

Long Non-Coding RNA DLEU1 Up-Regulates BIRC6 Expression by Competitively Sponging miR-381-3p to Promote Cisplatin Resistance in Nasopharyngeal Carcinoma

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Background: Cisplatin (DDP) resistance has become an obstacle to chemotherapy for nasopharyngeal carcinoma (NPC) patients. Recent evidences indicate that long noncoding RNAs (lncRNAs) are involved in tumorigenesis and chemoresistance. However, the potential role of lncRNAs in NPC progression remains largely unknown.

Methods: First, lncRNA expression profiling in NPC was performed via microarray analysis. To explore the involvement of DLEU1 in DDP resistance, loss-of-function experiments were employed in vitro and in vivo. Bioinformatics analysis, luciferase reporter assay, qRT-PCR, and Western blot assays were used to investigate the underlying mechanisms.

Results: Here, we identified 153 differentially expressed lncRNAs. Among them, DLEU1 was remarkably up-regulated in NPC tissues and associated with worse outcome. Knock-down of DLEU1 could sensitize NPC cells to DDP in vitro and in vivo. Further investigations revealed that DLEU1 positively regulated BIRC6 expression via its competing endogenous RNA (ceRNA) activity on miR-381-3p. We also observed that BIRC6 over-expression or miR-381-3p silence could significantly reverse DLEU1-dependent DDP resistance.

Conclusion: Our data suggest that DLEU1 acts as an oncogene to promote DDP resistance and BIRC6 expression in NPC through interacting with miR-381-3p, which may help to develop new strategy against NPC chemoresistance.

Keywords: DLEU1, cisplatin resistance, nasopharyngeal carcinoma, BIRC6

Introduction

Nasopharyngeal carcinoma (NPC) is a head and neck carcinoma with a very unique pattern of geographical distribution. In 2018, there were about 129, 000 newly diagnosed NPC cases, and 70% were in east and southeast Asia.¹ Cisplatin (DDP) is often the first choice in chemotherapy regimens for NPC patients.² However, long-term treatment with DDP will lead to drug resistance which is a main reason for chemotherapy failure.³ Hence, it is of great importance to investigate the molecular mechanisms underlying the DDP resistance and develop effective therapeutic strategies for NPC treatment.

Long noncoding RNAs (lncRNAs) are RNA molecules longer than 200 nucleotides. lncRNAs could function as competing endogenous RNA (ceRNA) to competitively bind with miRNA response elements and reduce their effect on mRNAs.^{4,5} For

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instance, up-regulated lncRNA LOC100129148 was associated with unfavorable outcome in patients with NPC and silencing of LOC100129148 suppressed cell proliferation and KLF12 expression through sponging miR-539-5p.⁶ Wang et al reported that lncRNA CCAT1 enhanced paclitaxel resistance of NPC cells via miR-181a/CPEB2 axis.⁷ These findings indicate that dysregulated lncRNAs play crucial roles in NPC pathogenesis and chemoresistance.

lncRNA DLEU1 (deleted in lymphocytic leukemia 1) is located on chromosome 13q14.3 which is frequently deleted in B-cell chronic lymphocytic leukemia.⁸ Recent studies demonstrated that DLEU1 was highly expressed and contributed to tumorigenesis and development in a variety of cancers, such as ovarian, colorectal and endometrial cancer.^{9–11} In Burkitt lymphoma, however, overexpression of DLEU1 decreased cell proliferation, increased programmed cell death and improved survival, indicating DLEU1 may function as a tumor suppressor.¹² Taken together, the functions and molecular basis of DLEU1 are complex, yet its function in NPC progression is still unclear.

In this study, we investigated the expression and role of DLEU1 in NPC. We reported the finding that DLEU1 expression was increased in NPC tissues and cell lines, and associated with poor overall survival. Functionally, DLEU1 promoted DDP resistance and up-regulated BIRC6 through reducing miR-381-3p expression. These findings indicate that DLEU1 could be a new therapeutic target and prognostic marker in NPC.

Materials and Methods

Cell Culture and Tissue Samples

Human NPC cell lines (5-8F, 6-10B, C666-1, CNE1, CNE2, HNE-1, HONE-1, and SUNE-1) were purchased from American Type Culture Collection (ATCC) and maintained in RPMI-1640 (Invitrogen, Grand Island, NY, USA). Human nasopharyngeal epithelial cell line NP69 was obtained from the Cell Bank of Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences (Shanghai, China) and cultured in keratinocyte-SFM (Invitrogen). HEK-293T cells were purchased from ATCC and grown in DMEM (Invitrogen). A total of 67 NPC tissues were obtained from patients who received primary surgery at Zhuji People's Hospital. Twelve normal nasopharyngeal epithelial tissues were collected from patients who had nasal operations. Our study protocol was approved by the research ethics committee of Zhuji

People's Hospital (ZJSRMY-2017H-052). Written informed consent was signed by all participants.

