

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



**ScienceDirect** 



## **Monogenic autoimmunity and infectious diseases: the double-edged sword of immune dysregulation** Tarin M Bigley and Megan A Cooper



The study of monogenic autoimmune diseases has provided kev insights into molecular mechanisms involved in development of autoimmunity and immune tolerance. It has also become clear that such inborn errors of immunity (IEIs) frequently present clinically not only with autoimmune diseases, but also frequently have increased susceptibility to infection. The genes associated with monogenic autoimmunity influence diverse functional pathways, and the resulting immune dysregulation also impacts the complex and coordinated immune response to pathogens, for example type I interferon and cytokine signaling, the complement pathway and proper differentiation of the immune response. The SARS-CoV-2 pandemic has highlighted how monogenic autoimmunity can increase risk for serious infection with the discovery of severe disease in patients with pre-existing antibodies to Type I IFNs. This review discusses recent insight into the relationship between monogenic autoimmunity and infectious diseases.

## Address

Department of Pediatrics, Division of Rheumatology/Immunology, Washington University in St. Louis, St. Louis, MO, 63110, United States

Corresponding author: Cooper, Megan A (Cooper\_m@wustl.edu)

## Current Opinion in Immunology 2021, 72:230-238

This review comes from a themed issue on **Host pathogen** 

Edited by Helen C Su and Jean-Laurent Casanova

For a complete overview see the  $\underline{\mathsf{Issue}}$  and the  $\underline{\mathsf{Editorial}}$ 

Available online 12th July 2021

### https://doi.org/10.1016/j.coi.2021.06.013

0952-7915/© 2021 Elsevier Ltd. All rights reserved.

## Introduction

The increased availability and utilization of high-throughput DNA sequencing has brought about a rapid increase in the identification of genes associated with monogenic autoimmunity, which are classified as inborn errors of immunity (IEI) by the International Union of Immunological Sciences Expert Committee [1]. Indeed, the transition in terminology from 'primary immunodeficiency' to 'IEI' reflects the understanding that these diseases are more complex than simply susceptibility to infection. Immune dysregulation with autoimmunity, autoinflammation, cancer susceptibility, and bone marrow failure can all be clinical features of IEIs. Identification of specific genes associated with monogenic autoimmunity and observation of the clinical consequences have accelerated our understanding of the balance between control of infections and autoimmunity [2].

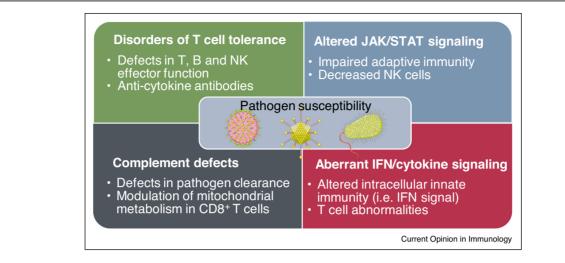
Autoimmune diseases are characterized by abnormal activation of the innate and adaptive immune system leading to self-reactive T and B cells and development of autoantibodies, which lead to inflammation and tissue damage. The genetic risk factors associated with many monogenic autoimmune diseases can also result in increased risk of infection, although there is significant clinical diversity with regards to autoimmune manifestations and pathogen susceptibility [2]. This highlights the diverse pathways that are important for control of both immunologic tolerance and pathogen defense. In this review we discuss recent discoveries in monogenic autoimmunity, highlighting examples of defects in genes involved in T cell tolerance, the complement pathways, interferon signaling, and STAT signaling and their contribution to autoimmunity and pathogen susceptibility (Figure 1), including recent evidence for susceptibility to SARS-CoV-2 associated with monogenic autoimmunity.

# Monogenic autoimmunity and susceptibility to infection

Genes associated with monogenic autoimmunity are involved in diverse immunologic processes. Defects in genes involved in monogenic autoimmunity result in specific patterns of autoimmunity based on their role in different aspects of the immune system. While many patients with monogenic autoimmunity may be at increased risk for infection due to immunomodulatory treatment, we highlight recent discoveries that provide mechanistic insight into how genes involved in monogenic autoimmunity also contribute to increased susceptibility to infection (Figure 1).

## **Disorders of T cell tolerance**

Both central and peripheral mechanisms of T cell tolerance are important for control of self-reactive T cell responses and autoimmunity. Autoimmune endocrinopathies are a common pattern of autoimmunity in disorders of T cell tolerance [1]. Central tolerance of developing T cells occurs in the thymus when thymocytes interact with thymic antigen presenting cells (eg. medullary thymic epithelial cells and thymic dendritic cells) that present a wide array of antigens expressed through the body, known as tissue-restricted antigens (TRAs) [3]. The



Mechanisms of pathogen susceptibility in different groups of monogenic autoimmune disorders highlighted here.

autoimmune regulator (AIRE) transcription factor stimulates expression of TRAs [3]. Deficiency of AIRE is associated with monogenic autoimmunity, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), also known as autoimmune polyendocrine syndrome type 1 (APS-1), due to defects in negative selection of T cells. In addition to autoimmunity due to defects in T cell tolerance, patients with AIRE deficiency suffer from chronic mucocutaneous candidiasis (CMC). In humans, susceptibility to CMC is associated with impaired Th17 immunity, as demonstrated by patients with monogenic defects in IL17F and IL17RA/C and in IEI with impaired Th17 immunity (e.g. STAT1 GOF) [4]. Patients with AIRE deficiency have a high rate of anti-cytokine autoantibodies, including anti-IL-17A, IL17F, and IL22, thought to be due to autoreactive T cell stimulation of B cells [5,6]. These Th17-associated cytokines autoantibodies are associated with susceptibility to CMC in APECED patients, providing a mechanistic link between T cell selection and infectious susceptibility (Figure 2). A recent study in Aire deficient mice suggested that there was an increased Th1 response to *Candida* infection, with similar responses seen in oral mucosa from APECED patients [7<sup>•</sup>]. In this mouse model, CMC was dependent on T cells and IFNy, suggesting an additional mechanism that may contribute to CMC susceptibility. There may also be non-T cell effects of AIRE, with a direct role for AIRE in pathogen response proposed through effects on Dectin-1 and Dectin-2 in macrophages, which are important receptors for recognition and phagocytosis of *Candida* (Figure 2) [8].

