

Contrasting functions of the epithelial-stromal interaction 1 gene, in human oral and lung squamous cell cancers

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Abstract. The epithelial-stromal interaction 1 gene (*EPSTII*) is known to play multiple roles in the malignant progression of breast cancer and also in some aspects of the immune responses to the tumor. However, the relevance of the gene in the onset/progression of oral squamous cell carcinoma (OSCC) and lung squamous cell carcinoma (LSCC) is not yet known. The present study was aimed at revealing the roles of *EPSTII* in conferring malignant characteristics to OSCC and LSCC, and the underlying mechanisms. Quantitative real-time polymerase chain reaction (PCR) and western blot analyses demonstrated significant upregulation of *EPSTII* in all four OSCC cell lines (HSC2, HSC3, HSC3-M3 and HSC4), and significant downregulation of *EPSTII* in all three LSCC cell lines (LK-2, EBC-1 and H226) used in the present study, as compared to the expression levels in the corresponding control cell lines. Both knockdown of *EPSTII* in OSCC and overexpression of the gene in LSCC suppressed cell proliferation, and induced cell-cycle arrest in the G1 phase, with upregulation of *p21* and downregulation of *CDK2* and cyclin D1. Furthermore, these alterations of *EPSTII* gene expression in the OSCC and LSCC cell lines suppressed the cell migration ability and reversed the EMT phenotype of the tumor cells. Collectively, while *EPSTII* appears to have oncogenic roles in OSCC, it appears to exert tumor-suppressive roles in LSCC. PCR array analyses revealed some genes whose expression levels were altered along with the modified *EPSTII* expression in both the OSCC and LSCC cell lines. These findings suggest that *EPSTII* may be a therapeutic target for both OSCC and LSCC.

Introduction

Airborne carcinogens, including tobacco smoke, are strongly implicated in the etiology of both squamous cell carcinomas of the head and neck and squamous cell carcinoma of the lung (1). Similar to the case for all head and neck cancers (HNC), squamous cell carcinoma is the most common histologic subtype of oral cancer (OSCC), accounting for nearly 90% of all cases of oral cancer (2). OSCC ranks sixth in cancer incidence worldwide, with approximately 300,400 newly diagnosed cases and 145,400 deaths each year, and the cancer is often diagnosed at an advanced stage (3,4). The two most important risk factors for OSCC, accounting for over 80% of all patients, are tobacco and alcohol (5). Smoking is also a strong risk factor for lung squamous cell carcinoma (LSCC), which accounts for 15 to 30% of all cases of lung cancer; a trend towards decrease in the incidence rates associated with a decrease in smoking rates has been reported in developed countries (6). Due to the occurrence of lung cancer in epidemic proportions worldwide, however, urgent development of countermeasures is still required for LSCC.

Recent advances in molecular-targeted therapies have brought about significant improvements in the treatment outcomes of some advanced cancers (7). Especially, tyrosine-kinase inhibitors (TKIs) have proven to be significantly beneficial for patients with advanced adenocarcinomas of the lung harboring driver oncogene mutations, including activating epidermal growth factor receptor (*EGFR*) mutations and rearrangements of the anaplastic lymphoma kinase gene (8). On the other hand, in the case of LSCC, no driver mutations or effective TKIs have been identified yet, thus no effective molecular-targeted therapies against this cancer are available to date. Although cetuximab, an anti-EGFR antibody, administered in combination with radiation (9) or cytotoxic chemotherapy (10) has been revealed to be effective for advanced HNC, including OSCC, and another anti-EGFR antibody, necitumumab, administered in combination with cytotoxic chemotherapy (11) has been revealed to be effective for advanced LSCC, most patients with advanced OSCC or LSCC eventually exhibit disease progression. Therefore, novel molecular targets must be sought for developing novel molecular-targeted therapies for OSCC and LSCC.

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The epithelial stromal interaction 1 gene (*EPSTII*) is a gene mapped to human chromosome 13q13.3, that has received increasing attention in recent years. It has been revealed to be expressed in some normal tissues, including the spleen, small intestine, salivary glands, testes, germinal centers of lymph nodes, and placenta (12-15). In addition, overexpression of the gene has been reported in primary cancers of the breast, colorectal and pancreas (12,16,17). In breast cancers, high expression levels of *EPSTII* have been revealed to be strongly associated with enhanced tumor cell migration ability and tumor invasiveness, a high propensity for metastasis, and escape from apoptosis, suggesting that *EPSTII* expression may be an independent prognostic marker in patients with breast cancer (13,18,19). Furthermore, accumulated evidence has revealed multiple roles of *EPSTII* in the immune response; it has been demonstrated to be associated with the establishment of immune privilege, development of autoimmune diseases, control of viral infections, and activation of macrophages. The gene has been revealed to have a role in the immune privilege of the testes (20), and a genome-wide association study has linked the gene to male fertility (21). Overexpression of *EPSTII* has been reported in the peripheral blood cells of patients with systemic lupus erythematosus (22), and such overexpression was revealed to be associated with abnormal B-cell activation via the NF- κ B signaling pathway in primary Sjogren syndrome (23); expression of the gene induced by HCV infection has been revealed to effectively inhibit viral replication and exert antiviral effects through upregulation of IL-28A (24). Finally, the gene has also been revealed to play a role in modulating macrophage activation and polarization (15). However, despite these already known functions of *EPSTII*, its roles in human OSCC and LSCC remain unclear.

Materials and methods

Cell lines. A human normal oral keratinocyte cell line, HNOKs, derived from a mixture of healthy gingival specimens of 22- to 35-year-old patients, was established and maintained as previously described (25). Human OSCC cell lines HSC2, HSC3, and HSC4 were purchased from RIKEN Bioresource Center, and HSC3-M3 was purchased from the Human Science Resources Bank. The cells were cultured in DMEM (Sigma-Aldrich; Merck KGaA). A normal human lung cell line, BEAS-2B, and the LSCC cell lines LK-2, EBC-1 and H226 were purchased from JCRB cell bank. BEAS-2B, LK-2 and H226 were cultured in PRMI-1640 medium, and EBC-1 was cultured in MEM (both from Sigma-Aldrich; Merck KGaA). All the culture media were supplemented with 10% fetal bovine serum and 1% antibiotics, including penicillin and streptomycin (all from Life Technologies; Thermo Fisher Scientific, Inc.). All the cell lines were cultured in a humidified incubator at 37°C in a 5% CO₂ atmosphere.

