993.
- [166] G. E. Weitsman, W. Weebadda, K. Ung, and L. C. Murphy, "Reactive oxygen species induce phosphorylation of serine 118 and 167 on estrogen receptor alpha," *Breast Cancer Research and Treatment*, vol. 118, no. 2, pp. 269–279, 2009.
- [167] P. C. O'Leary, M. Terrile, M. Bajor et al., "Peroxiredoxin-1 protects estrogen receptor  $\alpha$  from oxidative stress-induced suppression and is a protein biomarker of favorable



- prognosis in breast cancer,” *Breast Cancer Research*, vol. 16, no. 4, p. R79, 2014.
- [168] C. Polytaichou and E. Papadimitriou, “Antioxidants inhibit human endothelial cell functions through down-regulation of endothelial nitric oxide synthase activity,” *European Journal of Pharmacology*, vol. 510, no. 1-2, pp. 31–38, 2005.
- [169] E. A. Jaimes, C. Sweeney, and L. Raij, “Effects of the reactive oxygen species hydrogen peroxide and hypochlorite on endothelial nitric oxide production,” *Hypertension*, vol. 38, no. 4, pp. 877–883, 2001.
- [170] S. Choi, J. Kim, J. H. Kim et al., “Carbon monoxide prevents TNF- $\alpha$ -induced eNOS downregulation by inhibiting NF- $\kappa$ B-responsive miR-155-5p biogenesis,” *Experimental & Molecular Medicine*, vol. 49, no. 11, p. e403, 2017.
- [171] S. Choi, S. Park, G. Liang, J. Kim, and S. Suh, “Superoxide generated by lysophosphatidylcholine induces endothelial nitric oxide synthase downregulation in human endothelial cells,” *Cellular Physiology and Biochemistry*, vol. 25, no. 2-3, pp. 233–240, 2010.
- [172] A. A. Eid, D. Y. Lee, L. J. Roman, K. Khazim, and Y. Gorin, “Sestrin 2 and AMPK connect hyperglycemia to Nox4-dependent endothelial nitric oxide synthase uncoupling and matrix protein expression,” *Molecular and Cellular Biology*, vol. 33, no. 17, pp. 3439–3460, 2013.
- [173] D. Y. Lee, F. Wauquier, A. A. Eid et al., “Nox4 NADPH oxidase mediates peroxynitrite-dependent uncoupling of endothelial nitric-oxide synthase and fibronectin expression in response to angiotensin II: role of mitochondrial reactive oxygen species,” *The Journal of Biological Chemistry*, vol. 288, no. 40, pp. 28668–28686, 2013.
- [174] S. Srinivasan, M. E. Hatley, D. T. Bolick et al., “Hyperglycemia-induced superoxide production decreases eNOS expression via AP-1 activation in aortic endothelial cells,” *Diabetologia*, vol. 47, no. 10, pp. 1727–1734, 2004.
- [175] X. J. Zhou, N. D. Vaziri, X. Q. Wang, F. G. Silva, and Z. Laszik, “Nitric oxide synthase expression in hypertension induced by inhibition of glutathione synthase,” *Journal of Pharmacology and Experimental Therapeutics*, vol. 300, no. 3, pp. 762–767, 2002.
- [176] R. S. Barlow and R. E. White, “Hydrogen peroxide relaxes porcine coronary arteries by stimulating BK<sub>Ca</sub> channel activity,” *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 275, no. 4, pp. H1283–H1289, 1998.
- [177] Y. Hayabuchi, Y. Nakaya, S. Matsuoka, and Y. Kuroda, “Hydrogen peroxide-induced vascular relaxation in porcine coronary arteries is mediated by Ca<sup>2+</sup>-activated K<sup>+</sup> channels,” *Heart and Vessels*, vol. 13, no. 1, pp. 9–17, 1998.
