

# Significance of Interferon- $\gamma$ in Coronary Artery Bypass Surgery

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## ABSTRACT

**Background:** Gamma interferon (IFN- $\gamma$ ) is produced by activated natural killer and T cells under pathologic circumstances. The objective of our study was to compare the level of IFN- $\gamma$  in open and endoscopic methods of vein harvesting for coronary artery bypass surgery (CABG).

**Method:** Ninety samples of human saphenous veins harvested from patients prepared for CABG. Pre- and post-procedure sera of the patients, in addition to supernatants of 3-day endothelial cell culture, were analyzed for IFN- $\gamma$ .

**Results:** The mean preoperative IFN- $\gamma$  level ( $0.09 \pm 0.03$  pg/mL) and that for postoperative sera ( $0.08 \pm 0.02$  pg/mL) were not significantly different ( $P = 0.2$ ). The mean IFN- $\gamma$  level in endothelial cell culture from the endoscopic ( $0.18 \pm 0.21$  pg/mL) and the open method ( $0.19 \pm 0.39$  pg/mL) were not significant ( $P = 0.89$ ).

**Conclusion:** We recommend the endoscopic method of vein harvesting because of its lower morbidity and earlier hospital discharge.

**Key Words:** IFN- $\gamma$ , CABG, Endoscopic and open saphenectomies.

## INTRODUCTION

Vascular endothelial cells constitute the interface between the bloodstream and tissues.<sup>1,2</sup> In this strategic function, endothelial cells (EC) play an active role in all phases of immunologic and nonimmunologic inflammation through cytokine networks.<sup>3-10</sup>

EC produce various cytokines, such as interleukin 1 (IL-1), interleukin 6 (IL-6), colony stimulating factors (CSF), and chemotactic factors, and express adhesion molecules.<sup>11-14</sup> These data on the function of the endothelial cells in the immune response were obtained from EC culture experiments. These experiments found that EC can also be modulated by interleukins, interferons, and tumor necrosis factors.<sup>15-19</sup>

Interferons of all 3 types ( $\alpha$ ,  $\beta$ , and  $\gamma$ ) interact with specific receptors on virtually all cells of the body, 1 receptor for  $\alpha$  and  $\beta$  and another for  $\gamma$ .<sup>20</sup> Gamma interferon (IFN- $\gamma$ ) is a 45-KD homodimeric glycosylated protein detectably produced only under pathologic circumstances (trauma, infection, cancer, and autoimmunity) by particular lymphocyte populations, namely, activated natural killer (NK) cells and T cells. IFN- $\gamma$  suppresses cultured human vascular endothelial cell proliferation,<sup>21</sup> induces these cells to express MHC class II antigen,<sup>22</sup> and causes them to change shape and cell organization.<sup>23,24</sup>

The impact of trauma on IFN- $\gamma$  production is our major concern in saphenous vein preparation for coronary artery bypass graft (CABG) surgery. The present study was undertaken to evaluate and compare the levels of IFN- $\gamma$  in the endoscopic technique of vein harvesting with the standard open method, as the new technique is preferred now in CABG, due to the lower incidence of postoperative complications.

## MATERIALS AND METHODS

The protocols used in this project were reviewed and approved by the Institutional Review Board of the Maimonides Medical Center. Patients undergoing coronary artery bypass graft (CABG) surgery provided informed consent before enrolling as subjects in this research protocol. A total of 45 patients were enrolled in the project.

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### **Saphenous Vein Harvest**

Saphenous vein harvest (SVH) was performed using both the open and endoscopic techniques on each patient undergoing elective CABG surgery. Therefore, both an open technique sample and an endoscopic technique sample were obtained from each patient enrolled in the study. Standard endoscopic instrumentation, consisting of a subcutaneous tissue dissector, a retractor, and a modified vein stripper, was used for the endoscopic SVH (Endopath, Ethicon Endosurgery Inc., Cincinnati, OH). In addition, standard endoscopic equipment including a television monitor, light source, and fiberoptic camera with a 5-mm lens were used.

Prior to the initial skin incision, a preoperative blood sample was obtained to determine baseline systemic cytokine levels. The blood sample was immediately centrifuged at 3,000 rpm for 10 minutes at 28°C, and the supernatants were collected.

