



Solid Tumor Opioid Receptor Expression and Oncologic Outcomes: Analysis of the Cancer Genome Atlas and Genotype Tissue Expression Project

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Background: Opioid receptors are expressed not only by neural cells in the central nervous system, but also by many solid tumor cancer cells. Whether perioperative opioids given for analgesia after tumor resection surgery might inadvertently activate tumor cells, promoting recurrence or metastasis, remains controversial. We analysed large public gene repositories of solid tumors to investigate differences in opioid receptor expression between normal and tumor tissues and their association with long-term oncologic outcomes.

Methods: We investigated the normalized gene expression of μ , κ , δ opioid receptors (MOR, KOR, DOR), Opioid Growth Factor (OGFR), and Toll-Like 4 (TLR4) receptors in normal and tumor samples from twelve solid tumor types. We carried out mixed multivariable logistic and Cox regression analysis on whether there was an association between these receptors' gene expression and the tissue where found, i.e., tumor or normal tissue. We also evaluated the association between tumor opioid receptor gene expression and patient disease–free interval (DFI) and overall survival (OS).

Results: We retrieved 8,780 tissue samples, 5,852 from tumor and 2,928 from normal tissue, of which 2,252 were from the Genotype Tissue Expression Project (GTEx) and 672 from the Cancer Genome Atlas (TCGA) repository. The Odds Ratio (OR) [95%CI] for gene expression of the specific opioid receptors in the examined tumors varied: MOR: 0.74 [0.63–0.87], KOR: 1.27 [1.17–1.37], DOR: 1.66 [1.48–1.87], TLR4: 0.29 [0.26–0.32], OGFR: 2.39 [2.05–2.78]. After controlling all confounding variables, including age and cancer stage, there was no association between tumor opioid receptor expression and long–term oncologic outcomes.

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Conclusion: Opioid receptor gene expression varies between different solid tumor types. There was no association between tumor opioid receptor expression and recurrence. Understanding the significance of opioid receptor expression on tumor cells remains elusive.

Keywords: opioid receptors, perioperative opioid, cancer, surgery, neoplasm, tumor, immunohistochemistry

INTRODUCTION

Surgery remains a primary treatment for 70% of solid tumors (1) but analgesia after resection is challenging. Pain and nociceptive transmission involve neuronal networks with various receptor types that elicit either activation or suppression of the stimuli (2). Opioids are still the mainstay of postoperative pain management. Their primary site of action is the μ opioid receptor (MOR) which is expressed at various central nervous system (CNS) locations along the pain pathway. Opioid drugs activate MORs to suppress ascending nociception and enhance descending pain inhibition (3, 4). However, one-dimensional reliance on opioid medication has disadvantages. First, MOR is expressed in other tissues such as the brain stem and bowel, leading to undesired side effects such as respiratory depression, nausea, and ileus. Second, repeated, prolonged opioid administration results in hyperalgesia and has been linked to the ongoing problem of opioid dependence (5-8). Third, opioids suppress cell-mediated immunity and directly activate tumor angiogenesis, thereby potentially facilitating residual tumor cell spread (9).

Opioid drugs act as agonists not only at MOR but also at δ opioid receptors (DOR), and both can be expressed by tumor cells (10). Cancer metastasis and proliferation may be associated with the activation of these opioid receptors through different pathways (11). However, MOR, DOR, and κ opioid receptors (KOR), in addition to opioid growth factor receptor (OGFR) and Toll-like receptor 4 (TLR4), have been shown to promote tumor cell migration (12–14). Previous studies aiming to elucidate the role of these receptors in cancer differ widely in their methodology, e.g., immunohistochemistry (IHC) or nucleic acid polymerase chain reaction (PCR) amplification, as well as in the studied samples, e.g., tissue or cell lines, and in targeted receptors (**eTable 1** in the **Supplementary Digital Content**) (15–24).

The evolution of genetic sequencing technologies and the drive to unravel the mechanisms underlying many human diseases has led to the appearance of large repositories of genetic data such as the Genotype Tissue Expression Project (GTEx) and the Cancer Genome Atlas (TCGA). The TCGA is a National Institute of Health (NIH) sponsored project that aims to discover significant cancer-causing genome alterations in large cohorts of tumors through large-scale genome sequencing (25, 26).

Our study objective was to analyse opioid receptor gene expression in tumors compared to normal tissue and to evaluate the association between this and long-term oncologic outcome, defined as overall survival (OS) and disease-free interval (DFI). Gene expression data was obtained from GTEx and TCGA.

