



ORIGINAL ARTICLE

Non-invasive evaluation of coronary heart disease in patients with chronic kidney disease using photoplethysmography

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ABSTRACT

Background. Chronic kidney disease (CKD) patients have an increased risk for coronary artery disease (CAD) and myocardial infarction. Therefore, there is a need to identify CKD patients at high risk of CAD. Coronary angiography, the gold standard for detecting CAD, carries a risk of serious adverse events.

Methods. Here, we assessed the validity of a novel non-invasive reflectance mode photoplethysmography (PPG) sensor for the evaluation of CAD in patients with advanced CKD. PPG signals were generated using green and infrared wavelengths and recorded from fingers of 98 patients. The detected signal has the shape of the pulse wave contour carrying information about the vascular system, that is, arterial stiffness. We studied four patient groups: (i) controls—patients without CKD or CAD; (ii) CKD alone; (iii) CAD alone (confirmed by coronary angiography); and (iv) CKD and CAD combined.

Results. With advancing age, we observed a steeper ascending signal during systole and greater signal decline during diastole (infrared wavelength: Slopes 4–6, $P = 0.002$, $P = 0.003$ and $P = 0.014$, respectively; green wavelength: Slopes 2–3, $P = 0.006$ and $P = 0.005$, respectively). Presence of CAD was associated with a slower signal decline during diastole in CKD patients compared with those without CAD (infrared wavelength: Slope 1, $P = 0.012$). CKD was associated with lower blood volume amplitude during each cardiac cycle compared with those without CKD (R-value, $P = 0.022$).

Conclusions. PPG signal analyses showed significant differences between our groups, and it may be a potentially useful tool for the detection of CAD in CKD patients.

Keywords: chronic kidney disease, coronary artery disease, photoplethysmography, vascular stiffness

Received: 13.9.2018; Editorial decision: 7.12.2018

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INTRODUCTION

Cardiovascular diseases (CVD) such as coronary artery disease (CAD) represent the main causes of morbidity and mortality in patients with chronic kidney disease (CKD) [1, 2]. There is significant interest in cardiac screening tests and their ability to identify CAD in CKD patients [3]. Although coronary angiography is the gold standard for detecting CAD, it is invasive, costly and carries the risk of nephrotoxicity (incidence of 3.3–16.5%), arrhythmia (incidence of 0.1–4.3%), peri-procedural myocardial infarction (defined by elevation of cardiac biomarkers; incidence of 5–30%), embolic events (incidence of 0.6–1.9%) and artery injury (incidence of 0.2–1%) [4–7].

Arterial stiffness secondary to arteriosclerosis and endothelial dysfunction has been recognized as an independent predictor of incident CAD and all-cause mortality in the general population and in patients with CKD [8, 9]. Arterial stiffness is characterized by a reduced capability of an artery to expand and contract in response to blood pressure changes [10]. It can be determined by digital volume pulse analysis with photoplethysmography (PPG) [11]. PPG, a low-cost non-invasive optical technique, is easy to setup and operator-independent—all great advantages for direct clinical application and reproducibility. The detected signal has the shape of the pulse wave contour and contains information about arterial stiffness [10]. By analysing the PPG signal, various vital signs [e.g. heart rate (HR), respiratory rate (RR), blood oxygenation, blood pressure and vasomotion] can be measured in parallel to diagnose and monitor diseases such as CVD, sleep apnoea or diabetes mellitus [12–15].

Light in the range red to near infrared is usually used for transmission mode PPG sensors since it better penetrates tissue than green light [16]. However, when reflectance mode PPG sensors are used, green light might also be suitable for estimation of arterial stiffness [17]. The aim of the study was to identify PPG signal parameters, which are significantly influenced by the presence of angiographically confirmed CAD in patients with CKD. For this purpose, we used a novel reflective PPG sensor to detect separately the reflection of green and infrared light and to analyse PPG signal.

