



# Article Discrepancy between the Actions of Glucagon-like Peptide-1 Receptor Ligands in the Protection of the Heart against Ischemia Reperfusion Injury

Ali Ismaeil<sup>1</sup>, Fawzi Babiker<sup>1,\*</sup> and Suleiman Al-Sabah<sup>2,\*</sup>

- <sup>1</sup> Department of Physiology, Faculty of Medicine, Kuwait University, P.O. Box 24923, Safat 13110, Kuwait; ali.moh.esmaeil@gmail.com
- <sup>2</sup> Department of Pharmacology & Toxicology, Faculty of Medicine, Kuwait University, P.O. Box 24923, Safat 13110, Kuwait
- \* Correspondence: fawzi.babiker@ku.edu.kw (F.B.); sulaiman.alsabah@ku.edu.kw (S.A.-S.)

Abstract: Tirzepatide is a dual glucagon-like peptide-1 (GLP-1) and glucose-dependent insulinotropic polypeptide (GIP) receptor agonist and a promising therapy for type 2 diabetes mellitus (T2DM). GLP-1 is an incretin hormone with therapeutic potential beyond type 2 diabetes mellitus. However, GLP-1 is rapidly degraded by dipeptdyl peptidase-IV (DPP-IV) to GLP-1 (9-36). Exendin-4 (Ex-4) is a DPP-IV-resistant GLP-1 receptor agonist which, when truncated to Ex-4 (9-39), acts as a GLP-1 receptor antagonist. In the present study, hearts isolated from Wistar rats (n = 8 per group) were perfused with a modified Langendorff preparation. Left ventricular (LV) contractility and cardiovascular hemodynamics were evaluated by a data acquisition program and infarct size was evaluated by 2,3,5-Triphenyl-2H-tetrazolium chloride (TTC) staining and cardiac enzyme levels. Hearts were subjected to 30 min regional ischemia, produced by ligation of the left anterior descending (LAD) coronary artery followed by 30 min reperfusion. Hearts were treated during reperfusion with either the non-lipidated precursor of tirzepatide (NLT), GLP-1, GLP-1 (9-36), or Ex-4 in the presence or absence of Ex-4 (9-39). Infusion of GLP-1 (9-36) or Ex-4 protected the heart against I/R injury (p > 0.01) by normalizing cardiac hemodynamic and enzyme levels. Neither GLP-1, NLT, nor Ex-4 (9-39) showed any protection. Interestingly, Ex-4 (9-39) blocked Ex-4-mediated protection but not that of GLP-1 (9-36). These data suggest that Ex-4-mediated protection is GLP-1-receptor-dependent but GLP-1 (9-36)-mediated protection is not.

Keywords: ischemia reperfusion; GLP-1; GLP-1 (9-36); exendin-4

# 1. Introduction

Cardiovascular diseases (CVDs) are the most common cause of death worldwide [1]. Ischemic heart disease (IHD) is the leading cause of morbidity and mortality globally [2]. Myocardial reperfusion therapy is currently the only standard therapy for the protection of the ischemic heart. However, the development of complications in the first few years following infarction persists [3]. These complications include in-hospital mortality, recurrent myocardial infarction, and fatal left ventricular (LV) remodelling leading to heart failure [4]. The mechanisms responsible for ischemia/reperfusion (I/R) injury include opening of the mitochondrial permeability transition pore (mPTP), changes in intracellular pH, intracellular calcium overload, the activation of proteases, and the generation of oxidative stress by reactive oxygen species (ROS) and inflammation mediated by cytokines and complements [5–7]. I/R results in notable cardiomyocyte necrosis and apoptosis, which are proven causes of I/R injury [8]. Several therapeutic interventions to prevent myocardial I/R injury have been investigated. Unfortunately, the translation of such cardio-protective interventions to clinical trials has been rather disappointing [5,9]. Therefore, there is an urgent need for more effective regimens [10–12].



Citation: Ismaeil, A.; Babiker, F.; Al-Sabah, S. Discrepancy between the Actions of Glucagon-like Peptide-1 Receptor Ligands in the Protection of the Heart against Ischemia Reperfusion Injury. *Pharmaceuticals* 2022, *15*, 720. https://doi.org/ 10.3390/ph15060720

Academic Editor: Gill Diamond

Received: 26 April 2022 Accepted: 2 June 2022 Published: 6 June 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Glucagon-like peptide-1 (GLP-1) is an incretin hormone released from the gastrointestinal tract in response to feeding and functions primarily to potentiate insulin secretion from pancreatic  $\beta$ -cells in a glucose-dependent manner [13]. As a result, the GLP-1 receptor (GLP-1R) has become an attractive target in the treatment of type 2 diabetes mellitus (T2DM) [14,15]. Unfortunately, native GLP-1 has a very short half-life (1–2 min). The enzyme dipeptdyl peptidase-IV (DPP-IV) cleaves GLP-1 at its N-terminus to give GLP-1 (9-36) [16]. Exendin-4 (Ex-4), a peptide first discovered in the venom of the Gila monster *Heloderma suspectum*, acts as potent GLP-1R agonist. An amino acid substitution at position 2 (alanine to glycine) in the reptilian peptide results in DPP-IV-resistance and a longer circulating half-life. Removal of the first eight amino acids of Ex-4 results in Ex-4 (9-39), a GLP-1R antagonist that has been used extensively in the study of the physiology of GLP-1 [17].

GLP-1R is widely expressed and GLP-1 is now understood to be a pleiotropic hormone with therapeutic potential beyond type 2 diabetes mellitus. GLP-1R agonists have been shown to be both neuro- and cardio-protective [13]. Interestingly, the GLP-1 metabolite GLP-1 (9-36) has been shown to protect the heart against I/R reperfusion injury. Furthermore, the cardio-protective actions of GLP-1 (9-36) appear to be independent of the GLP-1R [18]. Recently, tirzepatide, a dual GLP-1R and GIP receptor agonist, is undergoing clinical trials for the treatment of type 2 diabetes [19]. This drug may have potential as a therapy for IHD. Hence, in the present study we investigated the potential role of GLP-1, Ex-4, GLP-1 (9-36), and the non-lipidated precursor of tirzepatide (NLT) in the protection of the heart against I/R injury during reperfusion to further understand the cardio-protective potential of these peptides.

### 2. Results

The end point for the evaluation of the effect of I/R and its treatments on the heart varies between laboratories. Some research groups use LV hemodynamics and contractility as an end point to evaluate the effect of I/R and its treatment on the heart muscle and its performance. Other groups use the infarct size, which is crucial for the prognosis of the disease, as an end point. Coronary and coronary vascular resistance were also used to evaluate the perfusion of the flow. In this study, for a full picture of the effect, we used LV hemodynamics, contractility, and coronary dynamics as end points. Furthermore, we used infarct size and cardiac enzyme levels as a confirmation of our findings. Heart hemodynamics during stabilization, ischemic, and reperfusion periods were evaluated. Left ventricular dynamics were determined throughout the experiment by measuring LVEDP, maximum DPmax, and LV  $\pm$ dP/dt. Coronary vascular dynamics were evaluated by measuring CF and CVR. Body weights and LV weights were not significantly different between the experimental groups. The values for cardiovascular dynamics in all study groups were presented as percentages of the baseline data.

