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Parasitic infections in HIV infected individuals: Diagnostic & therapeutic challenges

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After 30 years of the human immunodeficiency virus (HIV) epidemic, parasites have been one of the most common opportunistic infections (OIs) and one of the most frequent causes of morbidity and mortality associated with HIV-infected patients. Due to severe immunosuppression, enteric parasitic pathogens in general are emerging and are OIs capable of causing diarrhoeal disease associated with HIV. Of these, Cryptosporidium parvum and Isospora belli are the two most common intestinal protozoan parasites and pose a public health problem in acquired immunodeficiency syndrome (AIDS) patients. These are the only two enteric protozoan parasites that remain in the case definition of AIDS till today. Leismaniasis, strongyloidiasis and toxoplasmosis are the three main opportunistic causes of systemic involvements reported in HIV-infected patients. Of these, toxoplasmosis is the most important parasitic infection associated with the central nervous system. Due to its complexity in nature, toxoplasmosis is the only parasitic disease capable of not only causing focal but also disseminated forms and it has been included in AIDS-defining illnesses (ADI) ever since. With the introduction of highly active anti-retroviral therapy (HAART), cryptosporidiosis, leishmaniasis, schistosomiasis, strongyloidiasis, and toxoplasmosis are among parasitic diseases reported in association with immune reconstitution inflammatory syndrome (IRIS). This review addresses various aspects of parasitic infections in term of clinical, diagnostic and therapeutic challenges associated with HIV-infection.

Key words AIDS - anti-parasitic treatment - diagnosis - HAART - HIV-infected individuals - IRIS - parasitic opportunistic infections

Introduction

In 1981, the first patient diagnosed with acquired immunodeficiency syndrome (AIDS) in the USA was reported. The growing number of people living with HIV has constantly been detected the world over and in particular from the three main continents, Asia, South America and sub-Saharan Africa. In a global report on AIDS epidemic, it is estimated that there are 33.3 million (31.4-35.3 million) adults and children living with HIV around the world. The data from epidemiological surveillances have further shown that there are an estimated 2.6 million (2.3-2.8 million) people newly infected with HIV. About 370,000 (230,000-510,000) children are newly infected with HIV. It is also believed that there are 1.8 million (1.6-2.1 million) annually AIDS-related deaths worldwide¹. Opportunistic infections (OIs) are one of the existing identified causes that aggravate the condition of HIVinfected patients. Of these, parasites play an important role as OIs and are one of the most common causes of morbidity and mortality in HIV/AIDS patients. The spectrum of parasitic opportunistic infections (POIs) affecting HIV-infected individuals is divided into protozoa and helminthes (nematodes, cestodes, and trematodes). These POIs are not only associated with symptomatic HIV-infected patients but also are more evident with decreasing immune state (CD4⁺ cell count < 200 cells/µl), which constitutes an ADI. Among POIs, cryptosporiosis, isosporidiosis and microsporidiosis are the main enteric/intestinal parasitic infections, while leishmaniasis and toxoplasmosis are the main systemic POIs reported in HIV-infected patients. Of these, cryptosporiosis, isosporidiosis, and most importantly toxoplasmosis with brain involvement are the three key parasitic diseases which have been included in the Centers for Disease Control and prevention (CDC) case definitions for AIDS². This review therefore aims to highlight the important issues pertaining to these POIs in term of clinical, diagnostic and therapeutic challenges in HIV-infected patients. It also provides a better understanding of clinical scenario and future management of these POIs at the time of widely access to the HAART, particularly in resource limited settings.

Intestinal or enteric parasites in HIV/AIDS patients

Diarrhoea in patients with AIDS is significantly caused by intestinal parasites^{2,3}. The main aetiologic agents are intracellular protozoa, *Isospora belli*, *Cryptosporidium parvum*, and *Cyclospora* spp⁴. However, infection with other extracellular parasites is also related to diarrhoeal disease in AIDS patients. Among these parasites, *Entamoeba histolytica*, *Giardia intestinalis* and *Strongyloides stercoralis* are the most important⁵.

Several studies have been carried out to determine the presence of intestinal parasites in HIV patients. A higher prevalence of intracellular parasites, particularly *Cryptosporidium* spp. and *S. stercoralis* was found in HIV positive cases, than extracellular parasites⁶. Since cellular immunity is the major defense mechanism against intestinal parasitic infections⁷, the association between intestinal parasites and individuals with reduced immunity due to CD4⁺ T-lymphocyte reduction in HIV/AIDS is well predictable, particularly from cases presented with diarrhoea⁸.

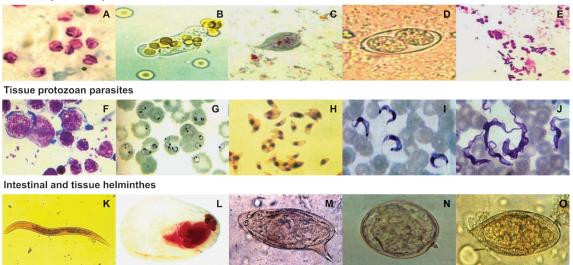
Intestinal protozoan parasites

C. parvum, *E. histolytica*, *G. intestinalis*, *I. belli* and *Microsporidium* spp. (Fig. 1) are among intestinal protozoan infections which have commonly

been reported in HIV-infected patients. *Ascaris lumbricoides*, hookworms, *Trichuris trichiura* and *S. stercoralis* are intestinal helminthic infections, which have also been reported in these patients⁹. These enteric parasitic infections usually produce diarrhoeal disease by infecting the small or large intestine, or both. They are also often found in children and adults in tropical climates. Amoebiasis caused by infection with *E. histolytica* leads to bloody diarrhoea and hepatic disease. The other protozoa produce persistent diarrhoea with or without malnutrition. Severe enteritis and chronic diarrhoea in HIV infected patients are often documented as a consequence of multiple opportunistic intestinal protozoa infections and can result in significant morbidity and mortality¹⁰.

Cryptosporidium spp., *Isospora belli* and *Microsporidium* spp.

Cryptosporidiosis and isosporiasis have been categorized by CDC as ADI2. These are the two most common intestinal protozoan parasites causing diarrhoea and pose a public health problem in AIDS patients. Recent reports from France indicated the increase of isosporiasis from 0.4 per 1000 patients in the pre-HAART era (1995-1996) to 4.4 per 1000 patients in the HAART era (2001-2003)¹¹. The prevalence of cryptosporidiosis varied according to geographical locations. In Southeast Asia, the infective rate of C. parvum is up to 40 per cent¹². In clinical cryptosporidiosis, chronic diarrhoea with watery stools, weight loss and dehydration are the prominent features in symptomatic patients¹³. Cryptosporidiosis occurs in AIDS patients when the CD4⁺ cell count is < 200cells/µl¹⁴. Microsporidium is an obligate intracellular microorganism that was recently reclassified from protozoa to fungi. It has emerged as the causes of OIs associated with diarrhoea and wasting in AIDS patients especially in developing countries where combination antiretroviral therapy (ART) is not always accessible¹⁵. Interestingly, most studies found that Cryptosporidium spp. was the most common intestinal parasitic co-infection with Microsporidium spp^{14,16}. Clinical symptoms and disease associated with microsporidiosis vary with the status of the host's immune system. Persistent diarrhoea, abdominal pain, and weight loss are common clinical symptoms¹⁵. Higher prevalent rates of these intestinal parasitic infections are found related to CD4⁺ cell count of <100 cells/µl¹⁷. With greater awareness and implementation of better diagnostic methods, it was demonstrated that microsporidia contribute to a wide range of clinical



Intestinal protozoan parasites

Fig. 1. Microscopic images from clinical samples. (A) Oocysts of *Cryptosporidium parvum* from a stool sample (Modified acid-fast stain).
(B) Trophozoite of *Entamoeba histolytica* from a fresh stool sample. (C) Trophozoite of *Giardia intestinalis* from a stool sample (Trichrome stain). (D) Oocyst of *Isospora belli* from a stool sample. (E) Spores of *Microsporidiam* spp. from a stool sample (Gram-chromotrope stain).
(F) Oval-shaped *Leishmania donovani* amastigotes from a bone marrow aspirate (Giemsa stain). (G) Ring form of *Plasmodium falciparum* from a blood smear (Giemsa stain). (H) Trophozoites of *Toxoplasma gondii* from intra-peritoneal fluid (Giemsa stain). (I) Monomorphic trypomastigotes of *Trypanosoma cruzi* in a blood smear (Giensa stain). (J) Polymorphic trypomastigotes of *Trypanosoma brucei gambiense* in a blood smear (Giemsa stain). (K) Rhabditiform larva of *Strongyloides stercoralis* from a stool sample. (L) *Cysticercus cellulosae* (Carmine and fastgreen stain). (M) Egg of *Schistosoma mansoni* from a stool sample. (N) Egg of *Schistosoma haematobium* from a stool sample. (A) Egg of *Schistosoma haematobium* from a stool sample. (A) Egg

syndromes in HIV-infected individuals¹⁸. Therefore, primary effectiveness of prevention and control strategies against these parasitic infections should be implemented in HIV/AIDS patients particularly in resource limited settings.

Entamoeba histolytica

E. histolytica is one of the most important parasitic diseases in developing and developed countries. It infects approximately 50 million people worldwide. Although, most subjects infected with *E. histolytica* are asymptomatic, it accounts for more than 100,000 deaths annually¹⁹. *E. histolytica* is unique among amoeba in its ability to disrupt the intestinal mucosa causing colitis and the capability for further haematogenous spread, producing potentially fatal abscesses in the absence of appropriate therapy²⁰.

E. histolytica infection is common in homosexual men, particularly those sexually active HIV-infected men who have sex with men (MSM) are at greater risk of developing amoebic disease than HIV-uninfected MSM and the general population²¹. Transmission in these men occur by oral-genital or oral-rectal sexual practices²¹.

A high prevalence of *E. histolytica* infection in HIV/AIDS patients in various countries has been reported²². *E. histolytica* seropositivity in HIV-infected individuals was found to be higher in patients with a CD4⁺ cell count of <200 cells/ μ l²². It is unclear whether HIV infection is a risk factor for *E. histolytica* infections but it has been suggested that these are more susceptible to an invasive form of the disease than are normal population²³. The common clinical symptoms and signs of amoebic liver abscess (abdominal pain, fever and chills, and abdominal tenderness) show a more insidious onset of illness in term of longer duration of hospital stay in HIV patients than in normal population²⁴ but in most cases there are no significantly differences between the two groups²⁵.

Giardia intestinalis

G. intestinalis is one of the most common pathogenic intestinal protozoan parasites of humans causing acute and chronic diarrhoea throughout the world. It is becoming increasingly important among HIV/AIDS patients. Although giardiasis was not considered a major cause of enteritis in AIDS patients, there are reports that some cases of acute and chronic diarrhoea may be associated with *G. intestinalis* infection^{4,26}. In HIV patient as well as in healthy individuals, infection with *G. intestinalis* is frequently asymptomatic but can also cause abdominal cramps, diarhoea with flatulence and weight $loss^{27}$. These symptoms generally begin 1-2 wk after infection, and may last 2-6 wk. Studies from Brazil²⁶ indicate a high prevalence of *G. intestinalis* infection among HIV/AIDS patients, whereas studies form Ethiopia found no significant increase in infection rates of *G. intestinalis*²⁸. However, HIV patients did have an increased probability of more severe symptoms when infected with this organism, especially in the most advanced stage of disease²⁹.

Diagnosis: The diagnosis of amoebiasis and giardiasis is usually made by examination of stool under microscope for evidence of trophozoites or cysts. For cryptosporidiosis, faecal sample may need acid fast staining and view under microscope for oocyst.