Oligonucleotide and Plasmid Transfection

The siRNA for DLEU1 (si-DLEU1), negative control siRNA (si-NC), miR-381-3p mimics, miR-381-3p inhibitor (miR-381-3p-in), miRNA negative control (miRNA-NC) and pcDNA-BIRC6 were commercially synthesized by GenePharma (Shanghai, China). Transient transfection was performed with 50 nM oligonucleotides using Lipofectamine 2000. Subsequent experiments were performed at 48 hrs post-transfection. Short hairpin RNA (shRNA) targeting DLEU1 (sh-DLEU1) and non-specific control (sh-NC) were synthesized by GenePharma and cloned into pSuper-retro-puromycin vectors. For a stable cell transfection, SUNE-1 cells were transfected with sh-DLEU1 or sh-NC and then selected by 0.5 µg/mL puromycin.

MTT Assay

Cells were plated into 96-well plates (5000 cells/well) and treated with different concentrations of DDP (0, 5, 10, 15 and 20 µg/mL). Forty-eight hours after incubation, cell viability was assessed by MTT assay. The spectrophotometric absorbance was measured at 490 nm.

qRT-PCR

Total RNA was extracted from tissues and cell lines using TRIzol reagent (Ambion, Life Technologies, USA). cDNA of miRNA and mRNA or lncRNA was synthesized using TaqMan miR Reverse Transcriptase Kit (Applied Biosystems-Life Technologies) and ImProm-II Reverse Transcription System (Promega), respectively. qPCR assays were performed using the SYBR Premix EX Taq™ (Takara). Their relative expression levels were calculated using $2^{-\Delta\Delta CT}$ and normalized to U6 and β-actin.

Western Blot

Cell lysate was prepared using RIPA lysis buffer with protease inhibitors. Equal amounts of protein were separated by 10% SDS-PAGE, transferred to PVDF membranes, and then probed with anti-BIRC6 (Cell Signaling Technology) at 4°C overnight followed by incubation with HRP-conjugated secondary antibody for 1 hr at room temperature. Protein was normalized with β-actin (Cell Signaling Technology).

Microarray Analysis

Total RNA was extracted from three NPC tissues and three normal nasopharyngeal epithelial tissues using TRIzol

Reagent (Invitrogen) and purified by an RNeasy Mini Kit (Qiagen, Valencia, CA). cDNA was synthesized, labeled, and hybridized to the LncRNA Expression Microarray (Arraystar, Rockville, MD). After washing, slides were analyzed on the lncRNAs microarray. Differentially expressed lncRNAs were identified with thresholds of $P < 0.05$ and fold change ≥ 2.0 using GeneSpring GX v12.1 software package (Agilent Technologies, Santa Clara, CA, USA).

Xenografts

Twenty 5-week-old female BALB/c nude mice were randomly divided into sh-DLEU1 group and sh-NC group. About 1×10^7 SUNE-1 cells were inoculated subcutaneously

into each mouse. When the tumors reached a volume of approximately 100 mm^3 , each group of mice were intraperitoneally injected with DDP (5 mg/kg) every 2 days. After 4 weeks of treatment, all mice were sacrificed and the tumors were surgically removed. The animal assay was approved by the animal management committee of Zhuji People's Hospital, and all experimental procedures and animal care were in accordance with the institutional ethics guidelines for animal experiments (ZJSRMY-2017D-068).

Luciferase Reporter Assay

A fragment of the wild type (wt) or mutant (mut) DLEU1 and BIRC6 3'UTR that contained the putative miR-381-3p