Left ventricular dynamics, contractility, and coronary vascular dynamics were not significantly different between the groups when at baseline heart perfusion. Ischemia, which followed the baseline perfusion, resulted in a considerable deterioration in the heart functions which was reversed by GLP-1 (9-36) and Ex-4 but not by GLP-1 (Figure 1).

The same effect was observed in the LV  $\pm$ dP/dt, which was significantly improved by the infusion of GLP-1 (9-36) and Ex-4 (Table 1). These results were confirmed by the decrease in the infarct size (Figure 2) and the levels of cardiac enzyme release, which were decreased in the presence of GLP-1 (9-36) and Ex-4 (Table 2).

We then used NLT, the non-lipidated precursor of tirzepatide, to investigate any cross-talk between the GLP-1 and GIP receptors or possible synergy in their protection. Surprisingly, the infusion of NLT before ischemia or reperfusion did not protect the heart against I/R (Figure 3A–D). These results were confirmed by a lack of decrease in the infarct size (Figure 4) and increased cardiac enzyme levels (Table 2).



**Figure 1.** GLP-1 protection of the heart against ischemia reperfusion injury (n = 8 per group). Postischemic recovery in myocardial hemodynamics (**A**) DPmax, and (**B**) LVEDP, (**C**) CF and (**D**) CVR. The data were computed at 30 min reperfusion and are expressed as means  $\pm$  SEM. Ctr, control; DPmax, maximum developed pressure; LVEDP, left ventricular end diastolic pressure; CF, coronary flow; CVR, coronary vascular resistance. \* p < 0.001 compared to respective control. <sup>†</sup> p < 0.001compared to ischemic period.

**Table 1.** The effect of GLP-1 receptor ligands on heart contractility in presence or absence of the GLP-1R antagonist Ex-4 (9-39).

	+dP/dt		-dP/dt	
Ireatment	Ischemia	Reperfusion	Ischemia	Reperfusion
Ctr	$57.30 \pm 4.81$	$55.62\pm5.05$	$55.41 \pm 3.68$	$48.27 \pm 4.03$
Ctr + EX-4	$46.65 \pm 1.38$	$73.53 \pm 3.20$ * <sup>+</sup>	$42.73 \pm 1.83$	$71.36 \pm 3.61$ * <sup>+</sup>
Ctr + NLT Rep	$52.90 \pm 4.82$	$48.55\pm2.22$	$51.16 \pm 2.66$	$44.17 \pm 1.27$
Ctr + NLT pre	$54.18 \pm 3.75$	$44.90\pm5.41$	$56.94 \pm 4.11$	$54.65 \pm 4.71$
Ctr + GLP-1	$46.63 \pm 2.44$	$51.13 \pm 2.96$	$44.90 \pm 1.66$	$51.68 \pm 3.71$
Ctr + GLP-1 (9-36)	$48.33 \pm 4.12$	$71.28 \pm 1.98$ * <sup>+</sup>	$48.10\pm2.79$	66.11 $\pm$ 1.91 * <sup>+</sup>
Ctr + Ex-4 (9-39)	$65.56 \pm 5.36$	$42.22\pm7.56$	$59.58 \pm 2.04$	$51.10\pm2.83$
Ctr + Ex-4 + Ex-4 (9-39)	$63.44 \pm 5.10$	$68.32 \pm 4.61$	$51.22\pm 6.05$	$55.13 \pm 8.33$
Ctr + GLP-1 (9-36) + Ex-4 (9-39)	$60.86\pm3.60$	$76.04 \pm 1.88 * ^{\dagger}$	$59.02\pm3.52$	73.63 $\pm$ 2.32 * <sup>+</sup>

Ctr, control; DPmax, maximum developed pressure; LVEDP, left ventricular end diastolic pressure; CF, coronary flow; CVR, coronary vascular resistance. \* p < 0.001 compared to respective control. \* p < 0.001 compared to ischemic period.

**Table 2.** Effects of GLP-1 isoforms on the heart contractility in presence or absence of GLP-1R antagonists EX-(9-39) on the cardiac enzyme levels.

Treatment	CK (IU/L)	p Value	LDH (IU/L)	p Value
Ctr	$15.78\pm0.46$	-	$10.95 \pm 1.02$	-
Ctr + EX-4	$10.73 \pm 0.56$ * <sup>†</sup>	0.001	$7.45\pm0.47$ * $^{+}$	0.01
Ctr + NLT Rep	$10.11\pm0.46$	0.369	$10.40\pm0.17$	0.558
Ctr + NLT pre	$14.95\pm0.84$	0.448	$15.02\pm0.76$	0.471
Ctr + GLP-1	$13.78\pm1.40$	0.215	$9.43\pm0.52$	0.131
Ctr + GLP-1 (9-36)	$10.58 \pm 0.85$ * <sup>†</sup>	0.001	$6.25 \pm 0.82$ * <sup>†</sup>	0.01
Ctr + Ex-4 (9-39)	$15.33\pm0.95$	0.635	$10.88\pm0.37$	0.937
Ctr + Ex-4 + Ex-4 (9-39)	$14.70\pm1.59$	0.427	$10.33\pm0.44$	0.487
Ctr + GLP-1 (9-36) + Ex-4 (9-39)	$10.48 \pm 0.78$ * <sup>†</sup>	0.01	$6.28\pm0.49$ * <sup>+</sup>	0.01

CK, creatine kinase; LDH, lactate dehydrogenase; Ctr, control; DPmax, maximum developed pressure; LVEDP, left ventricular end diastolic pressure; CF, coronary flow; CVR, coronary vascular resistance. \* p < 0.001 compared to respective control. <sup>†</sup> p < 0.001 compared to ischemic period.



**Figure 2.** Post-ischemic infarct size in different treatment groups. Post-infarct size is significantly reduced with the introduction of Ex-4, GLP-1(9-36), and GLP-1 (9-36) + Ex-4 (9-39). Ctr, control; GLP-1, glucagon-like peptide-1; GLP-1 (9-36), glucagon-like peptide-1 (9-36); Ex-4 (9-39), exendin-4 (9-39); Ex-4, exendin-4. \* p < 0.01 compared to the control group.



**Figure 3.** Post-ischemic recovery of CVR in the presence of exendin-4 and NLT infused before ischemia or at reperfusion. (A) DPmax, and (B) LVEDP, (C) CF and (D) CVR. No significant improvement in CVR was appreciated in the displayed treatment groups. The data were collected after 25 min of ischemia and 30 min of reperfusion and are presented as means  $\pm$  SE. CVR, coronary vascular resistance; Ex-4, exendin-4; NLT Pre, non-lipidated tirzepatide pre-ischemia; NLT Post, non-lipidated tirzepatide post-ischemia, \* *p* < 0.05 compared to the control group. \* *p* < 0.05 compared to the respective ischemic period.