Expression of mRNA and protein. The cells were washed three times with phosphate-buffered saline (PBS), followed by extraction of total RNA and protein. The mRNA expression levels were quantified by real-time reverse transcription-quantitative polymerase chain reaction (RT-qPCR) with

SYBR green expression assays (Roche Diagnostics), in accordance with the manufacturer's instructions. In brief, total RNA was extracted using TRIzol reagent (Invitrogen; Thermo Fisher Scientific, Inc.) when the cells reached 80% confluence in the 10-cm culture plates. Reverse transcription was performed using the ReverTra Ace qPCR RT Master Mix (Toyobo Life Science). Gene expression was measured using LightCycler 480 Instrument (Roche Diagnostics). The thermocycling conditions for the PCR were as follows: initial denaturation at 95°C for 10 min, followed by 45 cycles of amplification at 95°C (10 sec) for denaturation, 60°C (10 sec) for annealing and 72°C for extension, followed by a cooling step at 40°C for 30 sec. GAPDH was used as the internal control. Transcript amounts were estimated from respective standard curves and normalized to GAPDH. Primers were designed using the Universal Probe Library Assay Design Center (<http://lifescience.roche.com/>). The primers for *EPSTII* were 5'-CCGGAGAAATGAGATACAAAGAAT-3' (forward) and 5'-GGTGAACCGTTTGTAGCTCTG-3' (reverse), and the primers for *GAPDH* were 5'-AACATCATCCCTGCCTCTACTGG-3' (forward) and 5'-TTGAAGTCAGAGGAGACC ACTG-3' (reverse).

For the evaluation of the protein expression, the cells were washed twice with cold phosphate-buffered saline (PBS) and collected with lysis buffer [7 M urea, 2M thiourea, 4% (w/v) CHAPS, and 10 mM Tris] with a proteinase inhibitor cocktail (Roche Diagnostics). The protein concentration was first assessed by the Bradford method, followed by western blot analysis. Protein extracts (20 μ g/lane) were separated by a 4-12% Bis-Tris gel (Thermo Fisher Scientific, Inc.), and transferred to nitrocellulose membranes. Subsequently, the membrane was blocked for 1 h at room temperature in 5% skim milk, and then incubated with primary antibodies (1:1,000) overnight at 4°C. The membranes were washed with 0.1% Tween-20 in Tris-buffered saline, three times (15 min for each wash), followed by incubation with a secondary antibody with anti-mouse (cat. no. S3721) or anti-rabbit IgG (cat. no. S3731) (1:2,000; Promega Corporation) for 1.5 h at room temperature. Finally, the bands were detected using clarity Western ECL Substrate (Bio-Rad Laboratories, Inc.), and immunoblotting images were visualized by exposing the membranes to the BioRad ChemiDoc™ XRS System (BioRad Laboratories, Inc.). The densitometric analysis of the protein expression was performed using Image Lab software 6.0.1 (Bio Rad Laboratories, Inc.).

The antibodies used were mouse anti-EPSTII monoclonal antibody (cat. no. AT1934a; Abcepta; Abgent, Inc.), rabbit anti-E-cadherin (product no. 3195), anti-N-cadherin (product no. 13116), anti-p21 (product no. 2947), anti-cyclin D1 (product no. 55506), and anti-CDK2 (product no. 18048) monoclonal antibodies (all from Cell Signaling Technology, Inc.), and mouse anti-fibronectin (product no. SAB4200760), anti- β -actin (product no. SAB1305567) (both from Sigma Aldrich; Merck KGaA), and anti- α -tubulin monoclonal antibodies (cat. no. sc-5286; Santa Cruz Biotechnology, Inc.).

Transfection of plasmids. OSCC-derived cells (HSC3-M3 and HSC4) were transfected with *EPSTII*-suppressing shRNA or the control plasmid, shMock (cat. no. sc-105335-SH and cat. no. sc-108060, respectively, Santa Cruz Biotechnology,

Inc.), at a concentration of 2.5 $\mu\text{g}/\text{well}$, using Lipofectamine 3000 (Thermo Fisher Scientific, Inc.), and then incubated overnight at 37°C. After the transfection, the cells exhibiting stable transfection were selected by taking advantage of the selection marker and its inhibitor, puromycin (Santa Cruz Biotechnology, Inc.). LSCC-derived cells (LK-2 and EBC-1) were transfected with *EPSTII*-overexpressing plasmid (OriGene Technologies, Inc.) or the control plasmid (OriGene Technologies, Inc.) at a concentration of 2.5 $\mu\text{g}/\text{well}$, using Lipofectamine 3000 and incubated overnight at 37°C. Following the transfection, the cells exhibiting stable transfection were selected by utilizing the selection marker and its inhibitor G-418 Solution (Roche Diagnostics). Then, incubation of the selected cells with the inhibitors for 2 to 3 weeks yielded individual clones from each of the *EPSTII*-suppressed OSCC cell lines and *EPSTII*-overexpressing LSCC cell lines; these clones and the control cell lines were used for further experiments.

Proliferation assay. To evaluate the cell proliferation ability, shEPSTII and shMock cells (HSC3-M3 and HSC4), oeEPSTII and OeMock cells (5×10^4 cells/plate) were seeded on to a 6-cm plate, and the number of cells were counted every 24 h in triplicate using a hemocytometer to measure the chronological changes.

Wound healing assay. For evaluating the cell migration ability, 2×10^5 cells/well were seeded on to a 6-well plate overnight until they reached 95% confluence; the surface of the plate was then scratched with the tip of a 1,000- μl micropipette and the culture medium was immediately washed off with PBS; the cells were then cultured in serum-free medium for a further 24 h. The scratch scars were viewed under an inverted microscope (x10 magnification of objective) (ECLIPSE TS100; Nikon Corporation), and the scratched areas were quantified using the Lenaraf220b free software (available at <http://www.vector.co.jp/soft/dl/win95/art/se312811.html>). The wound closure rates were determined as a percentage of the total repaired area per hour and normalized to the control.