- [178] D. L. Dong, P. Yue, B. F. Yang, and W. H. Wang, “Hydrogen peroxide stimulates the Ca<sup>2+</sup>-activated Big-conductance K channels (BK) through cGMP signaling pathway in cultured human endothelial cells,” *Cellular Physiology and Biochemistry*, vol. 22, no. 1-4, pp. 119–126, 2008.
- [179] S. Brakemeier, I. Eichler, A. Knorr, T. Fassheber, R. Kohler, and J. Hoyer, “Modulation of Ca<sup>2+</sup>-activated K<sup>+</sup> channel in renal artery endothelium in situ by nitric oxide and reactive oxygen species,” *Kidney International*, vol. 64, no. 1, pp. 199–207, 2003.
- [180] X. D. Tang, H. Daggett, M. Hanner et al., “Oxidative regulation of large conductance calcium-activated potassium channels,” *The Journal of General Physiology*, vol. 117, no. 3, pp. 253–274, 2001.
- [181] X. D. Tang, M. L. Garcia, S. H. Heinemann, and T. Hoshi, “Reactive oxygen species impair Slo1 BK channel function by altering cysteine-mediated calcium sensing,” *Nature Structural & Molecular Biology*, vol. 11, no. 2, pp. 171–178, 2004.
- [182] Y. Liu, K. Terata, Q. Chai, H. Li, L. H. Kleinman, and D. D. Gutterman, “Peroxynitrite inhibits Ca<sup>2+</sup>-activated K<sup>+</sup> channel activity in smooth muscle of human coronary arterioles,” *Circulation Research*, vol. 91, no. 11, pp. 1070–1076, 2002.
- [183] A. K. Brzezinska, D. Gebremedhin, W. M. Chilian, B. Kalyanaraman, and S. J. Elliott, “Peroxynitrite reversibly inhibits Ca<sup>2+</sup>-activated K<sup>+</sup> channels in rat cerebral artery smooth muscle cells,” *American Journal of Physiology. Heart and Circulatory Physiology*, vol. 278, no. 6, pp. H1883–H1890, 2000.
- [184] R. Zhu, X. Huang, X. Q. Hu, D. L. Xiao, and L. Zhang, “Gestational hypoxia increases reactive oxygen species and inhibits steroid hormone-mediated upregulation of Ca<sup>2+</sup>-activated K<sup>+</sup> channel function in uterine arteries,” *Hypertension*, vol. 64, no. 2, pp. 415–422, 2014.
- [185] S. Choi, H. Y. Na, J. A. Kim, S. E. Cho, and S. H. Suh, “Contradictory effects of superoxide and hydrogen peroxide on K<sub>Ca</sub>3.1 in human endothelial cells,” *The Korean Journal of Physiology & Pharmacology*, vol. 17, no. 3, pp. 181–187, 2013.
- [186] T. Lu, Q. Chai, L. Yu et al., “Reactive oxygen species signaling facilitates FOXO-3a/FBXO-dependent vascular BK channel  $\beta$ 1 subunit degradation in diabetic mice,” *Diabetes*, vol. 61, no. 7, pp. 1860–1868, 2012.
- [187] S. Choi, J. A. Kim, H. Y. Li et al., “Altered redox state modulates endothelial K<sub>Ca</sub>2.3 and K<sub>Ca</sub>3.1 levels in normal pregnancy and preeclampsia,” *Antioxidants & Redox Signaling*, vol. 30, no. 4, pp. 505–519, 2019.
- [188] S. Choi, J. A. Kim, H. Y. Na et al., “NADPH oxidase 2-derived superoxide downregulates endothelial K<sub>Ca</sub>3.1 in preeclampsia,” *Free Radical Biology & Medicine*, vol. 57, pp. 10–21, 2013.
- [189] L. M. Zhao, Y. Wang, Y. Yang, R. Guo, N. P. Wang, and X. L. Deng, “Metformin restores intermediate-conductance calcium-activated K<sup>+</sup> channel- and small-conductance calcium-activated K<sup>+</sup> channel-mediated vasodilatation impaired by advanced glycation end products in rat mesenteric artery,” *Molecular Pharmacology*, vol. 86, no. 5, pp. 580–591, 2014.