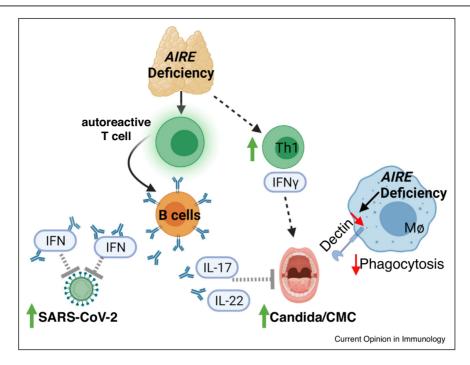
Peripheral tolerance occurs when autoreactive T and B cells escape central tolerance but are inhibited from inducing harm through tolerance checkpoints such as quiescence, ignorance, anergy, exhaustion, senescence

and death (reviewed in detail in Ref. [9]). Several disorders of peripheral tolerance have demonstrated overlap with requirements for pathogen defense. Deficiencies in the CTLA-4 pathway, including regulators of CTLA-4 trafficking, LRBA and DEF6, are associated with autoimmune disease due to defects in T cell tolerance resulting from loss of inhibitory signaling [10]. They are also associated with recurrent bacterial and fungal infections, and increased susceptibility to herpesvirus infections [10-12]. The mechanism for this infectious susceptibility is yet unknown. Patients with CTLA-4 deficiency have decreased CD8<sup>+</sup> T cells and potentially a trend toward increased expression of the senescence marker, CD57 on CD8<sup>+</sup> T cells [13,14]. Interestingly, CTLA-4 was shown to be expressed on activated NK cells, with NK cells from CTLA-4 haploinsufficiency patients showing decreased effector function [15].

IL-2 is an important cytokine for regulatory T cell (Treg) development and effector T cell proliferation, and deficiency of IL-2R $\alpha$  (CD25) results in autoimmune polyendocrinopathy and increased susceptibility to bacterial infections and CMV [16]. IL-2 signaling through IL-2R $\beta$  is known to be important for CD8<sup>+</sup> T cell cytotoxic function and Treg proliferation in mice [17]. Defects in IL-2R $\beta$  had not been observed in humans until two separate groups recently reported human IL-2R $\beta$  deficiency resulting in autoimmunity, atopy and increased EBV and CMV disease in humans [18°,19°]. These studies demonstrate the pleiotropic effects of IL-2 on T cell tolerance and effector function.

Finally, ITCH is a U3 ubiquitin ligase that suppresses inflammation and promotes tolerance through its impact on T cells, especially Th2 cells and Tregs (through enhancement of the *FOXP3* transcription factor), and B





Mechanisms of AIRE deficiency-associated pathogen susceptibility.

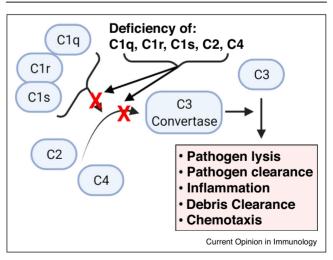
*AIRE* deficiency in the thymus results in altered T cell selection and autoreactive antibodies, including anti-IL-17 antibodies, which are associated with susceptibility to chronic mucocutaneous candidiasis (CMC), and anti-Type I IFN antibodies, which increase disease severity with SARS-CoV-2 infection. Studies in an animal model suggests that a dysregulated Th1 response in the mucosa may also be associated with CMC. *AIRE* deficiency in macrophages (Mø) decreases Dectin-1 and Dectin-2 expression, and may also contribute to defects in phagocytosis of *Candida* and CMC.

cells [20,21]. Autoimmunity associated with a nonsense variant in *ITCH* in a patient was first reported in 2010 [22]. In 2019, two groups reported cases of polyautoimmunity, including autoimmune endocrinopathies, and recurrent respiratory tract infection associated with *ITCH* variants [23,24]. Interestingly, ITCH was shown to be important in the antibody response to viral infection in mice, suggesting a potential mechanism for increased infection in ITCH deficiency [25].

## **Complement deficiencies**

The complement cascade is an important part of the innate immune system, contributing to opsonization, clearance of microbes and cellular debris, and promoting inflammation [26]. Deficiency in early complement proteins is now well-described to be associated with monogenic autoimmunity, and this has been suggested to be due to increased inflammation related to poor pathogen clearance and defective clearance of autoantigens after apoptosis and tissue damage. Deficiencies in C1q, C2 and C4, but not C3, can present with systemic lupus erythematosus (SLE)-like disease and/or recurrent bacterial infections (Figure 3) [1,26]. A recent study in C1q

### Figure 3



Defects in early complement components result in pathogen susceptibility.

Deficiency of early complement components result in impaired formation of the C3 convertase that is required for complementmediated response to pathogens that can also contribute to inflammation in autoimmunity.

deficiency resulted in increased type I IFN response,

which was suggested to be due to loss of STIM1-mediated retention of STING in the ER [39<sup>••</sup>]. How defects in

COPA and STIM1 that result in increased type I IFN

signaling impact susceptibility to infection is unclear, but

a mouse model of STING gain-of-function suggests that

constitutive activation of STING may increase suscepti-

deficient mice showed that C1q can regulate the response of CD8<sup>+</sup> T cells to autoantigens and LCMV infection through modulation of mitochondrial metabolism, establishing a non-complement pathway mediated mechanism for immunodeficiency [27<sup>•</sup>]. C1q may also inhibit type I interferons (IFN) and other cytokines through interaction with Ig-like receptor (LAI) 1 on dendritic cells, inhibiting TLR7 and TLR9-mediated IFN $\alpha$ , IL-6, IL-8 and TNF $\alpha$ expression [28]. The impact of C1q, as well as C1r, on IFN signaling is supported by high type I IFN scores reported in a patient with C1q deficiency-associated SLE [29]. Together, these studies underscore the relevance of the non-canonical complement pathway alterations in autoimmunity and infection.

## Autoimmunity resulting from aberrant interferon and cytokine signaling

Type I IFN signaling is stimulated in response to pathogens and mediates important pathogen responses. Increased type I IFN signaling is observed in polygenic autoimmune disease such as SLE, as well as monogenic disorders of the type I IFN pathway, called interferonopathies [30]. Interferonopathies are typically associated with autoinflammatory manifestations rather than autoimmune disease, however autoimmune features are observed in some interferonopathies as is the case for *IFIH1* and *TREX1* which are associated with Aicardi-Goutières syndrome and lupus-like disease [1]. Several recent studies have evaluated how type I IFN signaling is dysregulated in specific monogenic autoimmune diseases and given insight into susceptibility to infection.

