#### **Conference** Case



# A difficult case of angioimmunoblastic T-cell lymphoma with Epstein-Barr virus-negative large mononuclear atypical cells

Keywords: Angioimmunoblastic T-cell lymphoma, Classic Hodgkin lymphoma, Composite lymphoma, HRS-like cell, HRS cell

### **CASE REPORT**

A 74-year-old man with no history of a major medical condition presented with left neck lymphadenopathy that slowly progressed over approximately 2 months. He reported no fever, night sweats, or weight loss. A physical examination revealed generalized lymph node swelling, including the bilateral neck, axillary, and inguinal lymph nodes. On laboratory examination, the complete blood count suggested mild thrombocytopenia at  $14.1 \times 10^4/\mu$ L. The results of a chemistry panel were normal, except for a lactate dehydrogenase level of 294 U/L, soluble interleukin-2 receptor level of 2140 U/mL, and hypergammaglobulinemia (IgG, IgA, and IgM levels were 2568, 236, and 547 mg/dL, respectively). Staging positron emission tomography/computed tomography (PET/CT) revealed hypermetabolic lymph nodes in the bilateral cervical, bilateral supraclavicular, paraaortic, and bilateral iliac regions. Moreover, hypermetabolic foci were detected in the mediastinum and spleen (Figure 1). Excisional biopsy of the left neck lymph node demonstrated that the lymph node architecture was affected by small to medium-sized lymphoid cells with a small number of interspersed large mono and binuclear atypical cells. Prominent vascularity and a variety of inflammatory cells were present in the background. On immunohistochemical analysis, large atypical cells were positive for CD30 and PAX5 (weak), and negative for CD15, CD20, and ALK. These results were consistent with the phenotype of Hodgkin and Reed/ Sternberg (HRS) cells. The small to medium-sized lymphoid cells exhibited a T follicular helper cell phenotype, and were positive for CD3, CD4, CD10, and PD-1. Both large mononuclear cells and lymphoid cells were negative on EBV-encoded small RNA1 in situ hybridization (EBER1-ISH). The histopathological findings are shown in Figure 2. Rearrangements of the immunoglobulin heavy chain (IGH), T-cell receptor  $\gamma$  (TRG), and T-cell receptor  $\beta$  (TRB) were detected using DNA obtained from whole sections of paraffin-embedded specimens by polymerase chain reaction analysis using the BIOMED-2 protocol at SRL, Inc. (Tokyo, Japan).<sup>1</sup> Although the diagnosis of AITL was made mainly based on the histopathological findings, it was difficult to differentiate HRS-like cells from true HRS cells. Cyclophosphamide, doxorubicin, vincristine, and prednisolone (CHOP)

chemotherapy was initiated. After the completion of six cycles of CHOP, restaging PET/CT demonstrated a complete metabolic response.

AITL is a subtype of the second most common peripheral T-cell lymphoma, followed by peripheral T-cell lymphoma (PTCL), not otherwise specified.<sup>2</sup> Due to the immunological dysfunctions associated with AITL, EBV-positive B lymphocytes can often expand. In rare cases, these reactive EBVpositive B lymphocytes are morphologically and immunophenotypically indistinguishable from true HRS cells. Therefore, they are termed HRS-like cells. HRS-like cells are often associated with EBV reactivation; however, EBVnegative HRS-like cells can exist.<sup>3</sup> On the other hand, in rare instances, two distinct subtypes of malignant lymphomas develop simultaneously or serially in the same patient. Such lymphomas are referred to as composite lymphomas (CLs). CLs account for 1-4% of all malignant lymphomas.<sup>4</sup> To date, eight cases of CL comprising classic Hodgkin lymphoma (HL) and PTCL have been reported. In three cases, HRS cells harbored EBV infection.<sup>5-7</sup> On the other hand, five cases of CL constituting PTCL and HL with EBV negative-HRS cells have been reported.8 In general, IGH rearrangements are found in approximately 20% of AITL cases. However, B-cell clonality is likely in AITL cases with expanded EBV-infected B lymphocytes.9

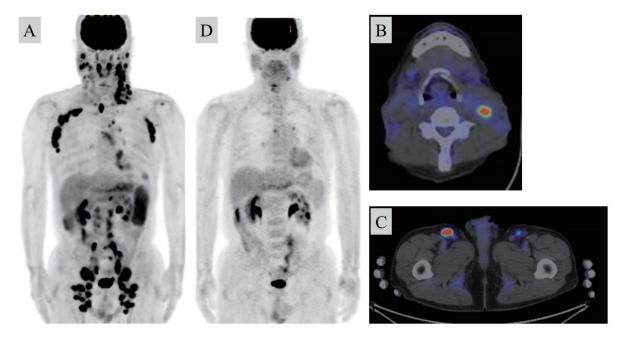
In summary, when determining the diagnosis, it is challenging to distinguish HRS-like cells from true HRS cells. Furthermore, in the present case, EBV-negative large mononuclear cells and the detection of clonal IGH gene rearrangements without evidence of EBV reactivation made the diagnosis more difficult. We would like to ask experts about how to diagnose the present case.

