


RESEARCH

Open Access



Therapeutic mechanism of Toujie Quwen granules in COVID-19 based on network pharmacology

Ying Huang^{1,2†}, Wen-jiang Zheng^{1†}, Yong-shi Ni³, Mian-sha Li⁴ , Jian-kun Chen², Xiao-hong Liu¹, Xing-hua Tan⁵ and Ji-qiang Li^{2*}

* Correspondence: lijiqiangjizhen@163.com

[†]Ying Huang and Wen-jiang Zheng contributed equally to this work.

²Integrative Dept.3 (Geriatrics Dept), Guangdong Provincial Hospital of Chinese Medicine, Guangzhou, China

Full list of author information is available at the end of the article

Abstract

Background: Chinese medicine Toujie Quwen granule (TJQW) has proven to be effective in the treatment of mild coronavirus disease 2019 (COVID-19) cases by relieving symptoms, slowing the progression of the disease, and boosting the recovery of patients. But the bioactive compounds and potential mechanisms of TJQW for COVID-19 prevention and treatment are unclear. This study aimed to explore the potential therapeutic mechanism of TJQW in coronavirus disease 2019 (COVID-19) based on an integrated network pharmacology approach.

Methods: TCMSP were used to search and screen the active ingredients in TJQW. The Swiss TargetPrediction was used to predict the potential targets of active ingredients. Genes co-expressed with ACE2 were considered potential therapeutic targets on COVID-19. Venn diagram was created to show correlative targets of TJQW against COVID-19. Cytoscape was used to construct a “drug-active ingredient-potential target” network, STRING were used to construct protein-protein interaction network, and cytoHubba performed network topology analysis. Enrichment of biological functions and signaling pathways of core targets was performed by using the clusterProfiler package in R software and ClueGO with CluePedia plugins in Cytoscape.

(Continued on next page)



© The Author(s). 2020 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

(Continued from previous page)

Results: A total of 156 active ingredients were obtained through oral bioavailability and drug-likeness screenings. Two hundred twenty-seven potential targets of TJQW were related to COVID-19. The top ten core targets are EGFR, CASP3, STAT3, ESR1, FPR2, F2, BCL2L1, BDKRB2, MPO, and ACE. Based on that, we obtained 19 key active ingredients: umbelliprenin, quercetin, kaempferol, luteolin, praeruptorin E, stigmasterol, and oroxylin A. And the enrichment analysis obtained multiple related gene ontology functions and signaling pathways. Lastly, we constructed a key network of “drug-component-target-biological process-signaling pathway”. Our findings suggested that TJQW treatment for COVID-19 was associated with elevation of immunity and suppression of inflammatory stress, including regulation of inflammatory response, viral process, neutrophil mediated immunity, PI3K-Akt signaling pathway, MAPK signaling pathway, Jak-STAT signaling pathway, Complement and coagulation cascades, and HIF-1 signaling pathway.

Conclusions: Our study uncovered the pharmacological mechanism underlying TJQW treatment for COVID-19. These results should benefit efforts for people around the world to gain more knowledge about Chinese medicine TJQW in the treatment of this vicious epidemic COVID-19, and help to address this pressing problem currently facing the world.

Keywords: COVID-19, Toujie Quwen granule, Potential therapeutic targets, Signaling pathways, Network pharmacology

Introduction

Since December 2019, a novel coronavirus pneumonia, namely coronavirus disease 2019 (COVID-19) has rapidly spread from Wuhan City to various provinces across China and other countries around the world [1]. Update to 29 July 2020, there were 16,341,920 confirmed cases, 650,805 confirmed deaths, and 216 countries, areas or territories with cases, according to the WHO. (<https://www.who.int/emergencies/diseases/novel-coronavirus-2019>). As an acute respiratory infectious disease, COVID-19 is transmitted through respiratory droplets and close contact, and the general population is susceptible to the infection. No specific drug but symptomatic supporting treatment is currently available for COVID-19. According to the traditional Chinese medicine (TCM) theory, COVID-19 belongs to the category of pestilence. Since the outbreak of COVID-19, the Chinese government has explicitly requested adherence to the integration of TCM and western medicine, enhancement of the research efforts, and acceleration of the development of drugs with good clinical efficacy for COVID-19. The Diagnosis and Treatment Program of Novel Coronavirus Pneumonia (trial editions 4 to 7) jointly issued by the National Health Commission of China and the National Administration of TCM of the People’s Republic also included a TCM treatment section [2].

Among all recommended TCM formulations for COVID-19 treatment, Guangzhou Eighth People’s Hospital (<http://www.gz8h.com.cn/>, Guangdong Province, China) introduced Toujie Quwen granules (TJQW, formerly known as pneumonia No. 1 formulation) based on the febrile disease theory of TCM and climate characteristics of Lingnan region, TJQW formulation has achieved good therapeutic results. TJQW have been mainly used to treat mild to moderate COVID-19 cases to effectively reduce fever, cough, and expectoration in the patients. On February 8, 2020, Guangdong Medical Products Administration officially recommended TJQW as COVID-19 treatment to 30

designated hospitals for COVID-19 in the Guangdong Province [3]. TJQW includes 16 TCM components, including *Forsythiae Fructus* (lian-qiao), edible tulip (shan-ci-gu), *Lonicera japonica* (Japanese honeysuckle flower, jin-yin-hua), *Radix Scutellariae baicalensis* (huang-qin), *Folium Isatidis* (da-qing-ye), *Bupleurum* root (chai-hu), *Artemisia apiacea* (qing-hao), *Periostracum Cicadae* (chan-tui), *Radix Peucedani* (qian-hu), *Fritillaria cirrhosa* (chuan-bei-mu), *Fritillaria thunbergii* (zhe-bei-mu), *Poria cocos* (fu-ling), *Fructus Mume* (wu-mei), radix *Scrophulariae* (xuan-shen), *Astragalus propinquus* (huang-qi), and radix *Pseudostellariae* (tai-zi-shen), which were mainly used for heat-clearing, detoxifying, relieving symptoms, expelling pathogenic factors, reinforcing *qi*, and nourishing *yin* according to the TCM theory [4].

TCM and Western medicine may come from two different medical systems and have different perspectives on health and diseases, but they are both based on the standard of factual clinical efficacy. Network pharmacology has emerged as a powerful tool to reveal active ingredients, biological targets and signaling pathways linked to certain diseases [5, 6]. The therapeutic target for COVID-19 remains unclear, while study has shown that the novel coronavirus infects alveolar type II cells through the mechanism of interaction of the spike protein with the human renin and type II angiotensin converting enzyme (ACE2), leading to pneumonia [7], therefore, genes co-expressed with ACE2 are considered as potential therapeutic targets on COVID-19 [8]. ACE2 is expressed by epithelial cells of the lungs at a high level, a major target of the disease, SARS-CoV-2 infects ACE2-expressing cells in the lung, blocking ACE2 interaction with the S protein of SARS-CoV-2 to curtail SARS-CoV-2 infection are becoming very attractive therapeutics potential for treatment and prevention of COVID-19, thus, we could target this interaction site with small molecules [9]. We analyzed the co-expressed genes with ACE2 in order to explore different target genes on which the small molecules could affect the SARS-CoV-2 activity [10].

This study selected the active ingredients of TJQW and used the network pharmacology approach to construct a common target for the drug and COVID-19, to further explore the effective ingredients and molecular mechanisms of this TCM formulation for COVID-19, and to provide further scientific basis for the study of TCM treatments for COVID-19, the overall workflow of this study was presented in Fig. 1.

Materials and methods

Screening of potential biologically active ingredients in TJQW

TJQW had a total of 16 TCM components, and the active ingredients in the related TCM components were searched through the TCM systems pharmacology database and the analysis platform (LSP, <https://tcmspw.com/tcmsp.php>) [11]. LSP captures relationships between drugs, targets, and diseases, includes pharmacokinetic properties for natural compounds involving oral bioavailability (OB), drug-likeness (DL), and this has started a trend in the search for candidate drugs in various types of traditional Chinese herbs. The TCM formulae must first overcome the barriers posed by absorption, distribution, metabolism, and excretion (ADME) processes, and only the molecules that pass through those barriers may be active. High OB is usually a key indicator for determining the DL index of bioactive molecules [12, 13]. Therefore, in this study, we screened bioactive molecules according to the pharmacokinetic parameters of OB $\geq 30\%$ and DL