To investigate whether the improvements in the hemodynamics, infarct size, and cardiac enzyme levels reported above were modulated via a signaling pathway involving GLP-1R, the GLP-1R antagonist Ex-4 (9-39) was used at reperfusion in the presence of GLP-1 (9-36) and Ex-4. Ex-4 (9-39) completely blocked the protection given by Ex-4 but not the protection produced by GLP-1 (9-36) (Figure 5A–D and Tables 1 and 2).

To study the possible effects of GLP-1 and NLT on the modulation of cytokines during I/R injury, the levels of TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 were measured in the cardiomyocyte lysate by ELISA. Exposure of the heart to GLP-1 (9-36) and Ex-4 resulted in a significant decrease in the protein levels of TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 compared to those of untreated controls (p < 0.05) (Figure 6). This effect was not observed in the treatment with NLT and was completely negated by Ex-4 (9-39) in the presence of Ex-4 but not when co-administered in presence of GLP-1 (9-36) (Figure 6A–C).



**Figure 4.** Post-ischemic infarct size in different treatment groups. Post-ischemic infarct size was not reduced by the introduction of NLT pre- and post-ischemia. Ctr, control; Ex-4, exendin-4; NLT Pre, non-lipidated tirzepatide pre-ischemia; NLT Post, non-lipidated tirzepatide post-ischemia. \* p < 0.01 compared to the control group.



**Figure 5.** Post-ischemic recovery of CVR in the presence of exendin-4 + exendin-4 (9-39) and GLP-1(9-36) + exendin-4 (9-39). (**A**) DPmax, and (**B**) LVEDP, (**C**) CF and (**D**) CVR. The data were collected after 25 min of ischemia and 30 min of repefusion and are presented as means  $\pm$  SE. LVEDP, left ventricular end diastolic pressure; DPmax, maximum developed pressure; Ex-4, exendin-4; GLP-1 (9-36), glucagon-like peptide-1 (9-36); Ex-4 (9-39), exendin-4 (9-39). \* *p* < 0.05 compared to the control group; \* *p* < 0.05 compared to the respective ischemic period.

To understand the potential pathways of the GLP-1 protection of the heart, the antioxidant enzymes SOD and CAT and were evaluated. Treatment with GLP-1 (9-36) and Ex-4 significantly (p < 0.01) increased SOD and CAT levels compared to controls. This was not seen when the heart was treated with NLT and was completely abrogated by Ex-4 (9-39) in the presence of Ex-4 but not when co-administered with GLP-1 (9-36) (Figure 7A,B).



**Figure 6.** Post-ischemic reduction of (**A**) TNF- $\alpha$ , (**B**) IL-1 $\beta$ , and (**C**) IL-6 levels. Ctr, control; GLP-1, glucagon-like peptide-1; GLP-1 (9-36), glucagon-like peptide-1 (9-36); Ex-4 (9-39), exendin-4 (9-39); Ex-4, exendin-4; NLT Pre, non-lipidated tirzepatide pre-ischemia; NLT Post, non-lipidated tirzepatide post-ischemia. \* *p* < 0.01 compared to the control group.



**Figure 7.** Post-ischemic recovery of (**A**) CAT and (**B**) SOD activity. Ctr, control; GLP-1 (9-36), glucagon-like peptide-1 (9-36); Ex-4 (9-39), exendin-4 (9-39); Ex-4, exendin-4; NLT Pre, non-lipidated tirzepatide pre-ischemia; NLT Post, non-lipidated tirzepatide post-ischemia. \* p < 0.01 compared to the control group.

## 3. Discussion

There are currently several GLP-1 receptor agonists used clinically for the treatment of both type 2 diabetes mellitus and obesity, for example, liraglutide, lixsenatide, semaglutide, dulaglutide and exenatide [13]. This class of medication has also been shown to display cardiovascular protection in high-risk patients with T2DM [20]. It is thought that GLP-1R agonists' cardiovascular effects are at least partially independent of their ability to control glycemia and body weight [21,22]. GLP-1R is expressed in the heart, although its exact location is unclear; therefore, it is possible that GLP-1R agonists exert direct, as well as indirect, effects upon the heart [23,24]. Initially thought of as simply the degradation product of GLP-1, GLP-1 (9-36) has since been shown to have distinct physiological effects compared to its parent peptide [25]. Furthermore, GLP-1 (9-36) has been shown to be cardio-protective properties were preserved in GLP-1 receptor knockout cardiomyocytes, suggesting GLP-1R-independent activity [18].

The present study also demonstrates the protective effects of GLP-1 (9-36) and Ex-4 on the heart against I/R injury. This involved the GLP-1R for Ex-4 but not for GLP-1 (9-36). Both Ex-4 and GLP-1 (9-36) have previously been shown to have cardio-protective effects [18,26,27]. However, the precise mechanism and the involvement of GLP-1R in the protection of the heart against I/R injury has yet to be determined. Furthermore, evidence for the potential protection of these peptides when infused at reperfusion, which is clinically more relevant than infusion before ischemia, is also lacking.

No protection of the heart against I/R injury by native GLP-1 could be demonstrated in the present study. This contrasts with previous studies which report GLP-1-mediated protection of the heart against I/R injury in both animal models of diabetes and diabetic patients [28,29]. The discrepancy between these studies and the results reported here could be due to GLP-1 protecting the diabetic heart under conditions of hyperglycemia or organ remodeling created by the presence of diabetes. The animal model, the dose and the time of infusion could all contribute to the reported discrepancies. A direct effect for GLP-1 in non-diabetic animals has also been previously reported; however, in these studies, GLP-1 was infused before the ischemic insult. These results can be explained by GLP-1 being protective to the heart against I/R injury in diabetic models or when infused before ischemia. The protocol used in the present study infuses GLP-1 at reperfusion, which is clinically more relevant than perfusion before ischemia.

Intriguingly, the present study demonstrates that the GLP-1 degradation product GLP-1 (9-36) displays GLP-1R-independent protection of the heart against I/R injury. As mentioned previously, GLP-1 (9-36) has been reported to be both an antagonist and a weak partial agonist of the GLP-1R as well as being biologically inactive [30–32]. Furthermore, Sauvé et al. demonstrated that genetic deletion and pharmacological inhibition of DPP IV, which inhibits the production of GLP-1 (9-36), improved cardiovascular outcomes following myocardial infarction in a murine model [33]. In contrast, other studies have shown GLP-1 (9-36) to be protective against I/R injury in various tissues and organs [24,34–36]. To our knowledge, the present study is the first to demonstrate that GLP-1 (9-36) is protective against I/R injury when infused at reperfusion.