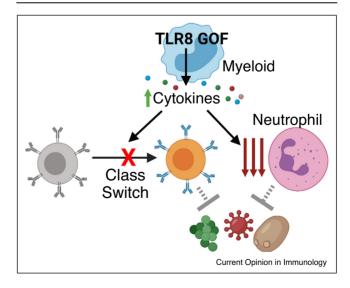
COPA syndrome is an autoimmune disease characterized by pulmonary hemorrhage, interstitial lung disease, arthritis, nephritis and autoantibodies caused by dominant variants in the COPA gene, the product of which is involved in transport of vesicle between with ER and Golgi [31]. Increased susceptibility to infection is not typically described, although a case series reported several patients who presented with multiple infections including acute pyelonephritis, meningitis and recurrent respiratory infections [32]. Recent studies have clarified that a key feature of this disorder is increased type I IFN signaling due to inability of mutant COPA protein to transport active STING from the Golgi to the endoplasmic reticulum  $[33^{\bullet}, 34^{\bullet}, 35]$ . Deng *et al.* showed that  $Copa^{E241K/+}$  knockin mice develop autoreactive T cells, decreased Tregs in the periphery, and autoimmune disease [36<sup>•</sup>]. Moreover, negative thymic selection of autoreactive thymocytes by thymic epithelial cells was impaired.

STIM1 deficiency manifests as recurrent bacterial infections, autoimmune hemolytic anemia, nephrotic syndrome, as well as dysmorphic features [37]. STIM1 is a calcium channel that was recently shown to influence Th17 cell metabolism, which is important for antifungal immunity [38]. Interestingly, similar to COPA, STIM1

TNFα bility to infection with a herpesvirus [40]. Additional studies of patient samples and *in vivo* models of COPA scores syndrome and STIM1 deficiency will increase our understanding of how dysregulated IFN responses influence susceptibility and response to infections.
 Toll-like receptor 8 (TLR8) is an endosomal sensor of single stranded RNA [41]. We recently described an IEI due to monogenic mosaic and germline variants in TLR8 associated with immune dysregulation, susceptibility to infection and bone marrow failure. Patients had elevated

Infection and bone marrow failure. Patients had elevated serum cytokines, including IFN- $\gamma$  and others such as IL-18 and IL12/23 p40, although interestingly these patients do not have a strong Type I IFN signature [42]. Patients with TLR8 GOF have neutropenia, and immune phenotyping demonstrated highly activated peripheral blood T cells and defects in class-switched B cells. Differentiation of macrophages from patient-derived induced pluripotent stem cells showed increased cytokine responses to TLR8 ligand stimulation. Patients with TLR8 GOF have infections related to these immunologic defects (Figure 4). While the mechanism of this immune suppression is uncertain, the phenotype in mosaic patients suggests a dominant and non-intrinsic effect of cytokines





Mechanisms of pathogen susceptibility in TLR8 GOF. TLR8 is expressed primarily in myeloid cells. Patients with TLR8 GOF have increased serum cytokines and production of pro-inflammatory cytokines in patient-derived macrophages. This cytokine expression is hypothesized to lead to impaired class switching of B cells and severe neutropenia, with resulting infectious susceptibility.

Table 1

Monogenic autoimmune disease with aberrant JAK/STAT signaling

### Molecular effects Immunologic defects Disease Gene Inheritance Autoimmunity Pathogen susceptibility JAK1 GOF [62] JAK1 AD (GOF) Increased STAT1 phosphorylation Eosinophilia with normal IgE, atopy AITD Recurrent viral infections as baseline, increased STAT3 phosphorylation after IL-6 stimulation SOCS1 haplo-insufficiency SOCS1 AD (LOF) Increased STAT1 and STAT5 Lymphoproliferation in some Respiratory infections Autoimmune cytopenias, [52°,53°,54°] phosphorylation after IFN<sub>y</sub>, IL-2, patients. Decreased Treg numbers. psoriasis, SLE, polyarthritis, and IL-4 stimulation Low switched memory B cells spondyloarthritis, celiac disease. AITD. AIH. autoimmune pancreatitis STAT1 deficiency STAT1 AD or AR Deficient STAT1 signaling including Impaired Th1 and Th17 immunity AITD, autoimmune CMC, recurrent bacterial [1,63] (LOF) **IFN** responses cytopenias, celiac disease infections (mycobacterium). herpes and fungal infections STAT1 GOF [43-45] STAT1 AD (GOF) STAT1 hyperphosphorylation and Impaired Th17 and IL-22 immunity. AITD, T1DM, autoimmune Increased bacteria and herpes transcriptional activity and impaired Decreased Candida-specific Th17 infections, Cryptococcus cytopenias, psoriasis, SLE, STAT3 activation and Th22 response. Rarely B cell scleroderma, AIH, IBD, infections, CMC lymphopenia and celiac disease hypogammaglobulinemia. STAT2 GOF [64] STAT2 AR (GOF) Abrogated ubiguitin-specific Decreased NK and CD8<sup>+</sup> T cell Autoinflammatory: fever, None reported protease 18 (USP18) interaction granule release. T cell lymphopenia cytopenias, nephropathy, resulting in elevated type I IFN and hypogammaglobulinemia in one hepatosplenomegaly, signaling and prolong JAK-STAT patient. elevated liver enzymes. sianalina. thrombotic microangiopathy, seizures, intracerebral hemorrhage. STAT3 GOF [46,47] STAT3 AD (GOF) Increased STAT3 transcriptional Reduced T and B cell numbers, Enteropathy, autoimmune Varied including viral respiratory hypogammaglobulinemia, activity with stimulation, possibly cytopenias, ILD, T1DM, tract infections, herpesvirus decreased STAT1 and STAT5 decreased peripheral Treg numbers, AITD, arthritis, AIH, infections. bacterial infections phosphorylation. in some patients increased Th17 scleroderma cells STAT5b deficiency STAT5B AR Defects in STAT5b signaling and Reduced Treg number/function, AITD, autoantibodies, Viral respiratory tract infections, [65] target genes including immune high IgG and IgE, reduced $\gamma \delta$ -T and arthritis, ITP lymphocytic Pneumocystis jirovecii pathways and IGF1 NK cells interstitial pneumonitis pneumonia, severe herpesvirus infections TOM1 LOF [50] TOM1 AD Impaired TOM1-TOLLIP interaction Low memory and class switch B Oligoarthritis, psoriasis, Recurrent respiratory infections, resulting in decreased STAT1. cells. hvpo-gammaglobulinemia. autoimmune enteropathy. EBV viremia STAT3, and STAT5 phosphorylation. Impaired Th1 and Th17 ILD differentiation. Deceased T effector memory.