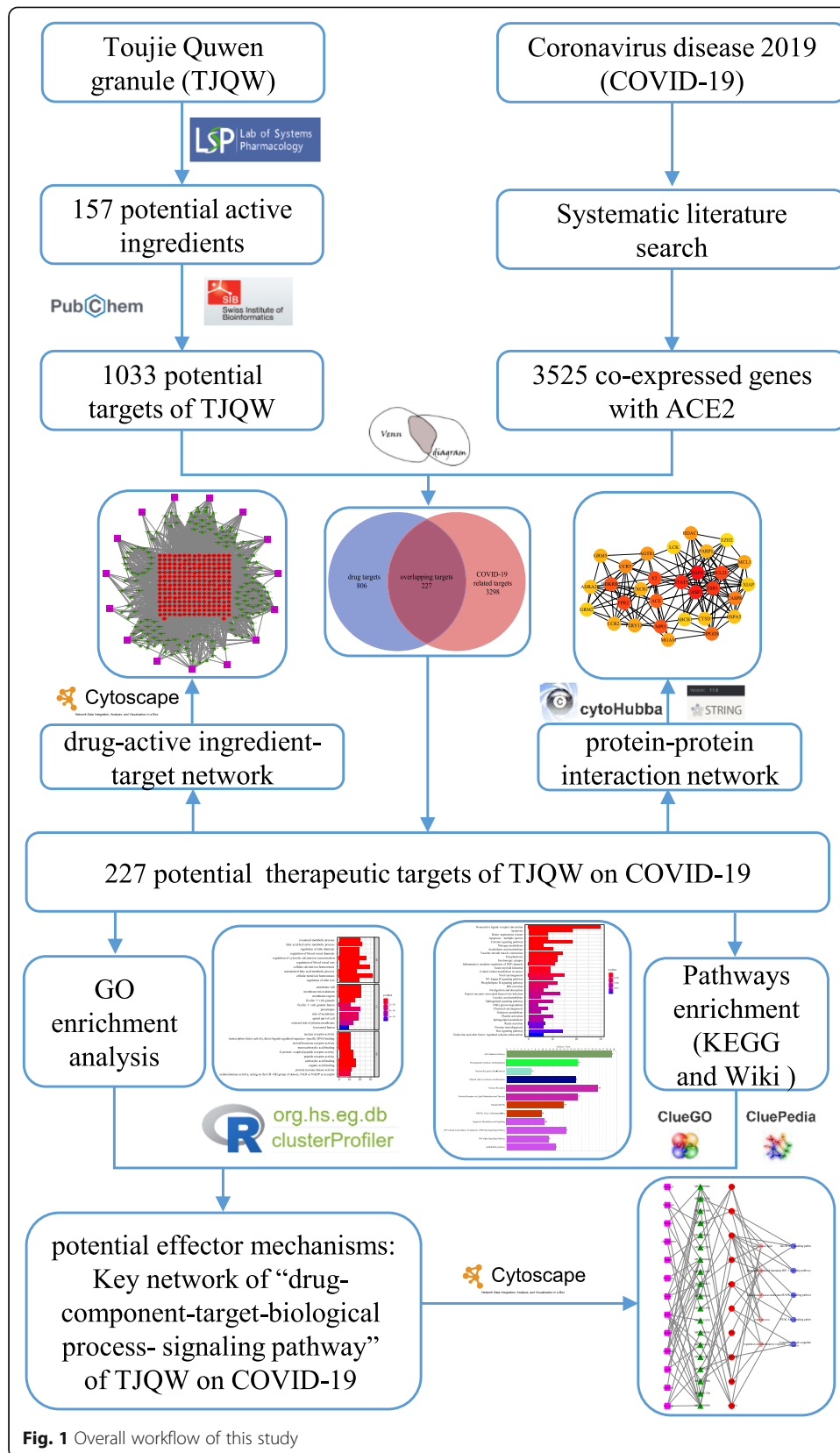


Fig. 1 Overall workflow of this study

≥ 0.18 screening conditions. Then we obtained the chemical abstracts service (CAS) number or the IUPAC International Chemical Identifier (InChIKey) of those molecules from TCMSP.

Prediction of potential targets for bioactive ingredients

The active ingredients obtained from screening were entered into the PubChem (<https://pubchem.ncbi.nlm.nih.gov/>) using their CAS number or InChIKey to acquire the corresponding molecular structures which were stored in canonical simplified molecular-input line-entry system (SMILES) format.

The Swiss TargetPrediction (<http://www.swisstargetprediction.ch/>, version 2019) [14] was then used to predict the potential targets of the active ingredients. We input the aforementioned potential active ingredients in SMILES format into this database, with “humans” (*Homo sapiens*) as the study species, to obtain the potential effector targets.

Selection of the intersection of the drug targets and disease targets

Our study was based on the results of single-cell sequencing of colonic epithelial cells in a previous study [15] to select the genes co-expressed with ACE2, which are considered as potential therapeutic targets on COVID-19, and the gene names in the original files were standardized to match the human-related targets. The targets of the active ingredients of the drug and the targets of the disease were intersected, the intersectional targets were considered potential therapeutic targets of TJQW in COVID-19.

Construction of a drug-active ingredient- potential target network

The components and the obtained active ingredients of TJQW and the target correspondence were imported into the Cytoscape (<https://cytoscape.org/>, version 3.7.0) [16] to construct the drug-active ingredient- potential target network.

Network topology analysis of potential therapeutic targets

The Search Tool for the Retrieval of Interacting Genes/Proteins (STRING) (<https://string-db.org/>, version:11.0) [17], was used to construct a protein-protein interaction (PPI) network, the species were set to “*Homo sapiens*”; the lowest mutual action threshold was set to “medium confidence” (> 0.4), and other parameters were the default settings. And the PPI results were imported into cytoHubba (<https://apps.cytoscape.org/apps/cytohubba>, version 0.1) [18] for network topology analysis. An important hub gene can be selected through calculation and analysis of the network structure and the weighted reconnection between nodes. For each node in the interaction network, we selected “Degree” to calculate topological features, which is defined as the number of edges to node i . When applying the degree algorithm, the results were sorted by the degree value.

Gene annotation analysis of potential therapeutic targets

The org. Hs.eg.db (<https://www.bioconductor.org/>, a package for the genome wide annotation of Human, primarily based on mapping using Entrez Gene identifiers, version 3.1.1) [19] and the clusterProfiler (<https://www.bioconductor.org>, a package implements methods to analyze and visualize functional profiles of gene and gene clusters, version

3.1.1) [20] on the R 3.5.2, were used to perform the gene ontology (GO) enrichment analysis [21] and the Kyoto Gene and Genome Encyclopedia (KEGG) enrichment analysis [22]. In our study, the R code is shown as below.

ClueGO (version 2.5.4) [23] and CluePedia (version 1.5.4) [24] plugins in Cytoscape were used to perform WikiPathways enrichment analysis. We chose the analysis mode of Functional analysis, load marker list is *Homo sapiens* (9606) and Symbol ID was input, then we selected “WikiPathways-503 terms/pathways with 6558 available unique genes-27.02.2019”. Regarding statistical options, we employed two-sided hypergeometric test and bonferroni step down for p value correction. With $P < 0.05$ were selected. The results of p value ≤ 0.05 were used to graph the GO and KEGG pathways, illustrating the roles of targets and signaling pathways in COVID-19 treatment with TJQW.

```
(1) enrich GO ego <- enrichGO(gene = gene$ENTREZID,
                               OrgDb=org.Hs.eg.db,
                               ont = "all",
                               pAdjustMethod = "BH",
                               minGSSize = 2,
                               pvalueCutoff = 0.05,
                               qvalueCutoff = 0.05,
                               readable = TRUE)
if(T){go <- enrichGO(gene = gene$ENTREZID, OrgDb = "org.Hs.eg.db", ont="all",
                    pAdjustMethod = "BH",
                    minGSSize = 2,
                    pvalueCutoff = 0.05,
                    qvalueCutoff = 0.05,
                    readable = TRUE)}

(2) enrich KEGG if(T){EGG <- enrichKEGG(gene= gene$ENTREZID,
                                         organism= 'hsa',
                                         pvalueCutoff = 0.05)}
if(T){EGG <- enrichKEGG(gene= gene$ENTREZID,
                        organism= 'hsa',
                        minGSSize = 5, maxGSSize = 500, qvalueCutoff = 0.05,
                        pvalueCutoff = 0.05)}
```

Results

Screening of the active ingredients in 16 components of TJQW

The TCMSP database was used to search for the active ingredients of TJQW, followed by using the parameters with $\geq 30\%$ OB and ≥ 0.18 DL as the criteria to screen the 15 components (among all components, *Periostracum Cicadae* did not have any relevant retrieval results, which ingredients did not match the parameters with $\geq 30\%$ OB and ≥ 0.18 DL) to obtain 238 active ingredients. Details of 15 components and its 238 active ingredients (without removing duplication) including herbal name, molecule ID, molecule name, OB information, DL information, and their CAS/InChIKey codes were shown in Supplementary material 1. After eliminating duplicates, a total of 194 active ingredients were obtained.

Screening of active ingredients for COVID-19 related targets

There are 38 out of 194 active ingredients that cannot be found in PubChem or cannot be predicted in Swiss TargetPrediction, so only 156 unique active ingredients and their

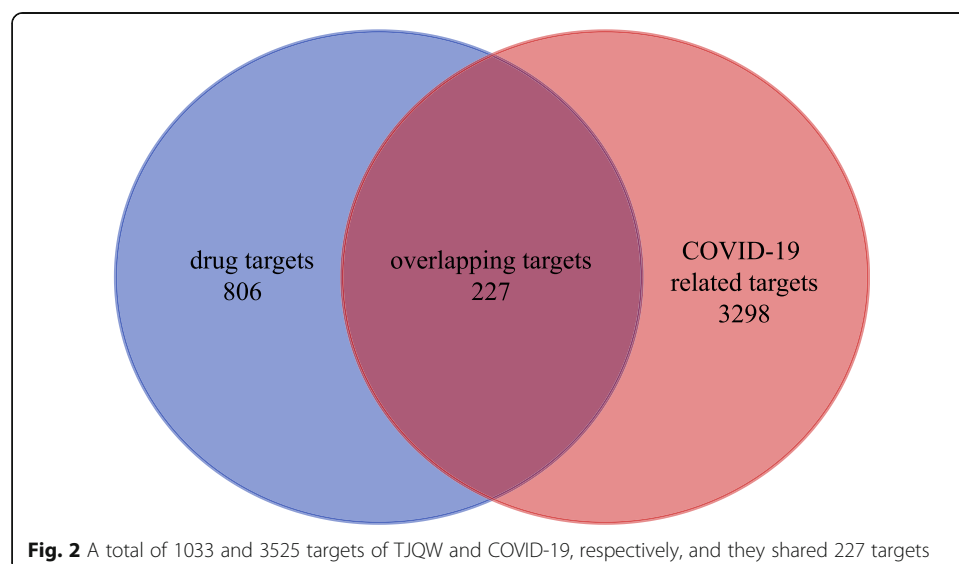
target information are obtained, including a total of 1033 potential targets. According to the single-cell sequencing results of colonic epithelial cells, 5556 genes, which were co-expressed with ACE2, were identified. After normalizing the gene names in the original files, a total of 3525 human targets were matched. Subsequently, 1033 ingredient targets and 3525 COVID-19 targets were used to draw a Venn diagram to obtain 227 coincident targets, as shown in Fig. 1, i.e., the relevant targets of the TCM active ingredients acted on the disease. Moreover, the targets of 152 ingredients have an intersection with COVID-19, and 4 ingredients do not. Therefore, 148 potential active ingredients may be related to COVID-19, as shown in Fig. 2.