MATERIALS AND METHODS

Study design

The study protocol was approved by the Local Clinical Research Ethics Committee and conforms to the principles outlined in the Declaration of Helsinki. Written informed consent of all patients was obtained. A total of 143 patients from the Cardio-Renal Inpatient Unit, University Hospital Aachen, Germany, were enrolled in the study (Figure 1). Data of six patients were excluded from all analyses due to contradictory statements made by patient or medical report about comorbidities. Data of one patient were excluded due to the withdrawal of the written informed consent. Data of another 10 patients were excluded due to technical problems (transfer of PPG signals to hardware). The remaining 126 patients were separated into four groups: (i) controls—patients without CKD or CAD ($n = 30$); (ii) CKD alone ($n = 28$); (iii) CAD alone ($n = 36$); and (iv) CKD and CAD combined ($n = 32$). CAD was defined as significant coronary stenosis ($\geq 70\%$ stenosis) as detected by coronary angiography [18]. Patients were considered negative for CAD when significant stenosis was excluded during coronary angiography. A maximum of 1 year between coronary angiography and PPG measurements was accepted in the present study. Patients with normal kidney function had an estimated glomerular filtration rate (eGFR) > 60 mL/min/1.73 m² and no albuminuria. All patients with CKD had an eGFR < 30 mL/min/1.73 m² (CKD

Stages 4 and 5). We chose Stages 4 and 5 because of a higher rate of CV events in these sub-populations [19]. Diabetes mellitus was defined as having an HbA_{1c} level of $\geq 6.5\%$ and/or the intake of any anti-diabetic medication. Hypertension was given having either a systolic blood pressure of ≥ 140 mmHg and/or a diastolic blood pressure of ≥ 90 mmHg and/or receiving antihypertensive medication.

PPG signals were measured simultaneously on the left and right index finger. All patients rested for 10 min and PPG recordings were carried out for at least 120 s. During the measurements, patients were kept in a quiet environment, and breathed normally while resting in a supine position. PPG recordings in dialysis patients were performed on non-dialysis days. Indices derived from signals were averaged over 120 s. The PPG signal is composed of alternating (AC) and static (DC) components (Figure 2A). The AC pulsatile waveform is attributed to synchronous cardiac changes in blood volume with each heartbeat, while the DC is the static component of the signal, also containing slow frequencies due to respiration, sympathetic nervous system activity and thermoregulation. The quality of the detected raw PPG signals was evaluated by calculation of the signal-to-noise ratio (SNR):

$$\text{SNR} = 20 \log \left(\frac{\sum_{0.15}^3 \text{Hz} \text{ PSD}(f)}{\sum_{30}^{\infty} \text{Hz} \text{ PSD}(f)} \right) \text{dB}$$

The integral of AC component (0, 15–3 Hz) is defined as signal. Signal components of the power spectral density (PSD) higher than 30 Hz are defined as noise [20]. Only signal sequences of an SNR > 12 dB for > 5 s were used for statistical analyses. PPG signals of 98 patients out of the remaining 126 patients fulfilled the SNR criteria and entered into the present analyses.

Sensor system

The reflectance mode PPG sensor has been developed by the Philips Chair for Medical Information Technology (RWTH Aachen, Germany) in collaboration with the CiS Institute for Microsystems and Photovoltaics (Erfurt, Germany), and was first described in 2009 [21], and has been validated in several human trials [20, 22–25].

The PPG sensor contains two inversely connected light-emitting diodes (LED) with wavelengths of $\lambda_1 = 525$ nm (green) and $\lambda_2 = 948$ nm (infrared). The light emitted by these two LEDs is reflected by skin, tissue and blood (Figure 2B). The raw data generated by reflected light were pre-processed by the microcontroller (MSP430F1609, Texas Instruments, Inc.) and sent to the computer for visualization of PPG signal by using customized software ('LAVIMO-System'). Algorithms as described below were programmed in MATLAB (The MathWorks Inc.) and were used for further automated signal processing [20, 24].

Monitoring

Heart rate. The magnitude of the Reflectance PPG signal (y -value; termed as *Data*), which decreases during systole and increases during diastole over the time (x -value), depends on the blood volume in the capillary. HR was determined by calculation of all Data_{max} per recorded time (Figure 2B).