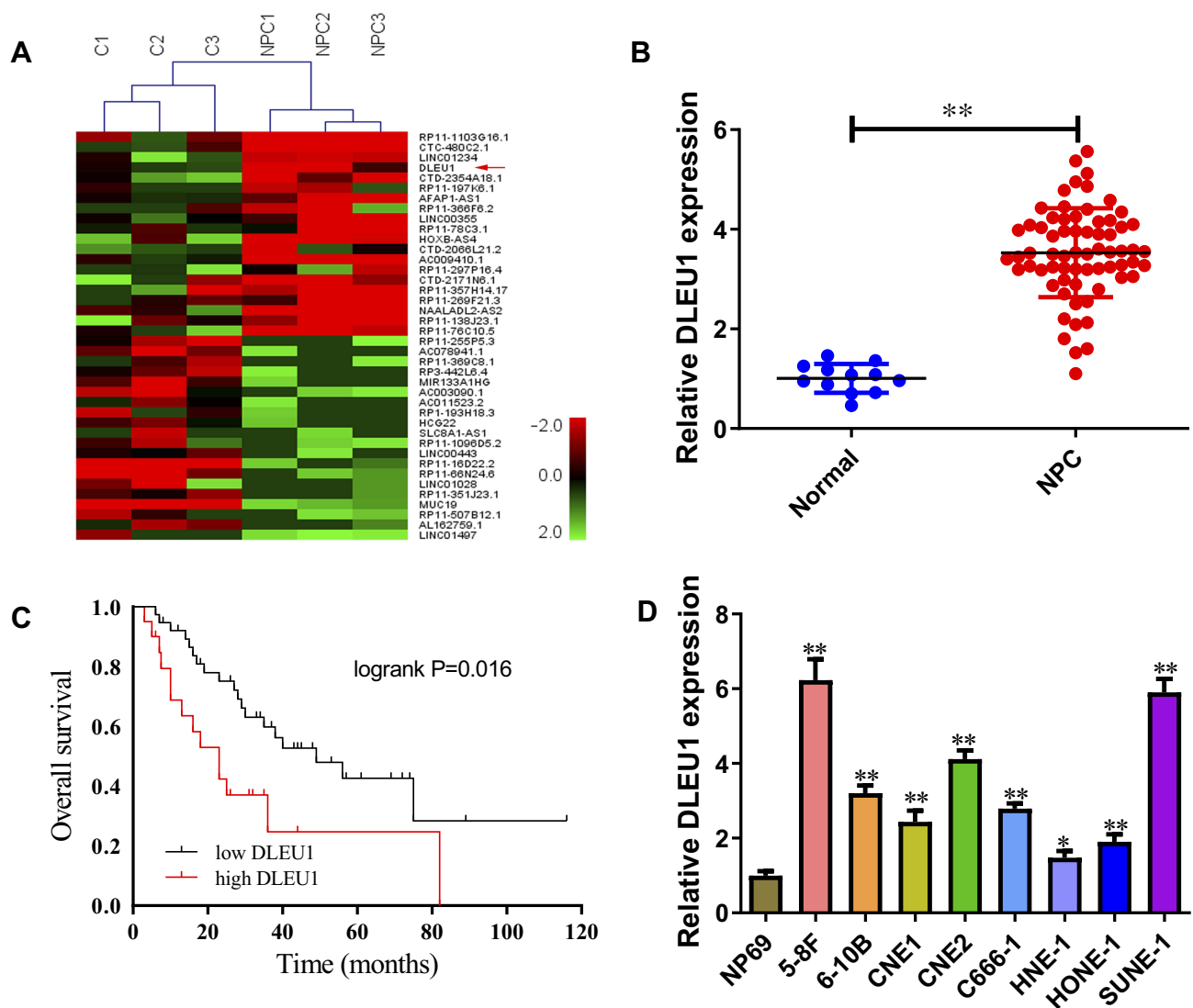


Figure 1 DLEU1 is overexpressed in NPC and predicts poor overall survival. (A) Heatmap of the top 20 up-regulated and 20 down-regulated lncRNAs between 3 NPC tissues and 3 normal nasopharyngeal epithelial tissues (fold change ≥ 2 , P-value < 0.05). (B) DLEU1 expression level in 67 NPC tissues and 12 normal nasopharyngeal epithelial tissues. (C) High DLEU1 expression was significantly associated with poor overall survival in NPC patients. (D) DLEU1 expression level in NPC cell lines and normal epithelium cells. * $P < 0.05$, ** $P < 0.01$.

binding sites was cloned into pGL3-REPORT luciferase vector. HEK293T cells (1×10^4) were seeded into 24-well plates and transiently co-transfected wt or mut reporter plasmid with miR-381 mimics or miRNA-NC. Dual luciferase reporter assays (Promega) were analyzed 48 hrs after transfection according to the manufacturer's instructions.

Statistical Analysis

Data are expressed as mean \pm SD of triplicate experiments. Statistical analysis was performed using Student's *t*-test on Graphpad Prism 6 software. Survival curves were assayed using the Kaplan-Meier method. P value < 0.05 was considered statistically significant.

Results

DLEU1 Is Overexpressed in NPC and Predicts Poor Overall Survival

To explore the potential involvement of lncRNAs in NPC progression, lncRNA microarray analysis was performed between NPC tissues and normal nasopharyngeal epithelial tissues. The result showed that 153 lncRNAs were differentially expressed (82 up-regulated and 71 down-regulated) with fold change >2.0 and $P < 0.05$ (Figure 1A). Of interest, DLEU1 expression was up-regulated in NPC tissues. The expression level of DLEU1 was validated to be up-regulated 3.53-fold in 67 NPC tissues compared with that of 12 normal nasopharyngeal epithelial tissues (Figure 1B). Clinicopathological features showed that up-expression of DLEU1 was strongly associated with clinical stage (Table 1; $P=0.019$), T stage ($P=0.032$) and chemotherapy response ($P=0.012$). Furthermore, Kaplan-Meier survival analysis revealed that high level of DLEU1 predicted poor outcome ($P=0.016$; Figure 1C). We also determined the expression profiles of DLEU1 in normal epithelium cells and NPC cell lines. As shown in Figure 1D, DLEU1 was up-regulated in all 8 NPC cell lines, especially in 5-8F and SUNE-1 cells. These findings suggest a crucial role of DLEU1 in NPC development.