Cell cycle analysis. Cells were harvested with trypsin and centrifuged for 5 min at 300 x g at room temperature, and then the concentration was adjusted to 1.0×10^6 cells/ml. Cell cycle analyses were performed using the Cycletest™ Plus DNA reagent kit (BD Biosciences) and the BD Accuri C6 Flow Cytometer (BD Biosciences), in accordance with the manufacturers' instructions. For data analysis FCS Express 4 (De Novo Software) was used.

PCR array. Differences in the mRNA expression of the *EPSTII*-related genes in the two *EPSTII*-downregulated OSCC cell lines, HSC3-M3 and HSC4, and two *EPSTII*-overexpressing LSCC cell lines, LK2 and EBC-1, as compared to the expression levels in the corresponding parental cells, were detected using the RT² Profiler PCR Array Human Cancer Pathway Finder (cat. no. PAHS-033ZF-6; Qiagen, Inc.). In brief, cDNA was synthesized from the total RNA extracted from each cell line using the RT² First Strand Kit (Qiagen, Inc.). RT² SYBR Green qPCR Master Mix (Qiagen, Inc.) was used for the PCR arrays, in accordance with the manufacturer's instructions.

Bioinformatics analysis. The gene expression in clinical samples was obtained from the Oncomine Platform (<http://www.oncomine.org>).

Statistical analysis. The unpaired Student's t-test was used to analyze the statistical significance of differences between two groups. One-way ANOVA with Dunnett's post hoc test calculated by GraphPad Prism software (version 8; GraphPad software, Inc.) were used to evaluate the differences of multiple comparisons. All experiments were performed in triplicate and repeated 3 times, unless otherwise specified. $P < 0.05$ was considered to indicate statistically significant differences. The data are expressed as the means \pm standard error (SE).

Results

***EPSTII* expression in the OSCC and LSCC cell lines.** The series of RT-qPCR and western blot analyses in the four OSCC (HSC2, HSC3, HSC3-M3 and HSC4) and three LSCC (EBC-1, LK-2 and H226) cell lines revealed significant overexpression of *EPSTII* in all of the OSCC cell lines (Fig. 1A and C), except for protein expression in HSC2, and significantly suppressed expression of the gene in all of the LSCC cell lines (with the exception of *EPSTII* mRNA expression in the LK-2 cell line) (Fig. 1B and D), as compared to the expression levels in the corresponding control cell lines, HNOKs and BEAS-2B, respectively (Fig. 1B).

Establishment of cells with EPSTII knockdown and EPSTII overexpression. Based on the results of the experiments aforementioned (Fig. 1C and D), the OSCC cell lines, HSC3-M3 and HSC4, and LSCC cell lines, LK-2 and EBC-1, were selected for further experiments because the former exhibited the strongest expression, among all the OSCC cell lines, of the EPSTII protein, and the latter exhibited the weakest expression, among all the LSCC cell lines, of the EPSTII protein. HSC3-M3 and HSC4, were transfected with *EPSTII* shRNA plasmid to suppress the gene expression, which yielded two clones for each cell line, whereas LK-2 and EBC-1 were transfected with the *EPSTII* overexpression plasmid, which yielded a single clone per cell line, to further investigate the roles of the gene. As transfection of the *EPSTII* overexpression plasmid into the LSCC cell lines was somewhat difficult, only a single clone was obtained for each cell line. The alterations in the expression levels of the gene were verified by RT-qPCR and western blot analysis. The results revealed significant downregulation of *EPSTII* at the mRNA and/or protein level in the shRNA-transfected OSCC cell lines as compared to the shMock-transfected cells (Fig. 2A and C); similarly, significant upregulation of the gene at the mRNA and/or protein level was confirmed in the LSCC cells transfected with the overexpression vector as compared to the oeMock-transfected cells (Fig. 2B and D).

Altered expression of EPSTII induces alterations in the cell proliferation ability and cell cycle progression. Cell propagation was significantly suppressed by downregulation of *EPSTII* in the two OSCC cell lines (Fig. 3A), and by upregulation of the gene in the two LSCC cell lines (Fig. 3B). Cell cycle analysis demonstrated accumulation of cells in the G1