R-value. The R-value takes the light intensity (I) of the reflected AC and the DC components of the green (λ_{green}) and infrared ($\lambda_{\text{infrared}}$) PPG signal into account [25, 26].

$$R = \frac{I_{\text{AC}} \lambda_{\text{green}}}{I_{\text{DC}} \lambda_{\text{green}}} / \frac{I_{\text{AC}} \lambda_{\text{infrared}}}{I_{\text{DC}} \lambda_{\text{infrared}}}$$

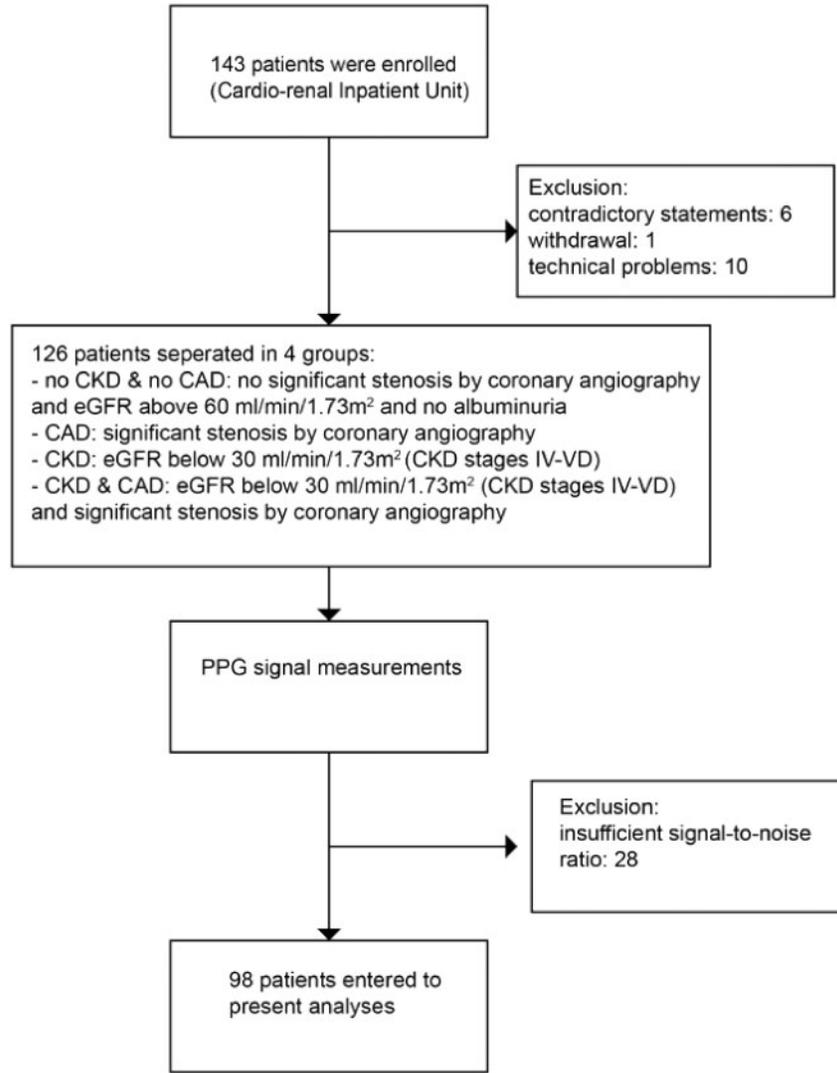


FIGURE 1: Flow chart of patients who met inclusion/exclusion criteria for the study population.