Stool examination - The introduction of stool examination for specific antigen tests has improved diagnostic capabilities. Immunochromatography card for the rapid detection of protozoal proteins from a small sample of stool are available from several different commercial sources and are more sensitive and more specific (>90%) than the traditional microscopec examinations. These kits are available from Meridian (Immuno*Card* STAT![®] *Cryptosporidium/Giardia*), Alexon-Trend (Xpect[®] *Giardia*), Tech-lab (PT5012-*Giardia* II). The benefits of antigen detection methods are that there are no requirement for expert microscopists and require less time than conventional microscopic method.

Serological diagnosis - Although the available commercial kits were claimed to be the best single test for diagnosing intestinal protozoa, microscopic examination of stool could identify other parasites, in addition to each specific detection kit, that can cause diarrhoea. Therefore, microscopic examination of stool has value beyond diagnosing a single disease.

Treatment: Treatment for amoebiasis and giardiasis involves nitroimidazoles such as metronidazole (Flagyl), which has been used successfully for more than four decades. Undesirable side effects including anorexia, nausea, vomiting, malaise, metallic taste, potential teratogenicity and failures in treatment in normal and HIV patients have been reported³⁰.

In the USA, the only drug approved by FDA for treating giardiasis is furazolidone (Furoxone) for 7-10 days. Nitazoxanide (Alinia) could become the preferred drug as it comes in a liquid form which may be easier

for children to swallow. Tinidazole (Tindamax) and ornidazole are alternatives to metronidazole. Although these have many of the same side effects as metronidazole, but can be given in a single dose. Diloxanide furoate (Furamide) was also recommended in case of failure treatment with metronidazole³¹.

Certain drugs such as co-trimoxazole or nitazoxanide seem to have partial treatment for cryptosporidiosis. There is also no effective drug to treat patients infected with *Microsporidium* spp. To date, no specific drug has been shown to be effective against these infections. One recent study found no oocysts of *Cryptosporidium* spp. were detected in stools of patients receive HAART³².

Diseases due to tissue parasites in HIV/AIDS patients

Leishmaniasis

Leishmaniasis is caused by Leishmania spp., an haemoflagellate, intracellular and zoonotic protozoan parasite. It is one of the most common neglected tropical diseases. Global epidemiological surveillance shows that infection with Leishmania spp. was found in more than 12 million people worldwide and 2 million new cases are reported annually³³. Of these, 350 million people are considered as "high risk" and the incidence of leishmaniasis is increasing³³. Overall, the disease has been reported from 88 developing countries, particularly found among people living in poor conditions. There are two basic forms of clinical manifestations, namely, cutaneous leishmaniasis (CL), a disfiguring and stigmatizing disease, and visceral leishmaniasis (VL) or kala azar, a lifethreatening but treatable condition if diagnosed early. VL is prevalent in approximately 70 countries and South Asia (estimated 300,000 cases in 2006); India is among the largest foci of the disease followed by East Africa (30,000 cases per year) and Brazil (4,000 cases reported in 2006)³⁴. There is an increase on the occurrence of new foci and the incidence of VL in east Africa. CL, cutaneous form of leishmaniasis, is geographical distributed among 82 countries whereby the predominant foci are countries in middle-east Asia, east Africa, South America and southern Europe³⁶. CL has several (localized, diffuse and mucocutaneous) clinical features which depend on the Leishmania species involved (Fig. 2). Mucocutaneous is the most severe form that causes disfiguring lesions and mutilation of the face.



Fig. 2. Clinical features of cutaneous leishmaniasis. Chronic ulcerative lesions of the lower limb as a result of infection with *Leishmania tropica*.

The impact of HIV/AIDS pandemic is not only changing the natural history of leishmaniasis³⁴ but also increasing the risk (100 to 1,000 times) of developing VL in endemic areas, reducing the therapeutic response, and increasing the possibility of relapse cases³⁷. Leishmaniasis in HIV-infected patients is mainly VL, however, some cases of CL have also been reported in HIV-infected patients from endemic countries such as in Africa³⁸ and South America³⁹. Since the introduction of HAART, the numbers of *Leishmania*-HIV co-infected cases from countries where they are endemic in Europe are rapidly declining.

Diagnosis: The diagnosis of leishmaniasis in HIVinfected patients may not be straightforward due to clinical variations from non-specific to atypical features found in majority of cases. Further, the presence of clinical signs such as hepatomegaly is typically less frequent in patients with this co-infection and about half of these patients present with clinical pictures that mimic with other common OIs making clinical diagnosis difficult. In suspected cases of leishmaniasis in HIV-infected patients, combined diagnostic investigations (if applicable) are recommended to confirm the diagnosis.

Parasitological diagnosis - Microscopic examination and culture are conventional methods that have historically been used to detect the present of *Leishmania* spp. from different biological specimens and considered as the gold standard. These methods may also help in the detection of treatment failures in cases of leishmaniasis in HIV-infected patients³⁵.

Serological diagnosis - Antibodies detection using several serological tests are available, including

immunofluorescent antibody test (IFAT), enzymelinked immunosorbent assay (ELISA) using crude antigen or rK39 antigen, immunoblotting, direct agglutination test (DAT), and the Rk39 dipstick test³⁵. The Rk39 dipstick test is a simple, rapid, inexpensive and non-invasive method with a sensitivity of >90 per cent for the detection of VL in India⁴⁰. However, sensitivities as low as 20 per cent have been reported for VL-HIV co-infected patients in Europe³⁵. In cases of severe immunodeficiency, these serological tests can be of limited use because antibodies to Leishmania spp. may be absent despite an acute leishmaniasis^{29,41}. Furthermore, antibodies to Leishmania spp. can crossreact with other parasitic infections such as those of Trypanosoma spp. which make the use of some serological tests problematic³⁵. Since the results of seropositivity vary according to serological tests performed, at least two different serological tests are recommended for each patient to enhance the sensitivity of antibody production⁴².

Molecular diagnosis - PCR is a useful technique for the detection and identification of different *Leishmania* species directly from clinical samples⁴³. PCR assays applied to bone marrow and peripheral blood specimens is a useful tool for the diagnosis of VL-HIV co-infected patients³⁵. By using non-invasive biological specimens, PCR assays also seem to be feasible for long-term monitoring the efficacy of treatment and the prediction of relapse cases⁴⁴.

Treatment: Antileishmanial therapy consists of the antimonial compounds (sodium stibogluconate and meglumine antimoniate), miltefosine, and amphotericin B is used for the treatment of leishmaniasis in HIV-infected patients. Patients with *Leishmania*-HIV co-infection generally have low cure rates, high drug toxicity and high mortality rates. Moreover, low CD4⁺ cells count and recurrent relapses may contribute to temporary recovery and resistance to the previous drug regimens used³⁵.

The antimonial compounds have been used for past decades as the first line drug for treatment because of their low cost and availability in most countries. However, the compounds are well known for their toxicities such as severe vomiting and emerging drug resistant⁴⁵. The compounds have high toxicity, high mortality rate and potentially stimulated HIV-1 replication in *in vitro*, therefore, it is not recommended for the first-line treatment of *Leishmania*-HIV co-infected patients³⁵.

Lipid formulations of amphotericin B (Liposomal amphotericin B; L-AmB) is the drug of choice for treating VL⁴⁶. Due to its cost, this therapeutic agent is more commonly used in developed than in resource-poor countries⁴¹. A European study demonstrated that L-AmB with a total dose of up to 30-40 mg/kg was well tolerated in small numbers of *Leishmania*/HIV-coinfected patients but this therapy did not prevent relapse cases⁴⁷. Data on AmB lipid complex or L-AmB used for secondary prophylaxis in co-infected patients is scanty; therefore, more study are needed to prevent the high possibility of recurrent leishmaniasis in these patients.

Malaria

Of the four *Plasmodium* species: *P. falciparum*, P. vivax, P. malariae, and P. ovale, P. vivax is the most common cause of malaria particularly found in endemic areas and P. falciparum is the main cause of severe malaria that leads to high morbidity and mortality. The major foci of malaria are found in many regions such as South America, Southeast Asia and sub-Saharan Africa⁴⁸. Approximately 300 to 500 million children are infected with malaria each year leading to more than 1 million deaths annually in sub-Saharan Africa⁴⁹. About 25 million pregnant women are at risk of infection of *P. falciparum* in this region⁴⁸. A higher parasites load was detected in HIV-infected multigravidae and also the increasing rates of fever due to malaria with the progressive immunosuppression in HIV-infected partients⁴⁹.

Diagnosis: Parasitological diagnosis - The diagnosis of clinical malaria is based on a high index of suspicion in both the patient's history of exposure and clinical presentation. The definite diagnosis requires the direct identification of malarial parasite species in the peripheral blood specimens (parasitaemia) from thick and thin blood smears using Giemsa staining technique.

Serological diagnosis - There are a few commercial diagnostic kits available in the detection of anti-malarial antibodies namely *Para*Sight-F is the simple and accurte diagnostic kits for *P. falciparum* infection⁵⁰.

Neuroimaging diagnosis - In cerebral malaria, brain computed tomography (CAT scan) shows cerebral oedema by effacement of the cortical sulci and small or slit-like ventricles, and if it occurs with hypoattenuation of the basal ganglia or cerebellum, a sign of poor prognosis⁵¹. Brain magnetic resonance image (MRI) demonstrates diffuse swelling with or without oedema, as well as multifocal cortical and subcortical lesions⁵². In fatal cases of cerebral malaria, brain CAT scan and MRI reveal a common finding of transtentorial herniation⁵³.

Treatment: The interaction between HIV infection and malaria has been well described over the past few years. As a standard guideline, co-trimoxazole is daily recommended in HIV-infected patients, including pregnant women⁵⁴. Apart from antimalarial treatment, implementation of insecticide-treated nets, cotrimoxazole prophylaxis, and ART play an important role in reducing the morbidity and mortality of clinical malaria in HIV-infected patients. There are many more treatment challenges that need to be answered regarding clinical malaria in HIV-infected patients: the role of co-trimoxazole prophylaxis in HIV-infected patients undergoing ART, the efficacy of antiretroviral drugs against malaria, the synergistic action between antimalarial and antiretroviral treatments, and the effect of antifolate resistance under co-trimoxazole prophylaxis54.

Neurocysticercosis

Neurocysticercosis (NCC), caused by a larval tapeworm Taenia solium, is the most common helminthic infection of the central nervous system and the most common cause of patients with acquired seizure. Cysticercosis is highly endemic in most developing countries such as Asia, Latin America, parts of Oceania, and sub-Saharan Africa where pigs are raised55. Faeco-oral contamination is the main route of transmission through accidentally ingestion of T. solium egg found among infected humans. Clinical presentations of patients with NCC depend on the number, size, location of CNS lesions, and the host immune response⁵⁶. Of these, seizures are the most common clinical presentation of NCC, but other syndromic features, such as headache, hemiparesis and ataxia may also be seen in these patients.

Diagnosis: Though, NCC is not that common compared to other OIs in HIV-infected patients, misdiagnosis can easily lead to fatal outcome in immunocompromised hosts. The definitive diagnosis of NCC is recommended based on combined histology, radio-imaging, serology, and clinical symptoms. The diagnosis of NCC is made on histopathologic evidence of NCC, CAT scan or MRI showing a scolex within a cystic lesion, or neuroimaging showing a lesion compatible with NCC or a clinical response to initial therapy of NCC, combined with serological evidence of infection with *T. solium* using CSF ELISA⁵⁷.

Treatment: Albendazole and praziquantel are the mainstay drugs for treatment of NCC, especially for meningeal infection and steroids should be administered to reduce oedema as a result of medical treatment⁵⁸.

Schistosomiasis (Bilharziasis)

Schistosomiasis, a neglected tropical disease or a man-made disease, is caused by five species of blood flukes of the genus *Schistosoma*: *S. mansoni*, *S. japonicum*, *S. haematobium*, *S. intercalatum*, and *S. mekongi*. This parasitic disease infects 200 million people worldwide and about 600 million people are at high risk to the infection in 72 endemic countries⁵⁹. The infection with *Schistosoma* spp. is mostly transmitted through water contact and requires an intermediate mollusk host to complete its life-cycle. Factors contribute to the occurrence of this systemic helminthic infections in new geographical foci are international travel, refugee and population migration, and the new development of water resources⁶⁰.