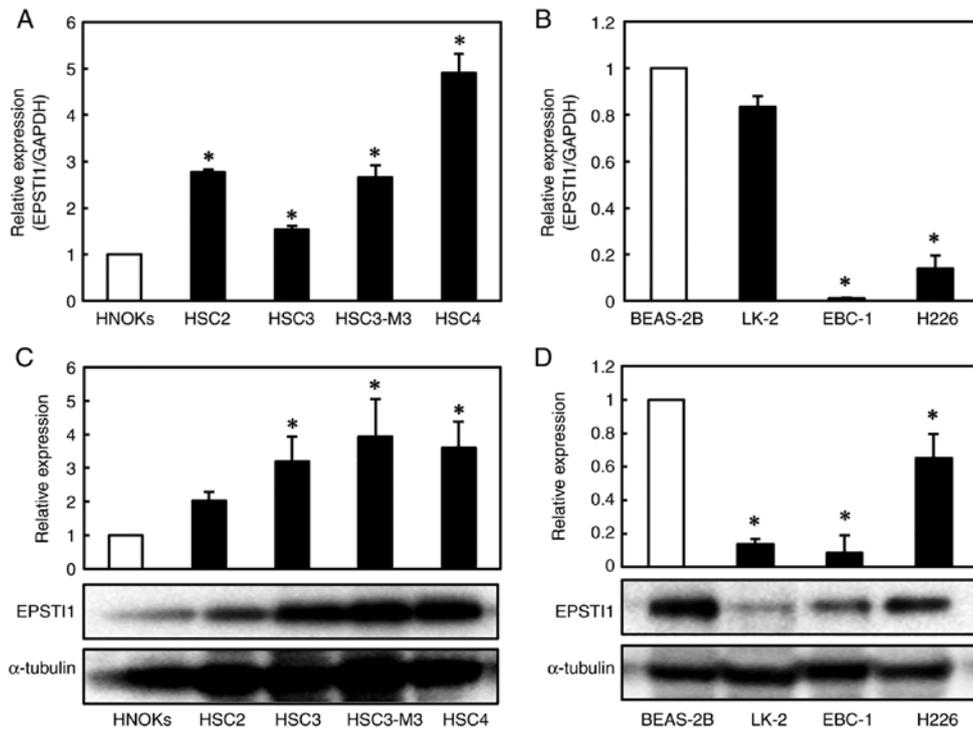


Figure 1. Expression of *EPSTI1* in human OSCC and LSCC cell lines. Quantification of *EPSTI1* mRNA expression levels by RT-qPCR revealed (A) significantly increased expression levels in the four human OSCC cell lines (HSC2, HSC3, HSC3-M3 and HSC4) as compared to the levels in the control cell line, HNOKs, and (B) significantly reduced expression levels in two of the three human LSCC cell lines (EBC-1 and H226, but not LK-2) as compared to the levels in the normal control cell line, BEAS-2B. Western blotting, using α -tubulin as the internal control, confirmed (C) significant upregulation of the gene in the OSCC cell lines and (D) significant downregulation of the gene in the LSCC cell lines, including LK-2. The values on the y-axes represent the ratios to the expression levels in the normal control cell lines. Each experiment was performed in triplicate and repeated three times, and data are presented as the means \pm SE. * $P < 0.05$. *EPSTI*, epithelial-stromal interaction 1; OSCC, oral squamous cell carcinoma; LSCC, lung squamous cell carcinoma; SE, standard error.

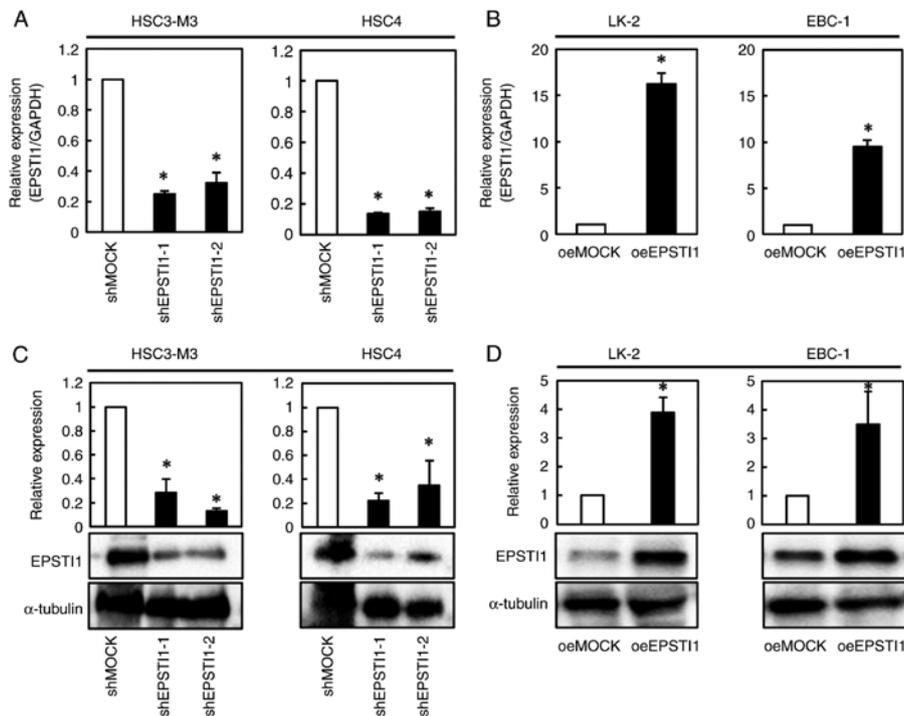


Figure 2. Confirmation of the modified gene expression of *EPSTI1* in the OSCC and LSCC cell lines. For further elucidation of the functions of *EPSTI1*, expression of the gene was knocked down in two OSCC cell lines, HSC3-M3 and HSC4, by transfection of shRNA, and overexpressed in two LSCC cell lines, LK-2 and EBC-1, using overexpressing vectors. Consequently, *EPSTI1* mRNA expression was (A) significantly suppressed in the two OSCC cells as compared to that in the mock transfectants, and (B) significantly overexpressed in the two LSCC cells as compared to that in the mock-transfected cells. In addition, western blotting confirmed (C) reduced *EPSTI1* protein expression in the shRNA-transfected OSCC cell lines, and (D) overexpression of *EPSTI1* protein in the overexpressing vector-transfected LSCC cell lines. Each experiment was performed in triplicate and repeated three times, and data are presented as the means \pm SE. * $P < 0.05$. *EPSTI*, epithelial-stromal interaction 1; OSCC, oral squamous cell carcinoma; LSCC, lung squamous cell carcinoma; SE, standard error.