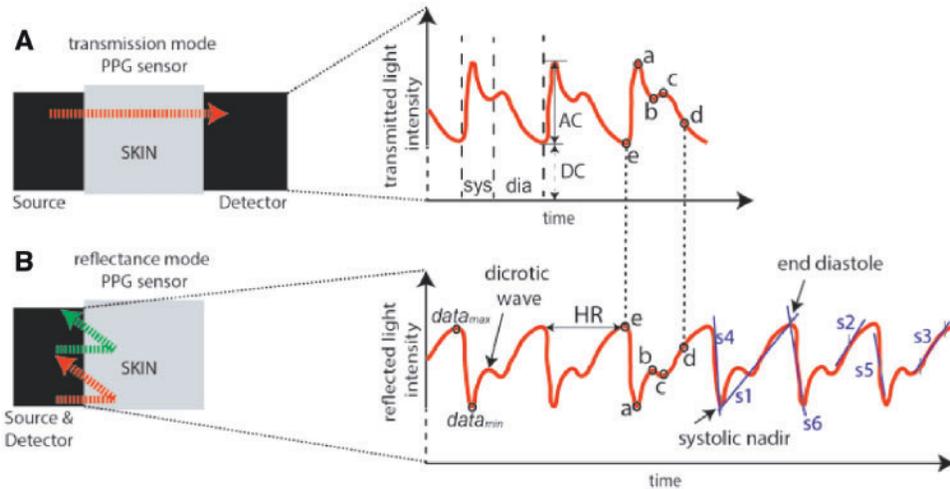


FIGURE 2: PPG sensor. (A and B) Comparison of transmission and reflectance mode PPG sensors. The signal intensity and shape of transmitted PPG signal (A) behaves inverse to PPG signal detected by a reflectance mode PPG sensor (B). sys, systole; dia, diastole; s1–s6, slope of the connecting lines 1–6.

with $AC = Data_{max} - Data_{min}$
and $DC = \frac{1}{2} \times (Data_{max} + Data_{min})$

Respiratory rate. The RR can be determined by the separation of amplitude fluctuations in PPG signal [25]. We chose a fifth-order Butterworth filter with cut-off frequencies 0.34 Hz and calculated all $Data_{min}$ per recorded time.

PPG segments/slopes, dicrotic wave. The systolic amplitude is an indicator of the pulsatile changes in blood volume, caused by arterial blood volume, which is directly proportional to local vascular distensibility. The dicrotic wave (b in Figure 2B) corresponds with the closing of aortic valve and separates systole from diastole [27]. We chose a fourth-order Butterworth filter with cut-off frequencies 3 Hz to calculate dicrotic wave. The wave is followed by a short dip of the PPG signal (c in Figure 2B), which is the result of the short increase in local blood pressure caused by the elastic recoil of the valves and aorta.

To analyse the effect of CAD on PPG signal in more detail, we divided the PPG curve in sections and measured the slope of each section (Figure 2B). We assume that all slopes are steeper when finger arterioles are stiffened (arterial stiffness causes both a greater signal peak during systole and a larger decline during diastole [10]).

Slope 1 (s1) is determined by the slope of the connecting line between minimum and maximum of reflected signal intensity. It correlates with the relative shift of blood volume during late systole and diastole.

$$\text{Slope 1} = \frac{AC}{x_{max} - x_{min}} \times 200 \text{ (per seconds)}$$

S2 and s3 are characterized by changes of reflected light intensity after closure of the aortic valve during diastole. S2 is determined by the slope around the centre of the ascending curve ($centre_{asc}$) (Figure 2B). The centre of the ascending curve was determined by following formula:

$$\text{Centre}_{asc} = \frac{AC}{2} + data(x_{min})$$

$$\text{Slope 2} = \frac{Data(x_{centre_{asc}} + 5) - Data(x_{centre_{asc}} - 5)}{11} \times 200 \text{ (per seconds)}$$

S3 is determined by the slope of the ascending part between one-third and two-thirds of the line between a and e (Figure 2B):

$$\text{Slope 3} = \frac{Data(x_{centre_{asc}} + 15) - Data(x_{centre_{asc}} - 15)}{31} \times 200 \text{ (per seconds)}$$

S4, 5 and 6 describe the reflected light intensity during early systole. S4 is determined by the slope of the connecting line between e and a (Figure 2B):

$$\text{Slope 4} = \frac{-AC}{x_{min} - x_{max}} \times 200 \text{ (per seconds)}$$

S5 is determined by the slope around the centre of the descending curve ($centre_{desc}$) (Figure 2B):

$$\text{Slope 5} = \frac{Data(x_{centre_{desc}} + 5) - Data(x_{centre_{desc}} - 5)}{11} \times 200 \text{ (per seconds)}$$

S6 is determined by the slope of the descending part between one-third and two-thirds of the line between e and a (Figure 2B):

$$\text{Slope 6} = \frac{Data(x_{centre_{desc}} + 15) - Data(x_{centre_{desc}} - 15)}{31} \times 200 \text{ (per seconds)}$$

The slope of those lines was normalized for HR and AC.