In HIV-infected patients, a couple of studies showed that up to 17 per cent of HIV-infected patients from sub-Saharan Africa had seropositive for schistosomiasis⁶¹. There are three main clinical diagnoses of schistosomiasis (gastrointestinal, disseminated and CNS) reported in HIV-infected patients. Gastrointestinal schistosomiasis is the most common form and patients typically present with weight loss, diarrhoea, abdominal pain and odynophagia⁶². Only a few cases of disseminated and neurkoschistosomiasis (NSS) have been reported in HIV-infected patients⁶³.

Diagnosis: The diagnosis of NSS is difficult because clinical CNS symptoms are non-specific and laboratory findings such as eosinophilia and the presence of *Schistosoma* eggs in clinical samples may not be present⁶⁴. NSS should be included in the differential diagnosis in any patient who is from endemic area of schistosomiasis and who displays an acute encephalopathy of unknown origin.

Parasitological diagnosis - The definite diagnosis is based on the direct identification of the eggs in tissue biopsy specimens of patients with NSS compared to the supportive evidence on direct detection of *Schistosoma* eggs in stool (*S. mansoni*) or urine (*S. haematobium*) samples for patients with schistosomiasis⁶⁵.

Serological diagnosis - In general, there are promising serological methods used for

antibodies detection such as IgG, IgM or IgE by different diagnostic kits of ELISA, IHA or immunofluorescence⁶⁶. These methods are sensitive and useful in the diagnosis of schistosomiasis of both affected people living in endemic areas and among travellers. However, there are some limitations to these commercial kits that need to be rectified in order to avoid misdiagnosis: the antibodies persist even after parasitological cure⁶⁵, or the possibility of cross-reactivity with other helminthic infections. Another promising method of the detection of circulating adult worm antigen and soluble egg antigen is in using monoclonal and polyclonal antibodies from various clinical samples⁶⁷.

Neuroimaging diagnosis - CAT scan and MRI are useful tools in the investigation of NSS. CAT scan finding typically shows single or multiple hyperdense lesions surrounded by oedema with variable contrast enhancement⁶⁸, while, MRI of the brain demonstrates a characteristic "arborized" appearance with linear enhancement surrounded by punctate enhancing nodules⁶⁹.

Treatment: The drug praziquantel, steroids and surgery are currently available as the mainstay for treatment of NSS. A 5-days course of praziquantel of 50 mg/ kg/day divided in two daily doses is recommended for treatment of NSS⁷⁰. Artemether, an antimalarial drug and a synergistic drug with praziquantel is effective against the immature migrating larvae (schistosomula)⁷¹. In non-compliance to praziquantel, oxamniquine of 30 mg/kg daily for 2 days can be alternatively used for treatment, particularly for the infection with *S. mansoni*⁷⁰. In HIV-infected patients, the treatment is effective for schistosomiasis but it shows a high reinfection rate and correlates with a low CD4⁺ cell count⁷².

Strongyloidiasis

Strongyloidiasis is caused by *Strongyloides stercoralis*, a human intestinal helminthic infection. This parasitic disease has been found geographically distributed in both tropical and subtropical foci. In HIV-infected patients, hyperinfection syndrome (HS) and disseminated infection (DI) are uncommon but usually occurs in HIV-infected patients with CD4⁺ counts of <200 cells/µl. So far, very few cases of systemic/disseminated strongyloidiasis have been reported in HIV-infected patients⁷³. The low incidence was due to indirect development of *S. stercoralis* infective larvae in the gut of HIV-infected patients

do not favor disseminated strongyloidiasis⁷⁴. Only a (<30 cases) of strongyloidiasis HS in HIV-infected patients have been reported⁷⁵.

Diagnosis: Parasitological diagnosis - Definitive diagnosis is confirmed by direct identification of *Strongyloides* spp. rhabditiform larvae in different clinical specimens: stool, sputum, serum/blood smears, bronchial aspirates, cerebrospinal fluid (CSF), peritoneal fluid or ascitic fluid⁷⁶. Stool examination is a simple and cheap method but the screening for *Strongyloides* infection cannot rely solely on this method particularly in cases of HS where there is low number of larvae in the stool. The Harada-Mori filter paper appears to be a less successful culture method for *Strongyloides* than blood agar plate culture⁷⁷.

Serological diagnosis - ELISA should be performed if strongyloidiasis is suspected but not detected by parasitological diagnosis⁷⁷. Serology may give false negative results in patients with disseminated disease, may cross-react with other helminthic infections, or cannot differentiate between recent and remote infections.

Skin testing - The sensitivity of skin test decreases in the immunocompromised and most likely found in HIV-infected patients due to the similar effect and this method is not easily accessible⁷⁵.

Treatment: Ivermectin 200 μ g/kg/day is the treatment of choice for strongyloidiasis, in cases of HS, and

DI, should be continued at least for 7-10 days or until the clinical symptoms are resolved⁵⁷. Secondary prophylaxis is recommended in immunocompromised individuals *e.g.*, a 2-day course of ivermectin every 2 wk may prevent HS or DI⁷⁵.

Toxoplasmosis

The coccidian Toxoplasma gondii is a ubiquitous, intracellular. protozoan parasite that causes toxoplasmosis. T. gondii infection can enhance the immunodeficiency found after HIV-1 infections. Up till the present day, toxoplasmosis is medically the most important POIs of both epidemiological and clinical aspects in association with HIV-infected patients, which has been reported worldwide. The high prevalent rates of latent Toxoplasma infection (41.9-72%) in patients infected with HIV were reported in South America and in approximately half of the studies $(\geq 40\%)$ from the Asian continent. In North America, however, the rate of Toxoplasma infection was low (Fig. 3). Latent stage of toxoplasmosis is still prevalent as an infection that coexists with HIV pandemic. In concurrence with HIV infection, more than 95 per cent of cerebral toxoplasmosis (CT) occurs primarily due to reactivation of latent Toxoplasma infection and CT is one of the most frequent OIs, particularly in patients with full-blown AIDS. CT is the most common clinical presentation of toxoplasmosis and is one of the most frequent causes of focal intracerebral lesions that complicates AIDS⁷⁸. When HIV-infected patients

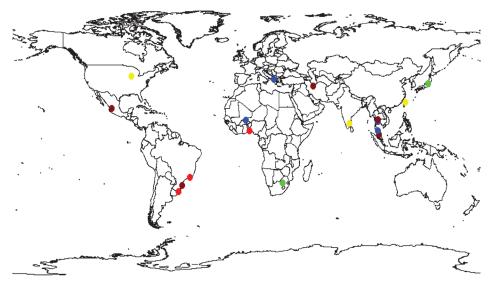


Fig. 3. Seroprevalence of toxoplasmosis in HIV-infected patients. Light red equals prevalence above 60%, brown equals 40-60%, blue 20-40%, yellow 10-20% and green equals prevalence <10%. White equals absence of data.

develop CT this poses many diagnostic and therapeutic challenges for clinicians⁷⁹.

In general, CT can be prevented by primary behavioural practices in avoiding acquisition of *Toxoplasma* infection such as consumption of well cooked meat, avoiding close contact with stray cats, contaminated soil and/or water, and receiving unscreened blood transfusions. Compliance to both therapeutic regimens for treatment and prophylaxis of toxoplasmosis is an imperative to prevent relapse of CT in areas where HAART is not fully accessible for people living with HIV/AIDS.

Diagnosis: Among patients with AIDS, cerebral involvement is more common and more serious than extracerebral toxoplasmosis. The definitive diagnosis is crucial for CT patients by directly demonstrating the presence of the tachyzoite form of *T. gondii* in the cerebral tissues. The presumptive diagnosis for CT, including the clinical presentations, radio-imaging findings, molecular and sero-diagnosis for *Toxoplasma* infection, and good response to anti-*Toxoplasma* therapy are widely accepted in clinical practice. The favourable outcome of CT is the improvement of clinical and radiological features after 2 to 3 wk of initiated empirical therapy.

Serological diagnosis - CT poses a diagnostic problem that relies on classical serological methods to detect anti-*Toxoplasma* immunoglobulins because blood samples from patients with immunodeficiency can fail to produce sufficient titres of specific antibodies. Seroevidence of *Toxoplasma* infection, independent of antibody levels, is generally seen in all patients before developing CT⁸⁰. Most cases have high titres of anti-*Toxoplasma* IgG antibodies with high IgG avidity that provides serological evidence of infection⁸¹ and this is the result of a secondary reactivation of latent or chronic *Toxoplasma* infection⁸¹. Therefore, it is important to determine the *Toxoplasma* serostatus in all HIV-infected patients to define the population at risk for CT.

Radiological diagnosis: Radio-imaging findings, either by CAT scan or MRI, are useful tools for the presumptive/empirical diagnosis of CT. CT usually causes unifocal, and more frequently multifocal lesions, or less likely diffuse encephalitis. These findings are however, not pathognomonic of CT. Radiological diagnosis⁸² can be classified as typical findings of hypodense lesions with ring-enhancing and perilesional oedema, and are observed in ~80 per cent of CT cases (Fig. 4). A typical pattern of hypodense lesions found without contrast enhancing. CAT scan without focal lesions and MRI demonstrating focal lesions, and diffuse cerebral encephalitis without visible focal lesions, are shown in about 20 per cent of CT cases. An unusual but highly suggestive image of CT is the 'eccentric target sign', which is a small asymmetric nodule along the wall of the enhancing ring⁸³. A CAT scan seems to be a sensitive diagnostic method for patients with focal neurological deficits; however it may underestimate the minimal inflammatory responses seen during early disease⁸⁴. MRI is recommended to be performed in patients with; neurological symptoms and positive serology to anti-Toxoplasma antibodies whose CAT scans show no or only a single abnormality, or persistent or worsening focal neurological deficits of disease if results of the initial procedure were negative⁸⁵.

Molecular diagnosis - The direct detection of *T. gondii* DNA in biological specimens by PCR has provided a major breakthrough for the diagnosis of toxoplasmosis⁸⁶. PCR techniques are suitable for patients with AIDS because these methods do not depend on the host immune responses and allow for direct detection of *T. gondii* DNA from a variety of clinical samples.

Other diagnostic methods - Brain biopsy is generally reserved for those patients with a diagnostic dilemma or do not fulfill the criteria for presumptive treatment and for those patients who fail to improve clinically or radiologically over the succeeding 10-14 days after being empirically treated for toxoplasmosis

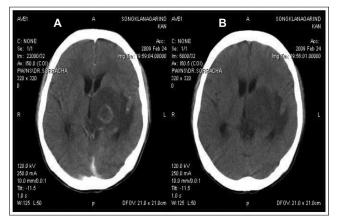


Fig. 4. Neuroimaging findings of cerebral toxoplasmosis in an AIDS patients. (A) Contrast enhanced CAT brain showed a ringenhancing lesion and marked surrounding vasogenic brain oedema. (B) Contrast enhanced CAT brain showed a resolution of cerebral lesion after 4 wk of anti-*Toxoplasma* therapy.

initially. Stereotactic brain biopsy (SBB) is then required to institute specific and appropriate therapy^{85,87}. Surprisingly, SBB is not commonly used for the diagnosis of CT or CT-associated other opportunistic CNS diseases in Asian countries⁸⁸ compared to other settings where CT cases have been reported in patients with AIDS. Brain biopsies do not influence survival of CT patients⁸⁹. SBB is an efficient, safe and important diagnostic procedure. In selected patients even expensive investigations should be undertaken before considering specific therapy and cost-effective homecare⁹⁰. This procedure should be performed early to achieve a prompt and accurate diagnosis and to guide the therapeutic scheme for AIDS patients with focal brain lesions⁹¹.