Abbreviations: ADautosomal dominant; AlHautoimmune hepatitis; AlHAautoimmune hemolytic anemia; AlTDautoimmune thyroid disease; ARautosomal recessive; GOFgain of function; ITPimmune thrombocytopenic purpura; JIAjuvenile idiopathic arthritis; LOFloss of function; CMCchronic mucocutaneous candidiasis; SLEsystemic lupus erythematosus; T1DMtype 1 diabetes mellitus.

in these patients, potentially within the bone marrow microenvironment.

## STAT signaling

The JAK/STAT pathway plays an important role in transducing signals from extracellular cytokines to intracellular signaling pathways that mediate a variety of responses. Alterations in JAK/STAT signaling are now well-described in monogenic autoimmunity including STAT1, STAT3, and STAT5b, with prominent T cell dysregulation, infectious susceptibility, and an autoimmune pattern including cytopenias, thyroid disease, and gastrointestinal disease being common features (Table 1). Gain-of-function (GOF) *STAT1* variants result in impairment of IL-17 and IL-22 mediated immunity and abnormal B cell differentiation, contributing to humoral defects and increased risk of candidiasis [43–45]. Patients with STAT3 GOF have early onset autoimmunity, but also susceptibility to infection associated with antibody deficiency [46,47].

TOM1 (target of Myb protein 1) is a lysosomal adapter protein thought to be involved maturation of the autophagosome and trafficking of endosomal cargo [48,49]. A newly described pathogenic variant in TOM1 (G307D) presented clinically as oligoarthritis, psoriasis, autoimmune enteropathy, interstitial lung disease, hypogammaglobulinemia, recurrent URIs, and EBV viremia [50]. This variant was associated with decreased STAT1. 3 and 5 signaling, poor Th1 and Th17 differentiation, low class switch memory B cells, as well as decreased NK cell numbers. This was attributed to impaired TOM1 interaction with TOLLIP. The mechanism resulting in altered STAT signaling in TOM1-associated disease is yet unknown, although TOLLIP deficiency has been demonstrated to decrease STAT5 expression, perhaps through its function in autophagy [51]

Another recent example of altered STAT signaling associated with autoimmunity and infections is that of haploinsufficiency of suppressor of cytokine signaling-1 (SOCS1). SOCS1 attenuates signaling of multiple STATs (particularly STAT1) and is upregulated as a compensatory response to JAK/STAT signaling. These patients have early onset autoimmunity including cytopenias and even systemic lupus erythematosus, highlighting the importance of tight regulation of this system in human immunity [52°,53°,54°]. Together, these studies demonstrate the relationship between autoimmunity and pathogen susceptibility in dysregulated STAT and cytokine signaling.

## Lessons learned from SARS-CoV-2

The SARS-CoV-2 pandemic has provided insight into the variability of the immune response to a pathogen infecting a heterogenous and immunologically naïve population. Type I IFN signaling has emerged as a critical requirement for an effective immune response to acute SARS-CoV2 infection. In addition, immune dysregulation has clearly

been induced in some COVID-19 patients as seen in multisystem inflammatory syndrome in children (MIS-C), with the characteristic features and clinical course suggestive of an underlying immunologic defect in these previously healthy children brought out by this unique virus [55,56].

Genomic sequencing of a cohort of 650 individuals with severe acute COVID-19 identified 3.5% of patients with monogenic defects in the type I IFN signaling pathway [57]. Functional studies proving pathogenicity of both known and novel defects demonstrated the importance of this pathway for efficient control of SARS-CoV2. In a complementary study of almost 1000 patients without genetic defects in the type I IFN pathway, neutralizing autoantibodies against type I IFNs were associated with severe COVID-19, illustrating a mechanism by which loss of tolerance can directly lead to infection susceptibility [58]. Patients with AIRE deficiency and APECED/APS-I almost uniformly develop neutralizing type I IFN autoantibodies, which previously were not recognized to be associated with infectious susceptibility [59]. The Casanova and Lionakis laboratories recently demonstrated that in contrast to most other patients with IEIs, patients with AIRE deficiency and COVID-19 develop severe pneumonia and suffer from high mortality associated with these pre-existing auto-antibodies (Figure 2) [60<sup>••</sup>]. This finding provides a direct link between monogenic autoimmunity, autoantibodies, and infectious susceptibility. While in MIS-C, there has not yet been a report of a monogenic defect clearly associated with susceptibility. the description of MIS-C in a patient with haploinsufficiency of SOCS1, a monogenic autoimmune/immune dysregulation syndrome, supports a role of cytokine/ STAT signaling in control of inflammation after infection in children [53<sup>•</sup>]. Early studies of SARS-CoV-2 infections in patients with IEIs did not report exacerbation of their autoimmune disease but did suggest that young patients with IEIs were more likely to develop severe symptoms than age-matched controls [61]. More detailed information and collection of immune phenotyping from SARS-CoV-2 infected patients with monogenic autoimmunity will be informative.

## Conclusion

Immune dysfunction in IEIs can lead to autoimmune diseases with auto-reactivity of T and B cells, increased activity of the innate immune response, and in some cases increased susceptibility to infection. While T and B cell dysfunction contributes to autoimmune and infectious sequelae, recent studies have brought to light molecular pathways such as abnormalities in IFN and cytokine signaling that contribute to pathogen susceptibility in monogenic autoimmunity. Although there are similarities in the immunologic aberrancies amongst monogenic autoimmune disease, future studies describing the differential effects of specific genes will be needed to resolve why certain monogenic autoimmune diseases are associated with unique or shared pathogen susceptibility. Certainly, the COVID-19 pandemic has and will continue to provide new insights into the interaction between pathogens and genes that regulate the immune system. New in vivo models and advancements in the study of human immunology will also expand our understanding of the impact of genes associated with monogenic autoimmunity on the complex relationship between maintaining self-tolerance and controlling infection.