Knock-Down of DLEU1 Sensitizes NPC Cells to DDP in vitro and vivo

DDP is an important chemotherapy regimen for NPC treatment. To investigate whether DLEU1 could affect DDP resistance in NPC development, loss-of-function experiment was performed via transfecting specific DLEU1 siRNA into 5-8F and SUNE-1 cells (Figure 2A).

Table 1 Correlation Analysis Between DLEU1 Expression and Clinicopathologic Features of NPC Patients

Features	Number	DLEU1 Expression		P value
		High	Low	
Gender				0.892
Male	31	22	9	
Female	36	25	11	
Age (years)				0.747
≥ 44	45	31	14	
<44	22	16	6	
Clinical stage				0.019
I-II	29	16	13	
III-IV	38	31	7	
T stage				0.032
T1-2	27	15	12	
T3-4	40	32	8	
N stage				0.072
N0-1	38	30	8	
N2-3	29	17	12	
Metastasis				0.228
Yes	24	19	5	
No	43	28	15	
Chemotherapy response				0.012
Yes	25	13	12	
No	42	34	8	

When exposing to increasing concentrations of DDP, cells transfected with si-DLEU1 were more sensitive to DDP than cells transfected with si-NC (Figure 2B and C). To further evaluate the effect of DLEU1 on DDP resistance in vivo, SUNE-1 cells stably expressing sh-DLEU1 or sh-NC were subcutaneously injected into nude mice following by treatment with DDP. Four weeks after treatment, both tumor volume and weight in sh-DLEU1 group were decreased than control group (Figure 2D-F). Collectively, these data indicate up-regulated DLEU1 may contribute to NPC development through increasing DDP resistance.

DLEU1 Binds to miR-381-3p and Suppresses Its Expression

lncRNAs mainly exert their biological functions through ceRNA mechanism. We speculated whether DLEU1 could interact with miRNAs to regulate DDP-resistance. Bioinformatics analysis revealed that DLEU1 contains