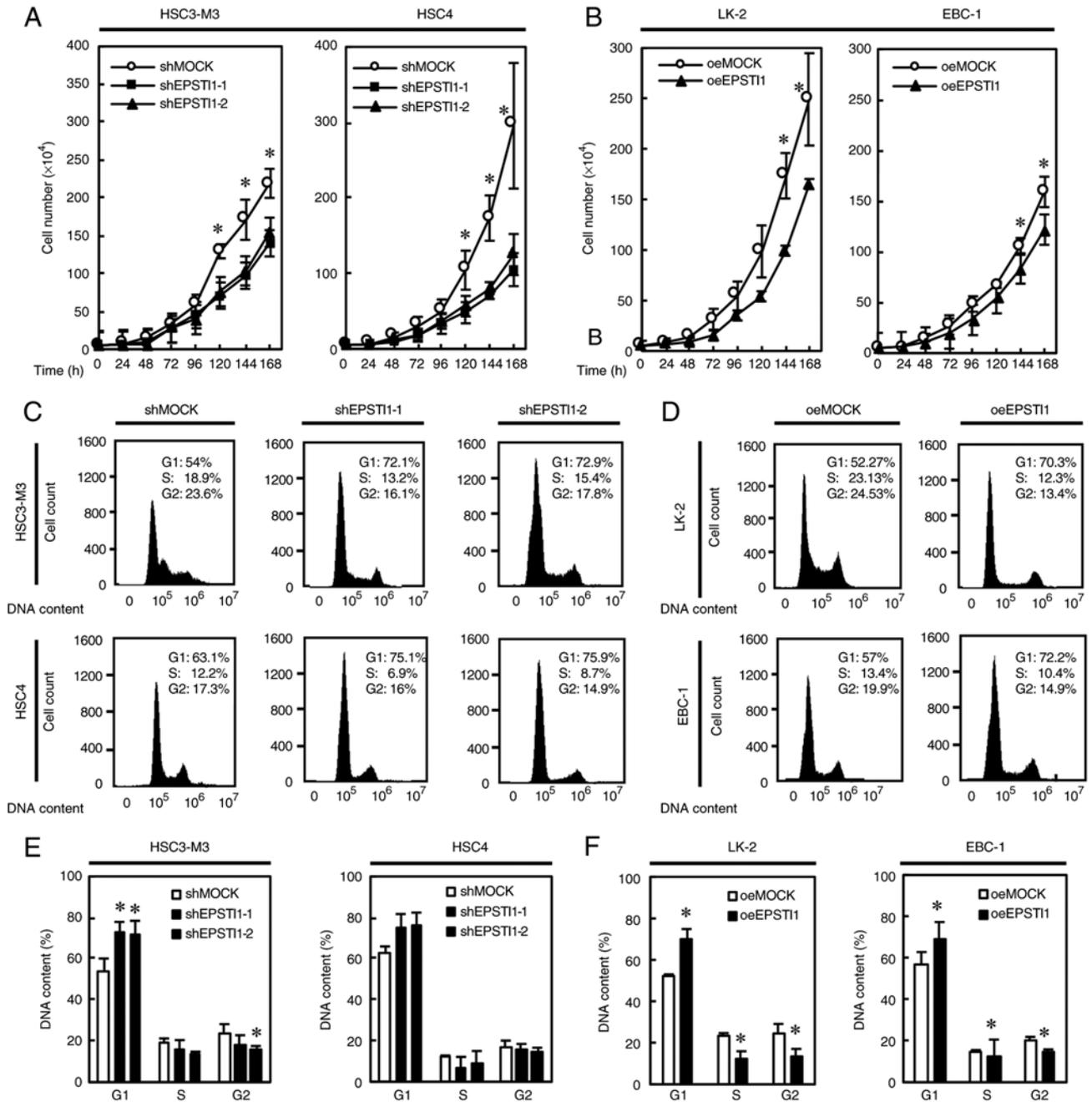


Figure 3. Inhibition of cell proliferation and induction of cell cycle arrest in the G1 phase is associated with modified *EPSTII* expression. Genetically modified expression of *EPSTII*; (A) downregulation of the gene in two OSCC cell lines was associated with significantly suppressed cell proliferation and (C and E) increase in the G1 population, although statistical significance was marginal in HSC4 (P=0.0533), suggesting cell cycle arrest. Similarly, (B) overexpression of the gene in two LSCC cell lines significantly suppressed cell proliferation and (D and F) induced cell cycle arrest in the G1 phase, with a significant decrease in the population of cells in the S and G2 phases in the LK-2 cell line, and in the population of cells in the G2 phase in the EBC-1 cell line. Each experiment was performed in triplicate and repeated six times for A and B, and three times for C-F. Data are presented as the means ± SE. *P<0.05. *EPSTI*, epithelial-stromal interaction 1; OSCC, oral squamous cell carcinoma; LSCC, lung squamous cell carcinoma; SE, standard error.

phase in every OSCC cell line with downregulated *EPSTII* expression and every LSCC cell line with upregulated *EPSTII* expression with statistical significance except for HSC4 which revealed a trend of G1 accumulation (P=0.053) (Fig. 3C-F). To confirm this finding, the expression levels of the cell cycle-related genes, *p21*, *CDK2*, and cyclin D1, were investigated. As anticipated, upregulation of *p21* and downregulation of *CDK2* and cyclin D1 were observed in every OSCC cell line with downregulated *EPSTII* expression (Fig. 4A and C) and in every LSCC cell line with upregulated *EPSTII* expression

(Fig. 4B and D), although statistical significance was marginal for *p21* (P=0.08) and cyclin D1 (P=0.06) in HSC3-M3 cells.

Altered expression of EPSTII induces altered cell migration ability and epithelial-mesenchymal transition. The wound healing assay revealed significantly decreased cell migration ability in OSCC cell lines with downregulated *EPSTII* expression, HSC3-M3 and HSC4 (Fig. 5A) and in the LSCC cell line EBC-1 with upregulated *EPSTII* expression (LK-2 was not used since the cells were scattered once the cells reached