The patient was placed in the “frog-leg” position, and a 3- to 4-cm longitudinal incision was made approximately 4-finger breadth posterior to the patella along the medial genicular region. The greater saphenous vein (SV) was identified with the sharp dissecting technique. The portion of the SV dissected below the skin incision was considered the open technique sample. This segment remained in place until after adjacent segments were harvested with the endoscopic method. Next, a 5-cm sample was retrieved by dissecting cranially with the endoscopic technique. This portion of the SV was clipped distally to identify the direction of blood flow. An additional 5-cm endoscopic technique sample was obtained by dissecting below the knee joint. Finally, the portion of the SV remaining at the site of the open incision was removed and labeled “open-technique sample.” Significant segments of the SV remained after collecting samples, and these were used as the grafts in the CABG surgery. At the conclusion of SV harvesting, during skin closure, a second blood sample was drawn. Vein samples were harvested and handled under sterile conditions according to the operating room protocol at Maimonides Medical Center.

### **Vein Preparation**

Vein samples were incubated in 10-mL endothelial cell culture medium (ECCM) consisting of Iscove's Modified Dullbecco's Medium (IMDM, Fischer Scientific,

Cincinnati, OH) with 200 mL penicillin-streptomycin solution (Fischer Scientific, Cincinnati, OH) during transport to the laboratory. In the laboratory, each vein sample was flushed and cannulated. They were then injected with plasmalyte solution (Baxter Scientific, St. Louis, MO) containing 60-mg/mL papaverine (Fischer Scientific, Cincinnati, OH), and the branches were ligated with 3.0 silk sutures. Plasmalyte solution contained sodium 140 mEq/L, potassium 5 mEq/L, magnesium 3 mEq/L, chloride 98 mEq/L, acetate 27 mEq/L, and gluconate 23 mEq/L. The pH was adjusted to 7.4 by addition of sodium hydroxide, and the final osmolarity was 294 mOsm/Kg.

### **Endothelial Cell Culture**

Endothelial cell cultures were prepared as described previously. Vein samples were placed into a culture dish (Fischer Scientific, Cincinnati, OH) containing 5 mL of ECCM with the addition of 130 U/mL heparin and 2 mM L-glutamine. Vein samples were slit open and placed flat with the luminal surface facing up. The luminal surface was gently scraped with a sterile scalpel blade, with light single strokes, covering each area only once. The cells, which collected on the scalpel blade, were shaken off into the ECCM in the culture dish. Next, the vein was rinsed with an aliquot of ECCM, and the rinsing fluid was added to the culture dish. The ECCM and scraped endothelial cells in the culture dish was collected and transferred to a 25 mL culture flask (Fischer Scientific, Cincinnati, OH). The cells were incubated at 37°C in humidified room air with 5% CO<sub>2</sub> for the next 72 hours.

### *IFN- $\gamma$ Measurement*

A commercially available enzyme-linked immuno-sorbent assay (ELISA) was used to measure the presence of human IFN- $\gamma$  (Biosource International, California). IFN- $\gamma$  levels were determined in samples of sera and ECCM. Blood sera were diluted 1:10 in plasmalyte solution before IFN- $\gamma$  levels were determined. The ECCM was aspirated from the cell cultures after 72 hours and diluted 1:50 in plasmalyte solution. The IFN- $\gamma$  level was then determined in the diluted samples.

### *Statistical Analysis*

Results for each group are expressed as mean  $\pm$  standard deviation of the quantity of IFN- $\gamma$  measured in the samples. Data were analyzed with the two-tailed Student

*t* test. A *P*-value of less than 0.05 was designated as a statistically significant difference between groups.

## RESULTS

The mean level of IFN- $\gamma$  for preoperative sera in the 45 patients was  $0.096 \pm 0.032$  pg/mL (mean  $\pm$  standard deviation) with a confidence interval (CI) = 0.087-0.105, and that for postoperative sera was  $0.089 \pm 0.019$  pg/mL, with a confidence interval (CI) = 0.083-0.095. The means were not statistically significant (*P* = 0.2).

The mean level of IFN- $\gamma$  measured in endothelial cell culture from the endoscopic technique was  $0.185 \pm 0.215$  with CI = 0.122-0.248, whereas that for the open method was  $0.194 \pm 0.39$  with CI = 0.079-0.308. The difference between the means of IFN- $\gamma$  for the 2 techniques was not statistically significant (*P* = 0.9) (**Figure 1**).