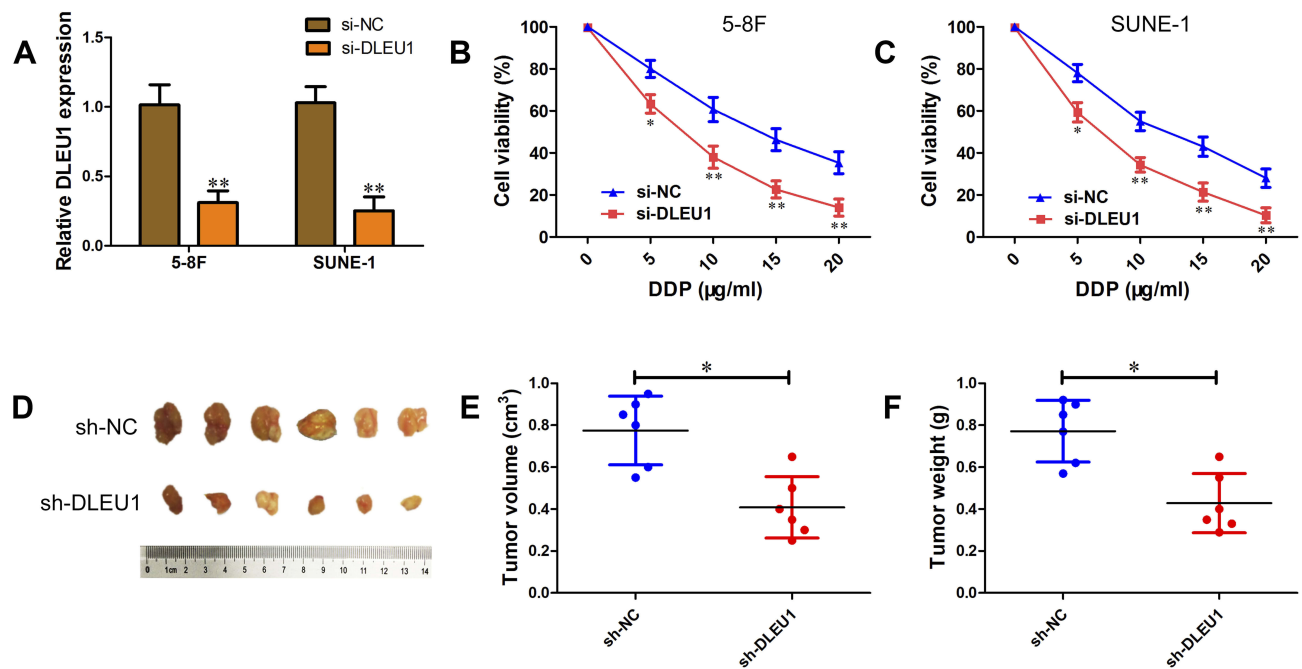


Figure 2 Knock-down of DLEU1 sensitizes NPC cells to DPP in vitro and vivo. (A) DLEU1 expression level in 5-8F and SUNE-1 cells transfected with si-DLEU1 or si-NC. (B, C) Cell viability was determined using MTT assay in 5-8F (B) and SUNE-1 (C) cells transfected with si-DLEU1 or si-NC. (D–F) SUNE-1 cells stably sh-DLEU1 or sh-NC were subcutaneously injected into nude mice (n=10 per group) following by treatment with DDP. Representative images (D), tumor volume (E) and weight (F) of the formed tumors. *P<0.05, **P<0.01, n=6.

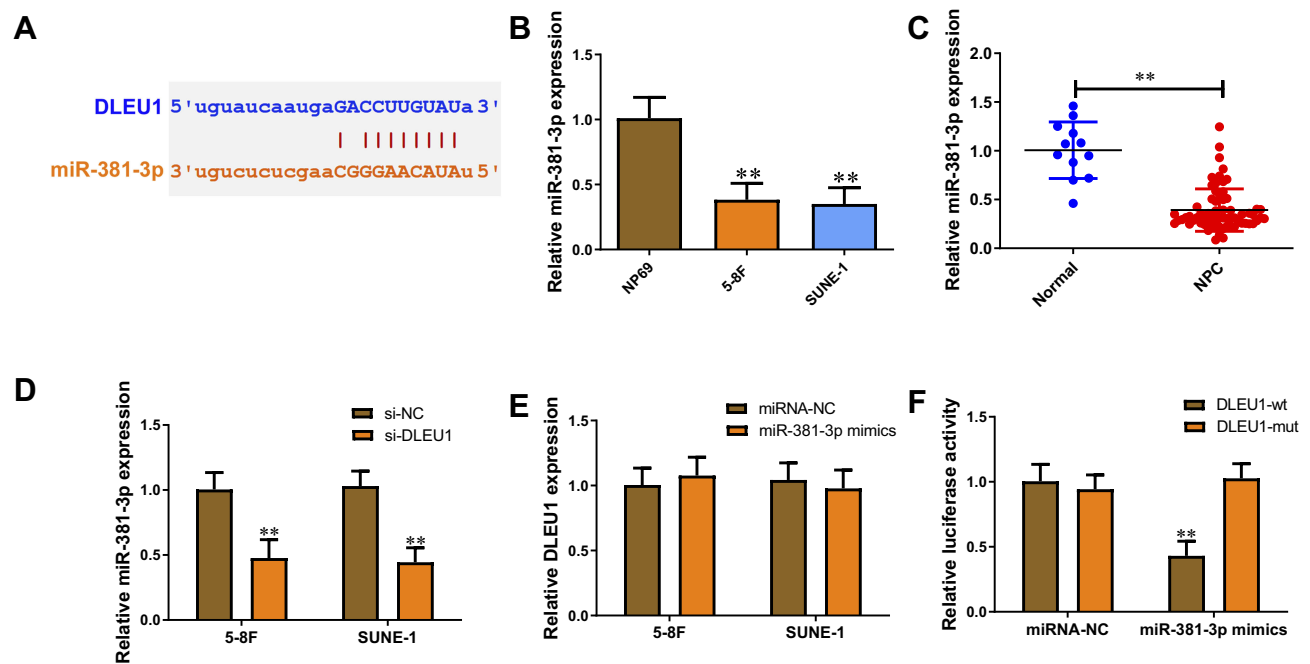


Figure 3 DLEU1 binds to miR-381-3p and suppresses its expression. (A) Putative binding site of miR-381-3p in DLEU1 using starBase v3.0. (B) miR-381-3p expression level in 5-8F, SUNE-1, and NP69 cells. (C) miR-381-3p expression level in 67 NPC tissues and 12 normal nasopharyngeal epithelial tissues. (D) miR-381-3p expression level in 5-8F and SUNE-1 cells transfected with si-DLEU1 or si-NC. (E) DLEU1 expression level in 5-8F and SUNE-1 cells transfected with miR-381-3p mimics or miRNA-NC. (F) The luciferase activities in HEK-293T cells co-transfected with wild type (wt) or mutant (mut) DLEU1 vector with miR-381-3p mimics or miRNA-NC. **P<0.01.

binding sites for miR-381-3p (Figure 3A). miR-381-3p expression in 5-8F and SUNE-1 cells was lower than that of NP69 cells (Figure 3B). Besides, miR-381-3p level was

down-regulated 2.55-fold in NPC tissues compared with normal tissues (Figure 3C). Then we explored the regulatory relationship between DLEU1 and miR-381-3p in 5-8F