MATERIALS AND METHODS

In this analysis we followed the recommendations on reporting results from observational studies (STROBE guidelines. https://www.equator-network.org/reporting-guidelines/strobe/).

Population

We analyzed data from normal and tumor tissues from bladder, breast, colon, liver, salivary gland, esophagus, prostate, stomach, thyroid, lung, and kidney tumors.

Data Collection

Unified GTEx and TCGA gene expression data for MOR, DOR, KOR, TLR4, and OGFR genes were obtained using an established technique (27) (Data record 3) for each tissue type. This dataset includes a strict selection of high-quality RNA-Seq samples processed with the same analysis pipeline and corrected for unwanted non-biological variation that affects comparative analyses. In addition, gene expression values were reported in Fragments per Kilobase Million (FPKM) units.

TCGA survival data were downloaded from the TCGA TARGET GTEx dataset deposited in Xenabrowser (https://xenabrowser.net/). We collected DFI and OS. The remaining clinical data for TCGA samples were obtained from TCGABiolinks (28). Information for GTEx individuals was directly downloaded from the GTEx project website (GTEx Analysis Release V8). Clinical information from the different sources and gene expression data in log2 (FPKM+1) scale were formatted and merged.

Definitions

OS is the length of time from either the date of cancer diagnosis or the start of treatment and death from any cause. DFI is the length of time between primary cancer treatment and any signs or symptoms reappearance (29).

Statistical Analysis

We used data of all available patients without formal sample size calculation. Also, as the purpose was to explore a pathophysiological hypothesis, we did not specify any *a priori* effect size. We reported continuous variables as median and 25th-75th percentiles and categorical variables as numbers and percentages. Distribution was assessed by inspecting quantile–quantile plots, and log-transformation was carried out if the variable distribution violated the normality assumption. Finally, descriptive analyses were performed to summarize patient characteristics.

To assess the association between opioid receptor gene expression and type of tissue, i.e., control *versus* tumor, we fitted mixed logistic models introducing MOR, KOR, DOR, OGFR, and

TLR4 genes as covariables, and primary tumor site as a random effect to consider the variability between different tumor sites. This model was fitted for tumors with data available for every receptor included in the analysis. We fitted a logistic model with all the receptor data available for each tumor type as a sensitivity analysis.

To assess the association between opioid receptor gene expression and DFI and OS, we fitted a mixed Cox model introducing MOR, KOR, DOR, OGFR, and TLR4 genes, age at diagnosis, and cancer stage as covariables. Primary site of tumor was a random effect to consider the variability between different tumor sites.

Statistical significance was set for two-tailed test at P<0.05. No missing values imputation and no correction for multiple comparisons was prespecified: thus, all the findings should be viewed as exploratory. All analyses were performed with R 4.0.3 (The R Foundation for Statistical Computing, www.r-project.org)

RESULTS

We retrieved 8,780 tissue samples: 5,852 from tumor and 2,924 from normal control tissues, of which 2,252 and 672 were from the GTEx and TCGA repository, respectively. Sample characteristics are shown in **Table 1**. OGFR gene expression was highest while MOR gene expression was lowest, with comparable values between control and tumor samples on every

overall gene expression. Median and percentiles values and violin and density plots (**Figure 1**) show considerable overlap. Opioid receptor expression by tumor primary site is reported in **Figure 2**.

The logistic model's estimates are reported in **Table 2**. Opioid receptors were significantly associated with tumor samples, albeit differently, as some genes associated positively and others negatively. The estimated standard deviation among tumors, i.e. 3.06, is bigger than the largest estimate among the fixed effects, i.e. OGFR estimate 2.39, suggesting considerable effect differences among tumors (**Figure 3**). Logistic models estimated for each tumor are reported in **Figure 4** and show considerable variability among opioid receptor estimates across tumor types.

Mixed Cox models estimates for DFI and OS are reported in **Table 3**. After controlling for age and cancer stage, we found no association overall between opioid receptor expression and long-term outcomes except a weak effect for KOR on OS. The cancer stage is by far the predominant effect in both DFI and OS models as was expected.

DISCUSSION

This study's' main findings can be summarized as follows: Firstly, higher or lower opioid receptor gene expression within tumors is variable depending on the specific tumor type; Secondly, single gene expression also varies depending on tumor type; Thirdly,

TABLE 1 Clinical and tumor characteristics.