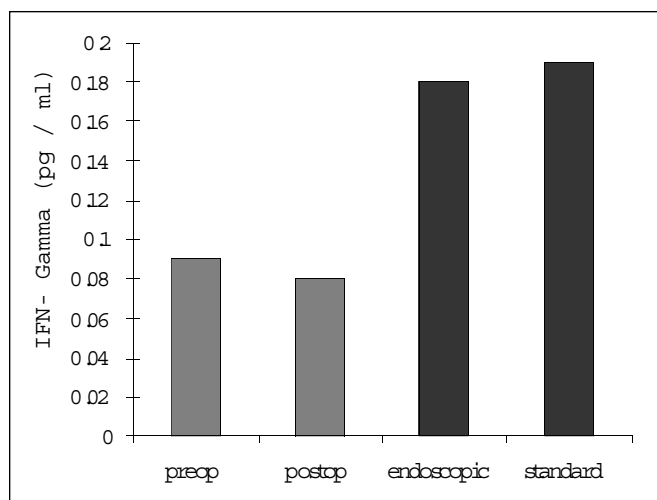
## DISCUSSION

The autologous greater saphenous vein is considered the best material for coronary artery revascularization. However, the morbidity associated with saphenous vein harvesting is significant with reported wound complications as high as 43%.<sup>25-30</sup>

Historically, the greater saphenous vein is exposed and harvested through a long continuous incision. This incision may be related to an increase in wound problems; therefore, there has been much interest in developing a more closed technique for saphenectomy.<sup>27-29</sup> More recently, a newly developed endoscopic technique for saphenous vein harvesting has been described with encouraging results.<sup>31-34</sup>

Studies of cytokines, leukocytes, and endothelial cell interaction, and the ensuing biologic events, have received much attention in recent years.<sup>35,36</sup> These studies found that vascular endothelium can produce several biologically active compounds, including nitric oxide, prostacyclin, and various cytokines.<sup>30</sup> IFN- $\gamma$  is a cytokine, produced by natural killer cells, that plays an important role in bringing about acute inflammation, mainly because of the activating effect of IFN- $\gamma$  on adhesive properties of endothelial cells and on mediator production by mononuclear phagocytes.<sup>37</sup>

Impairment of the immune response by surgery is suggested by clinical observations of the high rate of infection seen in postoperative patients. The principal



**Figure 1.** Graphic presentation of the level of interferon- $\gamma$  in preoperative, postoperative, endoscopic, and open techniques of saphenous vein harvesting.

immunological deficit after trauma and major surgery is decreased cell-mediated immunity from an impaired natural killer (NK) cell response and T helper (Th) 1 lymphocyte development. Cell-mediated immunity has been shown to be depressed for 3 to 10 days postoperatively in patients who have undergone major surgery.<sup>38</sup> In addition, many studies have shown that IFN- $\gamma$  is reduced in the early postoperative phase, reflecting impaired function of (Th) 1 cells and therefore of cell-mediated immune response.<sup>39</sup>

In contrast, our study showed that no significant difference exists in the level of IFN- $\gamma$  pre- and postoperatively. We also demonstrated that no significant difference exists in the level of IFN- $\gamma$  using both techniques of saphenous vein harvesting (StdSVH and EndoSVH). However, the number of patients in most of the other studies was small, and the measured levels of IFN- $\gamma$  varied widely. Therefore, conclusions about perioperative changes of INF- $\gamma$  are tentative at best.

The comparable expressions of IFN- $\gamma$  in the endoscopic and the traditional technique of vein harvesting suggests that vein manipulation and minor physical shears would have a minimal impact on the levels of IFN- $\gamma$  that may subsequently affect the performance of the vein conduit.

The limitation of our study is that open saphenectomy was not performed as a standard procedure on our

patients undergoing CABG, so we were unable to devise 2 separate groups of patients undergoing each technique alone.

The method of endothelial cell harvesting and culture might have falsely affected the level of IFN- $\gamma$ . Furthermore, in our study, we stored plasma at  $-2^{\circ}\text{C}$ , various investigators had used different temperatures for plasma storage in evaluating IFN- $\gamma$  expression. At the same time, we have used supernatants of cell culture to compare endoscopic with the open technique. Other studies used peripheral blood mononuclear cells to measure IFN- $\gamma$  level,<sup>40</sup> which might affect our results. Also, measurement of IFN- $\gamma$  in blood samples taken 3 to 10 days postoperatively would have incorporated the effect of CABG itself as a major surgery, and the individual role of vein harvesting could not be clarified.