Treatment of isolated hearts with either Ex-4 or GLP-1 (9-36) at reperfusion protected the heart against I/R injury and normalized LV dynamics and coronary vascular dynamics to those of controls. Whereas the cardio-protective effects of Ex-4 were inhibited by infusion of the GLP-1R antagonist Ex-4 (9-39), this was not the case for GLP-1 (9-36) (Figure 8). These data suggest that the cardio-protective effects of Ex-4 require the GLP-1R, which agrees with previous studies which demonstrate the involvement the GLP-1R in the protection of the heart [24,33]. Our data are also in agreement with other studies which demonstrate a GLP-1R-independent mechanism for GLP-1 (9-36) in the protection of the heart [24]. Although Ex-4 has been shown to be protective of the heart before ischemia, its infusion at reperfusion did not show protection in previous studies [22,29]. This contrasts with the data presented in the present study. This discrepancy may be due to either differences in the animal models or the procedures employed. The results of the study indicate acute protection of the heart against I/R injury by Ex-4 and GLP-1 (9-36) when infused at reperfusion by a GLP-1R mechanism for Ex-4 but not for GLP-1 (9-36). One of the strengths of this ex vivo preparation is that it shows the possible separation of the direct effects of the GLP-1 metabolites on the heart from the additional insulinotropic effects of GLP-1 that occur in vivo.

In addition, we used NLT in this study to investigate potential improvement in the protective effects compared to Ex-4 and GLP-1 (9-36). Interestingly, infusion with NLT before ischemia or at reperfusion did not protect the heart against I/R injury. NLT was proven to be effective as a therapy for type 2 diabetes, but its effect on the protection of the heart against I/R injury has not yet been reported [19]. Although NLT is a agonist at the GLP-1R, which plays a protective role in I/R [37], it does not show protection of the heart similar to the other agonists of the GLP-1R [38,39]. However, contrasting results were also reported [40,41]. The masking of the protective effect of NLT could be mainly due to its agonism of GIPR, which was reported to exacerbate ischemic injury, and its blockade is beneficial to the ischemic heart [40].

This study reported an anti-inflammatory effect for Ex-4 and GLP-1 (9-36), as treatment of the heart with these compounds resulted in a significant reduction in TNF- $\alpha$ , IL-1 $\beta$ , and IL-6. Although the presence of immune cells in the isolated heart was unlikely, several studies have shown that I/R causes its damage and cardiac dysfunction to the heart partly by promoting pro-inflammatory cytokine formation [12,42]. Treatment of the rat heart with Ex-4 and GLP-1 (9-36) showed a significant reduction in these pro-inflammatory cytokine protein levels. GLP-1R agonists were reported to have anti-inflammatory properties, which might explain their protective effects in this study [43]. These agonists showed a significant reduction in TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 [44,45]. Lowering the TNF- $\alpha$  [46], IL-1 $\beta$  [47,48], and IL-6 [47] protein levels was proven to be essential in the protection of the heart against I/R injury and was shown to decrease apoptosis in cardiac myocytes [47,48]. The results of this study are consistent with the above findings.



**Figure 8.** Schematic representation showing the experimental protocols used for the study (n = 8 per group). (**A**) Unprotected ischemia-reperfusion (control). (**B**) Infusion with GLP-1, GLP-1 (9-36), and Ex-4 (**C**) Infusion of non-lipidated precursor of tirzepatide. (**D**) Infusion with GLP-1, GLP-1 (9-36), and Ex-4 in presence of Ex-4 (9-39).

In this study, we reported an increase in the levels of the antioxidant enzymes SOD and CAT after the treatment with Ex-4 and GLP-1 (9-36), which proves the presence of antioxidant activity in these compounds. Although ROS are essential for many beneficial physiological processes, their contribution to pathological conditions is indispensable [49], and GLP-1R was proven to be a strong antioxidant [50]. Ex-4 [51] and (GLP- (9-36) [52] were reported to be essential effectors in the reduction of ROS levels in the context of hyper-glycemia. Reductions in ROS by Ex-4 and GLP-1 (9-36) in the protection of cardiomyocytes were also reported previously [53,54]. Nevertheless, it was in a pretreatment procedure, where Ex-4 and GLP-1 (9-36) protected the heart against hypoxic damage [55]. Similar protective effects by decreasing ROS levels were reported previously by our laboratory [56,57]. We have also previously reported increased levels of SOD and CAT in other procedures that showed protection of the heart against I/R injury [46].

#### 4. Materials and Methods

### 4.1. Ischemia Reperfusion Study

Age-matched male Wistar rats weighing between 270 and 300 g (n = 8 per group) were used in this study. Animal treatment and handling were carried out according to the guidelines of Animal Resource Center of Kuwait University, Kuwait. Rats were maintained at 22 °C on 12 h light/dark cycle (7 am–7 pm), and water and food were unrestrictedly available. Rats were anesthetized with intraperitoneal injections of sodium pentobarbital (60 mg/kg), and heparin (1000 U/kg) was administered through the femoral vein. Heart cannulation and perfusion were performed as described previously [10,58]. Briefly, isolated heart was retrograde perfused with a freshly prepared Krebs–Hensleit buffer, pH 7.35 to 7.45 at 37.0 ± 0.5 °C and was supplied with CO<sub>2</sub> (5%) and O<sub>2</sub> (95%). Heart instrumentation included placing pacing electrodes on the right atrium (RA) appendage to

maintain physiological heartbeat. Regional ischemia was induced by occluding left anterior descending (LAD) coronary artery for 30 min. The LAD was encircled by a snare at 0.5 cm below the atrioventricular groove, and a small, rigid plastic tube was positioned between the heart and the snare to ensure complete occlusion of the coronary artery. Subsequently, the heart was reperfused for 30 min. The flow of the perfusate was measured downstream of the aortic cannula using a "Statham pressure transducer" (P23 Db). Preload was held constant at 6 mmHg under basal controlled conditions, and the perfusion pressure (PP) was held constant at 50 mmHg throughout the experimental procedure in all protocols. Constant PP was ensured electronically by means of the perfusion assembly ("Module PPCM type 671 (Hugo Sachs Elektronik-Harvard Apparatus GmbH, March, Germany")), an effective system for the accurate adjustment of PP between 5 mmHg and 150 mmHg with  $\pm 1$  mmHg accuracy level.

#### 4.2. Study Protocol and Study Groups

A total of 72 hearts were subdivided into 9 groups (n = 8 per group) and subjected to four different experimental protocols. The control group was subjected to ischemia and reperfusion without any other treatment; the second group was treated with NLT (0.1 µM) started 15 min before and continued for 15 min; the third to seventh groups were subjected to control procedures with infusion of (0.1 µM each) Ex-4, NLT, GLP-1 (9-39), GLP-1 (9-36), and Ex-4 (9-39) 5 min before the start of reperfusion and continued for 10 min during reperfusion; and the eighth and ninth groups were treated with GLP-1 (9-36) or Ex-4 in presence of the GLP-1R antagonist Ex-4 (9-39) (0.1 µM), as summarized in Figure 1. The antagonists were always infused 5 min before the infusion of the drug in question. Animals were randomly assigned to groups and the treatments were applied randomly. All hearts were subjected to 30 min of ischemia produced by LAD coronary artery occlusion. Hearts were then reperfused for 30 min (Figure 8).