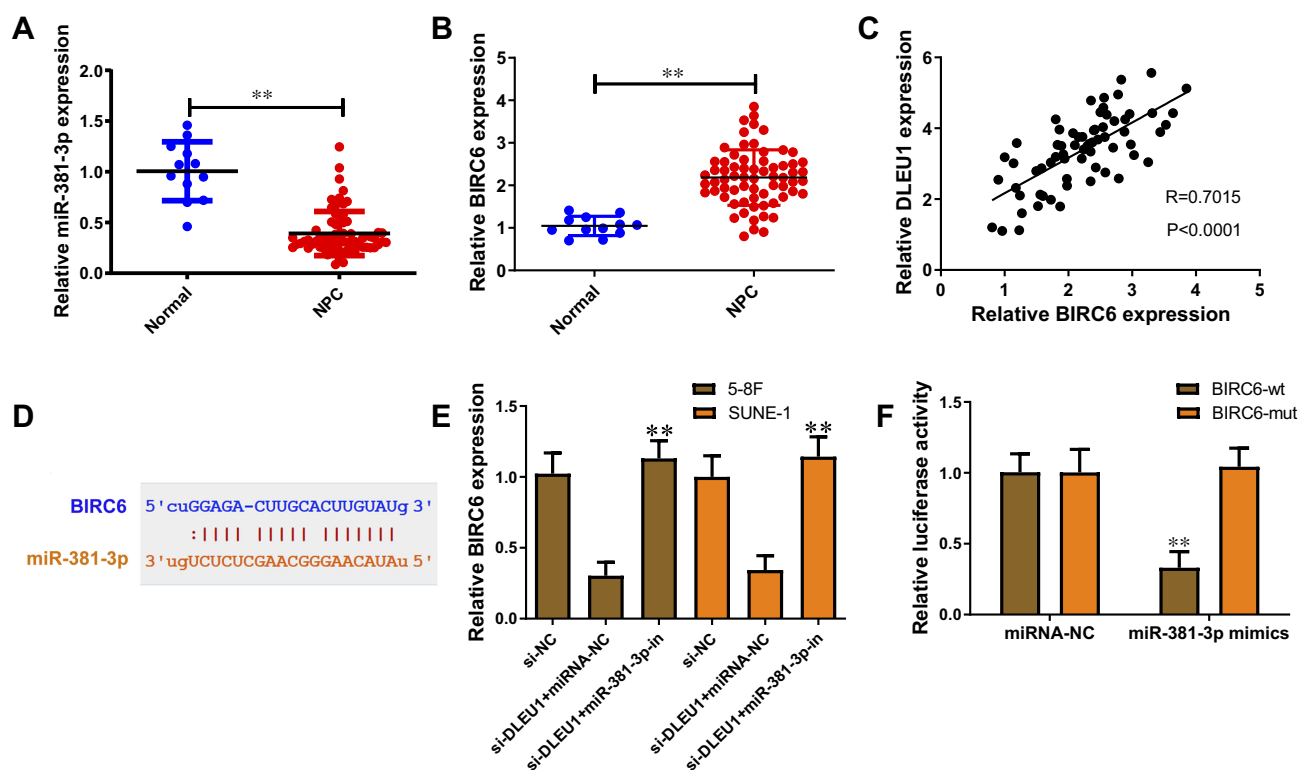


Figure 4 DLEU1 acts as a ceRNA to up-regulate BIRC6 expression via sponging miR-381-3p. (A) BIRC6 expression level in 5-8F, SUNE-1, and NP69 cells. (B) BIRC6 expression level in 67 NPC tissues and 12 normal nasopharyngeal epithelial tissues. (C) Pearson correlation analysis between DLEU1 and BIRC6 expression in NPC tissues. (D) Putative binding site of miR-381-3p in BIRC6 3'UTR using starBase v3.0. (E) BIRC6 protein expression level in 5-8F and SUNE-1 cells co-transfected with si-DLEU1 or si-NC with miR-381-3p-in or miRNA-NC. (F) The luciferase activities in HEK-293T cells co-transfected with wild type (wt) or mutant (mut) BIRC6 vector with miR-381-3p mimics or miRNA-NC. ** $P < 0.01$.

and SUNE-1 cells. Knock-down of DLEU1 negatively regulated miR-381-3p expression (Figure 3D). Meanwhile, miR-381-3p mimics have no significant influence on DLEU1 level (Figure 3E). Luciferase reporter assay showed that co-transfection of miR-381-3p mimics with DLEU1-wt, rather than the DLEU1-mut, significantly reduced luciferase activity (Figure 3F). These results indicate that miR-381-3p is a key downstream target of DLEU1 to promote DDP resistance in NPC.