## Conflict of interest statement

Nothing declared.

## Acknowledgements

This work was supported by the Jeffrey Modell Diagnostic and Research Center for Primary Immunodeficiencies at St. Louis Children's Hospital and the St. Louis Children's Hospital Foundation. Figures 2-4 were created with BioRender.com.

## References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- · of special interest
- •• of outstanding interest
- Tangye SG, Al-Herz W, Bousfiha A, Chatila T, Cunningham-Rundles C, Etzioni A, Franco JL, Holland SM, Klein C, Morio T *et al*.: 1. Human inborn errors of immunity: 2019 update on the classification from the International Union of Immunological Societies Expert Committee. J Clin Immunol 2020, 40:24-64
- Amaya-Uribe L, Rojas M, Azizi G, Anaya JM, Gershwin ME: 2. Primary immunodeficiency and autoimmunity: a comprehensive review. J Autoimmun 2019, 99:52-72.
- Cheng M, Anderson MS: Thymic tolerance as a key brake on 3. autoimmunity. Nat Immunol 2018, 19:659-664
- Okada S, Puel A, Casanova JL, Kobayashi M: Chronic mucocutaneous candidiasis disease associated with inborn errors of IL-17 immunity. *Clin Transl Immunol* 2016, **5**:e114. 4.
- Puel A, Döffinger R, Natividad A, Chrabieh M, Barcenas-5. Morales G, Picard C, Cobat A, Ouachée-Chardin M, Toulon A Bustamante J et al.: Autoantibodies against IL-17A, IL-17F, and IL-22 in patients with chronic mucocutaneous candidiasis and autoimmune polyendocrine syndrome type I. J Exp Med 2010, 207:291-297
- Kisand K, Bøe Wolff AS, Podkrajsek KT, Tserel L, Link M, 6. Kisand KV, Ersvaer E, Perheentupa J, Erichsen MM, Bratanic N et al.: Chronic mucocutaneous candidiasis in APECED or thymoma patients correlates with autoimmunity to Th17associated cytokines. J Exp Med 2010, 207:299-308.
- 7.
- Break TJ, Oikonomou V, Dutzan N, Desai JV, Swidergall M, Freiwald T, Chauss D, Harrison OJ, Alejo J, Williams DW *et al.*: Aberrant type 1 immunity drives susceptibility to mucosal fungal infections. *Science* 2021, **371**

Patients with *AIRE* deficiency are susceptible to chronic mucocutaneous candidasis (CMC), previously thought to be due to autoantibodies against IL-17 and Th17-pathway cytokines. This report used a mouse model and samples from patients to demonstrate intact Th17 responses at mucosal surfaces in response to Candida, and identified pathogenic Th1 and IFN- $\gamma$ responses associated with infection

de Albuquerque JAT, Banerjee PP, Castoldi A, Ma R, Zurro NB, 8 Ynoue LH, Arslanian C, Barbosa-Carvalho MUW, Correia-Deur JEM, Weiler FG et al.: The role of AIRE in the immunity against Candida albicans in a model of human macrophages. Front Immunol 2018, 9:567.

- ElTanbouly MA, Noelle RJ: Rethinking peripheral T cell 9. tolerance: checkpoints across a T cell's journey. Nat Rev Immunol 2021, 21:257-267.
- 10. Mitsuiki N, Schwab C, Grimbacher B: What did we learn from CTLA-4 insufficiency on the human immune system? Immunol Rev 2019, 287:33-49.
- 11. Garcia-Perez JE, Baxter RM, Kong DS, Tobin R, McCarter M, Routes JM, Verbsky J, Jordan MB, Dutmer CM, Hsieh EWY CTLA4 message reflects pathway disruption in monogenic disorders and under therapeutic blockade. Front Immunol 2019. 10:998
- Serwas NK, Hoeger B, Ardy RC, Stulz SV, Sui Z, Memaran N, 12. Meeths M, Krolo A, Yuce Petronczki O, Pfajfer L et al.: Human DEF6 deficiency underlies an immunodeficiency syndrome with systemic autoimmunity and aberrant CTLA-4 homeostasis. Nat Commun 2019, 10:3106.
- 13. Hoshino A, Tanita K, Kanda K, Imadome KI, Shikama Y, Yasumi T, Imai K, Takagi M, Morio T, Kanegane H: High frequencies of asymptomatic Epstein-Barr virus viremia in affected and unaffected individuals with CTLA4 mutations. Clin Immunol 2018, 195:45-48.
- 14. Schwab C, Gabrysch A, Olbrich P, Patino V, Warnatz K, Wolff D, Hoshino A, Kobayashi M, Imai K, Takagi M et al.: Phenotype, penetrance, and treatment of 133 cytotoxic T-lymphocyte antigen 4-insufficient subjects. J Allergy Clin Immunol 2018, 142:1932-1946.
- 15. Lougaris V, Tabellini G, Baronio M, Patrizi O, Gazzurelli L Mitsuiki N, Pozzi MR, Grimbacher B, Parolini S, Plebani A: CTLA-4 regulates human natural killer cell effector functions. Clin Immunol 2018, 194:43-45.
- 16. Goudy K, Aydin D, Barzaghi F, Gambineri E, Vignoli M, Ciullini Mannurita S, Doglioni C, Ponzoni M, Cicalese MP, Assanelli A et al.: Human IL2RA null mutation mediates immunodeficiency with lymphoproliferation and autoimmunity. Clin Immunol 2013, **146**:248-261
- 17. Suzuki H, Kundig TM, Furlonger C, Wakeham A, Timms E, Matsuyama T, Schmits R, Simard JJ, Ohashi PS, Griesser H et al.: Deregulated T cell activation and autoimmunity in mice lacking interleukin-2 receptor beta. Science 1995, 268:1472-1476

18. Fernandez IZ, Baxter RM, Garcia-Perez JE, Vendrame E,

Ranganath T, Kong DS, Lundquist K, Nguyen T, Ogolla S, Black J et al.: A novel human IL2RB mutation results in T and NK celldriven immune dysregulation. J Exp Med 2019. 216:1255-1267 Two siblings with polyautoimmunity and increased susceptibility to CMV were found to have a homozygous, hypomorphic variant in *IL2RB* that resulted in decreased surface expression. Both patients had a reduction on Tregs and dysregulated IL-2 and IL-15 signaling and STAT5 phos-phorylation, as well as increased CD56<sup>bright</sup>, immature, NK cells. Together with Ref. [13•], this study was the first to report *IL2RB* deficiency causing autoimmune disease and immunodeficiency.