	Overall (N= 8780)	GTEx Normal (N= 2256)	TCGA Normal (N= 672)	TCGA Tumor (N= 5852)
Tissue type (tumor) % (N)	66.7 (5852/8780)			100 (5852/8780)
MOR gene	0 [0 - 0]	0 [0 - 0.3]	0 [0 - 0.3]	0 [0 - 0]
KOR gene	0.3 [0 -1.1]	0.4 [0 - 1.0]	0.4 [0 - 1.1]	0.2 [0 - 1.2]
DOR gene	0 [0 - 0.4]	0 [0 - 0]	0 [0 - 0]	0 [0 - 0.5]
TLR4 gene	6.7 [6.5 – 7.1]	6.8 [6.5 – 7.0]	6.7 [6.5 – 7.0]	6.8 [6.5 – 7.1]
OGFR gene	4.7 [4.1 – 5.3]	4.7 [4.2 - 5.3]	4.7 [4.2 - 5.3]	4.5 [3.9 – 5.2]
Disease free time (days)			975 [538 – 1677]	724 [410 – 1340]
Relapse (yes) % (N)			16.5 (56/340)	16.8 (531/3167)
Overall survival time (days)			925 [491 - 1813]	738 [402 – 1409]
Death (yes) % (N)			37.4 (235/629)	25.9 (1412/5443)
AJCC stage % (N)			, , , , , , , , , , , , , , , , , , ,	
			198 (34.9)	1660 (34.3)
II			178 (31.3)	1452 (30.0)
III			123 (21.7)	1118 (23.1)
IV			69 (12.1)	616 (12.7)
Age at diagnosis	62 [53 – 71]		63 [52 - 72]	62 [53 - 71]
Gender (Male) % (N)	53.7 (4484/8780)	59.8 (1348/2256)	48.6 (306/629)	51.8 (2830/5467)
Primary site % (N)		, , , , , , , , , , , , , , , , , , ,		· · · · · ·
Bladder	4.4 (390/8780)	0.5 (11/2256)	2.5 (17/672)	6.2 (362/5852)
Breast	13.5 (1181/8780)	3.9 (89/2256)	16.4 (110/672)	16.8 (982/5852)
Colon	8.7 (762/8780)	15.0 (339/2256)	7.6 (51/672)	6.4 (372/5852)
Esophagus	9.7 (853/8780)	29.2 (659/2256)	1.6 (11/672)	3.1 (183/5852)
Kidney	10.6 (929/8780)	1.4 (32/2256)	18.8 (126/672)	13.2 (771/5852)
Liver	5.2 (458/8780)	5.1 (115/2256)	7.1 (48/672)	5.0 (295/5852)
Lung	16.1 (1415/8780)	13.9 (313/2256)	16.4 (110/672)	17.0 (992/5852)
Prostate	6.6 (580/8780)	4.7 (106/2256)	7.1 (48/672)	7.3 (426/5852)
Salivary Gland	5.7 (502/8780)	0 (0/2256)	6.2 (42/672)	7.9 (460/5852)
Stomach	6.9 (605/8780)	8.5 (192/2256)	4.9 (33/672)	6.5 (380/5852)
Thyroid	9.2% (812)	14.1 (318/2256)	7.9 (53/672)	7.5 (441/5852)
Uterus	3.3% (293)	3.6 (82/2256)	3.4 (23/672)	3.2 (188/5852)

Data are reported as median [25th – 75th percentile] or % (N). MOR, μ opioid receptor; KOR, κ opioid receptor; DOR, δ opioid receptor; TLR4, toll–like receptor 4; OGFR, opioid growth factor receptor; Age for GTEx samples is not reported because is recorded as a categorical variable with 10 years strata and not as a continuous variable.



(Log scale) is on the y axis. (B, D, F, H, J) Gene expression (Log scale) s on the x axis.

there was no association between opioid gene receptor expression and DFI or OS after controlling for age and tumor stage.

This analysis has several strengths. First, to our knowledge, this is the first analysis of large public genetic databases focusing specifically on tumor opioid receptor expression and their link to cancer outcome. Secondly, we selected normalized data through a previously published meticulous procedure that consistently removes the batch effect from samples. Thirdly, we controlled for confounding bias by performing a time-to-event analysis, including potential confounders such as age and tumor stage.



We also assessed the effect of specific cancer types by adding a random effect to the multivariable model to account for the hierarchical structure of the data. Fourthly, we included all previously studied opioid receptors known to be involved in perioperative opioid drug binding in our analysis.

The role of tumor opioid receptor expression on tumor growth and metastasis has generated considerable interest among researchers involved in surgical oncology (1). Because of the pivotal role of opioid analgesics in the perioperative process, it has been speculated that activation of these receptors could cause cancer cells to proliferate, migrate and escape immune control. Our findings add further to previous studies assessing opioid receptor expression in different tumors. Higher MOR expression was found on prostate cancer samples compared to unpaired control tissue (22). Likewise, in a study that compared human lung cancer samples with non-tumor adjacent tissue samples, MOR expression was significantly increased in tumor tissue from patients with metastatic lung cancer had an approximately twofold increase in MOR expression (21).