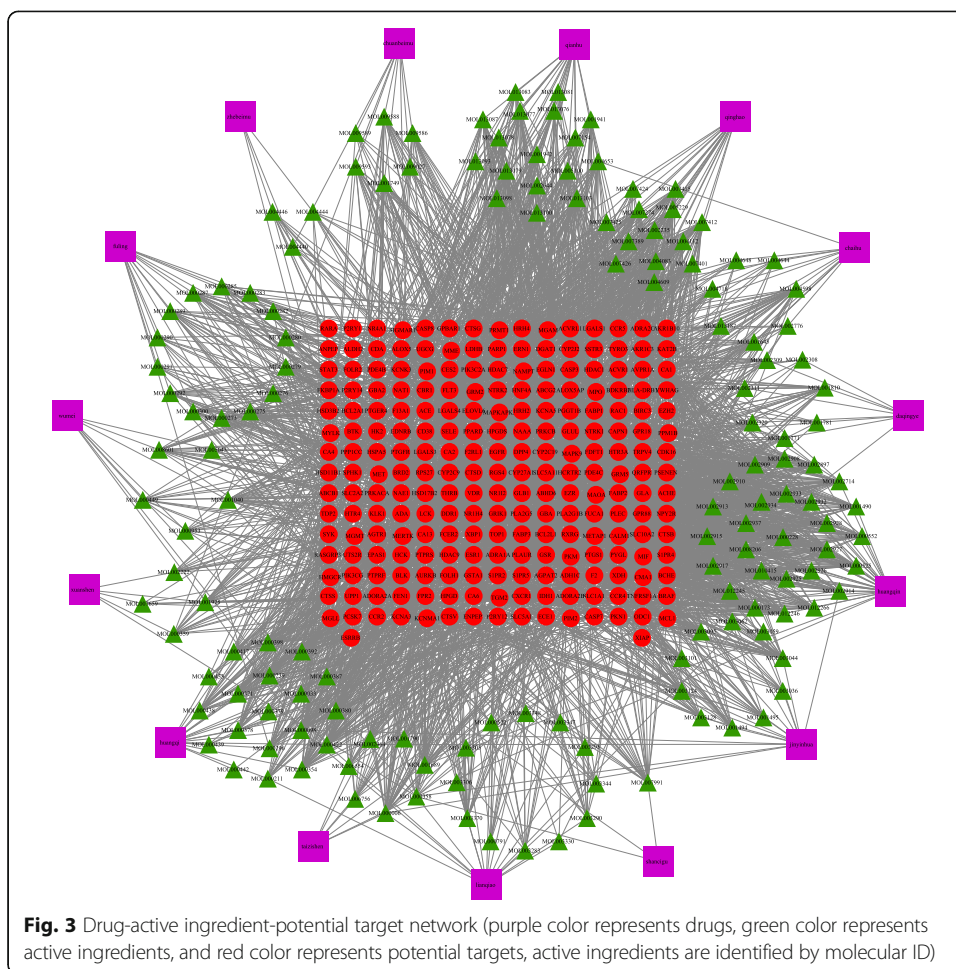
The drug-active ingredient-target network analysis

As shown in Fig. 3, Cytoscape was used to construct the drug-active ingredient-target network. On this network, different nodes represented drugs, potential active ingredients and effector targets of TJQW, and the network's edges showed relationships between these three factors. Rectangles with purple color represent 15 components of TJQW. The surrounding green triangles represent their active ingredients. And the 227 red circles in the center represents the potential targets of TJQW on COVID-19. This fully indicated the characteristics of multi-components and multi-targets of TCM. When hub genes were found, a new simplified network was built to show relationships between key ingredients, key targets, and key pathways.

Network topology analysis

The network topology analysis showed the network parameters with targets (Table 1 and Fig. 4), visually showing the network relationship of the top 30 targets with degree values. Figure 4 displays the connection status of these hub nodes in the network. The darker the color, the higher the value. There are 30 nodes and 155 edges in this network, average node degree is 28.8, the PPI enrichment p -value $< 1.0e-16$. The top ten targets are EGFR, CASP3, STAT3, ESR1, FPR2, F2, BCL2L1, BDKRB2, MPO, and ACE. Their source composition information was shown in Table 2.





GO enrichment analysis

GO enrichment analysis results included GO-molecular function (MF), GO-biological process (BP), and GO-cell component (CC), followed by the selection of the top 10 enrichment results. In Fig. 5, the y-axis represents the enriched categories and the x-axis represents the number of enrichment. The top 10 results of GO-BP included blood vessel diameter regulation, blood vessel inner diameter regulation, cytosolic calcium-ion concentration regulation, cellular calcium-ion homeostasis, intracellular metal-ion hemostasis, and unsaturated fatty-acid metabolism process. The top 10 enrichment results of GO-CC included membrane rafts, membrane micro-regions, membrane regions, presynapses, membrane sides, apical part of the cell, plasma membrane outer surface, and lysosome cavity. The top 10 enrichment results of GO-MF included nuclear receptor activity, steroid hormone receptor activity, monocarboxylic acid-binding, G protein-coupled receptor activity, peptide receptor activity, organic acid binding, protein tyrosine kinase activity, and oxidoreductase activity.

Signaling pathways enrichment analysis

KEGG pathways enrichment analysis was shown in Fig. 6, the y-axis represents the enrichment pathway name and the x-axis represents the numbers of enrichment. The

Table 1 Top 30 in network ranked by Degree method

| No. | Target | Degree | No. | Target | Degree |
|-----|--------|--------|-----|--------|--------|
| 1 | EGFR | 63 | 16 | MCL1 | 24 |
| 2 | CASP3 | 54 | 17 | MGAM | 24 |
| 3 | STAT3 | 46 | 18 | P2RY12 | 24 |
| 4 | ESR1 | 42 | 19 | PARP1 | 23 |
| 5 | BCL2L1 | 34 | 20 | GRM5 | 23 |
| 6 | F2 | 34 | 21 | CXCR1 | 23 |
| 7 | FPR2 | 34 | 22 | CCR2 | 23 |
| 8 | MPO | 31 | 23 | HSPA5 | 22 |
| 9 | BDKRB2 | 31 | 24 | CTSD | 22 |
| 10 | ACE | 30 | 25 | ADRA2C | 22 |
| 11 | HPGDS | 28 | 26 | XIAP | 21 |
| 12 | CASP8 | 27 | 27 | LCK | 21 |
| 13 | CCR5 | 26 | 28 | ABCB1 | 21 |
| 14 | HDAC1 | 25 | 29 | GRM2 | 21 |
| 15 | AGTR1 | 25 | 30 | EZH2 | 20 |

results suggested that the KEGG pathways of TJQW targets against COVID-19 were mainly involved in neuroactive ligand-receptor interaction, apoptosis, renin-angiotensin system, calcium signaling pathway, arachidonic acid metabolism, vascular smooth muscle contraction, toxoplasmosis, Inflammatory mediator regulation of transient receptor potential (TRP) channels, acute myeloid leukemia, central carbon metabolism in cancer, platelet activation, and nuclear factor kappa-B (NF-κB) signaling pathway.

WikiPathways enrichment analysis was shown in Fig. 7, the y-axis represents the enrichment pathway name and the x-axis represents the numbers of enrichment. The