In conclusion, our findings indicate that endoscopic and open saphenectomies are comparable for IFN- $\gamma$  expression in CABG patients. However, we still recommend the endoscopic method of vein harvesting due to fewer postoperative leg wound complications compared with traditional open saphenectomy, which ultimately results in less postoperative pain, reduced hospital stay, and reduction in health care costs.<sup>41</sup>

## References:

1. Holzinger C, Weissinger E, Zuckermann A, et al. Effects of interleukin-1,-2,-4,-6, and interferon gamma and granulocyte/macrophage colony stimulating factor on human vascular endothelial cells. *Immunol Lett.* 1993;35:109-118.
2. Harlan JM. Leukocyte-endothelial interaction. *Blood.* 1985;65:513.
3. Ruszczak Z, Detmar M, Imcke E, Orfan CE. Effects of rIFN Alpha, Beta, and Gamma on the morphology, proliferation, and cell surface antigen expression of human dermal microvascular endothelial cells in vitro. *Soc Invest Dermatol.* 1990; 95:693-699.
4. Cotran RS, Pober JS. Effects of cytokines on vascular endothelium: their role in vascular and immune injury. *Kidney Int.* 1989;35:969-975.
5. Cotran RS. New roles for the endothelium in inflammation and immunity. *Am J Pathol.* 1987;129:407-413.
6. Kupper TS. Production of cytokines by epithelial tissues. A new model for cutaneous inflammation. *Am J Dermatopathol.* 1989;11:69-73.
7. Pober JS, Collins T, Gimbrone MA Jr, Libby P, Reiss CS. Inducible expression of class II major histocompatibility complex antigens and the immunogenicity of vascular endothelium. *Transplantation.* 1986;41:141-146.
8. Pober JS, Gimbrone MA Jr, Lapierre LA, et al. Overlapping patterns of activation of human endothelial cells by interleukin-1, tumor necrosis factor and immune interferon. *J Immunol.* 1986;137:1893-1896.
9. Pober JS. Effects of tumor necrosis factor and related cytokines on vascular endothelial cells. *Ciba Foundation Symp.* 1987;131:170-184.
10. Pober JS. Cytokine-mediated activation of vascular endothelium. Physiology and Pathology. *Am J Pathol.* 1988;133:426-433.
11. Hiromatsu Y, Masayuki S, Kentaro Y, Kyohei N. Inhibitory effect of nicotinamide on recombinant human interferon-gamma-induced intercellular adhesion molecule-1 (ICAM-1) and HLA-DR antigen expression on cultured human endothelial cells. *Immunol Lett.* 1991;31:35-40.
12. Miossec P, Cavender D, Ziff M. Production of interleukin-1 by human endothelial cells. *J Immunology.* 1986;136:2486.
13. Mantovani A, Dejana E. Cytokines as communication signals between leukocytes and endothelial cells. *Immunol Today.* 1989;10:370.
14. Rothlein R, Dustin ML, Marlin SD, Springer TA. A human intercellular adhesion molecule [ICAM-1] distinct from LFA-1. *J Immunol.* 1986;137:1270-1274.
15. Montesano R, Orci L, Vassalli P. Human endothelial cell cultures: phenotypic modulation by leukocyte interleukins. *J Cell Physiol.* 1985;122:424-435.
16. Lapierre LA, Fiers W, Pober JS. Three distinct classes of regulatory cytokines control endothelial cell MHC antigen expression. *J Exp Med.* 1988;167:794-804.
17. Cavender DE, Edelbaum D, Ziff M. Endothelial cell activation induced by tumor necrosis factor and lymphotoxin. *Am J Pathol.* 1989;134:551-560.
18. Norioka K, Nakagawa K, Onozuka S, et al. The effect of lymphokines on growth and phenotype of human vascular endothelial cells. *J Natl Def Med Coll.* 1988;13:71-79.
19. Detmar M, Ruszczak ZB, Imcke E, Orfanos CE. Effects of tumor necrosis factor alpha (TNF) on cultured microvascular endothelial cells derived from human debris (abstr). *J Invest Dermatol.* 1989;93:546.
20. Mims C, Wakelin D, Playfair J, Williams R, Roitt I. Medical Microbiology: Natural Defenses in Action. London: Mosby-Year Book Europe Ltd; 1998.
21. Friesel JS, Komoriya A, Maciag T. Inhibition of endothelial cell proliferation by gamma interferon. *J Cell Biol.* 1987;104:689.
22. Pober JS, Gimbrone MA, Cotron RS, et al. Ia expression by vascular endothelium is inducible in vitro demonstration by monoclonal antibody binding & immunoprecipitation. *J Exp Med.* 1983;157:1339.
23. Montesano R, Orci L, Vassalli P. Human endothelial cell cul-