Treatment: Most patients with CT respond well to anti-*Toxoplasma* agents as demonstrated by findings from studies in various settings. However, about 10 per cent of CT cases died despite what was thought to be adequate treatment⁸². There are a few options other than anti-*Toxoplasma* regimens used as first-choice initial therapy; 6 wk with sulphadiazine (1.0-1.5 g per oral PO every 6 h) with pyrimethamine (100-200 mg [PO] loading dose, then 50 mg PO daily) and folinic acid (10-20 mg PO daily) that can reduce the haemato-toxicities related to pyrimethamine⁹².

Resistance to standard combination therapy (pyrimethamine and sulphadiazine) was reported in patients with CT⁹³. Atovaquone has consistently been found to be a promising therapeutic for salvage therapy in CT patients who were intolerant to or who failed standard regimens⁹⁴. However, the role of atovaquone in the treatment and prophylaxis of CT in AIDS patients is not well defined and more studies are required before a firm recommendation can be made⁹⁵.

Trypanosomiasis: American trypanosomiasis

American trypanosomiasis (Chagas' disease) is caused by *T. cruzi*, a hemoflagellate, intracellular and zoonotic protozoan parasite. It is a significant public health problem for not only people living in endemic areas (~15-16 million) in Latin American countries⁹⁶ but also in non-endemic countries due mainly to immigration to European countries, Canada or the United States⁹⁷. The clinical manifestation of Chagas' disease is divided into three stages: the acute stage, often remains unnoticed or misdiagnosed due to its initial clinical pictures are non-specific to Chagas' disease, intermediate stage, a period of asymptomatic carriage of parasites which remains life-long or chronic stage, multi-organs involvement after an incubation period of 10 to 30 years.

T. cruzi is one of the tropical pathogens that can coexist within HIV-infected individuals. T. cruzi/HIV co-infection in some areas of Mexico, Central and South American regions occur as a result of the geographical overlapping of these two pathogens, the occurrence of global migration and international travel⁹⁸. This dual infection occurs because of the secondary reactivation of chronic (latent) T. cruzi infection triggered by immunosuppression in HIV-infected patients98. In regions where Chagas' disease is endemic, the reactivation of this parasitic infection may be the first presentation of patients with HIV infection99,100 and it can cause a high mortality rate among these coinfected patients¹⁰¹. HIV-related immunosuppression changes the clinical manifestations of Chagas' disease including the central nervous system involvement¹⁰². Patients with reactivated CNS chagasic disease are more common in 75-80 per cent of patients with AIDS¹⁰³. Since 2004, reactivated Chagas' disease (meningoencephalitis and/ or myocarditis) was included in the AIDS in Brazil.

Diagnosis: In HIV-infected patients, the reactivation of Chagas' disease can be easily misdiagnosed as the clinical features and pathological appearance mimic with other brain diseases especially with CT. In such cases, Chagas' disease should be included in the differential diagnosis in HIV-infected patients presenting with focal neurological deficits which requires further investigations⁹⁸. Chagas' disease poses a diagnostic dilemma in clinical practice and clinician require highly suspicion of patients with chagasic encephalitis who arrive from endemic areas, has a history of blood transfusion or intravenous drug use, and has no response to empirically anti-*Toxoplasma* therapy¹⁰⁴.

Parasitological diagnosis - The definitive diagnosis is based on the direct detection of trypomastigote stage of *T. cruzi* in relevant clinical specimens using thick smears or Strout's concentration method, and CSF smears. In HIV-infected patients, the identification of trypomastigotes in CSF sample is the definitive diagnosis of chagasic encephalitis. Based on earlier studies, trypomastigotes are more frequently observed in the blood of HIV-infected patients with low level of CD4⁺ cell count¹⁰⁵. In cases of focal brain lesions with an initial empiric anti-*Toxoplasma* therapy, a stereotactic biopsy should be performed to identify of *T. cruzi* amastigotes in all brain specimens, to confirm the presence of parasites by using the immunohistochemical method and to differentiate reactivated Chagas' disease from CT¹⁰⁶. Therefore, parasitological studies and histopathology are the confirmatory diagnostic tools for chagasic encephalitis in HIV-infected patients¹⁰⁴.

Serological diagnosis - ELISA, indirect immunofluorescence, and indirect haemagglutination are the most common serological methods used to detect antibodies levels indicating the latent or recent infections with *T. cruzi*. The reactive results are usually obtained from at least two or three serological assays¹⁰⁷ and the negative result does not exclude the diagnosis especially for disease reactivation¹⁰⁸.

Neuroimaging diagnosis: Neuroimaging finding is the initial diagnosis especially for patients with chagasic encephalitis. The characteristic of space occupying lesions are found to be similar as to those of CT^{98,107}. Based on the lesion location, it can suggest the aetiological agent. CAT scan typically shows hypodense, single or multiple, irregular ring enhancing lesions predominantly located in the white matter of the brain lobes¹⁰⁸. However, the neuroimaging finding might be normal in a few cases¹⁰². MRI demonstrates a heterogenous expansive lesion with mass effect, hypointense in T1 and hyperintense in T2 and FLAIR sequences, with irregular or ring enhancement¹⁰⁶. Although Chagas' disease and toxoplasmosis rarely co-exist in the same patient, PCR or microscopy is essential in differentiating these two pathogens and in preventing disease progression as well as fatal outcome of these patients¹⁰⁹.

Molecular diagnosis - PCR is a useful tool for the detection of *T. cruzi* DNA from various biological specimens: peripheral blood for the early diagnosis of reactivated Chagas' disease¹¹⁰ and for evaluation of the efficacy of anti-*T. cruzi* therapy¹¹¹. The real-time PCR system is considered as an alternative to conventional PCR techniques as it is simpler, faster and more reliable. Moreover, the reduced risk of contamination and the possibility of quantification are added values to this method¹¹².

Treatment: In HIV-infected patients, benznidazole 5 mg/kg daily divided in two doses for 60-90 days is recommended treatment for reactivated Chagas' disease^{103,113} and nifurtimox 8-10 mg/kg daily divided in three doses for 60-120 days is an alternative option even though clinical experience is generally not encouraging¹⁰³. After resolution of the CNS Chagas' disease, benznidazole 5 mg/kg given three times

per week is recommended for a lifetime secondary prophylaxis⁵⁷. The exacerbation of HIV viral load simultaneous with asymptomatic reactivation of chronic Chagas' disease¹¹⁴. In HIV patients with seropositive for *T. cruzi*, it might be beneficial to administer a single course of treatment with benznidazole and nifurtimox. ART should be systematically initiated in Chagas/HIV-coinfected patients to further control parasites^{103,113}, reconstitute the immune system, and maintain high level of CD4⁺ cell count, hence reducing the incidence of reactivated Chagas' disease^{104,106}.

Trypanosomiasis: African trypanosomiasis

African trypanosomiasis (sleeping sickness) is caused by Trypanosoma brucei gambiense and Trypanosoma brucei rhodesiense, a haemoflagellate, extracellular and zoonotic protozoan parasite. These two parasite forms exist only in the lymph vessel, bloodstream and the CSF. The clinical course of African sleeping sickness consists of two stages: the early haemolymphatic, trypanosomal chancre and winterbottom's sign, and the CNS (encephalitic) stage, diversity of neurological deficits and severe condition. Human African trypanosomiasis (HAT) is a potentially fatal disease and continues to pose a major threat to 60 million people in 36 well defined regions of sub-Saharan Africa where HAT is endemic. It is estimated that there are currently 300,000-500,000 cases, with 50,000 deaths yearly¹¹⁵. It remains unclear whether HIV influences the clinico-epidemiology of HAT although both infections are endemic in sub-Saharan Africa and co-infections are also common¹¹⁶.

Diagnosis: Definite diagnosis is based on the direct detection of trypanosomes from various clinical samples stained smears such as, peripheral blood, CSF or tissue biopsy¹¹⁷. The demonstration of trypomastigotes of T. gambiense is more difficult than T. rhodesiense infections because of low parasite load in the blood as well as CSF samples¹¹⁸. In acute phase, the diagnosis can be done by demonstration of trypanosomes in a chancre or the site bitten by an infected tsetse fly. Microscopic examination is not sensitive to detect the presence of parasites during the late-stage of clinical disease. Serology and molecular (PCR assays) diagnosis are neither sensitive nor specific as compared to direct detection of the parasites through a conventional microscopy. However, a combined method of PCR assays and microscopy is recommended using CSF sample in highly clinical suspicion cases of HAT¹¹⁹.

Neuroimaging diagnosis - In CAT scan, no specific findings have been reported. MRI scan is often used to demonstrate the abnormalities in the brain¹²⁰: focal high signal abnormalities in the white matter, especially with T2 sequences. This appearance is resolved after treatment¹²¹.

Treatment: HAT is a life-threatening disease with high mortality rate (2%-7%) and uniformly fatal if left untreated. Pentamidine, suramin, melasoprol, effornithine, and benznidazole are the regimen uses for treatment of this parasitic disease. These drugs are recommended once confirmation of diagnosis is achieved. In cases of HAT with confirmed or suspected CNS involvement, melarsoprol should be administered immediately. This agent is potentially effective for both T. gambiense and T. rhodesiense infections, nevertheless, the emergence of melasoprol resistance has been reported¹²². If melasoprol fails, effornithine can be used to treat CNS infection by T. gambiense but not effective for T. rhodesiense¹²³. Suramin and pentamidine isethionate are recommended therapies for the early-stage¹²⁴ and systemic HAT but not effective against CNS infection by T. gambiense and T. rhodesiense¹²⁵. Moreover, treatment with suramin prevents HIV from penetrating CD4⁺ T cells and then inhibits reverse-transcriptase activity¹²⁵. HAT/HIV coinfected patients are at higher risk for treatment failure and unfavourable outcome than seronegative HIV patients126.

Immune reconstitution inflammatory syndrome (IRIS) associated with parasitic infections

ART partially restores the immune function of HIV-infected patients, thereby remarkably reducing morbidity and mortality of OIs in general and parasitic pathogen in particular. The incidence of parasitic diseases such as leishmaniasis35,127, cryptosporidiosis, and toxoplasmosis^{32,128} has been decreased especially in areas where ART, including HAART, is accessible. HAART has significantly reduced secondary reactivation and relapse in cases of toxoplasmosis and has improved survival in these HIV-infected patients. This is certainly due to the successful suppression of virus replications followed by an increase in CD4⁺ lymphocytes, a partial recovery of T-cell specific immune responses and decreased susceptibility to both local and systemic opportunistic pathogens¹²⁹. The IRIS constitutes a clinical deterioration of OIs in HIV-infected patients as a result of the enhancement of pathogen-specific immune responses due to the

use of HAART. IRIS has been widely recognized in associating with parasitic diseases following initiation of HAART and development of a paradoxical clinical deterioration despite an increased CD4⁺ cell count and decreased HIV viral load which leads to the rapid restoration of the immune system. So far, more than 20 cases of CT associated IRIS have been reported in the literature (Table). A low CD4⁺ cell count has been identified as a significant risk factor in AIDS patients with CT^{93,130-134} due to impaired proliferative response to Toxoplasma antigen135, a decreased production of interferon γ^{136} , and IRIS develops more commonly in HIV-infected patients¹³⁷. Therefore, monitoring of CD4⁺/CD8⁺ T cells in patients on HAART might serve as a better marker for the restoration of T. gondii-specific immune responses than the total number of CD4⁺ cells count¹³⁸. Immune reconstitution under HAART has been associated with a restoration of immune responses against T. gondii¹³⁹. In the case w here IRIS is suspected in CT patient, close observation for 7-15 days, a higher steroid dose to control IRIS¹⁴⁰, uninterrupted HAART, and continued treatment for toxoplasmosis can resolve this problem without biopsy¹⁴¹. Based on reported cases of CT-associated IRIS from different studies, this could verify its association that it can develop in a substantial numbers of HIV-infected patients receiving HAART. No case of IRIS-related toxoplasmosis has ever been reported among AIDS patients in Malaysia even though CT was one of the most common systemic OIs in AIDS patients^{78,128}. The infection with *Leishmania* spp. occurs anywhere between 10 to 100 times the expected rate, found in patients co-infected with HIV¹⁴². Three clinical variations of leishmaniasis with cutaneous findings in association with IRIS have been reported: diffuse cutaneous leishmaniasis, post-or para-kala-azar dermal leishmaniasis, sporotrichoid and subcutaneous nodules. The reports of unmasking and paradoxical reactions are also detected in these patients. Diffuse mucocutaneous leishmaniasis (New World disease) has been described in three cases from Brazil and one case in a Nicaraguan immigrant to the USA¹⁴³⁻¹⁴⁵. From the Old World, seven cases of IRIS-post-kala-azar dermal leishmaniasis have been reported¹⁴⁶⁻¹⁵¹.