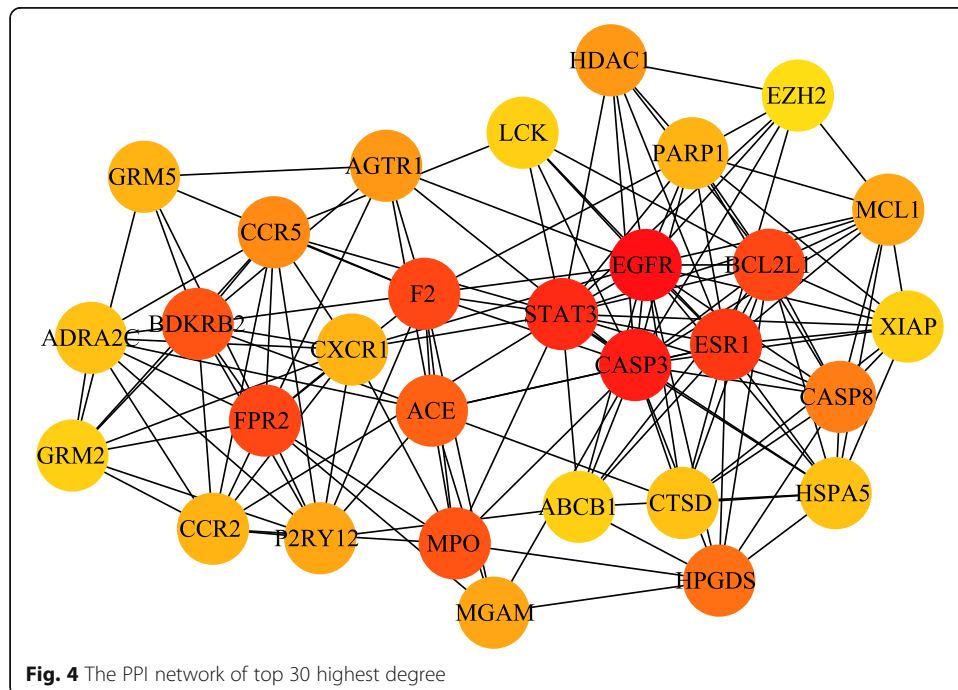


Table 2 Top ten targets and their source composition information

| No | Target | Degree | Mol ID | Molecule Name | Probability ^a | OB (%) | DL | CAS/InChIKey | pinyin name | herbal name |
|----|--------|--------|-----------|---|--------------------------|--------|------|---------------------------------|-------------|------------------------------------|
| 1 | EGFR | 63 | MOL000098 | quercetin | 1.000 | 46.43 | 0.28 | 117-39-5 | wumei | Mume Fructus |
| | | | | | | | | | qinghao | <i>Artemisia annua</i> L. |
| | | | | | | | | | qianhu | Peucedani Radix |
| | | | | | | | | | liangqiao | Forsythiae Fructus |
| | | | | | | | | | jinyinhua | Lonicerae Japonicae Flos |
| | | | | | | | | | huangqi | <i>Hedysarum multijugum</i> Maxim. |
| | | | | | | | | | chaihu | Radix Bupleuri |
| | | | | | 0.403 | 41.88 | 0.24 | 520-18-3 | wumei | Mume Fructus |
| | | | MOL000422 | kaempferol | | | | | qinghao | <i>Artemisia Annua</i> L. |
| | | | | | | | | | liangqiao | Forsythiae Fructus |
| | | | | | | | | | jinyinhua | Lonicerae Japonicae Flos |
| | | | | | | | | | huangqi | <i>Hedysarum Multijugum</i> Maxim. |
| | | | | | | | | | chaihu | Radix Bupleuri |
| | | | | | 0.271 | 36.16 | 0.25 | 491-70-3 | taizhishen | Pseudostellariae Radix |
| | | | MOL000006 | luteolin | | | | | qinghao | <i>Artemisia Annua</i> L. |
| | | | | | | | | | liangqiao | Forsythiae Fructus |
| | | | | | | | | | jinyinhua | Lonicerae Japonicae Flos |
| | | | | | | | | | chaihu | Radix Bupleuri |
| | | | | | 0.119 | 46.06 | 0.66 | PNTWXEIOXBRCP S-JLTQLCOSA-N | qianhu | Peucedani Radix |
| 2 | CASP3 | 54 | MOL004653 | (+)-Anomalin | 0.119 | 51.22 | 0.66 | UFUVJROSOXJGR -MKKZQTCBSA-N | qianhu | Peucedani Radix |
| | | | MOL013078 | praeruptorin E | 0.116 | 46.46 | 0.53 | 73,069-25-7 | qianhu | Peucedani Radix |
| | | | MOL013079 | di-praeruptorin a | 0.181 | 43.83 | 0.76 | 19,716-26-8 | jinyinhua | Lonicerae Japonicae Flos |
| | | | MOL003036 | Stigmasterol Glucoside | 0.097 | 43.09 | 0.78 | XXMJRBRPNZVNU R-UHFFFAOYSA-N | huangqin | Scutellariae Radix |
| | | | MOL002897 | epiberberine | 0.568 | 42.36 | 0.21 | 480-41-1 | wumei | Mume Fructus |
| 3 | STAT3 | 46 | MOL003036 | Stigmasterol Glucoside | 0.181 | 43.83 | 0.76 | 19,716-26-8 | jinyinhua | Lonicerae Japonicae Flos |
| | | | MOL002897 | epiberberine | 0.097 | 43.09 | 0.78 | XXMJRBRPNZVNU R-UHFFFAOYSA-N | huangqin | Scutellariae Radix |
| 4 | ESR1 | 42 | MOL001040 | (2R)-5,7-dihydroxy-2-(4-hydroxyphenyl)chroman-4-one | 0.568 | 42.36 | 0.21 | 480-41-1 | wumei | Mume Fructus |

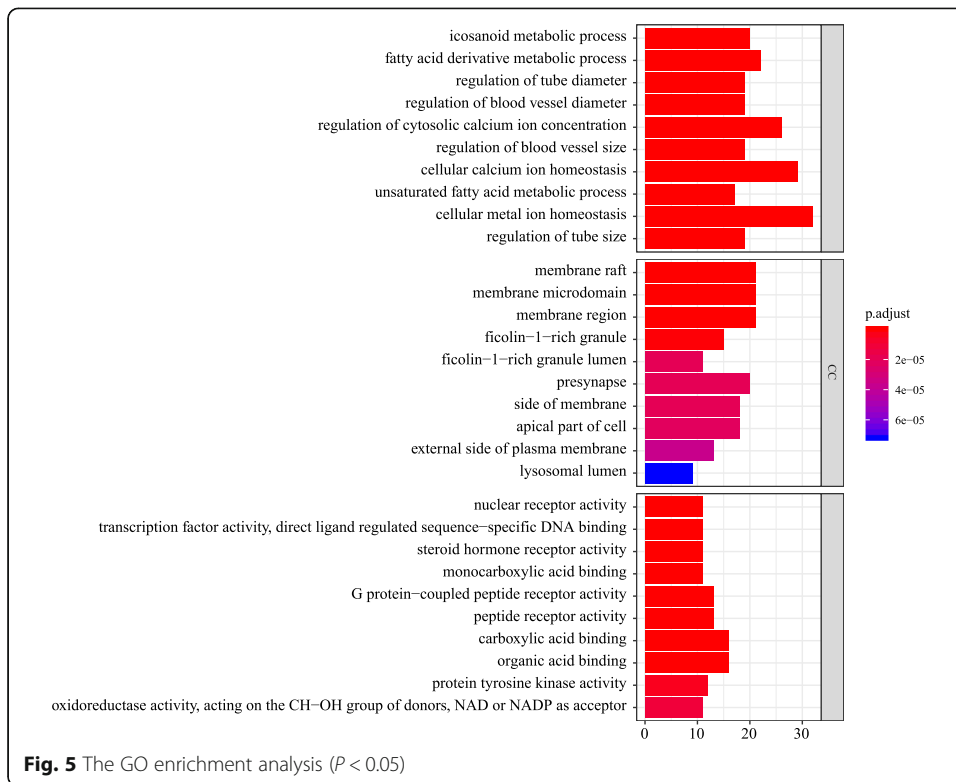
Table 2 Top ten targets and their source composition information (Continued)

| No | Target | Degree | Mol ID | Molecule Name | Probability ^a | OB (%) | DL | CAS/InChIKey | pinyin name | herbal name |
|----|--------|--------|-----------|---|--------------------------|--------|------|---------------------------------|-------------|---|
| | | | MOL000006 | luteolin | 0.394 | 36.16 | 0.25 | 491-70-3 | taizishen | Pseudostellariae Radix |
| | | | | | | | | | qinghao | Artemisia Annua L. |
| | | | | | | | | | liangjiao | Forsythiae Fructus |
| | | | | | | | | | jinyinhua | Lonicerae Japonicae Flos |
| | | | MOL000449 | Stigmasterol | 0.389 | 43.83 | 0.76 | 83-48-7 | shancigu | Pseudobulbus Cremastrae Seu Pleiones |
| | | | | | | | | | jinyinhua | Lonicerae Japonicae Flos |
| | | | | | | | | | huangqin | Scutellariae Radix |
| | | | | | | | | | chaihu | Radix Bupleuri |
| | | | | | | | | | qinghao | Artemisia Annua L. |
| | | | | | | | | | wumei | Mume Fructus |
| 5 | FPR2 | 34 | MOL013098 | [(9R)-8-dimethyl-2-oxo-9- 10-dihydropyrano[6,5-h]chromen- 9-yl](Z)-2-methylbut-2-enoate | 0.105 | 87.48 | 0.37 | 19427-82-8 | qianhu | Peucedani Radix |
| 6 | F2 | 34 | MOL000371 | 3,9-di-Omethylinissolin | 0.102 | 53.74 | 0.48 | RFNFQZKHNKOP O-BBRMVZONSAN | huangqi | Hedysarum Multijugum Maxim. |
| 7 | BCL2L1 | 34 | MOL001749 | ZINC03860434 | 0.116 | 43.59 | 0.35 | BJQHLKABXJVA M-WOJBXXKFSAN | chuanbeimu | Fritillariae Irrihosae Bulbus |
| | | | MOL001490 | bis[(2S)-2-ethylhexyl] benzene- 1,2-dicarboxylate | 0.116 | 43.59 | 0.35 | 117-81-7 | huangqin | Scutellariae Radix |
| | | | MOL013103 | Umbelliprenin | 0.113 | 46.57 | 0.44 | 532-16-1 | qianhu | Peucedani Radix |
| 8 | BDKRB2 | 31 | MOL013098 | [(9R)-8-dimethyl-2-oxo-9,10- dihydropyrano[6,5-h]chromen-9- yl](Z)-2-methylbut-2-enoate | 0.105 | 87.48 | 0.37 | 19427-82-8 | qianhu | Peucedani Radix |
| | | | MOL000371 | 3,9-di-Omethylinissolin | 0.102 | 53.74 | 0.48 | RFNFQZKHNKOP O-BBRMVZONSAN | huangqi | Hedysarum Multijugum Maxim. |
| | | | MOL000398 | isoflavanone | 0.049 | 109.99 | 0.3 | JNSVNRWHSLLCB G-LLVKOONJSA-N | huangqi | Hedysarum Multijugum Maxim. |
| 9 | MPO | 31 | MOL000098 | quercetin | 1.000 | 46.43 | 0.28 | 117-39-5 | wumei | Mume Fructus |
| | | | | | | | | | qinghao | Artemisia Annua L. |

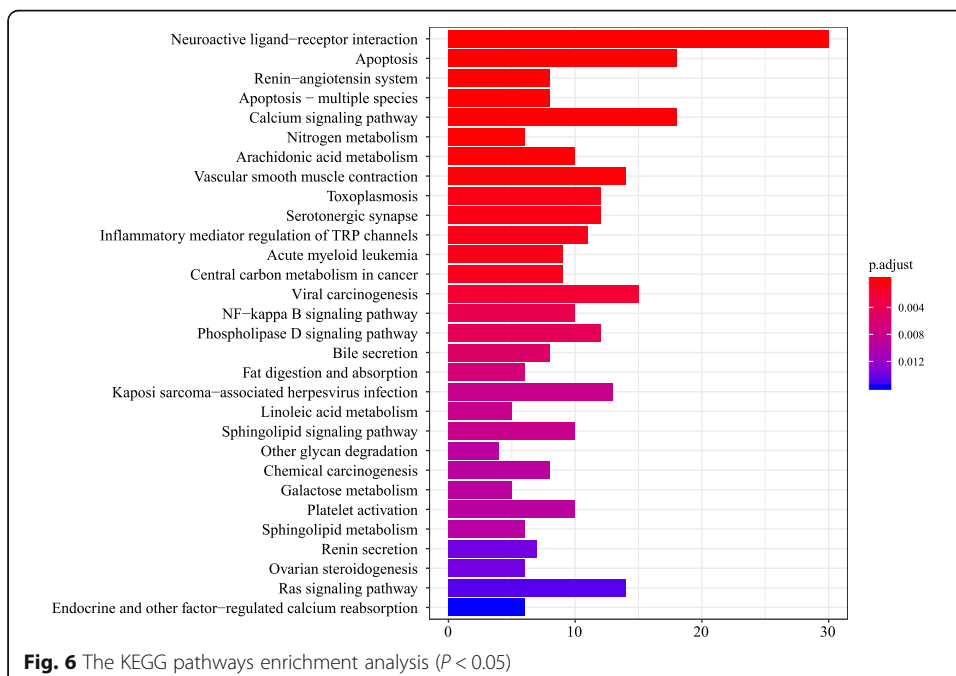
Table 2 Top ten targets and their source composition information (Continued)