Statistical analysis

The data were obtained and analysed by a single observer who was blinded in terms of the presence of CAD or CKD. Descriptive statistics were used to compare baseline clinical and demographic characteristics. The variables were tested for normality using graphic methods (histograms and quantile-quantile plot instead of Q-Q plot) and the Shapiro-Wilk test. Multifactorial repeated measures analysis of variance (ANOVA)/analysis of covariance (ANCOVA) were performed by accounting age, gender, site of measurement (left or right finger) and presence of diabetes mellitus as potential confounding factor. Post hoc analyses of parameters were performed when confounder-adjusted parameters differed between the groups. Paired t-test was performed to analyse differences between the arm with and without an arteriovenous (AV) fistula. $P < 0.05$ were considered significant. All statistical analyses were performed using SAS, version 9.4, and GraphPad Prism 6.01.

RESULTS

Demographic characteristics and comorbidity and influence of age on PPG signal parameters

Baseline clinical and demographic characteristics of the 98 patients entering the analyses are shown in Table 1. The mean age was significantly higher in patients with CKD and CAD compared with the group of patients with CKD ($P = 0.049$).

In all patients, age significantly and independently affected distinct sections of the PPG signal. In particular, using infrared wavelength, we observed advancing age with an augmented increase of PPG signal during systole (Table 2; s4-s6; $P = 0.002$, $P = 0.003$ and $P = 0.014$, respectively). By using the green wavelength, we detected an increase of age with a greater decline of PPG signal during diastole (Table 2; s2 and s3; $P = 0.006$ and $P = 0.005$, respectively).

Influence of CAD on PPG signal parameters

To determine whether the presence of CAD in CKD patients affects PPG signal parameters, PPG data were corrected for age, sex and diabetes mellitus. No differences in PPG signal parameters were detectable between the left and right index finger (data not shown). By using the infrared wavelength, the presence of CAD in patients with CKD was associated with a slower decline of signal during diastole (Table 3; Figure 3A; s1; $P = 0.012$). The PPG signal obtained by the green wavelength was not different between the groups (Table 4).

Influence of CKD and AV fistula on PPG signal parameters

Although the primary aim of the study was to evaluate CAD in CKD patients, we also asked whether CKD or the presence of an AV fistula alters the shape of the PPG signal. We observed that

Table 1. Basic characteristics of the participants who fulfilled the SNR requirements

Characteristic	No CKD and				
	All (n = 98)	no CAD (n = 27)	CAD (n = 31)	CAD and CKD (n = 21)	CKD (n = 19)
Age, mean ± SD, years	67.0 ± 13.4	62.3 ± 17.0	69.0 ± 11.3	72.3 ± 12.0	64.2 ± 13.3
BMI, mean ± SD, kg/m ²	26.5 ± 6.5	25.9 ± 7.9	27.4 ± 4.3	25.6 ± 4.8	27.6 ± 7.9
Male, %	65	52	94	52	53
eGFR, mean ± SD, mL/min/1.73 m ²	N/A	>60	>60	17 ± 11	17 ± 9
Systolic blood pressure, mean ± SD, mmHg	131 ± 21	127 ± 14	130 ± 16	135 ± 25	134 ± 31
Diastolic blood pressure, mean ± SD, mmHg	76 ± 14	76 ± 14	74 ± 10	74 ± 12	78 ± 15
Hypertension, %	75	48	81	91	84
Diabetes mellitus, %	26	11	16	52	32
Aortic stenosis, %	17	22	10	24	16
Chronic heart failure, %	48	52	52	33	53
AV fistula, %	9	0	0	24	21

BMI, body mass index; N/A, not applicable.

Table 2. Association between age and PPG signal

Parameter	P-value	
	λ green	λ infrared
HR	0.84	0.80
RR	0.94	0.35
AC	0.11	0.08
DC	0.43	0.54
R-value	0.27	0.28
Dicrotic wave	0.71	0.22
Slope 1/(AC×HR)	0.49	0.24
Slope 2/(AC×HR)	0.006*	0.36
Slope 3/(AC×HR)	0.005*	0.27
Slope 4/(AC×HR)	0.13	0.002*
Slope 5/(AC×HR)	0.15	0.003*
Slope 6/(AC×HR)	0.23	0.014*

*P < 0.05 were considered significant.

the R-values, which correlate with the blood volume amplitude during systole, were significantly lower in CKD patients compared with patients with normal kidney function (Tables 3 and 4). This was true for the comparison of CKD patients with both control or CAD patients (P = 0.021 and P = 0.036, respectively) (Figure 3B).