- [190] H. Y. Zoghbi and A. L. Beaudet, “Epigenetics and human disease,” *Cold Spring Harbor Perspectives in Biology*, vol. 8, no. 2, article a019497, 2016.
- [191] N. S. Chandel, D. S. McClintock, C. E. Feliciano et al., “Reactive oxygen species generated at mitochondrial complex III stabilize hypoxia-inducible factor-1 $\alpha$  during hypoxia: a mechanism of O<sub>2</sub> sensing,” *Journal of Biological Chemistry*, vol. 275, no. 33, pp. 25130–25138, 2000.
- [192] S. Kaewpila, S. Venkataraman, G. R. Buettner, and L. W. Oberley, “Manganese superoxide dismutase modulates hypoxia-inducible factor-1 $\alpha$  induction via superoxide,” *Cancer Research*, vol. 68, no. 8, pp. 2781–2788, 2008.
- [193] A. Sanjuan-Pla, A. M. Cervera, N. Apostolova et al., “A targeted antioxidant reveals the importance of mitochondrial reactive oxygen species in the hypoxic signaling of HIF-1 $\alpha$ ,” *FEBS Letters*, vol. 579, no. 12, pp. 2669–2674, 2005.
- [194] K. D. Mansfield, R. D. Guzy, Y. Pan et al., “Mitochondrial dysfunction resulting from loss of cytochrome c impairs cellular oxygen sensing and hypoxic HIF- $\alpha$  activation,” *Cell Metabolism*, vol. 1, no. 6, pp. 393–399, 2005.

- [195] M. Wang, J. S. Kirk, S. Venkataraman et al., "Manganese superoxide dismutase suppresses hypoxic induction of hypoxia-inducible factor-1 $\alpha$  and vascular endothelial growth factor," *Oncogene*, vol. 24, no. 55, pp. 8154–8166, 2005.
- [196] M. Calvani, G. Comito, E. Giannoni, and P. Chiarugi, "Time-dependent stabilization of hypoxia inducible factor-1 $\alpha$  by different intracellular sources of reactive oxygen species," *PLoS One*, vol. 7, no. 10, article e38388, 2012.
- [197] X. Liu, Y. Deng, J. Shang et al., "Effect of NADPH oxidase inhibitor apocynin on the expression of hypoxia-induced factor-1 $\alpha$  and endothelin-1 in rat carotid body exposed to chronic intermittent hypoxia," *Journal of Huazhong University of Science and Technology[Medical Sciences]*, vol. 33, no. 2, pp. 178–184, 2013.
- [198] J. K. Maranchie and Y. Zhan, "Nox4 is critical for hypoxia-inducible factor 2- $\alpha$  transcriptional activity in von Hippel-Lindau-deficient renal cell carcinoma," *Cancer Research*, vol. 65, no. 20, pp. 9190–9193, 2005.
- [199] I. Diebold, D. Flugel, S. Becht et al., "The hypoxia-inducible factor-2 $\alpha$  is stabilized by oxidative stress involving NOX4," *Antioxidants & Redox Signaling*, vol. 13, no. 4, pp. 425–436, 2010.
- [200] E. L. Bell, T. A. Klimova, J. Eisenbart et al., "The Qo site of the mitochondrial complex III is required for the transduction of hypoxic signaling via reactive oxygen species production," *The Journal of Cell Biology*, vol. 177, no. 6, pp. 1029–1036, 2007.
- [201] C. D. Allis and T. Jenuwein, "The molecular hallmarks of epigenetic control," *Nature Reviews Genetics*, vol. 17, no. 8, pp. 487–500, 2016.
- [202] A. C. E. Campos, F. Molognoni, F. H. M. Melo et al., "Oxidative stress modulates DNA methylation during melanocyte anchorage blockade associated with malignant transformation," *Neoplasia*, vol. 9, no. 12, pp. 1111–1121, 2007.