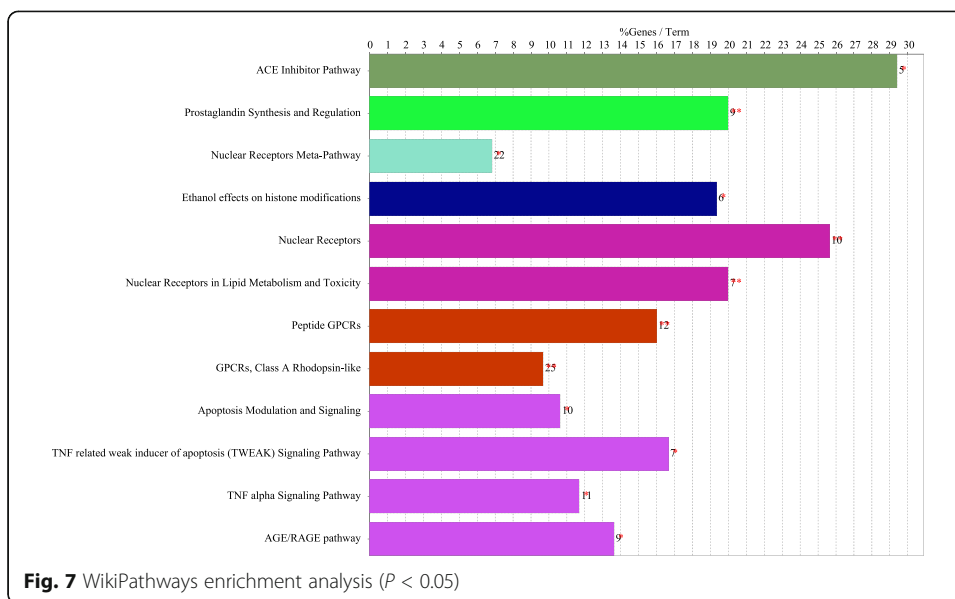
| No | Target | Degree | Mol ID | Molecule Name | Probability ^a | OB (%) | DL | CAS/InChIKey | pinyin name | herbal name |
|----|--------|--------|-----------|--|--------------------------|--------|------|---------------------------------|--|---|
| | | | MOL000422 | kaempferol | 0.394 | 41.88 | 0.24 | 520-18-3 | qianhu liangqiao jinyinhua huangqi chaihu wumei qinghao liangqiao jinyinhua huangqi | Peucedani Radix Forsythiae Fructus Lonicerae Japonicae Flos Hedysarum Multijugum Maxim. Radix Bupleuri Mume Fructus Artemisia Annua L. Forsythiae Fructus Lonicerae Japonicae Flos Hedysarum Multijugum Maxim. Radix Bupleuri |
| | | | MOL000006 | luteolin | 0.271 | 36.16 | 0.25 | 491-70-3 | chaitu taizishen qinghao liangqiao jinyinhua fuling | Pseudostellariae Radix Artemisia Annua L. Forsythiae Fructus Lonicerae Japonicae Flos Hedysarum Multijugum Maxim. Radix Bupleuri |
| 10 | ACE | 30 | MOL000273 | (2R)-2-[(3S,5R,10S,13R,14R,16R,17R)-3,16-dihydroxy-4,10,13,14-pentamethyl-2,3,5,6,12,15,16,17-octahydro-1H-cyclopentalalphenanthren-17-yl]-6-methylhept-5-enoic acid | 0.120 | 30.93 | 0.81 | XSLKAKROJMKMHI T-WIUKAADNSAN | fuling | Poria Cocos(Schw.) Wolf. |
| | | | MOL002308 | Indicaxanthin | 0.112 | 31.79 | 0.22 | KYJMYEJUHZA X-QOBSUCFISAN | daqingye | Isatisidis Folium |
| | | | MOL000280 | (2R)-2-[(3S,5R,10S,13R,14R,16R,17R)-3,16-dihydroxy-4,10,13,14-pentamethyl-2,3,5,6,12,15,16,17-octahydro-1H-cyclopentalalphenanthren-17-yl]-5-isopropylhex-5-enoic acid | 0.111 | 31.07 | 0.82 | 6754-16-1 | fuling | Poria Cocos(Schw.) Wolf. |

^aProbability value from Swiss TargetPrediction for the query molecule to have this protein as target



enriched WikiPathways ($P < 0.05$) were involved in ACE Inhibitor pathway, prostaglandin synthesis and regulation, nuclear receptors meta-pathway, ethanol effects on histone modifications, nuclear receptors in lipid metabolism and toxicity, apoptosis modulation and signaling, TNF related weak inducer of apoptosis (TWEAK) signaling pathway, TNF alpha signaling pathway, and AGE/RAGE pathway. Furthermore, based





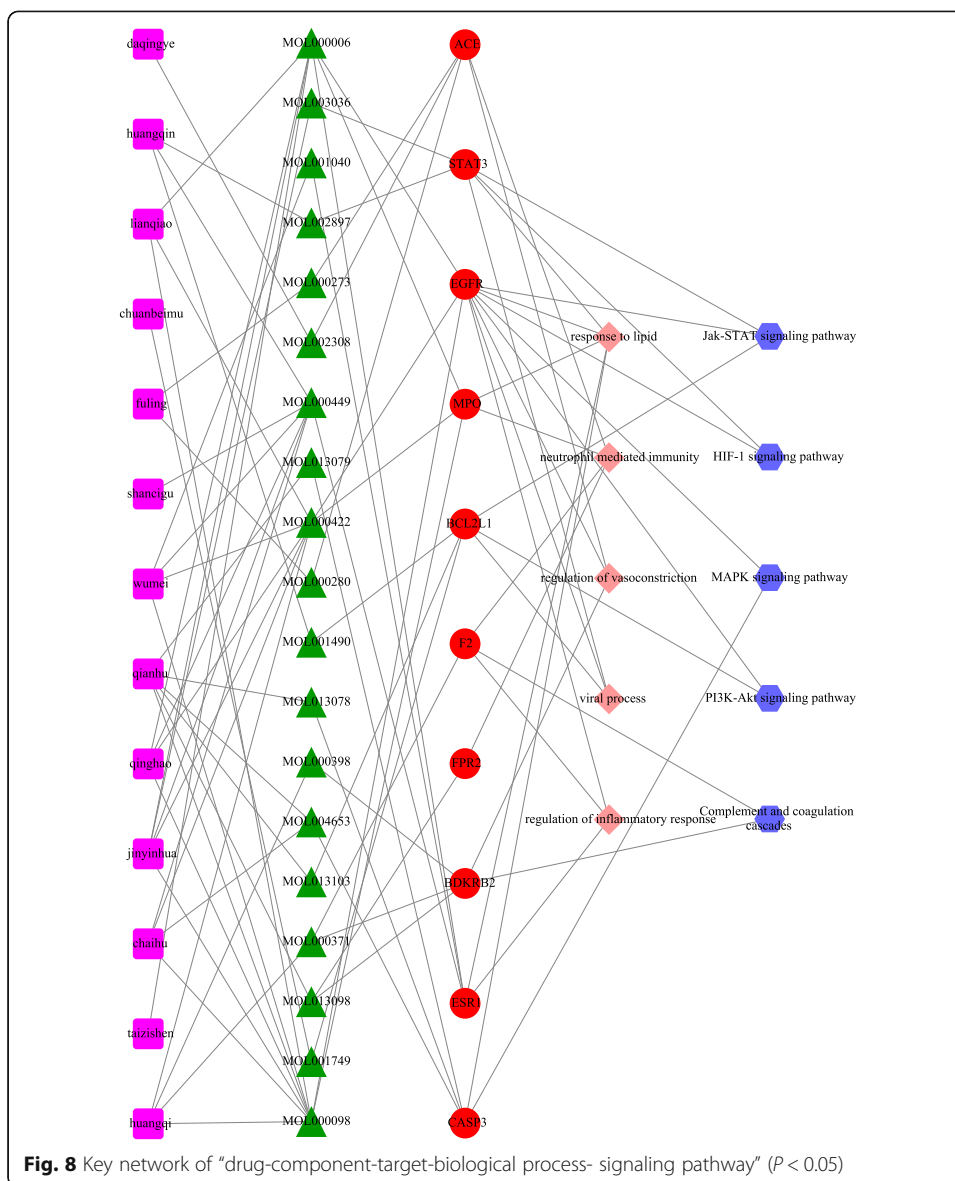
on the above comprehensive analysis, the key network of “drug-component-target-biological process- signaling pathway” was presented (Fig. 8).

Discussion

Clinical research shows that integrative therapy by combining TCM and western medicine for COVID-19 is significantly better at improving the symptoms and shortening the average length of hospital stay in COVID-19 patients than treatments using western medicine [25]. Early intervention using TCM reduces the transition from moderate to severe and critical COVID-19. The treatment strategy of TCM focuses on eliminating fever and expelling pathogenic factors, especially in relieving endogenous stagnated heat in the body to thus eliminate the pathogens [26]. Considering the characteristics of humid and hot climate of Lingnan area, supplementation with heat and damp-clearing agents is needed. In addition, TCM classic, *Huangdi 81 Nanjing* claims that “individuals with lung injuries are benefited from *qi* tonification.” Tonifying *qi* and nourishing *yin* in the early stage of COVID-19 to balance and maintain the vital energy is necessary.

Thus, the TJQW were formulated, with an emphasis on using heat-clearing and detoxifying herbs, such as *Forsythiae Fructus*, edible tulip, *Lonicera japonica*, *Radix Scutellariae baicalensis*, and *Folium Isatidis*, combined with *Fritillaria cirrhosa* and *Fritillaria thunbergii* for heat-clearing and phlegm removal, *Poria cocos* for spleen invigoration and dampness elimination, radix *Scrophulariae* for yin nourishment, and *Fructus Mume* as an astringent herb for stabilizing *qi* in the lungs. Combination of the two herbs was used to calm the body parts that were not affected by the pathogenic factors. *Radix Pseudostellariae* and *Astragalus propinquus* were used to balance and maintain the vital energy and eliminate the pathogenic factors. Supplementation of herbs against cold-nature and to maintain the vital energy also relieved the *qi* and *yin* deficiency in the later stage of the disease [3].

Prescription of TJQW has been incorporated into the TCM treatment program for COVID-19 in Guangdong Province [18]. At this time, no high-quality evidence



supports the clinical efficacy of TJQW treatment against COVID-19. However, we can perform bioinformatics analysis as a theoretical evaluation of potential anti-COVID-19 molecular mechanisms of TJQW to support future clinical trials. This study used network pharmacology to explore the potential mechanism of COVID-19 treatment with TJQW, and the findings are summarized below.