DLEU1 Acts as a ceRNA to Up-Regulate BIRC6 Expression via Sponging miR-381-3p

Recently, BIRC6 was identified as an oncogene involved in chemoradiotherapy resistance. Thus, we aimed to verify whether DLEU1 could regulate BIRC6 expression by interacting with miR-381-3p in NPC cells. Compared with NP69 cells, BIRC6 level was higher in 5-8F and SUNE-1 cells (Figure 4A). We also observed an increased level of BIRC6 in NPC tissues (Figure 4B). Our results showed that there was a strong correlation between DLEU1 and BIRC6 in NPC tissues ($R=0.7015$, $P < 0.0001$; Figure 4C). Bioinformatics

analysis showed that the 3'UTR sequence of BIRC6 was complementary to the seed sequence of miR-381-3p (Figure 4D). In addition, BIRC6 protein level was down-regulated when transfected with si-DLEU1, and miR-381-3p inhibitor could abolish BIRC6 inhibition induced by DLEU1 knock-down (Figure 4E). Furthermore, luciferase reporter assay showed BIRC6-wt and miR-381-3p mimics co-transfection significantly decreased luciferase activity, but BIRC6-mut failed to change luciferase activity (Figure 4F).

DLEU1 Promotes DDP Resistance via miR-381-3p and BIRC6-Dependent Manner

To further confirm the functional role of miR-381-3p and BIRC6 in DLEU1-modulated DDP resistance, miR-381-3p inhibitor or pcDNA-BIRC6 was co-transfected with si-DLEU1 into 5-8F and SUNE-1 cells. As shown in Figure 5A and B, cells co-transfected si-DLEU1 with miR-381-3p inhibitor exhibited a less sensitive ability to DDP compared with cells transfected with si-DLEU1 alone as determined using MTT assay. Consistently, BIRC6 over-expression significantly reversed the inhibition of DDP