- ture, Phenotype modulation by leukocyte interleukins. *J Cell Physiol.* 1985; 122:424.
24. Stolphen AH, Guinan EG, Fiers W, Pober JS. Recombinant tumor necrosis factor and immune interferon act singly and in combination to recognize human vascular endothelial cell monolayers. *Am J Pathol.* 1986;123:16.
25. Reifsnnyder T, Bandyk D, Seabrook G. Wound complications of the in situ saphenous vein bypass technique. *J Vasc Surg.* 1992;15: 843-850.
26. Wipke-Tevis DD, Stotts NA, Skov P, Carrieri-Kohlman V. Frequency, manifestations, and correlates of impaired healing of saphenous vein harvest incisions. *Heart Lung.* 1996;25:108-116.
27. Robbins MR, Hutchinson SA, Helmer SD. Endoscopic saphenous vein harvest in infrainguinal bypass surgery. *Am J Surg.* 1998;176:586-590.
28. Wengrovitz M, Atnip RG, Girrord RRM. Wound complications of autogenous subcutaneous infrainguinal arterial bypass surgery; predisposing factors and management. *J Vasc Surg.* 1990;11:156-163.
29. Schwartz ME, Harrington EB, Schanzer H. Wound complication after in situ bypass. *J Vasc Surg.* 1988;7:802-807.
30. Johnson JA, Cogbill TH, Strutt PJ, Gunderson AL. Wound complications after infrainguinal bypass. *Arch Surg.* 1988; 123:859-862.
31. Jordan W D Jr, Voellinger DC, Schroeder PT, McDowell HA. Video-assisted saphenous vein harvest: the evolution of a new technique. *J Vasc Surg.* 1997; 26:405-414.
32. Lumsden AB, Eaves JC III, Jordan WD. Subcutaneous, video-assisted saphenous vein harvest: report of the first 30 cases. *Cardiovasc Surg.* 1996;4:771-776.
33. Allen KB, Shaar CJ. Endoscopic saphenous harvesting. *Ann Thorac Surg.* 1997; 64:265-266.
34. Cable DC, Dearani JA. Endoscopic saphenous vein harvesting: minimally invasive video-assisted saphenectomy. *Ann Thorac Surg.* 1997;64:1183-1185.
35. Shalaby M, Waage A, Espevik T. Cytokine regulation of interleukin 6 production by human endothelial cells. *Cell Immunol.* 1989;121:372-382.
36. Cotran RS. American association of pathologists address new roles for the endothelium in inflammation and immunity. *Amer J Pathol.* 1987;129:407.
37. Billiau A, Heremans H, Vermeire K, Matthys P. Immunomodulatory properties of interferon- $\gamma$ . An update. *Ann NY Acad Sci.* 1998;856:22-32.
38. Helmy SA, Wahby MA, El-Nawaway M. The effect of anesthesia and on plasma cytokine production. *Anesthesia.* 1999;54:733-738.
39. Brune IB, Wilke W, Hensler T, Holzmann B, Siewert J. Downregulation of T helper type 1 immune response and altered pro-inflammatory and anti-inflammatory T cell cytokine balance following conventional but not laparoscopic surgery. *Am J Surg.* 1999;177:55-60.
40. Berguer R, Bravo N, Bowyer M, Egan C, Knolmayer T, Ferrick D. Major surgery suppresses maximal production of helper T-cell type 1 cytokines without potentiating the release of helper T- cell type 2 cytokines. *Arch Surg.* 1999;134:540-544.
41. Keith BA, Gary LG, David AH, et al. Endoscopic versus traditional saphenous vein harvesting: a prospective, randomized trial. *Ann Thorac Surg.* 1998;66:26-32.

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