Active ingredients

In the analysis of TJQW-active ingredient-potential target network, the active ingredients, including umbelliprenin, quercetin, kaempferol, luteolin, praeruptorin E, stigmasterol, and oroxylin A, have potential anti-inflammatory and antiviral effects.

Biological activities including anti-inflammatory, antioxidant, and antileishmanial activities have been reported from umbelliprenin (UMB) [27], UMB induced T(H)2 IL-4

and suppressed T(H)1 IFN γ secretion, and significantly suppressed LPS-induced production of NO and PGE (2) apparently and also led to reductions in inducible iNOS and COX-expression [28], showing anti-inflammatory and immunomodulatory properties in vitro [29]. Numerous studies on quercetin and quercetin glycosides have been proved that these compounds acts as potent drug candidate towards inhibition of influenza virus infection through inferring the viral cellular immune system, inhibition of viral cellular targets, interfering the viral replication and inhibiting the viral growth phases [30]. Kaempferol suppressed the expression of proinflammatory cytokine interleukin-6 and chemokines interleukin-8, monocyte chemoattractant protein-1, and regulated on activation, normal T-cell expressed and secreted [31]. Luteolin inhibits LPS-induced inflammatory responses through modulation of NF- κ B/AP-1/PI3K-Akt signaling cascades [32]. Luteolin decreased the expression of core genes at protein and mRNA levels (MMP9, MAPK1, HSP90AA1, EGFR, SRC and HRAS), and PI3K-AKT signaling pathway, Ras signaling pathway might be the critical pathways of luteolin against inflammation [33]. In silico, in vitro, in vivo, and clinical studies strongly suggest that the major pharmacological mechanism of luteolin is its anti-inflammatory activity, which derives from its regulation of transcription factors such as STAT3 and NF- κ B [34]. Praeruptorin E suppressed lipopolysaccharide (LPS) induced Nuclear Factor-kappa B (NF- κ B) pathway activation in the lung by decreasing the cytoplasmic loss of Inhibitor κ B- α (I κ B- α) protein and inhibiting the translocation of p65 from the cytoplasm to nucleus, which might be useful in the therapy of lung injury [35]. Treatment of stigmasterol significantly suppresses the expression of proinflammatory mediators (TNF- α , IL-6, IL-1 β , iNOS and COX-2) and increases the expression of anti-inflammatory cytokine (IL-10) [36]. Based on an in silico approach, an extensive molecular docking investigation of the phytochemicals at the active binding pockets of the viral proteins revealed the promising inhibitory potential on COVID-19 of stigmasterol [37]. Vascular dysfunction plays a critical role in the pathogenesis of sepsis in COVID-19. Oroxylin A regulates the vascular tone by inhibiting vascular hyporeactivity caused by NO overproduction and reverses the endothelial barrier dysfunction and inflammation by inhibiting the IRAK-4-mediated IKK α / β phosphorylation, suggesting that Oroxylin A administration as a potentially useful therapeutic approach for clinical interventions in septic shock [38].

Potential therapeutic mechanism

In terms of potential therapeutic targets, based on this study, all potential biotargets of TJQW action against COVID-19 were methodically obtained, and the potential core pharmacological targets were identified as EGFR, CASP3, STAT3, ESR1, FPR2, F2, BCL2L1, BDKRB2, MPO, and ACE, majority of them have been confirmed to be involved in pulmonary inflammation. EGFR-mediated signaling pathways are important for cell proliferation, differentiation, and survival in many tissues and cell types. Suppression of the EGFR pathway via the reduction of eIF2 α phosphorylation increases susceptibility to cellular oxidative stress [39]. The parameters including homocysteine, urea, creatinine and serum cystatin C were significantly higher in imaging progression patients, while EGFR decreased [40]. This suggested that low levels of EGFR may be a protective factor for COVID-19. Caspases are a family of enzymes that play a key role

in apoptosis. They are mediators and performers of apoptosis. When CASP8 interacts with CASP3, CASP3 is activated and it further induces apoptosis [41]. *Radix Scutellariae baicalensis* inhibits the apoptosis induced by influenza virus infection through regulating the expression of apoptosis-related genes in the CASP8-mediated exogenous pathway and the pathways of endoplasmic reticulum [42]. ACE2, a SARS-CoV-2 receptor, is upregulated by interleukin-6 via STAT3 signaling in synovial tissues [43]. In the later phase of COVID-19, the potential dysregulation of the AngII-AT1R pathway downstream of ACE2 could lead to cytokine release syndrome, and the IL-6-STAT3 axis may be the required targeting of cytokine pathways in the treatment in COVID-19 patients [44]. STAT3 and BCL2L1 are considered to have protein interactions with SARS-CoV [45]. A critical imbalance in RAS represented by decreased expression of ACE in combination with increases in ACE2 and both bradykinin receptors (BDKRB1, BDKRB2) [46].

Estrogen receptor decreases NF-kappa-B DNA-binding activity and inhibits NF-kappa-B-mediated transcription from the IL6 promoter [47]. The human FPR2, which is powerful neutrophils chemotactic factor, plays a crucial role in host defense and inflammation, and has been considered as a drug target for chronic inflammatory diseases [48]. Accumulating evidence demonstrates that FPR2 are critical mediators of myeloid cell trafficking in the sequential chemotaxis signal relays in microbial infection, inflammation, and immune responses [49]. As receptor, FPR2 displays potent mucosal protection and promotes catabasis after acute lung injury (ALI) [50], providing a novel therapeutic target to develop an effective treatment against ALI progression. F2 has functions in blood homeostasis, inflammation and wound healing [51]. Antiapoptotic proteins Bcl-2 and Bcl-X(L) may attenuate inflammation impairing NLRP1-inflammasome activation, reducing caspase-1 activation and interleukin-1beta (IL-1beta) production [52]. The increasing expression of Bcl-2 and Bcl-xL inhibits autophagy and bacterial killing in human macrophages, suggesting that targeting the Bcl-2/Bcl-xL-Beclin 1 interaction may improving resistance to infection in patients with advanced lung disease [53]. MPO is a part of the host defense system of polymorphonuclear leukocytes, and it is responsible for microbicidal activity against a wide range of organisms [54, 55].

The bioinformatics findings of this study suggested that the specific inactivation of signaling pathways of inflammatory stress and targeted modulation of intrapulmonary EGFR, CASP3, STAT3, ESR1, FPR2, F2, BCL2L1, BDKRB2, MPO, and ACE expressions, maybe potential pathways of TJQW action against COVID-19. These are the potential therapeutic mechanisms of active ingredients in TJQW and targets in the treatment of COVID-19. This study provided new predictions on the molecular-based therapeutic mechanism of TJQW in COVID-19 treatment and it offered new ideas for further study.

According to the results of GO enrichment analysis, the potential therapeutic targets of TJQW were mainly distributed in the regulation of inflammatory response, viral process, neutrophil mediated immunity, response to lipid, regulation of vasoconstriction, and were also involved in promotion/suppression of inflammation, cell division, abnormal cell proliferation, oxidation/reduction reactions, ion-channel regulation, and immune regulation. From the results of KEGG enrichment analysis, the pathways mainly involved in the treatment process, PI3K-Akt signaling pathway, MAPK signaling pathway, Jak-STAT signaling pathway, Complement and coagulation cascades, and HIF-1 signaling pathway, were important inflammatory or metabolism pathways.

This study implicated MAPK signaling as part of the potential mechanisms underlying TJQW antiviral function. MAPK cascades are crucial signaling pathways in the regulation of host immune response to infection [56]. MAPK phosphatase 1 regulates pro- and anti-inflammatory cytokines dynamically in innate immune responses [57]. Studies suggest that quercetin possesses antioxidant, anti-inflammatory, and antiplatelet properties, Quercetin-mediated antiplatelet activity involves PI3K/Akt inactivation, suppresses MAPK phosphorylations. Quercetin may have the potential to treat diseases involving aberrant platelet activation and inflammation [58]. Patients with severe COVID-19 might experience cytokine release syndrome, and several of the cytokines involved in COVID-19 employ a distinct intracellular signaling pathway mediated by Janus kinases (JAKs), therefore, JAK inhibition presents an attractive therapeutic strategy for CRS [59]. What's more, it has been reported that complement and HIF-1 are potential pathways of damage and targets for the cure of COVID-19-driven endothelial damage [60].

Besides, the arachidonic acid metabolic pathway is mainly used for the synthesis of inflammatory mediators, which mediate the production of various inflammatory cytokines, such as monocyte chemoattractant protein-1, TNF, IL, and interferon, which are related to the development, progression, and regression of inflammation [61]. In addition to regulating blood pressure and water and sodium balance in the renin-angiotensin system (RAS), a study has shown that the system is also involved in regulating various physiological activities, such as inflammation, immunity, apoptosis, and tissue repair in the body. In addition, it is also involved in the development and progression of lung diseases, such as pulmonary arterial hypertension, pulmonary fibrosis, and pulmonary thromboembolism, especially acute lung injury [62].

These findings suggested that TJQW may exert antioxidative, antiviral, and anti-inflammatory effects, along with immune system activation effects through the aforementioned targets, thereby playing an early protective role in the lungs. While in the TCM treatment program for COVID-19 in Guangdong Province, TJQW was mainly used for the treatment of COVID-19 in the early stage.

In summary, this study used network pharmacology techniques and approach to retrieve the active ingredients and potential targets of TCM in TJQW, by screening the common targets for TCM active ingredients and diseases and performing GO and KEGG enrichment analyses of the targets. As shown in the key network of "drug-component-target-biological process- signaling pathway", we speculated that the active ingredients in the TJQW treatment of COVID-19 may be umbelliprenin, quercetin, luteolin, praeruptorin E, stigmasterol. The potential effector targets were EGFR, CASP3, STAT3, ESR1, FPR2, F2, BCL2L1, BDKRB2, MPO, and ACE, through regulating signaling pathways, such as PI3K-Akt signaling pathway, MAPK signaling pathway, Jak-STAT signaling pathway, to suppress the inflammatory response, regulate the immune function, and reduce lung injury, thereby achieving the purpose of COVID-19 treatment.