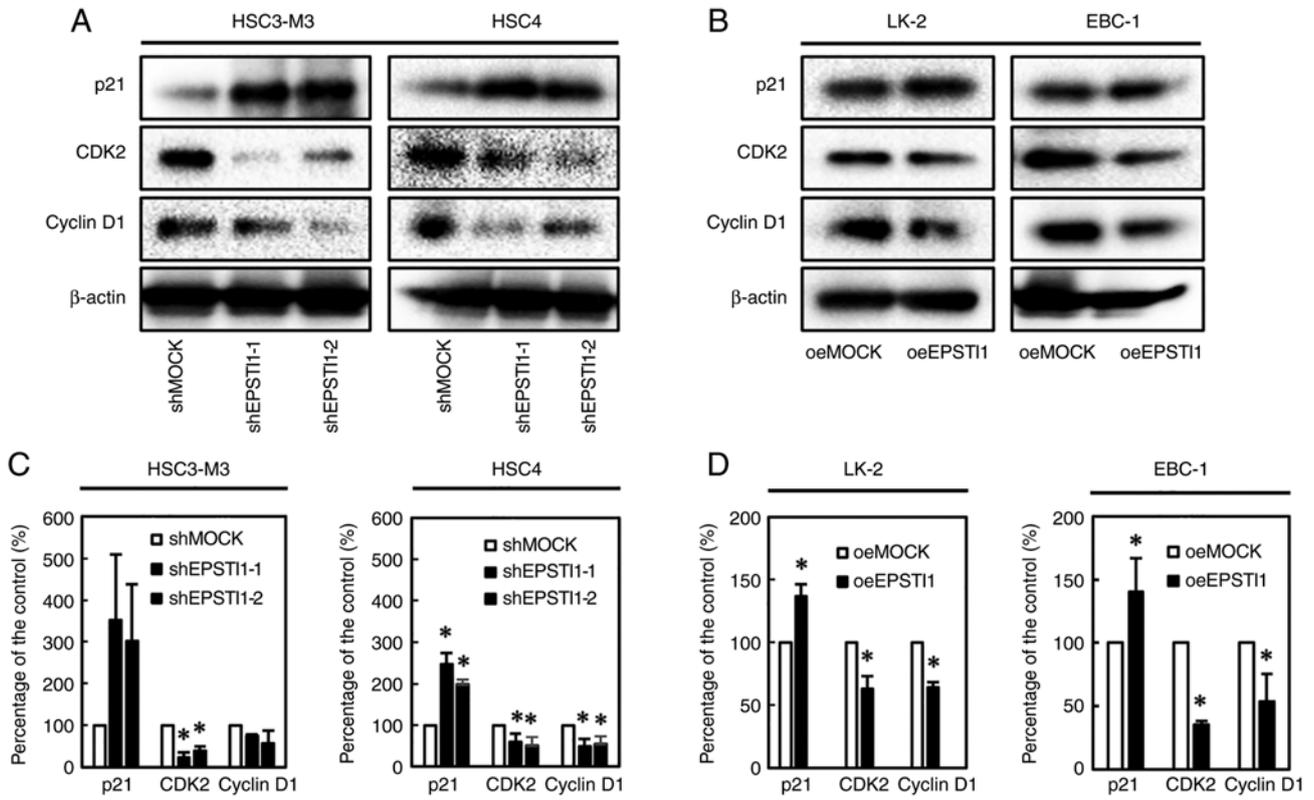


Figure 4. Expression of the cell-cycle-related genes, *p21*, *CDK2*, and Cyclin D1. Western blotting revealed enhanced expression of *p21*, and suppressed expression of *CDK2* and Cyclin D1 associated with modified *EPSTI1* expression (upregulation and downregulation, respectively) in the (A and C) OSCC and (B and D) LSCC cell lines, although statistical significance was marginal for *p21* ($P=0.08$) and cyclin D1 ($P=0.06$) in HSC3-M3, which appears consistent with cell cycle arrest in each of the cell lines. Each experiment was performed in triplicate and repeated three times, and data are presented as the means \pm SE. * $P<0.05$. *EPSTI*, epithelial-stromal interaction 1; OSCC, oral squamous cell carcinoma; LSCC, lung squamous cell carcinoma; SE, standard error.

confluence) (Fig. 5B). The expression levels of the epithelial-mesenchymal transition (EMT)-related proteins were evaluated by western blot analysis. Significant upregulation of E-cadherin and downregulation of N-cadherin and fibronectin, suggestive of a suppressed EMT phenotype, were observed in OSCC cell line with downregulated *EPSTI1* expression and LSCC cell line with upregulated *EPSTI1* expression with exception in fibronectin in HSC4 which exhibited marginal statistical significance ($P=0.08$) (Fig. 5C-F).

EPSTI1-related genes as shown by PCR arrays. PCR arrays identified some upstream and downstream genes in relation to the altered *EPSTI1* expression in the two OSCC and two LSCC cell lines, as summarized in Tables I and II. Overexpression of carbonic anhydrase 9 (*CA9*) and downregulation of C-C motif chemokine ligand 2 (*CCL2*) were identified in both the *EPSTI1*-suppressed OSCC cell lines, as compared to the corresponding control cell line (Table I), whereas downregulation of cyclin D2 (*CCND2*), baculoviral IAP repeat containing 3 (*BIRC3*) and insulin-like growth factor binding protein 7 (*IGFBP7*) were identified in all the *EPSTI1*-overexpressing LSCC cell lines, as compared to the corresponding control cell line (Table II).

Discussion

The present study revealed significantly higher expression levels of *ESPTI1* in all the four OSCC cell lines examined in

the present study as compared to the corresponding control cell line, and significantly lower expression levels of the *EPSTI1* gene in all the three LSCC cell lines examined as compared to the corresponding control cell line. Both OSCC cell lines with forced downregulation of the gene and LSCC cell lines with overexpression of the gene exhibited significantly suppressed tumor cell proliferation ability. Analyses of the cell cycle and of genes related to the cell cycle, i.e., *p21*, *CDK2* and cyclin D1, generally supported the finding of cell cycle arrest in the G1 phase in both the aforementioned OSCC and LSCC cell lines. OSCC cell lines with downregulation of *EPSTI1* and LSCC cell lines with upregulation of *EPSTI1* similarly revealed suppressed EMT, as assessed by the cell migration activity and cellular expression levels of E-cadherin, N-cadherin and fibronectin. Therefore, *EPSTI1* was demonstrated to play roles in enhancing the malignant phenotypes, including the cell proliferation ability and EMT in OSCC, whereas it played the opposite roles in LSCC.

Overexpression of *EPSTI1* was first identified in breast cancer, and overexpression of the gene is considered to drive the malignant phenotype, including the cell proliferation and migration abilities, invasiveness, EMT, and escape from apoptosis (12-14,18,19). It has also been identified as being upregulated in cell lines derived from peritoneal disseminations of colorectal cancer as compared to the cell lines derived from primary colorectal cancers (16). In pancreatic cancer, gene expression was associated with resistance to therapy with an oncolytic virus (17).