Higher expression of MOR was associated with tumor tissue in gastric cancer, hepatocellular carcinoma, and colorectal cancer samples (17–20, 24). In contrast, this association of MOR-1 with oncological results was not observed in other tumors. For instance, in esophageal squamous cell carcinoma (ESCC), MOR expression in the cytoplasm was associated with lymph node metastases. However, no link was found between MOR expression and the OS of patients with ESCC (18).

In a retrospective analysis among breast cancer patients, which analyzed the effect of anesthetic technique on MOR expression, the authors found that general anesthesia with opioid analgesia increased MOR expression in the resected tumor compared to anesthetic technique with locoregional analgesia (30), These results support the hypothesis that the opioid receptor genetic footprint varies with tumor type. This is consistent with recent data from a triple-negative breast tumor databank, which analyzed the same receptors as us and found that MOR, OPRD, and OPRK were overexpressed, while TLR4 was downregulated. Furthermore, these authors found that higher doses of intraoperative opioids were associated with somewhat worse oncologic outcomes than patients receiving lower doses during surgery (16). A thorough mapping of different receptors' expression is important because opposing effects have been described, with some receptor activation having protumor effects while others have potentially tumor suppressing effects. This is even more important since both exogenous and endogenous opioid receptor agonists may play a different role depending on the specific profile of receptor expression, while opioid receptor antagonists such as methylnaltrexone have found to be associated with longer median survival in an unplanned posthoc analysis of two clinical trials (31).

We found no association between opioid receptor expression and long-term outcomes such as DFI and OS. Existing data on this matter are diverging. For example, while some studies found an association between MOR expression in particular and cancer

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Gene expression (Log scale)	Odds Ratio [Lower-Upper 95%CI]	P- value
MOR	0.74 [0.63 – 0.87]	< 0.001
KOR	1.27 [1.17 – 1.37]	< 0.001
DOR	1.66 [1.48 – 1.87]	< 0.001
TLR4	0.29 [0.26 - 0.32]	< 0.001
OGFR	2.39 [2.05 – 2.78]	< 0.001
Random effect parameter (Tumor type): Standard deviation: 3.06		
ICC (Tumor_type): 0.74		

Primary site random effect Standard deviation for multivariable model: 1.34. MOR, μ opioid receptor; KOR, κ opioid receptor; DOR, δ opioid receptor; TLR4, toll-like receptor; OGFR, opioid growth factor receptor; CI, Confidence Interval; ICC, Intraclass correlation coefficient. The model has been estimated with all tumor types with available data for all receptors.







FIGURE 4 | Logistic model fit of opioid receptors association with tumor tissue by tumor type. Dotted red line represents no effect. Estimates are reported as red or blue when the odds ratio point estimate is lower or greater than one respectively. Dots are point estimates and bars 95% confidence intervals. Statistical significance is reported as * < 0.05, ** < 0.001, *** < 0.001.

recurrence (18, 19, 22), others did not (16, 24). Indeed, available research on this topic seems to point out that there is no one-size-fits-all explanation to this question. Further investigations on the specific receptor profile of each cancer strain should lay the foundation on whether opioid receptors can be included as oncologic prognostic markers.

Studies that assess opioid receptor expression in cancer rely on immunohistochemistry assay, while quantitative methods such as quantitative real-time polymerase chain reaction (qRT–PCR) are seldom reported, and with different procedures and primers (**eTable 1 Supplementary Material**) (15, 19, 20). Since there are no consensus guidelines on how to perform these assays (32), we consider that availing of large and validated databases such as the TCGA and GTEx is a powerful tool to draw a genetic footprint of opioid receptors in tumor cells. Furthermore, as our present results suggest, the impact of opioid receptors on cancer cells does not seem to be based on a simple pathway involving an individual receptor overexpression and is probably a more integrated mechanism involving several receptor targets with different effects that can vary depending on specific tumor type.

Furthermore, genetic content within tumors is variable and opioid receptors can present single nucleotide polymorphisms (SNPs). This type of polymorphisms on known oncogenes such as p53 and X-Ray Repair Cross Complementing 3 (XRRCC3) genes have been studied to elucidate their effect on cancer TABLE 3 | Opioid gene expression association with long term oncologic outcomes.