From the perspective of research limitations. Since the gene co-expressed with ACE2 was selected as a potential therapeutic target for COVID-19 in this study, it is not a specific therapeutic target and needs further study.

Conclusion

Our study uncovered the pharmacological mechanism underlying TJQW treatment for COVID-19. These results should benefit efforts for people around the world to gain

more knowledge about Chinese medicine TJQW in the treatment of this vicious epidemic COVID-19, and help to address this pressing problem currently facing the world.

Supplementary information

Supplementary information accompanies this paper at <https://doi.org/10.1186/s13040-020-00225-8>.

Additional file 1.

Acknowledgements

We thank LetPub for its linguistic assistance during the preparation of this manuscript.

Authors' contributions

Ji-qiang Li and Xing-hua Tan conceived and designed the research; Yong-shi Ni, Mian-sha Li and Jian-kun Chen collected and analyzed the data; Xiao-hong Liu provided useful suggestions on the methodology; Ying Huang and Wen-jiang Zheng wrote the paper. All authors read and approved the final version of the manuscript. Ying Huang and Wen-jiang Zheng contributed equally to this work and should be regarded as co-first authors.

Funding

This work is supported by Department of Science and Technology of Guangdong Province (No. 2020B111110001), COVID-19 Epidemic Prevention and Control Special Research Fund of the Department of Education of Guangdong Province (No. 2020KZDZX1054), Student Learning Team Incubation Project of the First Affiliated Hospital of Guangzhou University of Traditional Chinese Medicine (No.2018XXTD003) and 2020 National College Student Innovation and Entrepreneurship Training Program of Guangzhou University of Chinese Medicine (No. 202010572001).

Availability of data and materials

All data are available in the manuscript and they are showed in tables, figures.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no conflict of interests.

Author details

¹First College of Clinical Medicine, Guangzhou University of Chinese Medicine, Guangzhou, China. ²Integrative Dept.3 (Geriatrics Dept), Guangdong Provincial Hospital of Chinese Medicine, Guangzhou, China. ³The Second College of Clinical Medicine, Guangzhou University of Chinese Medicine, Guangzhou, China. ⁴Tianhe Shadong Street Community Healthcare Service Center, Guangzhou, China. ⁵Department of Traditional Chinese Medicine, Guangzhou Eighth People's Hospital, Guangzhou, China.

Received: 15 May 2020 Accepted: 8 September 2020

Published online: 24 September 2020

References

1. Wang C, Horby PW, Hayden FG, Gao GF. A novel coronavirus outbreak of global health concern. *Lancet*. 2020; 395(10223):470–3. [https://doi.org/10.1016/S0140-6736\(20\)30185-9](https://doi.org/10.1016/S0140-6736(20)30185-9).
2. Song P, Zhao L, Li X, Su J, Jiang Z, Song B, et al. Interpretation of the Traditional Chinese Medicine portion of the diagnosis and treatment protocol for corona virus disease 2019 (Trial Version 7). *J Tradit Chin Med*. 2020;40(3):497–508. <https://doi.org/10.19852/j.cnki.jtcm.2020.03.019>.
3. Xiaoxia F, Luping L, Xinghua T. Clinical Observation on Effect of Toujie Quwen Granules in Treatment of COVID-19. *Chin J Exp Tradit Med Formul*. 2020;26(12):44–8. <https://doi.org/10.13422/j.cnki.syfjx.20201314>.
4. Xiaoxia F, Luping L, Xinghua T. 2 cases reports of Corona Virus Disease 2019 treated by ToujieQuwen granules. *J Jinan Univ (Natural Science & Medicine Edition)*. 2020;41(02):151–6. <https://doi.org/10.11778/j.jdx.2020.02.009>.
5. Hopkins AL. Network pharmacology: the next paradigm in drug discovery. *Nat Chem Biol*. 2008;4(11):682–90. <https://doi.org/10.1038/nchembio.118>.
6. Li S, Zhang B. Traditional Chinese medicine network pharmacology: theory, methodology and application. *Chin J Nat Med*. 2013;11(2):110–20. [https://doi.org/10.1016/S1875-5364\(13\)60037-0](https://doi.org/10.1016/S1875-5364(13)60037-0).
7. Hoffmann M, Kleine-Weber H, Schroeder S, Kruger N, Herrler T, Erichsen S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell*. 2020;181(2):271–80 e8. <https://doi.org/10.1016/j.cell.2020.02.052>.
8. Huang YF, Bai C, He F, Xie Y, Zhou H. Review on the potential action mechanisms of Chinese medicines in treating coronavirus disease 2019 (COVID-19). *Pharmacol Res*. 2020;158:104939. <https://doi.org/10.1016/j.phrs.2020.104939>.

9. Datta PK, Liu F, Fischer T, Rappaport J, Qin X. SARS-CoV-2 pandemic and research gaps: understanding SARS-CoV-2 interaction with the ACE2 receptor and implications for therapy. *Theranostics*. 2020;10(16):7448–64. <https://doi.org/10.7150/thno.48076>.
10. Cava C, Bertoli G, Castiglioni I. In Silico Discovery of Candidate Drugs against Covid-19. *Viruses*. 2020;12(4). <https://doi.org/10.3390/v12040404>.
11. Ru J, Li P, Wang J, Zhou W, Li B, Huang C, et al. TCMSP: a database of systems pharmacology for drug discovery from herbal medicines. *J Cheminform*. 2014;6:13. <https://doi.org/10.1186/1758-2946-6-13>.
12. Alqahtani S. In silico ADME-Tox modeling: progress and prospects. *Expert Opin Drug Metab Toxicol*. 2017;13(11):1147–58. <https://doi.org/10.1080/17425255.2017.1389897>.
13. Khakar PS. Two-dimensional (2D) in silico models for absorption, distribution, metabolism, excretion and toxicity (ADME/T) in drug discovery. *Curr Top Med Chem*. 2010;10(1):116–26. <https://doi.org/10.2174/156802610790232224>.
14. Daina A, Michielin O, Zoete V. SwissTargetPrediction: updated data and new features for efficient prediction of protein targets of small molecules. *Nucleic Acids Res*. 2019;47(W1):W357–W64. <https://doi.org/10.1093/nar/gkz382>.
15. Wang J, Zhao S, Liu M, Zhao Z, Xu Y, Wang P, et al. ACE2 expression by colonic epithelial cells is associated with viral infection, immunity and energy metabolism. *medRxiv*. 2020:2020.02.05.20020545. <https://doi.org/10.1101/2020.02.05.20020545>.
16. Su G, Morris JH, Demchak B, Bader GD. Biological network exploration with Cytoscape 3. *Curr Protoc Bioinformatics*. 2014;47(8):13–24. <https://doi.org/10.1002/0471250953.bi0813s47>.
17. Szklarczyk D, Morris JH, Cook H, Kuhn M, Wyder S, Simonovic M, et al. The STRING database in 2017: quality-controlled protein-protein association networks, made broadly accessible. *Nucleic Acids Res*. 2017;45(D1):D362–D8. <https://doi.org/10.1093/nar/gkw937>.
18. Chin CH, Chen SH, Wu HH, Ho CW, Ko MT, Lin CY, et al. *BMC Syst Biol*. 2014;8(Suppl 4):S11. <https://doi.org/10.1186/1752-0509-8-S4-S11>.
19. Zhang C, Zheng Y, Li X, Hu X, Qi F, Luo J. Genome-wide mutation profiling and related risk signature for prognosis of papillary renal cell carcinoma. *Ann Transl Med*. 2019;7(18):427. <https://doi.org/10.21037/atm.2019.08.113>.
20. Yu G, Wang LG, Han Y, He QY. clusterProfiler: an R package for comparing biological themes among gene clusters. *OMICS*. 2012;16(5):284–7. <https://doi.org/10.1089/omi.2011.0118>.
21. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, et al. Gene ontology: tool for the unification of biology. *Gene Ontol Consortium Nat Genet*. 2000;25(1):25–9. <https://doi.org/10.1038/75556>.
22. Kanehisa M, Goto S, Kawashima S, Okuno Y, Hattori M. The KEGG resource for deciphering the genome. *Nucleic Acids Res*. 2004;32(Database issue):D277–80. <https://doi.org/10.1093/nar/gkh063>.
23. Bindea G, Mlecnik B, Hackl H, Charoentong P, Tosolini M, Kirilovsky A, et al. ClueGO: a Cytoscape plug-in to decipher functionally grouped gene ontology and pathway annotation networks. *Bioinformatics*. 2009;25(8):1091–3. <https://doi.org/10.1093/bioinformatics/btp101>.
24. Bindea G, Galon J, Mlecnik B. CluePedia Cytoscape plugin: pathway insights using integrated experimental and in silico data. *Bioinformatics*. 2013;29(5):661–3. <https://doi.org/10.1093/bioinformatics/btt019>.
25. Liu M, Gao Y, Yuan Y, Yang K, Shi S, Zhang J, et al. Efficacy and safety of integrated traditional Chinese and Western medicine for Corona virus disease 2019 (COVID-19): a systematic review and meta-analysis. *Pharmacol Res*. 2020;158:104896. <https://doi.org/10.1016/j.phrs.2020.104896>.
26. Zhao ZH, Zhou Y, Li WH, Huang QS, Tang ZH, Li H. Analysis of traditional Chinese medicine diagnosis and treatment strategies for COVID-19 based on "the diagnosis and treatment program for coronavirus Disease-2019" from Chinese authority. *Am J Chin Med*. 2020;48(5):1035–49. <https://doi.org/10.1142/S0192415X20050050>.
27. Shakeri A, Iranshahy M, Iranshahi M. Biological properties and molecular targets of umbelliprenin—a mini-review. *J Asian Nat Prod Res*. 2014;16(8):884–9. <https://doi.org/10.1080/10286020.2014.917630>.
28. Zamani Taghizadeh Rabe S, Iranshahi M, Mahmoudi M. In vitro anti-inflammatory and immunomodulatory properties of umbelliprenin and methyl galbanate. *J Immunotoxicol*. 2016;13(2):209–16. <https://doi.org/10.3109/1547691X.2015.1043606>.
29. Khaghanzadeh N, Samiei A, Mojtahedi Z, Ramezani M, Hosseinzadeh M, Ghaderi A. Umbelliprenin induced both anti-inflammatory and regulatory cytokines in C57/BL6 mice. *Iran J Basic Med Sci*. 2017;20(7):829–34. <https://doi.org/10.22038/IJBMS.2017.9021>.
30. Gansukh E, Nile A, Kim DH, Oh JW, Nile SH. New insights into antiviral and cytotoxic potential of quercetin and its derivatives - a biochemical perspective. *Food Chem*. 2020;334:127508. <https://doi.org/10.1016/j.foodchem.2020.127508>.
31. Devi KP, Malar DS, Nabavi SF, Sureda A, Xiao J, Nabavi SM, et al. Kaempferol and inflammation: from chemistry to medicine. *Pharmacol Res*. 2015;99:1–10. <https://doi.org/10.1016/j.phrs.2015.05.002>.
32. Park CM, Song YS. Luteolin and luteolin-7-O-glucoside inhibit lipopolysaccharide-induced inflammatory responses through modulation of NF-kappaB/AP-1/PI3K-Akt signaling cascades in RAW 264.7 cells. *Nutr Res Pract*. 2013;7(6):423–9. <https://doi.org/10.4162/nrp.2013.7.6.423>.
33. Huang XF, Zhang JL, Huang DP, Huang AS, Huang HT, Liu Q, et al. A network pharmacology strategy to investigate the anti-inflammatory mechanism of luteolin combined with in vitro transcriptomics and proteomics. *Int Immunopharmacol*. 2020;86:106727. <https://doi.org/10.1016/j.intimp.2020.106727>.
34. Aziz N, Kim MY, Cho JY. Anti-inflammatory effects of luteolin: a review of in vitro, in vivo, and in silico studies. *J Ethnopharmacol*. 2018;225:342–58. <https://doi.org/10.1016/j.jep.2018.05.019>.
35. Yu PJ, Li JR, Zhu ZG, Kong HY, Jin H, Zhang JY, et al. Preruptorin D and E attenuate lipopolysaccharide/hydrochloric acid induced acute lung injury in mice. *Eur J Pharmacol*. 2013;710(1–3):39–48. <https://doi.org/10.1016/j.ejphar.2013.03.050>.
36. Ahmad Khan M, Sarwar A, Rahat R, Ahmed RS, Umar S. Stigmasterol protects rats from collagen induced arthritis by inhibiting proinflammatory cytokines. *Int Immunopharmacol*. 2020;85:106642. <https://doi.org/10.1016/j.intimp.2020.106642>.
37. Kar P, Sharma NR, Singh B, Sen A, Roy A. Natural compounds from *Clerodendrum* spp. as possible therapeutic candidates against SARS-CoV-2: An in silico investigation. *J Biomol Struct Dyn*. 2020:1–12. <https://doi.org/10.1080/07391102.2020.1780947>.