- [203] S. O. Lim, J. M. Gu, M. S. Kim et al., "Epigenetic changes induced by reactive oxygen species in hepatocellular carcinoma: methylation of the E-cadherin promoter," *Gastroenterology*, vol. 135, no. 6, pp. 2128–2140.e8, 2008.
- [204] P. K. Mahalingaiah, L. Ponnusamy, and K. P. Singh, "Oxidative stress-induced epigenetic changes associated with malignant transformation of human kidney epithelial cells," *Oncotarget*, vol. 8, no. 7, pp. 11127–11143, 2017.
- [205] H. M. O'Hagan, W. Wang, S. Sen et al., "Oxidative damage targets complexes containing DNA methyltransferases, SIRT1, and polycomb members to promoter CpG Islands," *Cancer Cell*, vol. 20, no. 5, pp. 606–619, 2011.
- [206] A. J. Patterson, D. Xiao, F. Xiong, B. Dixon, and L. Zhang, "Hypoxia-derived oxidative stress mediates epigenetic repression of PKC $\epsilon$  gene in foetal rat hearts," *Cardiovascular Research*, vol. 93, no. 2, pp. 302–310, 2012.
- [207] F. Xiong, D. Xiao, and L. Zhang, "Norepinephrine causes epigenetic repression of PKC $\epsilon$  gene in rodent hearts by activating Nox1-dependent reactive oxygen species production," *The FASEB Journal*, vol. 26, no. 7, pp. 2753–2763, 2012.
- [208] Y. Niu, T. L. DesMarais, Z. Tong, Y. Yao, and M. Costa, "Oxidative stress alters global histone modification and DNA methylation," *Free Radical Biology & Medicine*, vol. 82, pp. 22–28, 2015.
- [209] K. Blaschke, K. T. Ebata, M. M. Karimi et al., "Vitamin C induces Tet-dependent DNA demethylation and a blastocyst-like state in ES cells," *Nature*, vol. 500, no. 7461, pp. 222–226, 2013.
- [210] A. Monfort and A. Wutz, "Breathing-in epigenetic change with vitamin C," *EMBO Reports*, vol. 14, no. 4, pp. 337–346, 2013.
- [211] B. E. Lee, Y. C. Hong, K. H. Lee et al., "Influence of maternal serum levels of vitamins C and E during the second trimester on birth weight and length," *European Journal of Clinical Nutrition*, vol. 58, no. 10, pp. 1365–1371, 2004.
- [212] J. H. Kim, S. G. Park, S. Y. Song, J. K. Kim, and J. H. Sung, "Reactive oxygen species-responsive miR-210 regulates proliferation and migration of adipose-derived stem cells via PTPN2," *Cell Death & Disease*, vol. 4, no. 4, article e588, 2013.
- [213] S. L. Archer, G. Marsboom, G. H. Kim et al., "Epigenetic attenuation of mitochondrial superoxide dismutase 2 in pulmonary arterial hypertension: a basis for excessive cell proliferation and a new therapeutic target," *Circulation*, vol. 121, no. 24, pp. 2661–2671, 2010.
- [214] J. Nanduri, V. Makarenko, V. D. Reddy et al., "Epigenetic regulation of hypoxic sensing disrupts cardiorespiratory homeostasis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 109, no. 7, pp. 2515–2520, 2012.
- [215] J. Y. Min, S. O. Lim, and G. Jung, "Downregulation of catalase by reactive oxygen species via hypermethylation of CpG island II on the catalase promoter," *FEBS Letters*, vol. 584, no. 11, pp. 2427–2432, 2010.
- [216] Y. J. Xin, B. Yuan, B. Yu et al., "Tet1-mediated DNA demethylation regulates neuronal cell death induced by oxidative stress," *Scientific Reports*, vol. 5, no. 1, p. 7645, 2015.