### ACKNOWLEDGMENTS

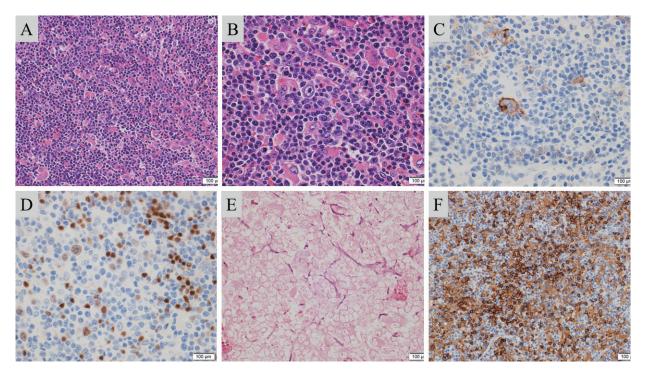
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### **CONFLICT OF INTEREST**

The authors declare the following competing interests: K.N. receives research funding from Novartis and Kyowa



**Fig. 1.** *A*, Baseline positron emission tomography/computed tomography (PET/CT) revealed hypermetabolic lymph nodes in the bilateral cervical, bilateral supraclavicular, para-aortic, and bilateral iliac regions. Hypermetabolic foci were also detected in the mediastinum and spleen. *B*, Left cervical lymph node. *C*, Right inguinal lymph node. *D*, Post-chemotherapy PET/CT demonstrated complete metabolic response.



**Fig. 2.** *A* and *B*, Lymph node biopsy revealed scattered large mononuclear lymphoid cells with infiltrating small to medium lymphocytes. Prominent vascularity and a variety of inflammatory cells, such as eosinophils, were also observed. (hematoxylineosin, original magnification  $\times 200$ ,  $\times 400$ ) *C* and *D*, Hodgkin/Reed-Sternberg (HRS) cells were positive for CD30 and weakly positive for PAX-5 compared with normal B-lymphocytes in the background. (original magnification  $\times 400$ ) *E*, HRS cells and lymphocytes in the background were negative on Epstein-Barr virus-encoded RNA1 *in situ hybridization*. (original magnification  $\times 400$ ) *F*, Neoplastic T-cells were dominantly positive for CD4. (original magnification  $\times 400$ )

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## REFERENCES

- van Krieken JHJM, Langerak AW, Macintyre EA, *et al.* Improved reliability of lymphoma diagnostics via PCR-based clonality testing: — Report of the BIOMED-2 Concerted Action BHM4-CT98-3936. Leukemia. 2007; 21 : 201-206.
- 2 Swerdlow SH, Campo E, Harris N, *et al.* World Health Organization Classification of Tumours of Haematopoietic and Lymphoid Tissue. 4th ed, Volume 2. Lyon, IARC Press. 2017.
- 3 Eladl AE, Satou A, Elsayed AA, *et al.* Clinicopathological study of 30 cases of peripheral T-cell lymphoma with Hodgkin and Reed-Sternberg-like B-cells from Japan. Am J Surg Pathol. 2017; 41 : 506-516.
- 4 Küppers R, Dührsen U, Hansmann ML. Pathogenesis, diagnosis, and treatment of composite lymphomas. Lancet Oncol. 2014; 15 : e435-e446.
- 5 Niedobitek G, Baumann I, Brabletz T, *et al.* Hodgkin's disease and peripheral T-cell lymphoma: composite lymphoma with evidence of Epstein-Barr virus infection. J Pathol. 2000; 191 : 394-399.
- 6 Sanchez S, Holmes H, Katabi N, *et al.* Composite lymphocyterich Hodgkin lymphoma and peripheral T-cell lymphoma associated with Epstein-Barr virus: a case report and review of the literature. Arch Pathol Lab Med. 2006; 130 : 107-112.
- 7 Gualco G, Chioato L, van den Berg A, Weiss LM, Bacchi CE. Composite lymphoma: EBV-positive classic Hodgkin lymphoma and peripheral T-cell lymphoma: a case report. Appl Immunohistochem Mol Morphol. 2009; 17 : 72-76.

- 8 Ichikawa A, Miyoshi H, Yamauchi T, *et al.* Composite lymphoma of peripheral T-cell lymphoma and Hodgkin lymphoma, mixed cellularity type; pathological and molecular analysis. Pathol Int. 2017; 67 : 194-201.
- 9 Attygalle AD, Chuang S-S, Diss TC, et al. Distinguishing angioimmunoblastic T-cell lymphoma from peripheral T-cell lymphoma, unspecified, using morphology, immunophenotype and molecular genetics. Histopathology. 2007; 50 : 498-508.

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