38. Tseng TL, Chen MF, Hsu YH, Lee TJF. OroxylinA reverses lipopolysaccharide-induced adhesion molecule expression and endothelial barrier disruption in the rat aorta. *Toxicol Appl Pharmacol*. 2020;400:115070. <https://doi.org/10.1016/j.taap.2020.115070>.
39. Kim MJ, Choi WG, Ahn KJ, Chae IG, Yu R, Back SH. Reduced EGFR Level in eIF2alpha PhosphorylationDeficient Hepatocytes Is Responsible for Susceptibility to Oxidative Stress. *Mol Cells*. 2020;43(3):264–75. <https://doi.org/10.14348/molcells.2020.2197>.
40. Yang Z, Shi J, He Z, Lu Y, Xu Q, Ye C, et al. Predictors for imaging progression on chest CT from coronavirus disease 2019 (COVID-19) patients. *Aging (Albany NY)*. 2020;12(7):6037–48. <https://doi.org/10.18632/aging.102999>.
41. Brasseur K, Auger P, Asselin E, Parent S, Sirois M. Parasporin-2 from a new bacillus thuringiensis 4R2 strain induces Caspases activation and apoptosis in human Cancer cells. *PLoS One*. 2015;10(8):e0135106. <https://doi.org/10.1371/journal.pone.0135106>.
42. Li R, Wang L. Baicalin inhibits influenza virus a replication via activation of type I IFN signaling by reducing miR146a. *Mol Med Rep*. 2019;20(6):5041–9. <https://doi.org/10.3892/mmr.2019.10743>.
43. Mokuda S, Tokunaga T, Masumoto J, Sugiyama E. Angiotensin-converting enzyme 2, a SARS-CoV-2 receptor, is upregulated by interleukin-6 via STAT3 signaling in synovial tissues. *J Rheumatol*. 2020. <https://doi.org/10.3899/jrheum.200547>.
44. Hirano T, Murakami M. COVID-19: a new virus, but a familiar receptor and cytokine release syndrome. *Immunity*. 2020;52(5):731–3. <https://doi.org/10.1016/j.immuni.2020.04.003>.
45. Vavougiou GD. A data-driven hypothesis on the epigenetic dysregulation of host metabolism by SARS coronavirus infection: potential implications for the SARS-CoV-2 modus operandi. *Med Hypotheses*. 2020;140:109759. <https://doi.org/10.1016/j.mehy.2020.109759>.
46. Garvin MR, Alvarez C, Miller JJ, Prates ET, Walker AM, Amos BK, et al. A mechanistic model and therapeutic interventions for COVID-19 involving a RAS-mediated bradykinin storm. *Elife*. 2020;9. <https://doi.org/10.7554/eLife.59177>.
47. Yamaguchi N, Nakayama Y, Yamaguchi N. Down-regulation of Forkhead box protein A1 (FOXA1) leads to cancer stem cell-like properties in tamoxifen-resistant breast cancer cells through induction of interleukin-6. *J Biol Chem*. 2017;292(20):8136–48. <https://doi.org/10.1074/jbc.M116.763276>.
48. Chen T, Xiong M, Zong X, Ge Y, Zhang H, Wang M, et al. Structural basis of ligand binding modes at the human formyl peptide receptor 2. *Nat Commun*. 2020;11(1):1208. <https://doi.org/10.1038/s41467-020-15009-1>.
49. Li L, Chen K, Xiang Y, Yoshimura T, Su S, Zhu J, et al. New development in studies of formyl-peptide receptors: critical roles in host defense. *J Leukoc Biol*. 2016;99(3):425–35. <https://doi.org/10.1189/jlb.2RI0815-354RR>.
50. Eickmeier O, Seki H, Haworth O, Hilberath JN, Gao F, Uddin M, et al. Aspirin-triggered resolvin D1 reduces mucosal inflammation and promotes resolution in a murine model of acute lung injury. *Mucosal Immunol*. 2013;6(2):256–66. <https://doi.org/10.1038/mi.2012.66>.
51. Madoiwa S, Nakamura Y, Mimuro J, Furusawa S, Koyama T, Sugo T, et al. Autoantibody against prothrombin aberrantly alters the proenzyme to facilitate formation of a complex with its physiological inhibitor antithrombin III without thrombin conversion. *Blood*. 2001;97(12):3783–9. <https://doi.org/10.1182/blood.v97.12.3783>.
52. Bruey JM, Bruey-Sedano N, Luciano F, Zhai D, Balpai R, Xu C, et al. Bcl-2 and Bcl-XL regulate proinflammatory caspase-1 activation by interaction with NALP1. *Cell*. 2007;129(1):45–56. <https://doi.org/10.1016/j.cell.2007.01.045>.
53. Casalino-Matsuda SM, Nair A, Beitel GJ, Gates KL, Sporn PH. Hypercapnia inhibits autophagy and bacterial killing in human macrophages by increasing expression of Bcl-2 and Bcl-xL. *J Immunol*. 2015;194(11):5388–96. <https://doi.org/10.4049/jimmunol.1500150>.
54. Aratani Y, Miura N, Ohno N, Suzuki K. Role of neutrophil-derived reactive oxygen species in host defense and inflammation. *Med Mycol J*. 2012;53(2):123–8. <https://doi.org/10.3314/mmj.53.123>.
55. Marcinkiewicz J, Walczewska M. Neutrophils as sentinel cells of the immune system: a role of the MPO-halide-system in innate and adaptive immunity. *Curr Med Chem*. 2020;27(17):2840–51. <https://doi.org/10.2174/0929867326666190819123300>.
56. Kirk SG, Samavati L, Liu Y. MAP kinase phosphatase-1, a gatekeeper of the acute innate immune response. *Life Sci*. 2020;241:117157. <https://doi.org/10.1016/j.lfs.2019.117157>.
57. Chi H, Barry SP, Roth RJ, Wu JJ, Jones EA, Bennett AM, et al. Dynamic regulation of pro- and anti-inflammatory cytokines by MAPK phosphatase 1 (MKP-1) in innate immune responses. *Proc Natl Acad Sci U S A*. 2006;103(7):2274–9. <https://doi.org/10.1073/pnas.0510965103>.
58. Oh WJ, Endale M, Park SC, Cho JY, Rhee MH. Dual roles of Quercetin in platelets: Phosphoinositide-3-kinase and MAP kinases inhibition, and cAMP-dependent vasodilator-stimulated Phosphoprotein stimulation. *Evid Based Complement Alternat Med*. 2012;2012:485262. <https://doi.org/10.1155/2012/485262>.
59. Luo W, Li YX, Jiang LJ, Chen Q, Wang T, Ye DW. Targeting JAK-STAT signaling to control cytokine release syndrome in COVID-19. *Trends Pharmacol Sci*. 2020;41(8):531–43. <https://doi.org/10.1016/j.tips.2020.06.007>.
60. Marchetti M. COVID-19-driven endothelial damage: complement, HIF-1, and ABL2 are potential pathways of damage and targets for cure. *Ann Hematol*. 2020;99(8):1701–7. <https://doi.org/10.1007/s00277-020-04138-8>.
61. Sala A, Proschak E, Steinhilber D, Rovati GE. Two-pronged approach to anti-inflammatory therapy through the modulation of the arachidonic acid cascade. *Biochem Pharmacol*. 2018;158:161–73. <https://doi.org/10.1016/j.bcp.2018.10.007>.
62. Kong J, Zhu X, Shi Y, Liu T, Chen Y, Bhan I, et al. VDR attenuates acute lung injury by blocking Ang-2-Tie-2 pathway and renin-angiotensin system. *Mol Endocrinol*. 2013;27(12):2116–25. <https://doi.org/10.1210/me.2013-1146>.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.