- [217] E. Nozik-Grayck, C. Woods, R. S. Stearman et al., "Histone deacetylation contributes to low extracellular superoxide dismutase expression in human idiopathic pulmonary arterial hypertension," *American Journal of Physiology. Lung Cellular and Molecular Physiology*, vol. 311, no. 1, pp. L124–L134, 2016.
- [218] D. Siuda, U. Zechner, N. El Hajj et al., "Transcriptional regulation of Nox4 by histone deacetylases in human endothelial cells," *Basic Research in Cardiology*, vol. 107, no. 5, p. 283, 2012.
- [219] S. Y. Chan, Y. Y. Zhang, C. Hemann, C. E. Mahoney, J. L. Zweier, and J. Loscalzo, "MicroRNA-210 controls mitochondrial metabolism during hypoxia by repressing the iron-sulfur cluster assembly proteins ISCU1/2," *Cell Metabolism*, vol. 10, no. 4, pp. 273–284, 2009.
- [220] C. Dasgupta, M. Chen, H. Zhang, S. Yang, and L. Zhang, "Chronic hypoxia during gestation causes epigenetic repression of the estrogen receptor- $\alpha$  gene in ovine uterine arteries via heightened promoter methylation," *Hypertension*, vol. 60, no. 3, pp. 697–704, 2012.
- [221] M. Chen, C. Dasgupta, F. Xiong, and L. Zhang, "Epigenetic upregulation of large-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channel expression in uterine vascular adaptation to pregnancy," *Hypertension*, vol. 64, no. 3, pp. 610–618, 2014.
- [222] X. Q. Hu, C. Dasgupta, M. Chen et al., "Pregnancy reprograms large-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channel in uterine arteries: roles of ten-eleven translocation methylcytosine dioxygenase 1-mediated active demethylation," *Hypertension*, vol. 69, no. 6, pp. 1181–1191, 2017.
- [223] A. Kulkarni, P. Chavan-Gautam, S. Mehendale, H. Yadav, and S. Joshi, "Global DNA methylation patterns in placenta

- and its association with maternal hypertension in pre-eclampsia,” *DNA and Cell Biology*, vol. 30, no. 2, pp. 79–84, 2011.
- [224] W. L. Gao, D. Li, Z. X. Xiao et al., “Detection of global DNA methylation and paternally imprinted H19 gene methylation in preeclamptic placentas,” *Hypertension Research*, vol. 34, no. 5, pp. 655–661, 2011.
- [225] X. Q. Hu, M. Chen, C. Dasgupta et al., “Chronic hypoxia upregulates DNA methyltransferase and represses large conductance  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channel function in ovine uterine arteries,” *Biology of Reproduction*, vol. 96, no. 2, pp. 424–434, 2017.
- [226] X. Xiao, Y. Zhao, R. Jin et al., “Fetal growth restriction and methylation of growth-related genes in the placenta,” *Epigenomics*, vol. 8, no. 1, pp. 33–42, 2016.
- [227] M. Ma, Q. J. Zhou, Y. Xiong, B. Li, and X. T. Li, “Preeclampsia is associated with hypermethylation of IGF-1 promoter mediated by DNMT1,” *American Journal of Translational Research*, vol. 10, no. 1, pp. 16–39, 2018.
- [228] Y. Tang, H. Liu, H. Li, T. Peng, W. Gu, and X. Li, “Hypermethylation of the HLA-G promoter is associated with preeclampsia,” *Molecular Human Reproduction*, vol. 21, no. 9, pp. 736–744, 2015.
- [229] M. Sun, M. M. Song, B. Wei et al., “5-Hydroxymethylcytosine-mediated alteration of transposon activity associated with the exposure to adverse in utero environments in human,” *Human Molecular Genetics*, vol. 25, no. 11, pp. 2208–2219, 2016.
- [230] X. Li, C. Wu, Y. Shen et al., “Ten-eleven translocation 2 demethylates the *MMP9* promoter, and its down-regulation in preeclampsia impairs trophoblast migration and invasion,” *Journal of Biological Chemistry*, vol. 293, no. 26, pp. 10059–10070, 2018.