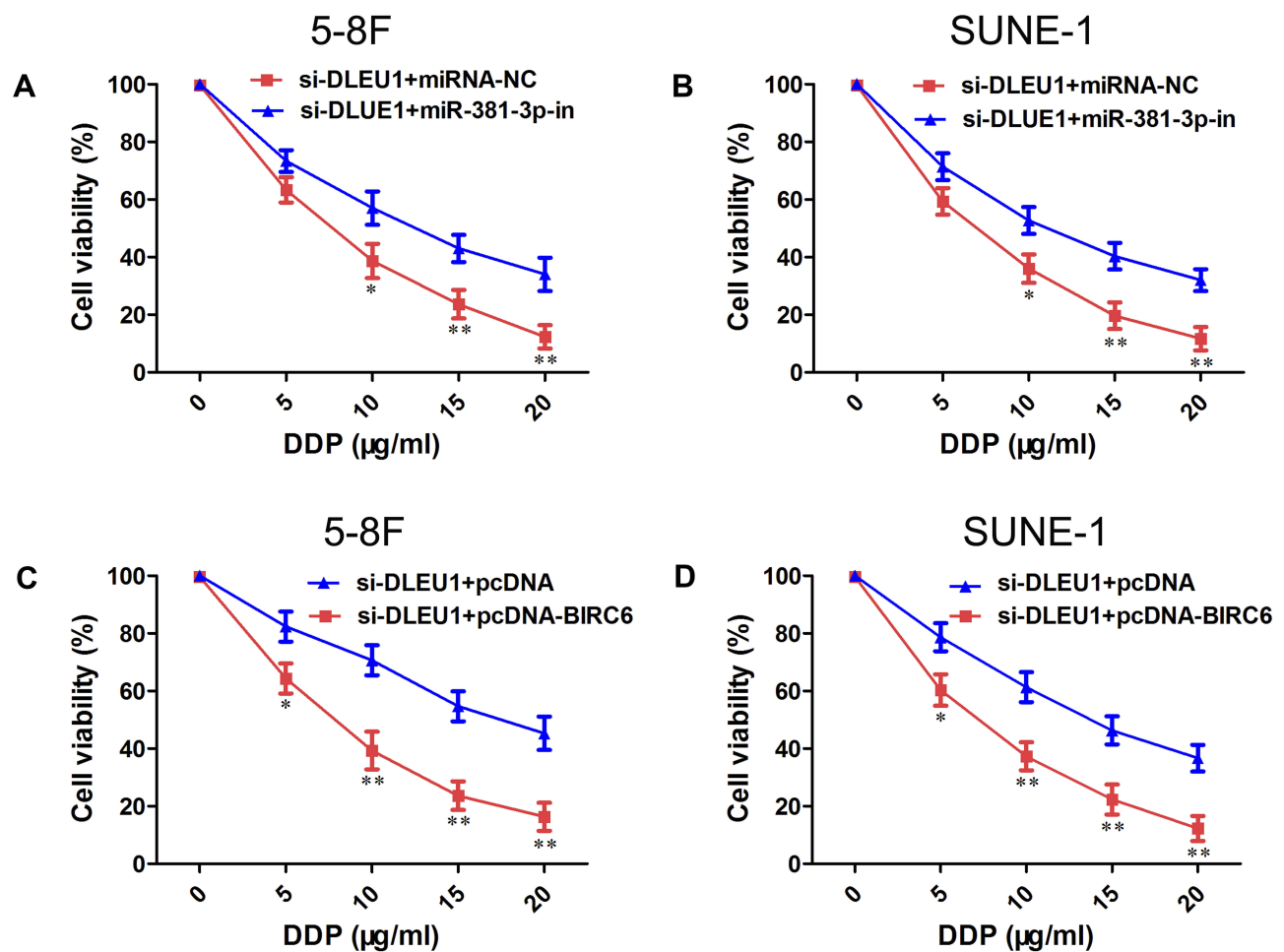


Figure 5 DLEU1 promotes DDP resistance via miR-381-3p and BIRC6-dependent manner. (A, B) Cell viability was determined using MTT assay in 5-8F (A) and SUNE-1 (B) cells co-transfected with si-DLEU1 or si-NC with miR-381-3p-in or miRNA-NC. (C, D) Cell viability was determined using MTT assay in 5-8F (C) and SUNE-1 (D) cells co-transfected with si-DLEU1 or si-NC with pcDNA-BIRC6 or pcDNA. * $P < 0.05$, ** $P < 0.01$.

resistance induced by DLEU1 knock-down in both into 5-8F and SUNE-1 cells (Figure 5C and D). These findings indicate that DLEU1 promotes DDP resistance at least partly through interacting with miR-381-3p and BIRC6.

Discussion

Abnormally expressed DLEU1 has been identified in several types of human cancer and could function as oncogene or tumor suppressor to affect cancer progression.⁹⁻¹² In our microarray analysis, we found that DLEU1 expression increased in NPC tissues. We also observed that high level of DLEU1 predicted an unfavorable prognosis of patients with NPC. In addition, knock-down of DLEU1 reduced DDP resistance in vivo and in vitro. These findings suggest that DLEU1 acts as an oncogene in NPC. Further mechanistic analysis demonstrated that DLEU1 exerted its role by sponging miR-381-3p and up-regulating BIRC6 expression

in NPC cells. Recently, DLEU1 was reported to enhance resistance to DDP in bladder cancer cells through regulating miR-99b/HS3ST3B1 axis, which is in line with our findings.¹³ Furthermore, DLEU1 overexpression was observed to be associated with rituximab and/or cyclophosphamide resistance in Burkitt lymphoma cells.¹² As a kind of lncRNA, DLEU1 can interact with many molecules, like miRNAs and RNA-binding proteins. There would be various underlying mechanisms of DLEU1-induced DDP resistance, which are worthy of further study.

A growing body of evidence has demonstrated that miRNAs play an important role in NPC progression.¹⁴ Recent studies have suggested that miR-381-3p level was decreased in several cancers and overexpression of miR-381-3p inhibited cell proliferation and metastasis, and induced cell cycle arrest and apoptosis.¹⁵⁻¹⁹ However, the functions and mechanisms of miR-381-3p in NPC are

still unclear. In our study, miR-381-3p was observed to be down-regulated in NPC. In addition, qRT-PCR and luciferase reporter assay showed that DLEU1 served as a ceRNA for miR-381-3p. Furthermore, silence of miR-381-3p reversed DDP sensitization-induced by DLEU1 knock-down. Taken together, these results suggest that miR-381-3p was a direct target of DLEU1.

BIRC6 gene encodes a 528 kDa protein that consists of N-terminal BIR domain and C-terminal ubiquitin-conjugating domain in mammals.²⁰ BIRC6 is a key regulator for multiple biological processes, such as anti-apoptosis, cytoprotection and regulation of cytokinesis.²¹ Increasing evidences indicated that BIRC6 is frequently overexpressed in various tumors and associated with poor outcome.^{22–25} Several studies also demonstrated that BIRC6 participated in tumor cell chemoresistance, including imatinib, enzalutamide and cisplatin.^{26–28} Here, we observed that BIRC6 level was higher in NPC tissues compared with normal controls and strongly correlated with DLEU1 expression. DLEU1 knock-down reduced BIRC6 expression while BIRC6 knock-down reversed DLEU1-modulated DDP resistance in NPC cells. These data indicate that DLEU1 promotes DDP resistance at least partly through regulating BIRC6 expression.

In summary, we demonstrated that DLEU1 was significantly up-regulated and predicted poor overall survival of NPC patients. Our data support that DLEU1 acts as oncogene to promote DDP resistance at least partly through miR-381-3p/BIRC6 axis. More importantly, DLEU1 might serve as a potential therapeutic sensitizer in NPC treatment.

Disclosure

The authors declare that they have no competing interests in this work.

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