#### 4.3. Data Collection and Processing

The function of LV was evaluated by the assessment of LV end diastolic pressure (LVEDP), LV developed pressure (DPmax), and LV contractility  $(\pm dP/dt)$ . The coronary-vascular dynamics were evaluated by the assessment of coronary vascular resistance (CVR) and coronary flow (CF). Cardiovascular functions were measured as described in the previous studies [56,59]. Briefly, a water-filled latex balloon was placed and secured in LV cavity. The balloon was attached to a pressure transducer and a "DC-Bridge amplifier (DC-BA)" of the pressure module (DC-BA type 660, Hugo-Sachs Electronik, March, Germany) and interfaced with a personal computer for online monitoring of DPmax. Left ventricular developed pressure was derived from online acquisition of "LVESP using Max-Min module (Number MMM type 668" (Hugo Sachs Elektronik-Harvard ApparatusGmbH, March, Germany) which could convert the output from DC bridge amplifier to DPmax by subtracting LVEDP from the LVEDP.

Coronary flow was continuously measured using an electromagnetic flow probe interfaced with a personal computer and attached to the inflow of the aortic cannula. The continuous monitoring of the coronary flow in ml/min was performed digitally using suitable software and was verified manually by timed collection of coronary effluent. The CVR and hemodynamics data were determined every 10 s using an "online data acquisition program" (Isoheart software V 1.524-S, Hugo-Sachs Electronik, March, Germany). By the end of each experiment, hearts were snap-frozen in liquid nitrogen, then stored in -80 °C for further analysis.

#### 4.4. Sample Collection and Storage

Coronary effluent was collected manually at the apex of the heart from the coronary outflow in small tubes at the end of the reperfusion phase. The hearts were also collected at the end of reperfusion. All heart samples and coronary effluents were frozen in liquid nitrogen and stored at -80 °C for further analysis.

#### 4.5. Infarct-Size Evaluation

Hearts were collected after 30 min of reperfusion and stored overnight at -20 °C. The next day, the hearts were sliced into 4–5 pieces from apex to base. The slices were then incubated in 1% 2,3,5-Triphenyl-2H-tetrazolium chloride (TTC) solution in isotonic (pH 7.4) phosphate buffer and then fixed in 4% formaldehyde. The red and pale non-stained areas of every slice were indicated manually on the image. The infarct size as a percentage of the LV was calculated for every heart. Quantification of the LV and infarct size was performed using ImageJ (Image J, Wayne Rasband and National Institute of Health, Bethesda, MD, USA). Cardiomyocyte injury was confirmed by measuring creatine kinase (CK) and lactate dehydrogenase (LDH) release in the coronary effluent during the reperfusion period, as described previously by Ferrera et al. [60].

#### 4.6. Protein Extraction from the Hearts

At the end of the experiments, the hearts were snap-frozen in liquid nitrogen and subsequently stored at -80 °C for further analysis. Left ventricle tissue was homogenized in a lysis buffer (MOPS, 20 mM; KCl, 150 mM; Mg Acetate, 4.5 mM; Triton X, 1%) containing protease inhibitor cocktail tablet (Roche Applied Science, Mannheim, Germany, Cat#05892970001) and centrifuged at 12,000× *g* for 15 min at 4 °C. The supernatant containing the proteins was then collected, aliquoted, and stored for further analysis. Protein levels were assayed using bicinchoninic acid protein assay (Pierce Chemical Co., Rockford, IL, USA, Cat# 23227).

### 4.7. Antioxidant Enzyme Superoxide Dismutase Status

Superoxide dismutase (SOD; EC.1.15.1.1) and Catalase (CAT; EC.1.11.1.6) activity were measured in cardiomyocyte lysate (n = 4) by UV spectrophotometer activity with RANDOX kits (Cat. No. SD 125; Randox Labs Ltd., Crumlin, UK) according to the manufacturer's instructions.

# 4.8. Estimation of Inflammatory Cytokines

The LV homogenate (n = 4) was used to evaluate the expression of the proinflammatory cytokines TNF- $\alpha$  (cat# MBS175904), IL-1 (cat# MBS825017), and IL-6 (cat# MBS355410) using enzyme-linked immunosorbent assay (ELISA). The protein content was measured in the samples, which were diluted as suggested by the manufacturer's assay protocols for the respective commercial kits (Biosource International, Camarillo, California, USA).

#### 4.9. Data Analysis

The data were expressed as the means  $\pm$  SE. Two-way analysis of variance (ANOVA) was used for repeated measures within each group and between the groups. In the case where analysis of variance illustrated a significant difference, the post hoc analysis with Tukey test was used for further comparison. Infarct size was analyzed using student *t*-test. Comparisons were made between group means and their respective controls. Values of *p* < 0.05 were considered statistically significant for the analysis.

## 5. Conclusions

The data presented demonstrate that Ex-4-mediated protection of the heart against I/R injury when infused at reperfusion is GLP-1-receptor-dependent. In contrast, GLP-1 (9-36) protection under the same conditions appears to be GLP-1-receptor-independent. These findings highlight the need for a greater understanding of the mechanisms of GLP-1 (9-36)'s GLP-1-receptor independence with the aim of translating these findings to the clinic (Figure 9).



**Figure 9.** Schematic representation showing the interactions between I/R injury and GLP-1 related peptides. Ischemia/reperfusion-induced cardiac injury characterized by impaired cardiac hemodynamics and enhanced release of cardiac enzymes, LDH, and CK. GLP-1 (9-36) and Ex-4 treatment improved cardiac hemodynamic and attenuated release of cardiac enzymes. Ex-4 protection involved downstream signaling via the GLP-1 receptor; however, the protection elicited by GLP-1(9-36) appears to be GLP-1-receptor independent.

**Author Contributions:** Conceptualization, F.B. and S.A.-S.; investigation, A.I. and F.B.; formal analysis, A.I. and F.B.; writing—original draft preparation, A.I., F.B. and S.A.-S.; writing—review and editing, F.B. and S.A.-S. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported and funded by Kuwait University research grant No. YM 01/20.

**Institutional Review Board Statement:** The study was conducted according to the guidelines of the Declaration of Helsinki, and approved by the Ethical Committee of the Research Health Science Center of Kuwait University (23/VDR/EC/3566).

Informed Consent Statement: Not applicable.

Data Availability Statement: Data is contained within the article.

**Acknowledgments:** The authors thank the technical assistance of Anju Devassy at the Department of Physiology, Kuwait University.

Conflicts of Interest: The authors have no conflict of interest to declare.