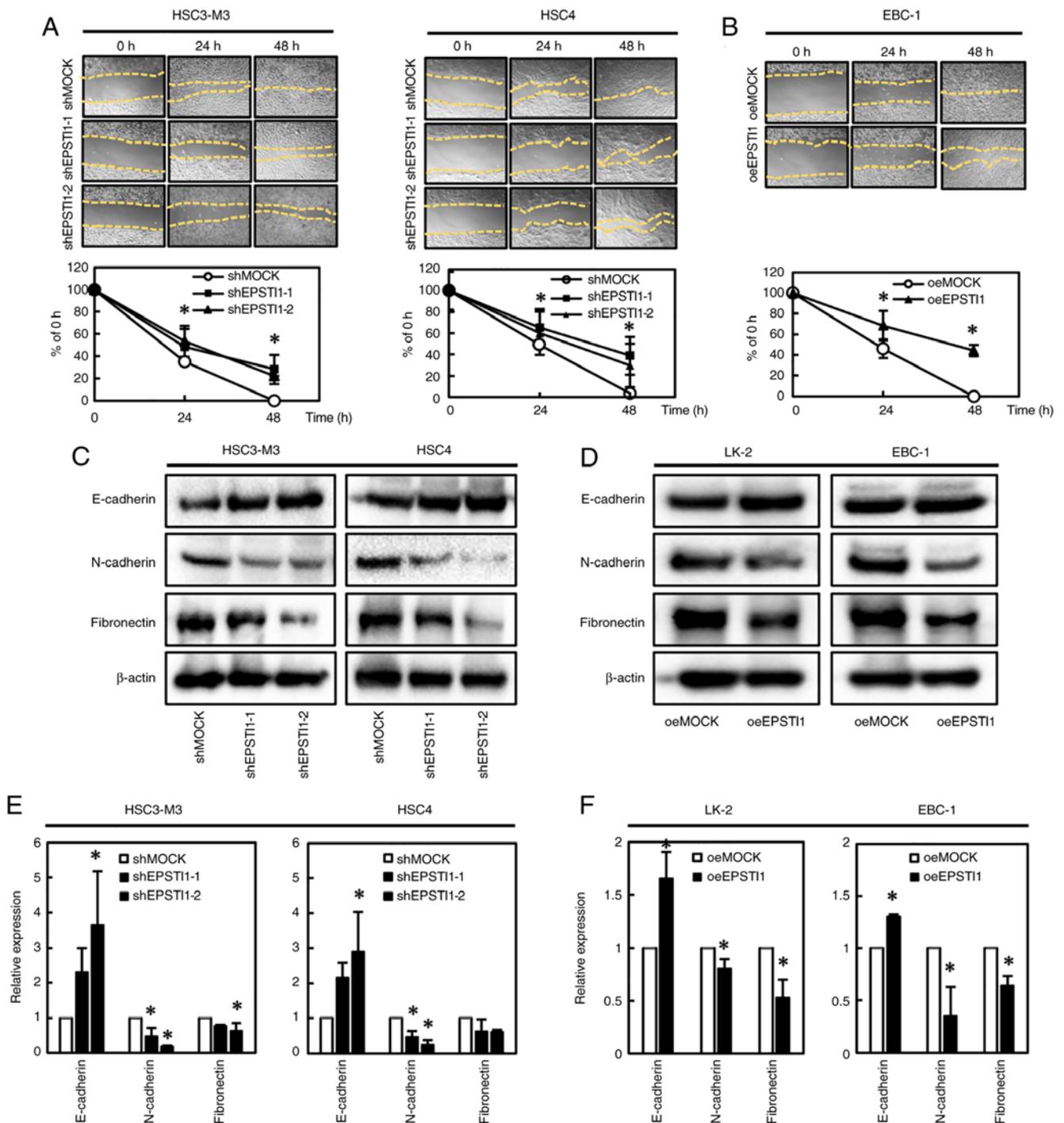


Figure 5. Decreased cell migration capability and reversal of the EMT phenotype in tumor cells with modified *EPSTII* expression. (A) Downregulation of *EPSTII* in OSCC and (B) upregulation of the gene in LSCC were associated with significant suppression of the cell migration ability, suggesting reversal of the EMT phenotype. Accordingly, (C and E) downregulation of *EPSTII* in OSCC and (D and F) upregulation of the gene in LSCC were also associated with significantly increased expression levels of E-cadherin and significantly suppressed expression levels of N-cadherin and fibronectin with exception in fibronectin in HSC4 which exhibited marginal statistical significance ($P=0.08$), again suggesting reversal of the EMT phenotype. The cell migration assay in LSCC was performed using only EBC-1, but not LK-2. Each experiment was performed in triplicate and repeated three times, and data are presented as the means \pm SE. * $P<0.05$. EMT, epithelial-mesenchymal transition; *EPSTII*, epithelial-stromal interaction 1; OSCC, oral squamous cell carcinoma; SE, standard error.

In the present study, *EPSTII* was demonstrated to enhance the malignant phenotype in OSCC, similar to breast (13,14,18,19) and colorectal cancers (16); notably, the gene played the opposite role in the LCSS cell lines. The gene expression in clinical samples obtained from the Oncomine database revealed higher expression levels of the gene in cancer

tissues as compared to normal tissues (26). Unfortunately, relevant data could not be obtained in the Oncomine database regarding *EPSTII* expression in LSCC (27). Although the contrasting roles of the same gene in different cancers, i.e., OSCC and LSCC, appear somewhat unusual, a few similar examples have been reported previously. Specifically, a T-box

Table I. Altered gene expression in OSCC with *EPSTII* knockdown as assessed by PCR array.

HSC3-M3		HSC4	
Gene symbol ^a (ratio ^b)		Gene symbol (ratio)	
Upregulated	Downregulated	Upregulated	Downregulated
<u>CA9</u> (7.2) ^c	<u>CCL2</u> (-3.4)	LDHA (79.9)	XIAP (-224.9)
SERPINF1 (4.2)	BIRC3 (-2.5)	MKI67 (15.0)	IGFBP3 (-38.5)
FGF2 (3.5)	SERPINB2 (-2.4)	SERPINB2 (14.7)	HMOX1 (-35.7)
TBX2 (3.0)		BMI1 (7.9)	<u>CCL2</u> (-26.5)
PGF (2.9)		PINX1 (5.8)	TBX2 (-8.3)
DDIT3 (2.1)		DKC1 (4.3)	DDIT3 (-7.3)
		<u>CA9</u> (3.9)	FLT1 (-5.8)
		WEE1 (2.5)	SOX10 (-5.8)
		LIG4 (2.4)	TEK (-5.8)
		AURKA (2.4)	EPO (-5.4)
		APAF1 (2.3)	IGFBP5 (-5.4)
		VEGFC (2.3)	LPL (-5.2)
		MAPK14 (2.1)	PPP1R15A (-4.6)
		CDH2 (2.1)	GADD45G (-3.4)
		CDC20 (2.0)	COX5A (-3.3)
			CASP7 (-3.1)
			GSC (-2.9)
			G6PD (-2.3)
			ANGPT2 (-2.2)
			KDR (-2.2)
			CASP9 (-2.2)
			TINF2 (-2.1)
			SNAI1 (-2.1)

^aGenes up- or downregulated by more than 2-fold are listed. ^bEach experiment was carried out in triplicate, and the mean ratios are presented in parentheses. ^cGene symbols in bold font and underlined are those that were common to the 2 cell lines. EPSTI, epithelial-stromal interaction 1; OSCC, oral squamous cell carcinoma; PCR, polymerase chain reaction.

family member, *TBX2*, was reported to be amplified and overexpressed in breast cancer (28) and its overexpression has been revealed to be associated with a high pathological grade in prostatic cancer (29); conversely, the gene has been revealed to be downregulated in non-small cell lung cancer (30); although further details have not yet been elucidated. Therefore, it is considered that the present study findings, provide further insight into understanding the multi-functional roles of *EPSTII*.