- [231] S. Alahari, M. Post, A. Rolfo, R. Weksberg, and I. Caniggia, “Compromised JMJD6 histone demethylase activity affects VHL gene repression in preeclampsia,” *The Journal of Clinical Endocrinology and Metabolism*, vol. 103, no. 4, pp. 1545–1557, 2018.
- [232] D. C. Lee, R. Romero, J. S. Kim et al., “miR-210 targets iron-sulfur cluster scaffold homologue in human trophoblast cell lines: siderosis of interstitial trophoblasts as a novel pathology of preterm preeclampsia and small-for-gestational-age pregnancies,” *The American Journal of Pathology*, vol. 179, no. 2, pp. 590–602, 2011.
- [233] S. Zamudio, Y. Wu, F. Ietta et al., “Human placental hypoxia-inducible factor-1 $\alpha$  expression correlates with clinical outcomes in chronic hypoxia in vivo,” *The American Journal of Pathology*, vol. 170, no. 6, pp. 2171–2179, 2007.
- [234] R. Tal, A. Shaish, I. Barshack et al., “Effects of hypoxia-inducible factor-1 $\alpha$  overexpression in pregnant mice: possible implications for preeclampsia and intrauterine growth restriction,” *The American Journal of Pathology*, vol. 177, no. 6, pp. 2950–2962, 2010.
- [235] X. Q. Hu, C. Dasgupta, J. Xiao, S. Yang, and L. Zhang, “Long-term high altitude hypoxia during gestation suppresses large conductance  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channel function in uterine arteries: a causal role for microRNA-210,” *The Journal of Physiology*, vol. 596, no. 23, pp. 5891–5906, 2018.
- [236] B. Thienpont, J. Steinbacher, H. Zhao et al., “Tumour hypoxia causes DNA hypermethylation by reducing TET activity,” *Nature*, vol. 537, no. 7618, pp. 63–68, 2016.
- [237] C. J. Watson, P. Collier, I. Tea et al., “Hypoxia-induced epigenetic modifications are associated with cardiac tissue fibrosis and the development of a myofibroblast-like phenotype,” *Human Molecular Genetics*, vol. 23, no. 8, pp. 2176–2188, 2014.
- [238] S. Antero, K. Kai, and K. Anu, “Hypoxia-inducible histone lysine demethylases: impact on the aging process and age-related diseases,” *Aging and Disease*, vol. 7, no. 2, pp. 180–200, 2016.
- [239] N. J. Mabeesh, D. Escuin, T. M. LaVallee et al., “2ME2 inhibits tumor growth and angiogenesis by disrupting microtubules and dysregulating HIF,” *Cancer Cell*, vol. 3, no. 4, pp. 363–375, 2003.
- [240] R. Luo, Y. Wang, P. Xu et al., “Hypoxia-inducible miR-210 contributes to preeclampsia via targeting thrombospondin type I domain containing 7A,” *Scientific Reports*, vol. 6, no. 1, article 19588, 2016.
- [241] R. Luo, X. Shao, P. Xu et al., “MicroRNA-210 contributes to preeclampsia by downregulating potassium channel modulatory factor 1,” *Hypertension*, vol. 64, no. 4, pp. 839–845, 2014.
- [242] A. A. Rawluszko, K. Horbacka, P. Krokowicz, and P. P. Jagodzinski, “Decreased expression of 17 $\beta$ -hydroxysteroid dehydrogenase type 1 is associated with DNA hypermethylation in colorectal cancer located in the proximal colon,” *BMC Cancer*, vol. 11, no. 1, p. 522, 2011.
- [243] X. C. Zeng, X. Ao, H. F. Yang et al., “Differential expression of stromal aromatase in obese females is regulated by DNA methylation,” *Molecular Medicine Reports*, vol. 10, no. 2, pp. 887–890, 2014.