## References

- Lopez, A.D.; Mathers, C.D.; Ezzati, M.; Jamison, D.T.; Murray, C.J. Global and regional burden of disease and risk factors, 2001: Systematic analysis of population health data. *Lancet* 2006, 367, 1747–1757. [CrossRef]
- GBD 2016 Mortality Collaborators. Global, regional, and national under-5 mortality, adult mortality, age-specific mortality, and life expectancy, 1970–2016: A systematic analysis for the Global Burden of Disease Study 2016. *Lancet* 2017, 390, 1084–1150. [CrossRef]
- Cung, T.T.; Morel, O.; Cayla, G.; Rioufol, G.; Garcia-Dorado, D.; Angoulvant, D.; Bonnefoy-Cudraz, E.; Guerin, P.; Elbaz, M.; Delarche, N.; et al. Cyclosporine before PCI in Patients with Acute Myocardial Infarction. *N. Engl. J. Med.* 2015, 373, 1021–1031. [CrossRef] [PubMed]

- 4. Hausenloy, D.J.; Yellon, D.M. The mitochondrial permeability transition pore: Its fundamental role in mediating cell death during ischaemia and reperfusion. *J. Mol. Cell Cardiol.* **2003**, *35*, 339–341. [CrossRef]
- Limalanathan, S.; Andersen, G.O.; Klow, N.E.; Abdelnoor, M.; Hoffmann, P.; Eritsland, J. Effect of ischemic postconditioning on infarct size in patients with ST-elevation myocardial infarction treated by primary PCI results of the POSTEMI (POstconditioning in ST-Elevation Myocardial Infarction) randomized trial. J. Am. Heart Assoc. 2014, 3, e000679. [CrossRef]
- Tsang, A.; Hausenloy, D.J.; Yellon, D.M. Myocardial postconditioning: Reperfusion injury revisited. Am. J. Physiol. Heart Circ. Physiol. 2005, 289, H2–H7. [CrossRef] [PubMed]
- Zhang, H.; Meng, J.; Li, X.; Zhou, S.; Qu, D.; Wang, N.; Jia, M.; Ma, X.; Luo, X. Pro-GLP-1, a Pro-drug of GLP-1, is neuroprotective in cerebral ischemia. *Eur. J. Pharm. Sci.* 2015, 70, 82–91. [CrossRef] [PubMed]
- 8. Al-Herz, W.; Babiker, F. Acute Intravenous Infusion of Immunoglobulins Protects Against Myocardial Ischemia-Reperfusion Injury Through Inhibition of Caspase-3. *Cell. Physiol. Biochem.* **2017**, *42*, 2295–2306. [CrossRef]
- Freixa, X.; Bellera, N.; Ortiz-Perez, J.T.; Jimenez, M.; Pare, C.; Bosch, X.; De Caralt, T.M.; Betriu, A.; Masotti, M. Ischaemic postconditioning revisited: Lack of effects on infarct size following primary percutaneous coronary intervention. *Eur. Heart J.* 2012, 33, 103–112. [CrossRef] [PubMed]
- Abwainy, A.; Babiker, F.; Akhtar, S.; Benter, I.F. Endogenous angiotensin-(1-7)/Mas receptor/NO pathway mediates the cardioprotective effects of pacing postconditioning. *Am. J. Physiol. Heart Circ. Physiol.* 2016, 310, H104–H112. [CrossRef]
- 11. Babiker, F.A. Pacing Postconditioning: Recent Insights of Mechanism of Action and Probable Future Clinical Application. *Med. Princ. Pract. Int. J. Kuwait Univ. Health Sci. Cent.* **2016**, 25 (Suppl. S1), 22–28. [CrossRef] [PubMed]
- 12. Babiker, F.A.; van Golde, J.; Vanagt, W.Y.; Prinzen, F.W. Pacing postconditioning: Impact of pacing algorithm, gender, and diabetes on its myocardial protective effects. *J. Cardiovasc. Transl. Res.* **2012**, *5*, 727–734. [CrossRef]
- 13. Muller, T.D.; Finan, B.; Bloom, S.R.; D'Alessio, D.; Drucker, D.J.; Flatt, P.R.; Fritsche, A.; Gribble, F.; Grill, H.J.; Habener, J.F.; et al. Glucagon-like peptide 1 (GLP-1). *Mol. Metab.* **2019**, *30*, 72–130. [CrossRef]
- Al-Sabah, S. Molecular Pharmacology of the Incretin Receptors. *Med. Princ. Pract. Int. J. Kuwait Univ. Health Sci. Cent.* 2016, 25 (Suppl. S1), 15–21. [CrossRef] [PubMed]
- Graaf, C.; Donnelly, D.; Wootten, D.; Lau, J.; Sexton, P.M.; Miller, L.J.; Ahn, J.M.; Liao, J.; Fletcher, M.M.; Yang, D.; et al. Glucagon-Like Peptide-1 and Its Class B G Protein-Coupled Receptors: A Long March to Therapeutic Successes. *Pharmacol. Rev.* 2016, 68, 954–1013. [CrossRef] [PubMed]
- 16. Kieffer, T.J.; McIntosh, C.H.; Pederson, R.A. Degradation of glucose-dependent insulinotropic polypeptide and truncated glucagon-like peptide 1 In Vitro and In Vivo by dipeptidyl peptidase IV. *Endocrinology* **1995**, *136*, 3585–3596. [CrossRef]
- 17. Thorens, B.; Porret, A.; Buhler, L.; Deng, S.P.; Morel, P.; Widmann, C. Cloning and functional expression of the human islet GLP-1 receptor. Demonstration that exendin-4 is an agonist and exendin-(9-39) an antagonist of the receptor. *Diabetes* **1993**, *42*, 1678–1682. [CrossRef]
- Ban, K.; Kim, K.H.; Cho, C.K.; Sauve, M.; Diamandis, E.P.; Backx, P.H.; Drucker, D.J.; Husain, M. Glucagon-like peptide (GLP)-1(9-36) amide-mediated cytoprotection is blocked by exendin (9-39) yet does not require the known GLP-1 receptor. *Endocrinology* 2010, 151, 1520–1531. [CrossRef] [PubMed]
- 19. Frias, J.P.; Davies, M.J.; Rosenstock, J.; Perez Manghi, F.C.; Lando, F.L.; Bergman, B.K.; Liu, B.; Cui, X.; Brown, K.; Investigators, S. Tirzepatide versus Semaglutide Once Weekly in Patients with Type 2 Diabetes. *N. Engl. J. Med.* **2021**, *385*, 503–515. [CrossRef]
- Sheahan, K.H.; Wahlberg, E.A.; Gilbert, M.P. An overview of GLP-1 agonists and recent cardiovascular outcomes trials. *Postgrad. Med. J.* 2020, *96*, 156–161. [CrossRef]
- Sivertsen, J.; Rosenmeier, J.; Holst, J.J.; Vilsboll, T. The effect of glucagon-like peptide 1 on cardiovascular risk. *Nat. Rev. Cardiol.* 2012, 9, 209–222. [CrossRef] [PubMed]
- 22. Ussher, J.R.; Drucker, D.J. Cardiovascular biology of the incretin system. Endocr. Rev. 2012, 33, 187–215. [CrossRef] [PubMed]
- 23. Baggio, L.L.; Yusta, B.; Mulvihill, E.E.; Cao, X.; Streutker, C.J.; Butany, J.; Cappola, T.P.; Margulies, K.B.; Drucker, D.J. GLP-1 Receptor Expression within the Human Heart. *Endocrinology* **2018**, *159*, 1570–1584. [CrossRef]
- Ban, K.; Noyan-Ashraf, M.H.; Hoefer, J.; Bolz, S.S.; Drucker, D.J.; Husain, M. Cardioprotective and vasodilatory actions of glucagon-like peptide 1 receptor are mediated through both glucagon-like peptide 1 receptor-dependent and -independent pathways. *Circulation* 2008, 117, 2340–2350. [CrossRef] [PubMed]
- 25. Tomas, E.; Stanojevic, V.; Habener, J.F. GLP-1-derived nonapeptide GLP-1(28-36) amide targets to mitochondria and suppresses glucose production and oxidative stress in isolated mouse hepatocytes. *Regul. Pept.* **2011**, *167*, 177–184. [CrossRef]
- 26. Matsubara, M.; Kanemoto, S.; Leshnower, B.G.; Albone, E.F.; Hinmon, R.; Plappert, T.; Gorman, J.H., 3rd; Gorman, R.C. Single dose GLP-1-Tf ameliorates myocardial ischemia/reperfusion injury. *J. Surg. Res.* **2011**, *165*, 38–45. [CrossRef] [PubMed]
- Xie, Y.; Wang, S.X.; Sha, W.W.; Zhou, X.; Wang, W.L.; Han, L.P.; Li, D.Q.; Yu, D.M. Effects and mechanism of glucagon-like peptide-1 on injury of rats cardiomyocytes induced by hypoxia-reoxygenation. *Chin. Med. J.* 2008, 121, 2134–2138. [CrossRef] [PubMed]
- Davidson, M.H. Cardiovascular effects of glucagonlike peptide-1 agonists. Am. J. Cardiol. 2011, 108, 33B–41B. [CrossRef] [PubMed]
- 29. Timmers, L.; Henriques, J.P.; de Kleijn, D.P.; Devries, J.H.; Kemperman, H.; Steendijk, P.; Verlaan, C.W.; Kerver, M.; Piek, J.J.; Doevendans, P.A.; et al. Exenatide reduces infarct size and improves cardiac function in a porcine model of ischemia and reperfusion injury. *J. Am. Coll. Cardiol.* **2009**, *53*, 501–510. [CrossRef] [PubMed]