To elucidate the mechanisms underlying these phenomena, a series of PCR array assays were performed. Upregulation of the *CA9* gene, observed in both the OSCC cell lines with forced downregulation of *EPSTII* expression, has been revealed to augment the cell proliferation and transformation ability under hypoxic conditions, under the regulation of the transcription factor HIF-1 α (31). Downregulation of the *CCL2* gene, a member of the chemokine superfamily, observed in the same cell lines, has been demonstrated to promote cancer progression through recruiting inflammatory cells in multiple cancers (32-34), and to promote EMT-associated cell migration ability in OSCC (35). Conversely, downregulation

of *CCND2*, *BIRC3*, and *IGFBP7*, observed in both the *EPSTII*-overexpressing LSCC cell lines, has been revealed to promote G1/S phase transition (*CCND2*) (36), suppress apoptosis by inhibiting caspase-3 (*BIRC3*) (37), and to promote TGF- β -induced EMT in human renal proximal tubular epithelial cells (*IGFBP7*) (38). These studies, except for the information concerning *CA9*, are all consistent with the phenomena observed in the OSCC and LSCC cell lines in the present study. There were other inconsistencies in the PCR array results, in addition to the case of *CA9*. Although western blotting revealed downregulation of N-cadherin in the HSC4 cell line with *EPSTII* knockdown, the PCR array disclosed upregulation of *CDH2* in the same cell line. Since the results of western blot analysis are likely to be more reliable, comprehensive gene analysis by PCR array may be susceptible to error. *MKI67*, as well as *CA9*, revealed to be upregulated in the HSC4 cells with *EPSTII* knockdown, reportedly augment cell proliferation, whereas knockdown of *EPSTII* in the HSC4 suppressed cell proliferation in the present research. The finding that there were no commonly altered gene expression between the OSCC and LSCC cell lines may indicate that

Table II. Altered gene expression in LSCC with *EPSTII* overexpression, as assessed by PCR array.

LK-2		EBC-1	
Gene symbol ^a (ratio ^b)		Gene symbol (ratio)	
Upregulated	Downregulated	Upregulated	Downregulated
TBXT2 (28.0)	<u>CCND2</u> (-17.3) ^c	HMOX1 (9.5)	CDC20 (-14.5)
LPL (16.2)	SNAI2 (-11.3)	DDIT3 (9.5)	MKI67 (-11.1)
IGFBP5 (5.3)	KRT14 (-6.2)	XIAP (3.4)	<u>IGFBP7</u> (-10.7)
CDH2 (3.9)	SERPINB2 (-4.1)	LIG4 (3.4)	LPL (-4.7)
KDR (3.7)	SLC2A1 (-2.4)	CASP9 (3.0)	STMN1 (-4.6)
HGDC (3.1)	<u>BIRC3</u> (-2.3)	ARNT (2.9)	TEK (-4.0)
SERPINF1 (3.1)	<u>IGFBP7</u> (-2.2)	ERCC5 (2.7)	CCL2 (-3.7)
GSC (2.6)	ACSL4 (-2.1)	IGFBP3 (2.7)	AURKA (-3.6)
GADD45G (2.4)		PPP1R15A (2.6)	<u>CCND3</u> (-2.9)
SOD1 (2.2)		SOX10 (2.2)	LDHA (-2.8)
SNAL1 (2.1)		TNKS (2.2)	CCND2 (-2.7)
ANGPT1 (2.1)		PINX1 (2.1)	PGF (-2.6)
		OCLN (2.0)	<u>BIRC3</u> (-2.3)
			SKP2 (-2.1)

^aGenes up- or downregulated by more than 2-fold are listed. ^bEach experiment was carried out in triplicate, and the mean ratios are presented in parentheses. ^cGene symbols in bold font and underlined are those that were common to the 2 cell lines. EPSTI, epithelial-stromal interaction 1; LSCC, lung squamous cell carcinoma; PCR, polymerase chain reaction.

EPSTII acts differently in the two cancers, which is also consistent with the findings of the present study.

The present study had some limitations. The results were obtained solely from studies using cell lines. The findings remain to be validated in tumor samples collected from patients with OSCC and LSCC. Expression of *EPSTII* in normal tissue and cancer tissue according to the patient characteristics, including the clinical stage of the cancer, tumor doubling time, and sensitivity to chemotherapy, are expected to further clarify the clinical relevance of *EPSTII* in these diseases. Although the comprehensive analyses with a PCR array were solely exploratory, the findings failed to explain mechanisms involved in the contrasting functions of the gene. New studies to elucidate the clinical relevance of the gene and to explore the mechanisms underlying the diverse functions of the gene according to the cancer type are warranted.

In conclusion, the present research revealed, for the first time, the opposite functions of *EPSTII*, namely, of promoting and suppressing cancer progression, according to the type of cancer; i.e., its overexpression promoted the malignant phenotype in OSCC, whereas its downregulation promoted the malignant phenotype in LSCC.

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Availability of data and materials

Data supporting the findings of the present study are available upon reasonable request from the corresponding author.

Authors' contributions

MF, MA, KU, MS, NK, HT and YT contributed to the design of the study. Cell culture and data acquisition were performed by MF, TC and RF. MF, MA, and AT performed statistical analyses, and interpretation of data was performed by all the authors. MF, MA and YT contributed to drafting the manuscript. All authors contributed to revising the manuscript critically for important intellectual content, as well as reading and approving the final version of the manuscript to be published.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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