Among intestinal soil-transmitted helminthes, IRIS-associated with strongyloidiasis caused by *S. stercoralis* has been reported as a new and unexpected presentation in HIV-infected patients. The first case of IRIS-associated strongyloidiasis was reported in a patient with fever, eosinophilia and hepatitis¹⁵². Another two cases of disseminated strongyloidiasis

		d as immune reconstitution disease associated with parasitic infections.	
Countries	No. of cases	Clinical presentation	Baseline CD4 ⁺ cell count/µl
Asia:			
India ¹⁵⁷	1 (child)	Cerebral toxoplasmosis	-
Israel ¹⁴⁸	1	Post kala-azar dermal leishmaniasis	150
Thailand ¹⁵⁸	2	Cerebral toxoplasmosis	-
Australia and New Zealand:			
Australia ¹⁵⁹	1	Visceral leishmaniasis	55
Europe:			
France ¹³⁴	1	Placental IRIS	7 (pregnancy)
France ¹⁶⁰	1	Uveitis	4
France ¹⁶¹	2	Visceral leishmaniasis	15, 186
France ¹⁶²	1	Sporotrichoid nodules-cutaneous leishmaniasis	150
France ¹⁶³	3	Cerebral toxoplasmosis	9, 23, 25
Germany ¹⁵¹	1	Para-kala-azar dermal leishmaniasis	-
Italy ⁴⁴	1	Post kala-azar dermal leishmaniasis	-
Italy ¹⁴⁷	1	Post kala-azar dermal leishmaniasis	35
Serbia & Montenegro ¹³⁷	1	Cerebral toxoplasmosis	<100
Spain ¹⁵⁶	1	Terminal ileitis (Cryptosporidium spp.)	10
Spain ¹⁶⁴	3	Cerebral toxoplasmosis	-
Spain ¹⁶⁵	3	Visceral leishmaniasis	30, 39, 94
Spain ¹⁶⁶	1	Cerebral toxoplasmosis	456
Switzerland ¹³²	1	Immune recovery vitritis with isolated toxoplasmic retinochoroiditis	11
The Netherlands ¹³¹	1	Cerebral toxoplasmosis	43
The Netherland ¹⁴⁹	2	Post-kala-azar dermal leishmaniasis	-
UK ¹⁶⁷	1	Colitis and polyposis with Schistosoma mansoni	-
UK ¹⁵³	1	Disseminated strongyloidiasis	135
UK ¹⁵⁴	1	Disseminated strongyloidiasis	30
UK ¹⁵⁵	1	Strongyloides stercoralis	96
UK ¹⁶⁸	1	Eosinophilia with Schistosoma mansoni	170
UK ¹⁶⁹	1	Eosinophilic enteritis with Schistosoma mansoni	170
North America and Canada:			
Canada ¹⁷⁰	1	Cerebral toxoplasmosis	2
USA ⁹³	1	Toxoplasmic myelopathy and myopathy	67
USA ¹³⁰	1	Cerebral toxoplasmosis	83
USA ¹³³	1	Cerebral toxoplasmosis	29
USA ¹⁴¹	1	Cerebral toxoplasmosis	14
USA^{144}	1	Diffuse cutaneous leishmaniasis	-
USA ¹⁵²	1	Eosinophilia with Strongyloides stercoralis	32
USA^{171}	1	Neurocysticercosis	103
USA ¹⁷²	1	Visceral leishmaniasis	-
South America:			
Brazil ¹⁴³	2	Cutaneous-mucosal leishmaniasis	23, 38
Brazil ¹⁴⁵	1	Tegumentary leishmaniasis	46
Brazil ¹⁷³	1	Cerebral toxoplasmosis	276
South Africa:			
Ethiopia ¹⁵⁰	1	Post-kala-azar dermal leishmaniasis	-
Ethiopia ¹⁷⁴	2	Cerebral toxoplasmosis	-
Ethiopia ¹⁷⁵	7	Cerebral toxoplasmosis	50-100

were reported in patients with Escherichia coli bacteraemia, eosinophilia and pneumonitis¹⁵³ and E. coli meningitis¹⁵⁴. The fourth case was described in a patient with Strongyloides enteritis with thrombocytopenia¹⁵⁵. IRIS-associated terminal ileitis caused by Cryptosporidium spp., the first case report of intestinal protozoan infection, was described in a patient with abdominal symptoms of watery, bloodless diarrhoea and epigastric pain¹⁵⁶. As for the increasing use of HAART worldwide, the care for patients receiving HAART will need to incorporate monitoring for and treating complications of IRIS including impaired CD4⁺ cell immune reconstitution upon HIV therapy in patients with opportunistic parasitic infections. In addition, as the number of HIV-associated parasitic diseases cases treated with HAART increases, the complications of IRIS-associated parasitic diseases may become more common and easier to recognize, particularly in areas where tropical parasitic infections are endemic.

Conclusion

POIs remain a major concern in HIV- infected individuals and are unlikely to be eradicated from these patients. Existing parasitic infections still occur in those not yet diagnosed with HIV or not in medical care, those not receiving prophylaxis, and those not taking or not responding to HAART. In addition, there are more emerging POIs such as onchocerciasis or lymphatic filariasis in HIV-infected patients that need to be closely monitored and further studies are recommended to verify any significant association with HIV particularly in tropical areas of endemicity. In most developing countries where ART is fairly limited, HIV-infected patients are certainly at high risk for POIs in general and CT in particular such as in China, India, South America, Southeast Asian region and most importantly sub-Saharan Africa. A better understanding of the clinico-epidemiology of POIs, and improved efforts in prevention, diagnosis and treatment, are urgently needed. The role of OIs with parasites require further studies, including as to whether infections, such as `cerebral toxoplasmosis have an impact on HIV/ AIDS patients.

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References

- 1. UNAIDS report on the global AIDS epidemic 2010 [homepage on the Internet]. Epidemic update. Available from: *http://www. unaids.org/documents/20101123_GlobalReport_Chap2_ em.pdf*, accessed on August 25, 2011.
- CDC. Revision of the CDC surveillance case definition for acquired immunodeficiency syndrome. *MMWR Surveill Summ* 1987; 36: 1-15.
- Akinbo FO, Okaka CE, Omoregie R. Prevalence of intestinal parasitic infections among HIV patients in Benin City, Nigeria. *Libyan J Med* 2010; 5 -doi: 10.3402/ljm.v5i0.5506.
- Mohandas, Sehgal R, Sud A, Malla N. Prevalence of intestinal parasitic pathogens in HIV-seropositive individuals in Northern India. *Jpn J Infect Dis* 2002; 55: 83-4.
- Fontanet AL, Sahlu T, Rinke de Wit T, Messele T, Masho W, Woldemichael T, *et al.* Epidemiology of infections with intestinal parasites and human immunodeficiency virus (HIV) among sugar-estate residents in Ethiopia. *Ann Trop Med Parasitol* 2000; 94 : 269-78.
- Lindo JF, Dubon JM, Ager AL, de Gourville EM, Solo-Gabriele H, Klaskala WI, *et al.* Intestinal parasitic infections in human immunodeficiency virus (HIV)-positive and HIVnegative individuals in San Pedro Sula, Honduras. *Am J Trop Med Hyg* 1998; 58: 431-5.
- Omalu ICJ, Yako AB, Duhlinska DD, Anyanwu GI, Pam VA, Inyama PU. First detection of intestinal microsporidia in Nigeria. *Online J Health All Sci* 2005; 3:4.
- Kulkarni SV, Kairon R, Sane SS, Padmawar PS, Kale VA, Thakar MR, *et al.* Opportunistic parasitic infections in HIV/ AIDS patients presenting with diarrhoea by the level of immunesuppression. *Indian J Med Res* 2009; *131*: 63-6.
- Asma I, Johari S, Sim BHL, Lim YAL. How common is intestinal parasitism in HIV-infected patients in Malaysia? *Trop Biomed* 2011; 28: 400-10.
- Siddiqui U, Bini EJ, Chandarana K, Leong J, Ramsetty S, Schiliro D, *et al.* Pravalence and impact of diarrhea on healthrelated quality of life in HIV-infected patients in the era of highly active antiretroviral therapy. *J Clin Gastroenterol* 2007; *41*: 484-90.
- Lagrange-Xélot M, Porcher R, Sarfati C, de Castro N, Carel O, Magnier JD, *et al.* Isosporiasis in patients with HIV infection in the highly active antiretroviral therapy era in France. *HIV Med* 2008; 9: 126-30.
- 12. Chhin S, Harwell JI, Bell JD, Rozycki G, Ellman T, Barnett JM, *et al.* Etiology of chronic diarrhea in antiretroviral-naive patients with HIV infection admitted to Norodom Sihanouk Hospital, Phnom Penh, Cambodia. *Clin Infect Dis* 2006; *43* : 925-32.
- 13. Kosek M, Alcantara C, Lima AA, Guerrant RL. Cryptosporidiosis: an update. *Lancet Infect Dis* 2001; *1* : 262-9.

- Wiwanitkit V. Intestinal parasitic infections in Thai HIVinfected patients with different immunity status. BMC Gastroenterol 2001; 1:3.
- Leelayoova S, Vithayasai N, Watanaveeradej V, Chotpitayasunondh T, Therapong V, Naaglor T, et al. Intestinal microsporidiosis in HIV-infected children with acute and chronic diarrhea. Southeast Asian J Trop Med Public Health 2001; 32: 33-7.
- Wanachiwanawin D, Chokephaibulkit K, Lertlaituan P, Ongrotchanakun J, Chinabut P, Thakerngpol K. Intestinal microsporidiosis in HIV-infected children with diarrhea. *Southeast Asian J Trop Med Public Health* 2002; 33 : 241-5.
- Sokolova OI, Demyanov AV, Bowers LC, Didier ES, Yakovlev AV, Skarlato SO, *et al.* Emerging microsporidian infections in Russian HIV-infected patients. *J Clin Microbiol* 2011; 49: 2102-8.
- 18. Didier ES, Weiss LM. Microsporidiosis: not just in AIDS patients. *Curr Opin Infect Dis* 2011; 24 : 490-5.
- 19. Stauffer W, Radvin JI. Entamoeba histolytica: an update. Curr Opin Infect Dis 2003; 16: 479-85.
- Haque R, Huston CD, Hughes M, Houpt E, Petri WA. Amebiasis. N Engl J Med 2003; 348: 1565-73.
- Hung CC, Wu PY, Chang SY, Ji DD, Sun HY, Liu WC, et al. Amebiasis among persons who sought voluntary counseling and testing for human immunodeficiency virus infection: a case-control study. *Am J Trop Med Hyg* 2011; 84: 65-9.
- 22. Chen Y, Zhang Y, Yang B, Qi T, Lu H, Cheng X, *et al.* Seroprevalence of *Entamoeba histolytica* infection in HIV-Infected patients in China. *Am J Trop Med Hyg* 2007; 77 : 825-8.
- 23. Hung CC, Wu PY, Chang SY, Ji DD, Sun HY, Liu WC, et al. Amebiasis among persons who sought voluntary counseling and testing for human immunodeficiency virus infection: a case control study. Am J Trop Med Hyg 2011; 84 : 65-9.
- Lee YJ, Chen KJ, Yang CH, Lin YC, Liu HY, Liao ST, et al. Characteristics of amebic liver abscess in patients with or without human immunodeficiency virus. J Microbiol Immunol Infect 2009; 42: 500-4.
- Wu KS, Tsai HC, Lee SS, Liu YC, Wann SR, Wang YH, et al. Comparison of clinical characteristics of amebic liver abscess in human immunodeficiency virus (HIV)-infected and non-HIV-infected patients. J Microbiol Immunol Infect 2008; 41: 456-61.
- Feitosa G, Bandeira AC, Sampaio DP, Badaró R, Brites C. High prevalence of giardiasis and stronglyloidiasis among HIV-infected patients in Bahia, Brazil. *Braz J Infect Dis* 2001; 5: 339-44.
- 27. Chui DW, Owen RL. AIDS and the gut. J Gastroenterol Hepatol 1994; 9: 291-303.
- Hailemariam G, Kassu A, Abebe G, Abate E, Damte D, Mekonnen E, *et al.* Intestinal parasitic infections in HIV/ AIDS and HIV seronegative individuals in a teaching hospital, Ethiopia. *Jpn J Infect Dis* 2004; *57* : 41-3.
- Angarano G, Maggi P, Di Bari MA, Larocca AM, Congedo P, Di Bari C, *et al.* Giardiasis in HIV: A possible role in patients with severe immune deficiency. *Eur J Epidemiol* 1997; *13*: 485-7.