- Zhang Z, Gothe F, Pennamen P, James JR, McDonald D, Mata CP,
   Modis Y, Alazami AM, Acres M, Haller W *et al.*: Human
- interleukin-2 receptor beta mutations associated with defects in immunity and peripheral tolerance. J Exp Med 2019, 216:1311-1327

Eight individuals from four consanguineous families presented with autoantibodies, bowel inflammation, lymphadenopathy, skin disease and CMV infections. Three different homozygous variants in *IL2RB* were identified by whole exome sequencing. T cells from patients lacked IL2-Rb surface expression and were unresponsive to IL-2 stimulation ex vivo. NK cells demonstrated partial IL2-Rb protein expression compared to controls. One patient was successfully treated with stem cell transplant, showing additional example of monogenic autoimmune dis-ease caused by IL2-Rb deficiency.

- Parravicini V, Field AC, Tomlinson PD, Basson MA, Zamoyska R: 20. Itch<sup>-/-</sup> alphabeta and gammadelta T cells independently contribute to autoimmunity in Itchy mice. Blood 2008, 111:4273-7282.
- 21. Venuprasad K, Huang H, Harada Y, Elly C, Subramaniam M, Spelsberg T, Su J, Liu YC: The E3 ubiquitin ligase Itch regulates expression of transcription factor Foxp3 and airway

inflammation by enhancing the function of transcription factor TIEG1. *Nat Immunol* 2008, **9**:245-253.

- Lohr NJ, Molleston JP, Strauss KA, Torres-Martinez W, Sherman EA, Squires RH, Rider NL, Chikwava KR, Cummings OW, Morton DH et al.: Human ITCH E3 ubiquitin ligase deficiency causes syndromic multisystem autoimmune disease. Am J Hum Genet 2010, 86:447-453.
- Brittain HK, Feary J, Rosenthal M, Spoudeas H, Deciphering Developmental Disorders S, Wilson LC: Biallelic human ITCH variants causing a multisystem disease with dysmorphic features: a second report. Am J Med Genet A 2019, 179:1346-1350.
- 24. Kleine-Eggebrecht N, Staufner C, Kathemann S, Elgizouli M, Kopajtich R, Prokisch H, Lainka E: Mutation in ITCH gene can cause syndromic multisystem autoimmune disease with acute liver failure. *Pediatrics* 2019, 143.
- 25. Xiao N, Eto D, Elly C, Peng G, Crotty S, Liu YC: The E3 ubiquitin ligase itch is required for the differentiation of follicular helper T cells. *Nat Immunol* 2014, **15**:657-666.
- 26. Lintner KE, Wu YL, Yang Y, Spencer CH, Hauptmann G, Hebert LA, Atkinson JP, Yu CY: Early components of the complement classical activation pathway in human systemic autoimmune diseases. Front Immunol 2016, 7:36.
- 27. Ling GS, Crawford G, Buang N, Bartok I, Tian K, Thielens NM,
   Bally I, Harker JA, Ashton-Rickardt PG, Rutschmann S et al.: C1q restrains autoimmunity and viral infection by regulating CD8 (+) T cell metabolism. Science 2018, 360:558-563

Using a mouse model of C1q deficiency, it was demonstrated that C1q, but not C3, reduces the CD8<sup>+</sup> T cell response to autoantigens while increasing the effector CD8<sup>+</sup> T cell response to chronic LCMV infection. This appears to be due to modulating expression of mitochondrial metabolism genes in memory precursor effector cells.

- Lood C, Gullstrand B, Truedsson L, Olin AI, Alm GV, Ronnblom L, Sturfelt G, Eloranta ML, Bengtsson AA: C1q inhibits immune complex-induced interferon-alpha production in plasmacytoid dendritic cells: a novel link between C1q deficiency and systemic lupus erythematosus pathogenesis. *Arthritis Rheum* 2009, 60:3081-3090.
- Bolin K, Eloranta ML, Kozyrev SV, Dahlqvist J, Nilsson B, Knight A, Ronnblom L: A case of systemic lupus erythematosus with C1q deficiency, increased serum interferon-alpha levels and high serum interferogenic activity. *Rheumatology (Oxford)* 2019, 58:918-919.
- Uggenti C, Lepelley A, Crow YJ: Self-awareness: nucleic aciddriven inflammation and the type I interferonopathies. Annu Rev Immunol 2019, 37:247-267.
- Watkin LB, Jessen B, Wiszniewski W, Vece TJ, Jan M, Sha Y, Thamsen M, Santos-Cortez RL, Lee K, Gambin T *et al.*: COPA mutations impair ER-Golgi transport and cause hereditary autoimmune-mediated lung disease and arthritis. *Nat Genet* 2015, 47:654-660.
- Taveira-DaSilva AM, Markello TC, Kleiner DE, Jones AM, Groden C, Macnamara E, Yokoyama T, Gahl WA, Gochuico BR, Moss J: Expanding the phenotype of COPA syndrome: a kindred with typical and atypical features. J Med Genet 2019, 56:778-782.
- Deng Z, Chong Z, Law CS, Mukai K, Ho FO, Martinu T, Backes BJ,
   Eckalbar WL, Taguchi T, Shum AK: A defect in COPI-mediated transport of STING causes immune dysregulation in COPA

syndrome. J Exp Med 2020, 217 Variants in COPA that are associated with COPA syndrome cause STING activation and retention in the Golgi due to failed retrieval by COPI complexes. This is potentially due to failed interaction between STING, COPA and SURF4. In Copa<sup>E241K/+</sup> knock-in mice, medullary thymic epithelial cells and thymocytes in the medulla show increased expression of ISGs and altered thymocyte development, both of which are dependent on STING expression. This report establishes a potential link between COPA, abnormal STING and IFN signaling, and thymic selection.

- 34. Lepelley A, Martin-Niclos MJ, Le Bihan M, Marsh JA, Uggenti C,
- Rice GI, Bondet V, Duffy D, Hertzog J, Rehwinkel J et al.:

Mutations in COPA lead to abnormal trafficking of STING to the Golgi and interferon signaling. *J Exp Med* 2020, 217

Increased type I IFN signaling was observed in the blood of COPA patients. *In vitro* expression of mutant COPA resulted in a STING-dependent increase in type I IFN signaling as well as accumulation of STING in the Golgi. This study shows a potential pathogenic mechanism whereby variants in COPA lead to type I IFN dysregulation through STING transport.