Using the green wavelength, PPG signals were not different between the arm with an AV fistula versus those without (Table 5). The infrared wavelength-derived PPG signal showed significantly steeper increase in blood volume during systole in the arm with AV fistula compared with the arm without AV fistula (Table 5; s2 and s3; P = 0.041 and P = 0.048, respectively).

Signal quality

We observed that our programme counted fewer PPG-derived values for HR, RR and R-value for each patient, when they were generated by green light as compared with infrared light (Supplementary data, Table S1). In contrast, fewer values for s1–s6 were counted when infrared light was used (Supplementary data, Table S1).

DISCUSSION

In this work, we evaluated the potential of a novel reflectance PPG sensor as a non-invasive tool to identify PPG signal

parameters, which are significantly influenced by the presence of angiographically confirmed CAD in patients with CKD.

Infrared wavelength is usually used to generate PPG signals, but green-wavelength PPG sensors are becoming increasingly popular for their higher specificity for detecting vascular alterations compared with infrared light [28]. The current study compared the performance of infrared and green wavelength and found significant alterations in PPG signal with advancing age and presence of CAD. Our findings suggest that the blood volume decreases more slowly during diastole in capillaries of CKD patients with CAD compared with CKD patients without CAD, and we speculate that this is explained by changes in arterial stiffness and/or changes in the tone of small vessels. However, this observation was limited to PPG signals generated by infrared LED. Sollinger et al. [29] demonstrated in a small subgroup analysis that patients with CKD had a greater PPG-derived stiffness index when they had a history of CAD compared with CKD patients without CAD. Although the presence of subclinical CAD in CKD patients was not angiographically evaluated and the sample group was only around a dozen, this study supports our findings. Previous reports also demonstrated an independent relationship between arterial stiffness and age [29–31]. In agreement with these reports, we observed in all patients that PPG slopes were significantly influenced by age.

We found that the presence of CKD was associated with lower blood volume amplitudes, as estimated by measuring the R-value. Especially, the lower AC values in CKD patients caused these differences. AC (blood volume amplitude in the finger capillaries during each cardiac cycle) is low in case of decreased blood volume shifts/pulsations, decreased venous blood volume or due to peripheral vasoconstriction [32]. Further studies are needed to clarify the reason for lower AC values in CKD patients. For example, a role of renin-angiotensin-aldosterone system herein is possible. So far, there is evidence that angiotensin II and aldosterone, both hormones that are linked to CKD, contribute significantly to arterial stiffness [33].

We tested whether the presence of a dialysis AV fistula influences the shape of the PPG signal. Using the infrared wavelength, we detected significantly steeper increase in blood volume during systole in the arm with an AV fistula compared with the arm without AV fistula, suggesting an increased small vascular tone in the shunt arm. AV fistula undergoes vascular remodelling and causes an altered haemodynamic situation and endothelial dysfunction in the dialysis arm [34]. Endothelial dysfunction is associated with reduced endothelial vasodilator

Table 3. Analysis of PPG signals (infrared wavelength)

Parameter	No CKD/no CAD	CAD	CAD/CKD	CKD
HR (beats per minute)	70 ± 10	68 ± 9	67 ± 9	72 ± 11
RR (breaths per minute)	15 ± 1	14 ± 1	15 ± 1	15 ± 1
AC	18 342 ± 12 909	24 731 ± 15 425	18 904 ± 14 477	16 106 ± 15 348
DC	1 298 369 ± 400 326	1 330 661 ± 411 236	1 355 604 ± 340 176	1 269 757 ± 356 132
R-value	0.32 ± 0.29*	0.30 ± 0.13*	0.21 ± 0.11*	0.21 ± 0.11*
Dicrotic wave	2116 ± 1564	2732 ± 1872	2116 ± 1442	1505 ± 1024
Slope 1/(AC×HR) (per second)	0.025 ± 0.004*	0.025 ± 0.003*	0.025 ± 0.003*	0.027 ± 0.003*
Slope 2/(AC×HR) (per second)	0.026 ± 0.008	0.028 ± 0.012	0.031 ± 0.015	0.029 ± 0.01
Slope 3/(AC×HR) (per second)	0.028 ± 0.08	0.029 ± 0.11	0.033 ± 0.015	0.032 ± 0.01
Slope 4/(AC×HR) (per second)	-0.047 ± 0.006	-0.0477 ± 0.0073	-0.049 ± 0.008	-0.045 ± 0.006
Slope 5/(AC×HR) (per second)	-0.07 ± 0.009	-0.072 ± 0.01	-0.07 ± 0.01	-0.069 ± 0.007
Slope 6/(AC×HR) (per second)	-0.67 ± 0.008	-0.0684 ± 0.009	-0.07 ± 0.011	-0.066 ± 0.008