- Phillips MA, Stanley Jr SL. Chemotherapy of protozoal infections: Amebiasis, giardiasis, trichomoniasis, trypanosomiasis, leishmaniasis, and other protozoal infections. In: Brunton LL, Chabner BA, Knollmann BC, editors. Goodman & Gilman's the pharmacological basis of therapeutics, 12th ed. New York: McGraw-Hill; 2001.
- Fujishima T, Nishise S, Ichihara M, Kobayashi S, Takeuchi T. Difficulties in the treatment of intestinal amoebiasis in mentally disabled individuals at a rehabilitation institution for the intellectually impaired in Japan. *Chemotherapy* 2010; *56* : 348-52.
- 32. Wiwanitkit V, Srisupanant M. Cryptosporidiosis occurrence in anti-HIV-seropositive patients attending a sexually transmitted diseases clinic, Thailand. *Trop Doct* 2006; *36* : 64.
- 33. WHO. Control of the leishmaniases. *WHO Technical Report* Series 2010; 949 : 1-104.
- Alvar J, Bashaye S, Argaw D, Cruz I, Aparicio P, Kassa A, et al. Kala-azar outbreak in Libo Kemkem, Ethiopia: epidemiologic and parasitologic assessment. Am J Trop Med Hyg 2007; 77 : 275-82.
- Alvar J, Aparicio P, Aseffa A, Den Boer M, Cañavate C, Dedet JP, *et al.* The relationship between leishmaniasis and AIDS: the second 10 years. *Clin Microbiol Rev* 2008; 21: 334-59.
- Reithinger R, Dujardin JC, Louzir H, Pirmez C, Alexander B, Brooker S. Cutaneous leishmaniasis. *Lancet Infect Dis* 2007; 7: 581-96.
- Paredes R, Munoz J, Diaz I, Domingo P, Gurgui M, Clotet B. Leishmaniasis in HIV infection. *J Postgrad Med* 2003; 49 : 39-49.
- Guiguemdé RT, Sawadogo OS, Bories C, Traore KL, Nezien D, Nikiema L, et al. Leishmania major and HIV co-infection in Burkina Faso. Trans R Soc Trop Med Hyg 2003; 97: 168-9.
- Rabello A, Orsini M, Disch J. Leishmania/HIV co-infection in Brazil: an appraisal. Ann Trop Med Parasitol 2003; 97 (Suppl 1)17-28.
- Chakravarty J, Kumar S, Kumar R, Gautam S, Rai M, Sundar S. Evaluation of rk39 immunochromatographic test with urine for diagnosis of visceral leishmaniasis. *Trans R Soc Trop Med Hyg* 2011; *105* : 537-9.
- 41. Desjeux P. Global control and *Leishmania* HIV co-infection. *Clin Dermatol* 1999; *17* : 317-25.
- Calza L, D'Antuono A, Marinacci G, Manfredi R, Colangeli V, Passarini B, *et al.* Disseminated cutaneous leishmaniasis after visceral disease in a patient with AIDS. *J Am Acad Dermatol* 2004; *50*: 461-5.
- Schönian G, Nasereddin A, Dinse N, Schweynoch C, Schallig HD, Presber W, *et al.* PCR diagnosis and characterization of *Leishmania* in local and imported clinical samples. *Diagn Microbiol Infect Dis* 2003; 47 : 349-58.
- 44. Antinori S, Longhi E, Bestetti G, Piolini R, Acquaviva V, Foschi A, et al. Post-kala-azar dermal leishmaniasis as an immune reconstitution inflammatory syndrome in a patient with acquired immune deficiency syndrome. Br J Dermatol 2007; 157 : 1032-6.
- 45. Ritmeijer K, Veeken H, Melaku Y, Leal G, Amsalu R, Seaman J, et al. Ethiopian visceral leishmaniasis: generic and proprietary sodium stibogluconate are equivalent; HIV coinfected patients have a poor outcome. Trans R Soc Trop Med Hyg 2001; 95: 668-72.

- 46. Rosenthal E, Marty P. Recent understanding in the treatment of visceral leishmaniasis. *J Postgrad Med* 2003; 49 : 61-8.
- Davidson RN, Russo R. Relapse of visceral leishmaniasis in patients who were coinfected with human immunodeficiency virus and who received treatment with liposomal amphotericin B. *Clin Infect Dis* 1994; *19* : 560.
- WHO 2009. Reported malaria cases and deaths [cited 2011 Aug 25] Available from: http://www.who.int/malaria/world_ malaria_report_2010/wmr2010_annex7a.pdf, accessed on August 25, 2011.
- 49. Idemyor V. Human immunodeficiency virus (HIV) and malaria interaction in sub-Saharan Africa: the collision of two Titans. *HIV Clin Trials* 2007; 8 : 246-53.
- 50. Cruciani M, Nardi S, Malena M, Bosco O, Serpelloni G, Mengoli C. Systematic review of the accuracy of the *Para*Sight-F test in the diagnosis of *Plasmodium falciparum* malaria. *Med Sci Monit* 2004; *10* : MT81-8.
- 51. Patankar TF, Karnad DR, Shetty PG, Desai AP, Prasad SR. Adult cerebral malaria: prognostic importance of imaging findings and correlation with postmortem findings. *Radiology* 2002; *224* : 811-6.
- Cordoliani YS, Sarrazin JL, Felten D, Caumes E, Lévêque C, Fisch A. MR of cerebral malaria. *AJNR Am J Neuroradiol* 1998; 19: 871-4.
- Mohsen AH, McKendrick MW, Schmid ML, Green ST, Hadjivassiliou M, Romanowski C. Postmalaria neurological syndrome: a case of acute disseminated encephalomyelitis? *J Neurol Neurosurg Psychiatry* 2000; 68 : 388-9.
- WHO. 2008. Essential prevention and care interventions for adults and adolescents living with HIV in resource-limited settings. Available from: http://www.who.int/hiv/pub/prev_ care/OMS_EPP_AFF_en.pdf, accessed on December 6, 2011.
- 55. Garcia HH, Pretell J, Gilman RH; Cysticercosis Working Group in Peru. Neurocysticercosis and the global world. *J Neurol* 2002; 249 : 1107-8.
- Garcia HH, Del Brutto OH; Cysticercosis Working Group in Peru. Neurocysticercosis: updated concepts about an old disease. *Lancet Neurol* 2005; 4: 653-61.
- 57. Walker M, Zunt JR. Parasitic central nervous system infections in immunocompromised hosts. *Clin Infect Dis* 2005; *40* : 1005-15.
- Takayanagui OM, Odashima NS, Bonato PS, Lima JE, Lanchote VL. Medical management of neurocysticercosis. *Expert Opin Pharmacother* 2011; 12: 2845-56.
- 59. WHO Expert Committee. Prevention and control of schistosomiasis and soil-transmitted helminthiasis. *World Health Organ Tech Rep Ser* 2002; *912* : i-vi, 1-57.
- Ross AG, Bartley PB, Sleigh AC, Olds GR, Li Y, Williams GM, et al. Schistosomiasis. N Engl J Med 2002; 346 : 1212-20.
- Smith C, Smith H, Seaton RA, Fox R. Seroprevalence of schistosomiasis in African patients infected with HIV. *HIV Med* 2008; 9: 436-9.
- 62. de Avila RE, Batista TS, Xavier MA, Nogueira AM, Lambertucci JR. *Schistosomiasis mansoni* and severe gastrointestinal cytomegalovirus disease in a patient with

acquired immunodeficiency syndrome. *Rev Soc Bras Med Trop* 2006; *39* : 379-82.

- Carod-Artal FJ. Neuroschistosomiasis. Expert Rev Anti Infect Ther 2010; 8: 1307-18.
- Carod-Artal FJ. Neurological complications of Schistosoma infection. Trans R Soc Trop Med Hyg 2008; 102: 107-16.
- Ferrari TC. Involvement of central nervous system in the schistosomiasis. *Mem Inst Oswaldo Cruz* 2004; 99 (5 Suppl 1): 59-62.
- 66. Zhu YC. Immunodiagnosis and its role in schistosomiasis control in China: a review. *Acta Trop* 2005; *96* : 130-6.
- 67. Van Gool T, Vetter H, Vervoort T, Doenhoff MJ, Wetsteyn J, Overbosch D. Serodiagnosis of imported schistosomiasis by a combination of a commercial indirect hemagglutination test with *Schistosoma mansoni* adult worm antigens and an enzyme-linked immunosorbent assay with *S. mansoni* egg antigens. *J Clin Microbiol* 2002; *40* : 3432-7.
- Fowler R, Lee C, Keystone JS. The role of corticosteroids in the treatment of cerebral schistosomiasis caused by *Schistosoma mansoni*: case report and discussion. *Am J Trop Med Hyg* 1999; *61*: 47-50.
- 69. Liu HQ, Feng XY, Yao ZW, Sun HP. Characteristic magnetic resonance enhancement pattern in cerebral schistosomiasis. *Chin Med Sci J* 2006; *21* : 223-7.
- Ferrari ML, Coelho PM, Antunes CM, Tavares CA, da Cunha AS. Efficacy of oxamniquine and praziquantel in the treatment of *Schistosoma mansoni* infection: a controlled trial. *Bull World Health Organ* 2003; *81* : 190-6.
- Xiao SH, You JQ, Gao HF, Mei JY, Jiao PY, Chollet J, et al. Schistosoma japonicum: effect of artemether on glutathione S-transferase and superoxide dismutase. Exp Parasitol 2002; 102: 38-45.
- 72. Karanja DM, Hightower AW, Colley DG, Mwinzi PN, Galil K, Andove J, *et al.* Resistance to reinfection with *Schistosoma mansoni* in occupationally exposed adults and effect of HIV-1 co-infection on susceptibility to schistosomiasis: a longitudinal study. *Lancet* 2002; 360 : 592-6.
- Rivero FD, Kremer LE, Allende L, Casero RD. Strongyloides stercoralis and HIV: a case report of an indigenous disseminated infection from non-endemic area. Rev Argent Microbiol 2006; 38 : 137-9.
- Viney ME, Brown M, Omoding NE, Bailey JW, Gardner MP, Roberts E, *et al.* Why does HIV infection not lead to disseminated strongyloidiasis? *J Infect Dis* 2004; *190*: 2175-80.
- Keiser PB, Nutman TB. Strongyloides stercoralis in the Immunocompromised Population. Clin Microbiol Rev 2004; 17: 208-17.
- Walker MD, Zunt JR. Neuroparasitic infections: nematodes. Semin Neurol 2005; 25 : 252-61.
- Siddiqui AA, Berk SL. Diagnosis of Strongyloides stercoralis infection. Clin Infect Dis 2001; 33 : 1040-7.
- Nissapatorn V, Lee C, Quek KF, Leong CL, Mahmud R, Abdullah KA. Toxoplasmosis in HIV/AIDS patients: a current situation. *Jpn J Infect Dis* 2004; 57 : 160-5.