- Volpi S, Tsui J, Mariani M, Pastorino C, Caorsi R, Sacco O, Ravelli A, Shum AK, Gattorno M, Picco P: Type I interferon pathway activation in COPA syndrome. *Clin Immunol* 2018, 187:33-36.
- 36. Deng Z, Law CS, Ho FO, Wang KM, Jones KD, Shin JS, Shum AK:
   A defect in thymic tolerance causes T Cell-mediated autoimmunity in a murine model of COPA syndrome. J Immunol 2020, 204:2360-2373
   A Copa<sup>E241K/+</sup> knock-in mouse developed interstitial lung disease similar

A Copa<sup>E241N+</sup> knock-in mouse developed interstitial lung disease similar to COPA patients as well as impaired thymic selection resulting in autoreactive T cells, impaired production of antigen-specific (OVA) Tregs, and decreased numbers of Tregs in the periphery. This report establishes the impact of COPA on T cell selection.

- Picard C, McCarl CA, Papolos A, Khalil S, Luthy K, Hivroz C, LeDeist F, Rieux-Laucat F, Rechavi G, Rao A et al.: STIM1 mutation associated with a syndrome of immunodeficiency and autoimmunity. N Engl J Med 2009, 360:1971-1980.
- Kahlfuss S, Kaufmann U, Concepcion AR, Noyer L, Raphael D, Vaeth M, Yang J, Pancholi P, Maus M, Muller J et al.: STIM1mediated calcium influx controls antifungal immunity and the metabolic function of non-pathogenic Th17 cells. EMBO Mol Med 2020, 12:e11592.
- 39. Srikanth S, Woo JS, Wu B, El-Sherbiny YM, Leung J, Chupradit K,
- Bitcaling two by Walby Electroning TW, Dealing G, Orlipitadirk,
   Rice L, Seo GJ, Calmettes G, Ramakrishna C et al.: The Ca(2+) sensor STIM1 regulates the type I interferon response by retaining the signaling adaptor STING at the endoplasmic reticulum. Nat Immunol 2019, 20:152-162

Deficiency of *STIM1* induces STING-dependent type I IFN signaling *in vitro* in mouse and human cells. STIM1 interacts with STING and inhibits its trafficking from the ER. In a mouse model, *STIM1* deficiency decreased HSV-1 genomic GFP expression and murine herpesevirus-68 (MHV68) early and late gene expression *in vitro*, and decreased HSV-1 induced mortality and viral load in the brain, suggesting that loss of *STIM1* impacts viral replication and host defense.

- Bennion BG, Ingle H, Ai TL, Miner CA, Platt DJ, Smith AM, Baldridge MT, Miner JJ: A human gain-of-function STING mutation causes immunodeficiency and gammaherpesvirusinduced pulmonary fibrosis in mice. J Virol 2019, 93.
- Moen SH, Ehrnstrom B, Kojen JF, Yurchenko M, Beckwith KS, Afset JE, Damas JK, Hu Z, Yin H, Espevik T et al.: Human toll-like receptor 8 (TLR8) is an important sensor of pyogenic bacteria, and is attenuated by cell surface TLR signaling. Front Immunol 2019, 10:1209.
- 42. Aluri J, Bach A, Kaviany S, Chiquetto Paracatu L, Kitcharoensakkul M, Walkiewicz MA, Putnam CD, Shinawi M, Saucier N, Rizzi EM et al.: Immunodeficiency and bone marrow failure with mosaic and germline TLR8 gain of function. Blood 2021, 137:2450-2462.
- Hiller J, Hagl B, Effner R, Puel A, Schaller M, Mascher B, Eyerich S, Eyerich K, Jansson AF, Ring J et al.: STAT1 gain-of-function and dominant negative STAT3 mutations impair IL-17 and IL-22 immunity associated with CMC. J Invest Dermatol 2018, 138:711-714.
- Nemoto K, Kawanami T, Hoshina T, Ishimura M, Yamasaki K, Okada S, Kanegane H, Yatera K, Kusuhara K: Impaired B-cell differentiation in a patient with STAT1 gain-of-function mutation. Front Immunol 2020, 11:557521.
- Tamaura M, Satoh-Takayama N, Tsumura M, Sasaki T, Goda S, Kageyama T, Hayakawa S, Kimura S, Asano T, Nakayama M et al.: Human gain-of-function STAT1 mutation disturbs IL-17 immunity in mice. Int Immunol 2020, 32:259-272.
- Fabre A, Marchal S, Barlogis V, Mari B, Barbry P, Rohrlich PS, Forbes LR, Vogel TP, Giovannini-Chami L: Clinical aspects of STAT3 gain-of-function germline mutations: a systematic review. J Allergy Clin Immunol Pract 2019, 7:1958-1969 e1959.

- Milner JD, Vogel TP, Forbes L, Ma CA, Stray-Pedersen A, Niemela JE, Lyons JJ, Engelhardt KR, Zhang Y, Topcagic N *et al.*: Early-onset lymphoproliferation and autoimmunity caused by germline STAT3 gain-of-function mutations. *Blood* 2015, 125:591-599.
- Tumbarello DA, Waxse BJ, Arden SD, Bright NA, Kendrick-Jones J, Buss F: Autophagy receptors link myosin VI to autophagosomes to mediate Tom1-dependent autophagosome maturation and fusion with the lysosome. Nat Cell Biol 2012, 14:1024-1035.
- Xiao S, Brannon MK, Zhao X, Fread KI, Ellena JF, Bushweller JH, Finkielstein CV, Armstrong GS, Capelluto DGS: Tom1 modulates binding of tollip to phosphatidylinositol 3-phosphate via a coupled folding and binding mechanism. *Structure* 2015, 23:1910-1920.
- Keskitalo S, Haapaniemi EM, Glumoff V, Liu X, Lehtinen V, Fogarty C, Rajala H, Chiang SC, Mustjoki S, Kovanen P et al.: Dominant TOM1 mutation associated with combined immunodeficiency and autoimmune disease. NPJ Genom Med 2019, 4:14.
- Kowalski E, Geng S, Rathes A, Lu R, Li L: Toll-interacting protein differentially modulates HIF1alpha and STAT5-mediated genes in fibroblasts. J Biol Chem 2018, 293:12239-12247.
- 52. Hadjadj J, Castro CN, Tusseau M, Stolzenberg MC, Mazerolles F,
- Aladjidi N, Armstrong M, Ashrafian H, Cutcutache I, Ebetsberger-Dachs G et al.: Early-onset autoimmunity associated with

**SOCS1 haploinsufficiency**. *Nat Commun* 2020, **11**:5341 Whole exome sequencing of patients with early onset autoimmunity revealed heterozygous loss-of-function variants in *SOCS1*. Lymphocytes from these patients demonstrated increased STAT activation after *in vitro* stimulation with IFN- $\gamma$ , IL-2 and IL-4, which was inhibited by a JAK1/2 inhibitor. This study describes the disease phenotype and immunologic dysfunction associated with *SOCS1* loss-of-function.