Disease	Free	interval
Disease	1166	interval

Gene expression (Log scale)	Hazard Ratio [Lower-Upper 95%CI]	P value
MOR	1.17 [0.89– 1.68]	0.203
KOR	1.07 [0.97 –1.18]	0.156
DOR	1.06 [0.91 – 1.24]	0.417
TLR4	1.01 [0.88 – 1.14]	0.929
OGFR	1.02 [0.85 - 1.23]	0.765
Age at diagnosis	0.99 [0.98 - 1.00]	0.203
AJCC Stage (Ref Stage I)*		
∥	1.32 [0.99 – 1.77]	0.057
	1.96 [1.43 – 2.69]	> 0.001
Random effect parameter (Tumor type): Standard deviation: 0.64		

Overall Survival

Gene expression (Log scale)	Hazard Ratio [Lower–Upper 95%CI]	P value
MOR	1.17 [0.97 – 1.41]	0.087
KOR	1.10 [1.01 – 1.21]	0.021
DOR	1.00 [0.94 – 1.07]	0.794
TLR4	0.98 [0.90 – 1.07]	0.731
OGFR	1.00 [0.90 – 1.11]	0.937
Age at diagnosis	1.02 [1.01 – 1.03]	< 0.001
AJCC Stage (Ref: Stage I)		
	1.49 [1.22 – 1.82]	< 0.001
	2.57 [2.10 - 3.14]	< 0.001
IV	4.69 [3.66 - 6.01]	< 0.001
Random effect parameter (Tumor type): Standard deviation: 0.62		

MOR, μ opioid receptor; KOR, κ opioid receptor; DOR, δ opioid receptor; TLR4, toll-like receptor; OGFR, opioid growth factor receptor; CI, Confidence Interval; AJCC, American Joint Committee on Cancer.

*AJCC stage IV data were not available for Disease free interval.

susceptibility with conflicting results to date (33–35). For instance, particular SNPs such as the A118G have been linked to reduced sensitivity to opioid medication in the patient suffering from chronic pain (36, 37) and cancer (19) and even cancer recurrence in specific tumor types and populations (38, 39), Also, TLR4 gene polymorphisms have also been studied and may play a role in proliferation and differentiation and multiple isoforms of receptor subtype resulting from alternative splicing of the pre-mRNA transcript have been identified albeit their functional role has yet to be clarified (40). Investigators are beginning to expand the horizon outside the genetic profile of opioid receptors and to include specific genetic alterations such as Cyclin-Dependent Kinase Inhibitor 2A (CDKN2A) mutations (41), although the influence that specific influence of individual receptor isoforms is still a matter of debate (42).

Several limitations must be acknowledged. For instance, differences in baseline characteristics from GTEx and TCGA repositories may be present. Also, although we controlled for age and cancer stage, the effect of other confounding factors not included in the analysis, such as type of surgery or pathologic stage or opioid agonists or antagonists administration, cannot be ruled out. These parameters could have a modifying effect on the association between opioid receptor expression and long-term cancer outcomes. Furthermore, we also acknowledge that while we assessed all the most common opioid receptor genes, other molecular pathways can be involved in the effect of opioids on cancer growth. Finally, because of the hypothesis-generating purpose of this study, we did not set any *a priori* effect

threshold or multiple comparisons correction; hence some results' statistical significance and the potential hypothesis derived from them should be confirmed in future trials.

In conclusion, the most common solid tumors express higher opioid receptor genes than normal tissue, but variably depending on the primary tumor analyzed. No association was found between disease-free and overall survival and opioid gene expression after controlling for age and tumor stage. Further studies are warranted to elucidate the specific genetic footprint of opioid receptors in each cancer type and the potential role of gene polymorphisms.

DATA AVAILABILITY STATEMENT

Publicly available datasets were analyzed in this study. This data can be found here: https://www.nature.com/articles/sdata201861#ref-CR22.

AUTHORS CONTRIBUTIONS

AB: This author conceived the idea, helped with data acquisition, critical review of the content, and manuscript preparation. SZ: This author coded the data acquisition from the repositories, provided a critical review of the content, and helped with manuscript preparation. IG-C: This author provided a critical

review of the content, and helped with manuscript preparation. PE: This author provided a critical review of the content, and helped with manuscript preparation. MA: This author provided a critical review of the content, and helped with manuscript preparation. DB: This author provided a critical review of the content, and helped with manuscript preparation. OD-C: This author provided a critical review of the content, and helped with manuscript preparation. GM: This author conceived the idea, helped with data acquisition, critical review of the content, and

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2022.801411/full#supplementary-material

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