- Israeski DM, Remington JS. AIDS-associated toxoplasmosis. In: Sande MA, Volberding PA, editors. *The medical management of AIDS*. Philadelphia, WB Saunders; 1992. p. 319-45.
- Carme B, M'Pele P, Mbitsi A, Kissila AM, Aya GM, Mouanga-Yidika G, *et al.* Opportunistic parasitic diseases and mycoses in AIDS. Their frequencies in Brazzaville (Congo). *Bull Soc Pathol Exot Filiales* 1988; 81 : 311-6.
- Colombo FA, Vidal JE, Penalva de Oliveira AC, Hernandez AV, Bonasser-Filho F, Nogueira RS, *et al.* Diagnosis of cerebral toxoplasmosis in AIDS patients in Brazil: importance of molecular and immunological methods using peripheral blood samples. *J Clin Microbiol* 2005; *43*: 5044-7.
- Vidal JE, Hernandez AV, de Oliveira AC, Dauar RF, Barbosa SP Jr, Focaccia R. Cerebral toxoplasmosis in HIV-positive patients in Brazil: clinical features and predictors of treatment response in the HAART era. *AIDS Patient Care STDS* 2005; *19*: 626-34.
- Pereira-Chioccola VL, Vidal JE, Su C. *Toxoplasma gondii* infection and cerebral toxoplasmosis in HIV-infected patients. *Future Microbiol* 2009; 4 : 1363-79.
- Luft BJ. Toxoplasma gondii. In: Walzer PD, Gertz R, editors. Parasitic infection in the compromised host. New York: Marcel Dekker; 1989. p. 179-279.
- 85. Luft BJ, Remington JS. Toxoplasmic encephalitis in AIDS. *Clin Infect Dis* 1992; 15 : 211-22.
- 86. Reischl U, Bretagne S, Krüger D, Ernault P, Costa JM. Comparison of two DNA targets for the diagnosis of Toxoplasmosis by real-time PCR using fluorescence resonance energy transfer hybridization probes. *BMC Infect Dis* 2003; 3:7.
- Antinori A. Evaluation and management of intracranial mass lesions in AIDS: Report of the Quality Standard Subcommittee of the American Academy of Neurology. *Neurology* 1998; *51*: 1233-4.
- Yeo KK, Yeo TT, Chan CY, Sitoh YY, Teo J, Wong SY. Stereotactic brain biopsies in AIDS patients-early local experience. *Singapore Med J* 2000; *41* : 161-6.
- Sadler M, Brink NS, Gazzard BG. Management of intracerebral lesions in patients with HIV: a retrospective study with discussion of diagnostic problems. *QJM* 1998; *91*: 205-17.
- Armbruster C, Alesch F, Budka H, Kriwanek S. Stereotactic brain biopsy in AIDS patients: a necessary patient-oriented and cost-effective diagnostic measure?. *Acta Med Austriaca* 1998; 25 : 91-5.
- Cota GF, Assad EC, Christo PP, Giannetti AV, Santos Filho JA, Xavier MA. Ventriculitis: a rare case of primary cerebral toxoplasmosis in AIDS patient and literature review. *Braz J Infect Dis* 2008; 12: 101-4.
- Portegies P, Solod L, Cinque P, Chaudhuri A, Begovac J, Everall I, *et al*. Guidelines for the diagnosis and management of neurological complications of HIV infection. *Eur J Neurol* 2004; *11*: 297-304.
- Kung DH, Hubenthal EA, Kwan JY, Shelburne SA, Goodman JC, Kass JS. Toxoplasmosis myelopathy and myopathy in an

AIDS patient: a case of immune reconstitution inflammatory syndrome? *Neurologist* 2011; *17* : 49-51.

- 94. Chirgwin K, Hafner R, Leport C, Remington J, Andersen J, Bosler EM, et al. Randomized phase II trial of atovaquone with pyrimethamine or sulfadiazine for treatment of toxoplasmic encephalitis in patients with acquired immunodeficiency syndrome: ACTG 237/ANRS 039 Study. AIDS Clinical Trials Group 237/Agence Nationale de Recherche sur le SIDA, Essai 039. Clin Infect Dis 2002; 34 : 1243-50.
- 95. Baggish AL, Hill DR. Antiparasitic agent atovaquone. Antimicrob Agents Chemother 2002; 46 : 1163-73.
- Coura JR. Chagas disease: what is known and what is neededa background article. *Mem Inst Oswaldo Cruz* 2007; *102* (Suppl 1): 113-22.
- 97. Jackson Y, Myers C, Diana A, Marti HP, Wolff H, Chappuis F, et al. Congenital transmission of Chagas disease in Latin American immigrants in Switzerland. Emerg Infect Dis 2009; 15: 601-3.
- 98. Vaidian AK, Weiss LM, Tanowitz HB. Chagas' disease and AIDS. *Kinetoplastid Biol Dis* 2004; *3*:2.
- Antunes AC, Cecchini FM, Bolli FB, Oliveira PP, Reboucas RG, Monte TL, et al. Cerebral trypanosomiasis and AIDS. Arq Neuropsiquiatr 2002; 60: 730-3.
- 100. Sica RE, Gargiullo G, Papayanis C. Tumour-like chagasic encephalitis in AIDS patients: an atypical presentation in one of them and outcome in a small series of cases. Arq Neuropsiquiatr 2008; 66: 881-4.
- 101. Madalosso G, Pellini AC, Vasconcelos MJ, Ribeiro AF, Weissmann L, Oliveira Filho GS, *et al.* Chagasic meningoencephalitis: case report of a recently included AIDSdefining illness in Brazil. *Rev Inst Med Trop Sao Paulo* 2004; 46 : 199-202.
- 102. Cordova E, Boschi A, Ambrosioni J, Cudos C, Corti M. Reactivation of Chagas disease with central nervous system involvement in HIV-infected patients in Argentina, 1992-2007. Int J Infect Dis 2008; 12: 587-92.
- 103. Ferreira MS, Borges AS. Some aspects of protozoan infections in immunocompromised patients - a review. *Mem Inst Oswaldo Cruz* 2002; 97 : 443-57.
- 104. DiazGranados CA, Saavedra-Trujillo CH, Mantilla M, Valderrama SL, Alquichire C, Franco-Paredes C. Chagasic encephalitis in HIV patients: common presentation of an evolving epidemiological and clinical association. *Lancet Infect Dis* 2009; 9: 324-30.
- 105. Burgos JM, Begher SB, Freitas JM, Bisio M, Duffy T, Altcheh J, et al. Molecular diagnosis and typing of *Trypanosoma cruzi* populations and lineages in cerebral Chagas disease in a patient with AIDS. Am J Trop Med Hyg 2005; 73 : 1016-8.
- 106. Pittella JE. Central nervous system involvement in Chagas disease: a hundred-year-old history. *Trans R Soc Trop Med Hyg* 2009; *103*: 973-8.
- 107. Montero A, Cohen JE, Martínez DP, Giovannoni AG. Empirical anti-*toxoplasma* therapy in cerebral AIDS and Chagas disease. Presentation of 2 cases, review of the literature and proposal of an algorithm. Medicina (B Aires) 1998; 58 : 504-6.

- Corti M. AIDS and Chagas' disease. AIDS Patient Care STDS 2000; 14: 581-8.
- Barratt JL, Harkness J, Marriott D, Ellis JT, Stark D. Importance of nonenteric protozoan infections in immunocompromised people. *Clin Microbiol Rev* 2010; 23: 795-836.
- 110. Maldonado C, Albano S, Vettorazzi L, Salomone O, Zlocowski JC, Abiega C, *et al.* Using polymerase chain reaction in early diagnosis of re-activated *Trypanosoma cruzi* infection after heart transplantation. *J Heart Lung Transplant* 2004; *23* : 1345-8.
- 111. Diez M, Favaloro L, Bertolotti A, Burgos JM, Vigliano C, Lastra MP, *et al.* Usefulness of PCR strategies for early diagnosis of Chagas' disease reactivation and treatment follow-up in heart transplantation. *Am J Transplant* 2007; 7:1633-40.
- 112. Piron M, Fisa R, Casamitjana N, López-Chejade P, Puig L, Vergés M, *et al.* Development of a real-time PCR assay for *Trypanosoma cruzi* detection in blood samples. *Acta Trop* 2007; *103* : 195-200.
- 113. Corti M, Yampolsky C. Prolonged survival and immune reconstitution after chagasic meningoencephalitis in a patient with acquired immunodeficiency syndrome. *Rev Soc Bras Med Trop* 2006; *39* : 85-8.
- 114. Sartori AM, Caiaffa-Filho HH, Bezerra RC, do S Guilherme C, Lopes MH, Shikanai-Yasuda MA. Exacerbation of HIV viral load simultaneous with asymptomatic reactivation of chronic Chagas' disease. *Am J Trop Med Hyg* 2002; 67 : 521-3.
- 115. Kennedy PG. The continuing problem of human African trypanosomiasis (sleeping sickness). *Ann Neurol* 2008; *64* : 116-26.
- 116. Barat C, Pepin J, Tremblay MJ. HIV-1 replication in monocytederived dendritic cells is stimulated by melarsoprol, one of the main drugs against human African trypanosomiasis. *J Mol Biol* 2011; *410*: 1052-64.
- CDC. Laboratory diagnosis of parasites of public health concern. Atlanta, GA: CDC, 2003.
- 118. Lejon V, Boelaert M, Jannin J, Moore A, Büscher P. The challenge of *Trypanosoma brucei gambiense* sleeping sickness diagnosis outside Africa. *Lancet Infect Dis* 2003; *3* : 804-8.
- 119. Lejon V, Buscher P. Stage determination and follow-up in sleeping sickness. *Med Trop (Mars)* 2001; *61* : 355-60.
- Kennedy PG. Human African trypanosomiasis of the CNS: current issues and challenges. *J Clin Invest* 2004; *113*: 496-504.
- Gill DS, Chatha DS, del Carpio-O'Donovan R. MR imaging findings in African trypansomiasis. *AJNR Am J Neuroradiol* 2003; 24: 1383-5.
- 122. Matovu E, Seebeck T, Enyaru JC, Kaminsky R. Drug resistance in *Trypanosoma brucei* spp., the causative agents of sleeping sickness in man and nagana in cattle. *Microbes Infect* 2001; *3* : 763-70.
- Docampo R, Moreno SN. Current chemotherapy of human African trypanosomiasis. *Parasitol Res* 2003; *90* (Supp 1): S10-3.