Lee PY, Platt CD, Weeks S, Grace RF, Maher G, Gauthier K,
 Devana S, Vitali S, Randolph AG, McDonald DR *et al.*: Immune dysregulation and multisystem inflammatory syndrome in children (MIS-C) in individuals with haploinsufficiency of SOCS1. J Allergy Clin Immunol 2020, 146:1194-1200 e1191

Two patients with autoimmune cytopenias, one of whom developed MIS-C after a SARS-CoV-2 infection, were found to have heterozygous variants in *SOCS1*. PBMCs from these patients exhibited increased STAT1 phosphorylation, type I and II IFN signaling and pro-apoptotic gene expression. This report suggests that disruption of regulation of JAK/STAT signaling is associated with risk of MIS-C.

54. Thaventhiran JED, Lango Allen H, Burren OS, Rae W, Greene D,
Staples E, Zhang Z, Farmery JHR, Simeoni I, Rivers E et al.: Whole-genome sequencing of a sporadic primary immunodeficiency cohort. Nature 2020, 583:90-95
Whole genome sequencing of 138 patients with primary immune defitional sequencing of the sequence of

Whole genome sequencing of 138 patients with primary immune deficiency, mainly adults, was utilized to identify disease-causing variants in genes and non-coding regulatory regions. Genome-wide association study was also used to establish potential contribution of common variants to monogenic IEI. Functional studies of variants from patients with compound heterozygous common and rare variants in *PTPN2* or *SOCS1*, both of which are negative regulators of STAT signaling, demonstrated potential interaction between common and pathogenic variants.

- Diorio C, Henrickson SE, Vella LA, McNerney KO, Chase J, Burudpakdee C, Lee JH, Jasen C, Balamuth F, Barrett DM et al.: Multisystem inflammatory syndrome in children and COVID-19 are distinct presentations of SARS-CoV-2. J Clin Invest 2020, 130:5967-5975.
- Sancho-Shimizu V, Brodin P, Cobat A, Biggs CM, Toubiana J, Lucas CL, Henrickson SE, Belot A, Mis CC, Tangye SG et al.: SARS-CoV-2-related MIS-C: a key to the viral and genetic causes of Kawasaki disease? J Exp Med 2021, 218.
- Zhang Q, Bastard P, Liu Z, Le Pen J, Moncada-Velez M, Chen J, Ogishi M, Sabli IKD, Hodeib S, Korol C et al.: Inborn errors of type I IFN immunity in patients with life-threatening COVID-19. Science 2020, 370.
- Bastard P, Rosen LB, Zhang Q, Michailidis E, Hoffmann HH, Zhang Y, Dorgham K, Philippot Q, Rosain J, Beziat V *et al.*: Autoantibodies against type I IFNs in patients with lifethreatening COVID-19. *Science* 2020, 370.
- Meager A, Visvalingam K, Peterson P, Möll K, Murumägi A, Krohn K, Eskelin P, Perheentupa J, Husebye E, Kadota Y *et al.*: Anti-interferon autoantibodies in autoimmune polyendocrinopathy syndrome type 1. *PLoS Med* 2006, 3:e289.
- Bastard P, Orlova E, Sozaeva L, Levy R, James A, Schmitt MM,
   Ochoa S, Kareva M, Rodina Y, Gervais A et al.: Preexisting autoantibodies to type I IFNs underlie critical COVID-19 pneumonia in patients with APS-1. J Exp Med 2021. 218

pneumonia in patients with APS-1. J Exp Med 2021, 218 This publication reported 22 patients with APS-1 (also known as APECED) due to AIRE deficiency who were infected with SARS-CoV-2. Patients with APS-1 are known to have neutralizing antibodies against type I IFNs, and the patients tested in this cohort all had autoantibodies to IFN- $\alpha$  and/ or IFN- $\omega$ . In this cohort, a striking 86% of patients were hospitalized for COVID-19 pneumonia and 18% died, suggesting high morbidity and mortality with pre-existing type I IFN antibodies.

- Meyts I, Bucciol G, Quinti I, Neven B, Fischer A, Seoane E, Lopez-Granados E, Gianelli C, Robles-Marhuenda A, Jeandel PY et al.: Coronavirus disease 2019 in patients with inborn errors of immunity: an international study. J Allergy Clin Immunol 2020, 147:520-531.
- 62. Del Bel KL, Ragotte RJ, Saferali A, Lee S, Vercauteren SM, Mostafavi SA, Schreiber RA, Prendiville JS, Phang MS, Halparin J et al.: JAK1 gain-of-function causes an autosomal dominant immune dysregulatory and hypereosinophilic syndrome. J Allergy Clin Immunol 2017, 139:2016-2020 e2015.
- Kim HS, Kim DC, Kim HM, Kwon HJ, Kwon SJ, Kang SJ, Kim SC, Choi GE: STAT1 deficiency redirects IFN signalling toward suppression of TLR response through a feedback activation of STAT3. Sci Rep 2015, 5:13414.
- Duncan CJA, Thompson BJ, Chen R, Rice GI, Gothe F, Young DF, Lovell SC, Shuttleworth VG, Brocklebank V, Corner B et al.: Severe type I interferonopathy and unrestrained interferon signaling due to a homozygous germline mutation in STAT2. Sci Immunol 2019, 4.
- 65. Kanai T, Jenks J, Nadeau KC: The STAT5b pathway defect and autoimmunity. Front Immunol 2012, 3:234.