PPG signal-derived values were obtained from each group using the infrared wavelength. Values are presented as mean ± SD. Slope of the connecting lines 1–6 were adjusted for HR and AC. Between-group comparisons were made by two-way repeated measures ANOVA.

*P < 0.05 were considered significant.

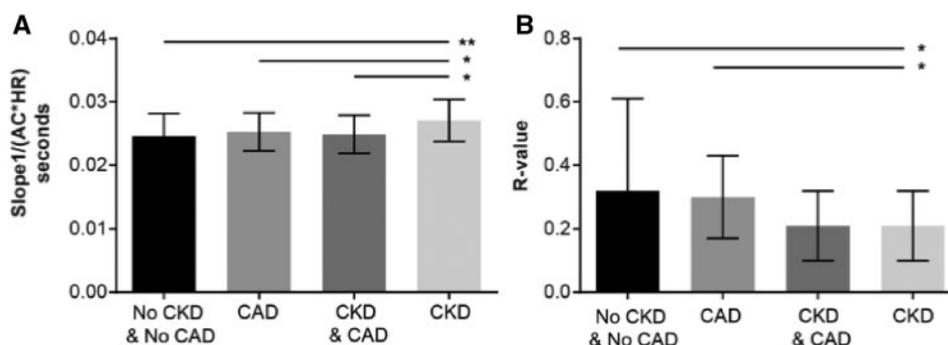


FIGURE 3: PPG signal-derived values for $s1/(AC \times HR)$ and R-value using infrared wavelength. Values for $s1/(AC \times HR)$ (A) and R-value (B) are presented as mean ± SD. Between-group comparisons were made by two-way repeated measures ANOVA and post hoc analysis. From each patient, $n = 118$ – 124 or $n = 87$ – 103 data points were recorded to quantify $s1/(AC \times HR)$ or R-value, respectively. $P < 0.05$ or 0.01 were considered significant (*) or highly significant (**). See also Table 3.

Table 4. Analysis of PPG signals (green wavelength)

Parameter	No CKD and No CAD	CAD	CAD and CKD	CKD
HR (beats per minute)	71 ± 10	67 ± 9	67 ± 9	72 ± 11
RR (breaths per minute)	14 ± 2	14 ± 1	14 ± 1	15 ± 1
AC	6826 ± 5937	7542 ± 7556	7535 ± 6355	6479 ± 8864
DC	116 766 ± 64 382	97 294 ± 63 317	101 091 ± 52 948	79 569 ± 45 502
R-value	0.32 ± 0.29*	0.30 ± 0.13*	0.21 ± 0.11*	0.21 ± 0.11*
Dicrotic wave	851 ± 752	793 ± 882	878 ± 852	509 ± 814
Slope 1/(AC×HR) (per second)	0.022 ± 0.004	0.023 ± 0.003	0.023 ± 0.003	0.032 ± 0.04
Slope 2/(AC×HR) (per second)	0.022 ± 0.007	0.022 ± 0.006	0.026 ± 0.01	0.022 ± 0.006
Slope 3/(AC×HR) (per second)	0.024 ± 0.007	0.024 ± 0.006	0.027 ± 0.009	0.024 ± 0.006
Slope 4/(AC×HR) (per second)	-0.044 ± 0.010	-0.047 ± 0.009	-0.046 ± 0.008	-0.044 ± 0.007
Slope 5/(AC×HR) (per second)	-0.067 ± 0.015	-0.071 ± 0.013	-0.069 ± 0.012	-0.067 ± 0.013
Slope 6/(AC×HR) (per second)	-0.067 ± 0.014	-0.067 ± 0.012	-0.066 ± 0.011	-0.06 ± 0.013

PPG signal-derived values were obtained from each group using the green wavelength. Values are presented as mean ± SD. Slope of the connecting lines 1–6 were adjusted for HR and AC. Between-group comparisons were made by two-way repeated measures ANOVA.