- Knudsen, L.B.; Pridal, L. Glucagon-like peptide-1-(9-36) amide is a major metabolite of glucagon-like peptide-1-(7-36) amide after in vivo administration to dogs, and it acts as an antagonist on the pancreatic receptor. *Eur. J. Pharmacol.* 1996, 318, 429–435. [CrossRef]
- Li, N.; Lu, J.; Willars, G.B. Allosteric modulation of the activity of the glucagon-like peptide-1 (GLP-1) metabolite GLP-1 9-36 amide at the GLP-1 receptor. *PLoS ONE* 2012, 7, e47936. [CrossRef]
- Montrose-Rafizadeh, C.; Yang, H.; Rodgers, B.D.; Beday, A.; Pritchette, L.A.; Eng, J. High potency antagonists of the pancreatic glucagon-like peptide-1 receptor. J. Biol. Chem. 1997, 272, 21201–21206. [CrossRef] [PubMed]
- Sauve, M.; Ban, K.; Momen, M.A.; Zhou, Y.Q.; Henkelman, R.M.; Husain, M.; Drucker, D.J. Genetic deletion or pharmacological inhibition of dipeptidyl peptidase-4 improves cardiovascular outcomes after myocardial infarction in mice. *Diabetes* 2010, 59, 1063–1073. [CrossRef]
- 34. Chen, F.; Wang, W.; Ding, H.; Yang, Q.; Dong, Q.; Cui, M. The glucagon-like peptide-1 receptor agonist exendin-4 ameliorates warfarin-associated hemorrhagic transformation after cerebral ischemia. *J. Neuroinflamm.* **2016**, *13*, 204. [CrossRef] [PubMed]
- DeNicola, M.; Du, J.; Wang, Z.; Yano, N.; Zhang, L.; Wang, Y.; Qin, G.; Zhuang, S.; Zhao, T.C. Stimulation of glucagon-like peptide-1 receptor through exendin-4 preserves myocardial performance and prevents cardiac remodeling in infarcted myocardium. *Am. J. Physiol. Endocrinol. Metab.* 2014, 307, E630–E643. [CrossRef] [PubMed]
- Gupta, N.A.; Kolachala, V.L.; Jiang, R.; Abramowsky, C.; Romero, R.; Fifadara, N.; Anania, F.; Knechtle, S.; Kirk, A. The glucagonlike peptide-1 receptor agonist Exendin 4 has a protective role in ischemic injury of lean and steatotic liver by inhibiting cell death and stimulating lipolysis. *Am. J. Pathol.* 2012, *181*, 1693–1701. [CrossRef] [PubMed]
- Bose, A.K.; Mocanu, M.M.; Carr, R.D.; Yellon, D.M. Myocardial ischaemia-reperfusion injury is attenuated by intact glucagon like peptide-1 (GLP-1) in the in vitro rat heart and may involve the p70s6K pathway. *Cardiovasc. Drugs Ther.* 2007, 21, 253–256. [CrossRef] [PubMed]
- Almutairi, M.; Gopal, K.; Greenwell, A.A.; Young, A.; Gill, R.; Aburasayn, H.; Al Batran, R.; Chahade, J.J.; Gandhi, M.; Eaton, F.; et al. The GLP-1 Receptor Agonist Liraglutide Increases Myocardial Glucose Oxidation Rates via Indirect Mechanisms and Mitigates Experimental Diabetic Cardiomyopathy. *Can. J. Cardiol.* 2021, 37, 140–150. [CrossRef]
- Aravindhan, K.; Bao, W.; Harpel, M.R.; Willette, R.N.; Lepore, J.J.; Jucker, B.M. Cardioprotection Resulting from Glucagon-Like Peptide-1 Administration Involves Shifting Metabolic Substrate Utilization to Increase Energy Efficiency in the Rat Heart. *PLoS* ONE 2015, 10, e0130894. [CrossRef]
- Ussher, J.R.; Baggio, L.L.; Campbell, J.E.; Mulvihill, E.E.; Kim, M.; Kabir, M.G.; Cao, X.; Baranek, B.M.; Stoffers, D.A.; Seeley, R.J.; et al. Inactivation of the cardiomyocyte glucagon-like peptide-1 receptor (GLP-1R) unmasks cardiomyocyte-independent GLP-1R-mediated cardioprotection. *Mol. Metab.* 2014, *3*, 507–517. [CrossRef] [PubMed]
- Wohlfart, P.; Linz, W.; Hubschle, T.; Linz, D.; Huber, J.; Hess, S.; Crowther, D.; Werner, U.; Ruetten, H. Cardioprotective effects of lixisenatide in rat myocardial ischemia-reperfusion injury studies. *J. Transl. Med.* 2013, *11*, 84. [CrossRef] [PubMed]
- 42. Babiker, F.A.; Joseph, S.; Juggi, J.S. What's new in salvage of the ischemic myocardium: Estrogen postconditioning. *Med. Princ. Pract. Int. J. Kuwait Univ. Health Sci. Cent.* **2011**, *20*, 495–496. [CrossRef] [PubMed]
- Bunck, M.C.; Diamant, M.; Eliasson, B.; Corner, A.; Shaginian, R.M.; Heine, R.J.; Taskinen, M.R.; Yki-Jarvinen, H.; Smith, U. Exenatide affects circulating cardiovascular risk biomarkers independently of changes in body composition. *Diabetes Care* 2010, 33, 1734–1737. [CrossRef]
- 44. Chaudhuri, A.; Ghanim, H.; Vora, M.; Sia, C.L.; Korzeniewski, K.; Dhindsa, S.; Makdissi, A.; Dandona, P. Exenatide exerts a potent antiinflammatory effect. *J. Clin. Endocrinol. Metab.* **2012**, *97*, 198–207. [CrossRef] [PubMed]
- Hogan, A.E.; Gaoatswe, G.; Lynch, L.; Corrigan, M.A.; Woods, C.; O'Connell, J.; O'Shea, D. Glucagon-like peptide 1 analogue therapy directly modulates innate immune-mediated inflammation in individuals with type 2 diabetes mellitus. *Diabetologia* 2014, 57, 781–784. [CrossRef] [PubMed]
- 46. Gouda, E.; Babiker, F. Micronized flavonoid fraction Daflon 500 protects heart against ischemia-reperfusion injury: An old medicine for a new target. *All Life* 2020, *13*, 556–568. [CrossRef]
- Ridker, P.M.; Howard, C.P.; Walter, V.; Everett, B.; Libby, P.; Hensen, J.; Thuren, T.; Group, C.P.I. Effects of interleukin-1beta inhibition with canakinumab on hemoglobin A1c, lipids, C-reactive protein, interleukin-6, and fibrinogen: A phase IIb randomized, placebo-controlled trial. *Circulation* 2012, 126, 2739–2748. [CrossRef] [PubMed]
- Suzuki, K.; Murtuza, B.; Smolenski, R.T.; Sammut, I.A.; Suzuki, N.; Kaneda, Y.; Yacoub, M.H. Overexpression of interleukin-1 receptor antagonist provides cardioprotection against ischemia-reperfusion injury associated with reduction in apoptosis. *Circulation* 2001, 104, I-308–I-313. [CrossRef] [PubMed]
- Sugamura, K.; Keaney, J.F., Jr. Reactive oxygen species in cardiovascular disease. Free Radic. Biol. Med. 2011, 51, 978–992. [CrossRef] [PubMed]
- 50. Pujadas, G.; Drucker, D.J. Vascular Biology of Glucagon Receptor Superfamily Peptides: Mechanistic and Clinical Relevance. *Endocr. Rev.* 2016, *37*, 554–583. [CrossRef]
- 51. Kim, J.W.; Park, S.Y.; You, Y.H.; Ham, D.S.; Lee, S.H.; Yang, H.K.; Jeong, I.K.; Ko, S.H.; Yoon, K.H. Suppression of ROS Production by Exendin-4 in PSC Attenuates the High Glucose-Induced Islet Fibrosis. *PLoS ONE* **2016**, *11*, e0163187. [CrossRef] [PubMed]
- Giacco, F.; Du, X.; Carratu, A.; Gerfen, G.J.; D'Apolito, M.; Giardino, I.; Rasola, A.; Marin, O.; Divakaruni, A.S.; Murphy, A.N.; et al. GLP-1 Cleavage Product Reverses Persistent ROS Generation after Transient Hyperglycemia by Disrupting an ROS-Generating Feedback Loop. *Diabetes* 2015, 64, 3273–3284. [CrossRef] [PubMed]