- 124. Bouteille B, Oukem O, Bisser S, Dumas M. Treatment perspectives for human African trypanosomiasis. *Fundam Clin Pharmacol* 2003; *17*: 171-81.
- 125. Nok AJ. Arsenicals (melarsoprol), pentamidine and suramin in the treatment of human African trypanosomiasis. *Parasitol Res* 2003; *90* : 71-9.
- 126. Blum J, Nkunku S, Burri C. Clinical description of encephalopathic syndromes and risk factors for their occurrence and outcome during melarsopol treatment of human African trypanosomiasis. *Trop Med Int Health* 2001; 6: 390-400.
- 127. de La Rosa R, Pineda JA, Delgado J, Macías J, Morillas F, Mira JA, *et al.* Incidence of and risk factors for symptomatic visceral leishmaniasis among human immunodeficiency virus type 1-infected patients from Spain in the era of highly active antiretroviral therapy. *J Clin Microbiol* 2002; *40* : 762-7.
- 128. Lian YL, Heng BS, Nissapatorn V, Lee C. AIDSdefining illnesses: a comparison between before and after commencement of highly active antiretroviral therapy (HAART). *Curr HIV Res* 2007; *5* : 484-9.
- 129. Silva MT, Araújo A. Highly active antiretroviral therapy access and neurological complications of human immunodeficiency virus infection: impact versus resources in Brazil. *J Neurovirol* 2005; *11* (Suppl 3): 11-5.
- Tsambiras PE, Larkin JA, Houston SH. Case report. *Toxoplasma* encephalitis after initiation of HAART. *AIDS Read* 2001; *11*: 608-10.
- 131. de Boer J, Wulffraat N, Rothova A. Visual loss in uveitis of childhood. *Br J Ophthalmol* 2003; *87* : 879-84.
- 132. Sendi P, Sachers F, Drechsler H, Graber P. Immune recovery vitritis in an HIV patient with isolated toxoplasmic retinochoroiditis. *AIDS* 2006; 20 : 2237-8.
- 133. Chen KC, Chen JY, Tung GA. Case 149: Immune reconstitution inflammatory syndrome. *Radiology* 2009; *252*: 924-8.
- 134. Caby F, Lemercier D, Coulomb A, Grigorescu R, Paris L, Touafek F, *et al.* Fetal death as a result of placental immune reconstitution inflammatory syndrome. *J Infect* 2010; *61* : 185-8.
- 135. Belanger F, Derouin F, Grangeot-Keros L, Meyer L. Incidence and risk factors of toxoplasmosis in a cohort of human immunodeficiency virus-infected patients: 1988-1995. HEMOCO and SEROCO Study Groups. *Clin Infect Dis* 1999; 28: 575-81.
- 136. Ullum H, Victor J, Katzenstein TL, Gerstoft J, Gøtzsche PC, Bendtzen K, *et al.* Decreased short-term production of tumor necrosis factor-alpha and interleukin-1beta in human immunodeficiency virus-seropositive subjects. *J Infect Dis* 1997; *175* : 1507-10.
- 137. Jevtović DJ, Salemović D, Ranin J, Pesić I, Zerjav S, Djurković-Djaković O. The prevalence and risk of immune restoration disease in HIV-infected patients treated with highly active antiretroviral therapy. *HIV Med* 2005; 6 : 140-3.
- Furco A, Carmagnat M, Chevret S, Garin YJ, Pavie J, De Castro N, *et al.* Restoration of *Toxoplasma gondii*-specific immune responses in patients with AIDS starting HAART. *AIDS* 2008; 22 : 2087-96.

- 139. Fournier S, Rabian C, Alberti C, Carmagnat MV, Garin JF, Charron D, *et al.* Immune recovery under highly active antiretroviral therapy is associated with restoration of lymphocyte proliferation and interferon-gamma production in the presence of *Toxoplasma gondii* antigens. *J Infect Dis* 2001; *183* : 1586-91.
- Venkataramana A, Pardo CA, McArthur JC, Kerr DA, Irani DN, Griffin JW, *et al.* Immune reconstitution inflammatory syndrome in the CNS of HIV-infected patients. *Neurology* 2006; 67: 383-8.
- 141. Tremont-Lukats IW, Garciarena P, Juarbe R, El-Abassi RN. The immune inflammatory reconstitution syndrome and central nervous system toxoplasmosis. *Ann Intern Med* 2009; *150*: 656-7.
- Desjeux P, Alvar J. Leishmania/HIV co-infections: epidemiology in Europe. Ann Trop Med Parasitol 2003; 97 (Suppl 1): 3-15.
- 143. Posada-Vergara MP, Lindoso JA, Tolezano JE, Pereira-Chioccola VL, Silva MV, Goto H. Tegumentary leishmaniasis as a manifestation of immune reconstitution inflammatory syndrome in 2 patients with AIDS. *J Infect Dis* 2005; *192*:1819-22.
- 144. Sinha S, Fernández G, Kapila R, Lambert WC, Schwartz RA. Diffuse cutaneous leishmaniasis associated with the immune reconstitution inflammatory syndrome. *Int J Dermatol* 2008; 47 : 1263-70.
- 145. Chrusciak-Talhari A, Ribeiro-Rodrigues R, Talhari C, Silva RM Jr, Ferreira LC, Botileiro SF, et al. Tegumentary leishmaniasis as the cause of immune reconstitution inflammatory syndrome in a patient co-infected with human immunodeficiency virus and Leishmania guyanensis. Am J Trop Med Hyg 2009; 81: 559-64.
- 146. Neva FA. Biology and immunology of human strongyloidiasis. *J Infect Dis* 1986; *153* : 397-406.
- 147. Ridolfo AL, Gervasoni C, Antinori S, Pizzuto M, Santambrogio S, Trabattoni D, *et al.* Post-kala-azar dermal leishmaniasis during highly active antiretroviral therapy in an AIDS patient infected with *Leishmania infantum. J Infect* 2000; *40* : 199-202.
- 148. Gilad J, Borer A, Hallel-Halevy D, Riesenberg K, Alkan M, Schlaeffer F. Post-kala-azar dermal leishmaniasis manifesting after initiation of highly active anti-retroviral therapy in a patient with human immunodeficiency virus infection. *Isr Med Assoc J* 2001; 3 : 451-2.
- 149. Belay AD, Asafa Y, Mesure J, Davidson RN. Successful miltefosine treatment of post-kala-azar dermal leishmaniasis occurring during antiretroviral therapy. *Ann Trop Med Parasitol* 2006; *100* : 223-7.
- 150. Tadesse A, Hurissa Z. Leishmaniasis (PKDL) as a case of immune reconstitution inflammatory syndrome (IRIS) in HIV-positive patient after initiation of anti-retroviral therapy (ART). *Ethiop Med J* 2009; 47 : 77-9.
- 151. Gelanew T, Amogne W, Abebe T, Kuhls K, Hailu A, Schönian G. A clinical isolate of *Leishmania donovani* with ITS1 sequence polymorphism as a cause of para-kala-azar dermal leishmaniasis in an Ethiopian human immunodeficiency virus-positive patient on highly active antiretroviral therapy. *Br J Dermatol* 2010; *163*: 870-4.

- 152. Kim AC, Lupatkin HC. *Strongyloides stercoralis* infection as a manifestation of immune restoration syndrome. *Clin Infect Dis* 2004; *39* : 439-40.
- Lanzafame M, Faggian F, Lattuada E, Antolini D, Vento S. Strongyloidiasis in an HIV-1-infected patient after highly active antiretroviral therapy-induced immune restoration [letter]. J Infect Dis 2005; 191 : 1027.
- 154. Brown M, Miiro G, Nkurunziza P, Watera C, Quigley MA, Dunne DW, *et al. Schistosoma mansoni*, nematode infections, and progression to active tuberculosis among HIV-1-infected Ugandans. *Am J Trop Med Hyg* 2006; 74 : 819-25.
- 155. Taylor CL, Subbarao V, Gayed S, Ustianowski AP. Immune reconstitution syndrome to *Strongyloides stercoralis* infection. *AIDS* 2007; 21: 649-50.
- 156. Plasencia LD, Socas Mdel M, Valls RA, Fernández EM, Higuera AC, Gutierrez AB. Terminal ileitis as a manifestation of immune reconstitution syndrome following HAART. *AIDS* 2006; 20 : 1903-5.
- 157. Shah I. Immune Reconstitution Syndrome in HIV-1 infected children a study from India. *Indian J Pediatr* 2011; 78 : 540-3.
- Subsai K, Kanoksri S, Siwaporn C, Helen L, Kanokporn O, Wantana P. Neurological complications in AIDS patients receiving HAART: a 2-year retrospective study. *Eur J Neurol* 2006; 13: 233-9.
- 159. Auyeung P, French MA, Hollingsworth PN. Immune restoration disease associated with *Leishmania donovani* infection following antiretroviral therapy for HIV infection. *J Microbiol Immunol Infect* 2010; 43 : 74-6.
- 160. Blanche P, Gombert B, Rivoal O, Abad S, Salmon D, Brezin A. Uveitis due to *Leishmania major* as part of HAART-induced immune restitution syndrome in a patient with AIDS. *Clin Infect Dis* 2002; *34* : 1279-80.
- 161. Berry A, Abraham B, Dereure J, Pinzani V, Bastien P, Reynes J. Two case reports of symptomatic visceral leishmaniasis in AIDS patients concomitant with immune reconstitution due to antiretroviral therapy. *Scand J Infect Dis* 2004; *36* : 225-7.
- 162. Kérob D, Bouaziz JD, Sarfati C, Pavie J, Vignon-Pennamen MD, Menotti J, *et al.* First case of cutaneous reconstitution inflammatory syndrome associated with HIV infection and leishmaniasis.*Clin Infect Dis* 2006; *43* : 664-6.
- 163. Martin-Blondel G, Alvarez M, Delobel P, Uro-Coste E, Cuzin L, Cuvinciuc V, et al. Toxoplasmic encephalitis IRIS in HIV-infected patients: a case series and review of the literature. J Neurol Neurosurg Psychiatry 2011; 82: 691-3.
- Rodriguez-Rosado R, Soriano V, Dona C, González-Lahoz J. Opportunistic infections shortly after beginning highly active antiretroviral therapy. *Antivir Ther* 1998; 3: 229-31.
- Jiménez-Expósito MJ, Alonso-Villaverde C, Sardà P, Masana L. Visceral leishmaniasis in HIV-infected patients with nondetectable HIV-1 viral load after highly active antiretroviral therapy. *AIDS* 1999; *13*: 152-3.
- 166. González-Castillo J, Blanco F, Soriano V, Barreiro P, Concepción Bravo M, Jiménez-Nácher I, *et al.* Opportunistic episodes in patients infected with the human immunodeficiency virus during the first 6 months of HAART. *Med Clin (Barc)* 2001; *117* : 81-4.

- 167. Javid B, Aliyu SH, Save VE, Carmichael AJ, Lever AM. Schistosomal colonic polyposis in an HIV-positive man. *AIDS* 2007; 21: 386-8.
- Fernando R, Miller R. Immune reconstitution eosinophilia due to schistosomiasis. Sex Transm Infect 2002; 78: 76.
- 169. de Silva S, Walsh J, Brown M. Symptomatic Schistosoma mansoni infection as an immune restoration phenomenon in a patient receiving antiretroviral therapy. Clin Infect Dis 2006; 42: 303-4.
- McCombe JA, Auer RN, Maingat FG, Houston S, Gill MJ, Power C. Neurologic immune reconstitution inflammatory syndrome in HIV/AIDS: outcome and epidemiology. *Neurology* 2009; 72: 835-41.
- 171. Serpa JA, Moran A, Goodman JC, Giordano TP, White AC Jr. Neurocysticercosis in the HIV era: a case report and review of the literature. *Am J Trop Med Hyg* 2007; 77: 113-7.

- Albrecht H. Leishmaniosis-new perspectives on an underappreciated opportunistic infection. *AIDS* 1998; *12*: 2225-6.
- Cabral RF, Valle Bahia PR, Gasparetto EL, Chimelli L. Immune reconstitution inflammatory syndrome and cerebral toxoplasmosis. *AJNR Am J Neuroradiol* 2010; *31*: E65-6.
- 174. Huruy K, Mulu A, Mengistu G, Shewa-Amare A, Akalu A, Kassu A, *et al.* Immune reconstitution inflammatory syndrome among HIV/AIDS patients during highly active antiretroviral therapy in Addis Ababa, Ethiopia. *Jpn J Infect Dis* 2008; *61* : 205-9.
- 175. Klotz SA, Aziz Mohammed A, Girmai Woldemichael M, Worku Mitku M, Handrich M. Immune reconstitution inflammatory syndrome in a resource-poor setting. J Int Assoc Physicians AIDS Care (Chic) 2009; 8: 122-7.

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