*P-values < 0.05 were considered significant.

activity (mediated by nitric oxide, prostacyclin and bradykinin) and increased synthesis of vasoconstrictors (endothelin, angiotensin II and free oxygen radicals) [35, 36], resulting in an increased small vascular tone.

Our study has some limitations. The sample size was small, which limits our ability to detect a cut-off value for a blinded

detection of CAD. However, we minimized the sample size effect by adjusting the parameters for age, sex and presence of diabetes mellitus. Despite PPG's wide range of applications, there are technical limitations. The PPG sensor can only detect dynamic changes of the blood volume at one spot per probe, thus PPG signal interpretation has to be discussed with

Table 5. PPG signal parameters recorded from the arm with AV fistula and arm without AV fistula

Parameter	Green wavelength		Infrared wavelength	
	AV fistula	No AV fistula	AV fistula	No AV fistula
HR (beats per minute)	71	70	70	70
RR (breaths per minute)	15	15	16	15
AC	4262	5242	14 148	20 339
DC	74 327	83 279	1 192 405	1 326 814
R-value	0.22	0.22	0.22	0.22
Dicrotic wave	568	743	2020	1949
Slope 1/(AC×HR) (per second)	0.023	0.23	0.027	0.025
Slope 2/(AC×HR) (per second)	0.028	0.029	0.039*	0.032*
Slope 3/(AC×HR) (per second)	0.029	0.030	0.040*	0.034*
Slope 4/(AC×HR) (per second)	-0.043	-0.048	-0.046	-0.049
Slope 5/(AC×HR) (per second)	-0.065	-0.071	-0.071	-0.073
Slope 6/(AC×HR) (per second)	-0.062	-0.067	-0.067	-0.069

Mean PPG signal-derived values were obtained from each group using the green and infrared wavelength. Slope of the connecting lines 1–6 were adjusted for HR and AC. Asterisks marks values that were significantly different between arm with fistula and arm without fistula. $P < 0.05$ were considered as significant.

caution. A remote imaging PPG technology may overcome this limitation by detecting signal simultaneously from multiple sites [37]. Another technical limitation is that PPG is susceptible to signal artefacts. Therefore, we evaluated the quality of the detected PPG signal by calculation of the signal-to-noise ratio and had to exclude 22% of all enrolled patients. While PPG may detect CAD-associated changes in the vessels of investigation, we are aware that there are other factors that influence cardiac morbidity and mortality that we were unable to evaluate with PPG [38]. Patients with CAD and CKD have complex and severe microcirculatory alterations (i.e. abnormal coronary ischaemic events in the absence of significant stenosis) [39, 40], and randomized controlled trails are needed to assess the predictive capacity of PPG in determining CAD in patients with CKD.

In summary, PPG is an interesting technology due to its simplicity, low-cost and non-invasiveness, and PPG signal was affected by the presence of angiographically confirmed CAD in patients with advanced CKD. Although the use of PPG in CKD patients may represent a promising diagnostic tool and could help physicians to choose appropriate treatment, it requires further validation in larger prospective clinical cohorts.

SUPPLEMENTARY DATA

Supplementary data are available at ckj online.

ACKNOWLEDGEMENTS

The authors thank Dr D. Starke[†] and Dr O. Brodersen from the Institute for Microsystems and Photovoltaik (CiS), Erfurt, Germany, for providing sensor hardware.

FUNDING

T.S. was supported by a START grant (691433), by a Rotationsstellen-program of the Faculty of Medicine of the RWTH Aachen University and by a grant of the Else-Kröner-Fresenius Foundation (2015/A197). V.G.P. was awarded a CJ Martin Fellowship by the National Health and Medical Research Council of Australia (NHMRC 1128582).

CONFLICT OF INTEREST STATEMENT

None declared.

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