- Nuamnaichati, N.; Mangmool, S.; Chattipakorn, N.; Parichatikanond, W. Stimulation of GLP-1 Receptor Inhibits Methylglyoxal-Induced Mitochondrial Dysfunctions in H9c2 Cardiomyoblasts: Potential Role of Epac/PI3K/Akt Pathway. *Front. Pharmacol.* 2020, 11, 805. [CrossRef]
- 54. Wei, H.; Bu, R.; Yang, Q.; Jia, J.; Li, T.; Wang, Q.; Chen, Y. Exendin-4 Protects against Hyperglycemia-Induced Cardiomyocyte Pyroptosis via the AMPK-TXNIP Pathway. J. Diabetes Res. **2019**, 2019, 8905917. [CrossRef] [PubMed]
- 55. Tsutsumi, Y.M.; Tsutsumi, R.; Hamaguchi, E.; Sakai, Y.; Kasai, A.; Ishikawa, Y.; Yokoyama, U.; Tanaka, K. Exendin-4 ameliorates cardiac ischemia/reperfusion injury via caveolae and caveolins-3. *Cardiovasc. Diabetol.* **2014**, *13*, 132. [CrossRef] [PubMed]
- Babiker, F.; Al-Kouh, A.; Kilarkaje, N. Lead exposure induces oxidative stress, apoptosis, and attenuates protection of cardiac myocytes against ischemia-reperfusion injury. *Drug Chem. Toxicol.* 2019, 42, 147–156. [CrossRef] [PubMed]
- 57. Babiker, F.A.; Al-Jarallah, A.; Joseph, S. Understanding pacing postconditioning-mediated cardiac protection: A role of oxidative stress and a synergistic effect of adenosine. *J. Physiol. Biochem.* **2017**, *73*, 175–185. [CrossRef] [PubMed]
- 58. Khalaf, A.; Babiker, F. Discrepancy in calcium release from the sarcoplasmic reticulum and intracellular acidic stores for the protection of the heart against ischemia/reperfusion injury. *J. Physiol. Biochem.* **2016**, *72*, 495–508. [CrossRef] [PubMed]
- 59. Babiker, F.A.; Joseph, S.; Juggi, J. The protective effects of 17beta-estradiol against ischemia-reperfusion injury and its effect on pacing postconditioning protection to the heart. *J. Physiol. Biochem.* **2014**, *70*, 151–162. [CrossRef]
- 60. Ferrera, R.; Benhabbouche, S.; Bopassa, J.C.; Li, B.; Ovize, M. One hour reperfusion is enough to assess function and infarct size with TTC staining in Langendorff rat model. *Cardiovasc. Drugs Ther.* **2009**, *23*, 327–331. [CrossRef] [PubMed]