

RESEARCH ARTICLE

Open Access

Thyroid hormone increases fibroblast growth factor receptor expression and disrupts cell mechanics in the developing organ of corti

Katherine B Szarama^{1,2,3,5*}, Núria Gavara^{2,6}, Ronald S Petralia⁴, Richard S Chadwick² and Matthew W Kelley¹

Abstract

Background: Thyroid hormones regulate growth and development. However, the molecular mechanisms by which thyroid hormone regulates cell structural development are not fully understood. The mammalian cochlea is an intriguing system to examine these mechanisms, as cellular structure plays a key role in tissue development, and thyroid hormone is required for the maturation of the cochlea in the first postnatal week.

Results: In hypothyroid conditions, we found disruptions in sensory outer hair cell morphology and fewer microtubules in non-sensory supporting pillar cells. To test the functional consequences of these cytoskeletal defects on cell mechanics, we combined atomic force microscopy with live cell imaging. Hypothyroidism stiffened outer hair cells and supporting pillar cells, but pillar cells ultimately showed reduced cell stiffness, in part from a lack of microtubules. Analyses of changes in transcription and protein phosphorylation suggest that hypothyroidism prolonged expression of fibroblast growth factor receptors, and decreased phosphorylated Cofilin.

Conclusions: These findings demonstrate that thyroid hormones may be involved in coordinating the processes that regulate cytoskeletal dynamics and suggest that manipulating thyroid hormone sensitivity might provide insight into the relationship between cytoskeletal formation and developing cell mechanical properties.

Keywords: Young's modulus, Hair cell, Pillar cell, Hypothyroid, Cell mechanics

Background

The cytoskeleton plays a key role in modulating the morphological changes of cells and tissues, impacting both the resistance to deformation and the exertion of force by the cell [1,2]. In the central nervous system, thyroid hormones are thought to act as master regulators of tissue growth and cytoskeletal development [3,4]. Thyroid hormone action can be mediated through traditional genomic mechanisms such as the regulation of transcription [5,6] in association with co-activators and co-repressors [7]. However, a direct transcriptional regulation of cytoskeletal elements by thyroid hormone receptors has not been described. This suggests that

thyroid hormones might act indirectly by controlling intermediary signaling cascades, which could coordinate the regulatory components of cytoskeletal formation [8,9].

The sensory epithelium of the mammalian inner ear is an excellent model system to understand basic questions of cytoskeletal organization, as structural elements, such as actin and microtubules, facilitate the organization and architecture of this tissue. The auditory epithelium of the inner ear contains one row of inner hair cells and three rows of outer hair cells (OHCs), which detect and amplify sound, and several types of non-sensory supporting cells, which withstand the mechanical stress of sound vibrations. In particular, supporting pillar cells (PCs), which form the fluid-filled tunnel of Corti [10], have a highly organized cytoskeletal network composed primarily of microtubules [11,12]. Disruptions in the formation of this tunnel have been shown to negatively impact hearing function [13-15]. Therefore, it is imperative that we understand the regulatory cues that facilitate normal structural development.

* Correspondence: Katherine.Szarama@stjude.org

¹Section on Developmental Neuroscience, Laboratory of Cochlear Development, National Institute on Deafness and other Communication Disorders, National Institutes of Health, Bethesda, MD, USA

²Section on Auditory Mechanics, Laboratory of Cellular Biology, National Institute on Deafness and other Communication Disorders, National Institutes of Health, Bethesda, MD, USA

Full list of author information is available at the end of the article

There is reason to suggest that thyroid hormone signaling has an important role in development of the cochlea. Both the sensory and supporting cells in this epithelium have unique expression patterns of thyroid hormone receptors [16,17], and deiodinase enzymes [18,19], which control the availability of thyroid hormone. More recently, thyroid hormone transporters have also been localized to this sensory epithelium [20], and the developmental expression profile contributes some control over the accessibility of ligand to the cell-specific targets. Consistent with this hypothesis, deletion of all known thyroid hormone receptors leads to hearing impairment and inner ear cell structural defects, including malformation of cells in the organ of Corti, and a collapsed tunnel of Corti [14]. However, the factors that mediate development of the inner ear structures are largely unknown. In the cochlea, cytoskeletal formation, including the actin-based cuticular plate in OHCs [11] and the dense 15-prot filament microtubule network in PCs [21] that is mostly acetylated [12], follows cell differentiation [10-12]. However, the signaling pathways mediating cell structural development are not well understood.

Interestingly, mutations in some components of the Fibroblast growth factor (Fgf) signaling pathway cause inner ear structural defects that appear similar to those of the hypothyroid phenotype. In particular, mutations in Fgf receptor 3 (Fgfr3) lead to disruptions in tissue structure, including a collapsed tunnel of Corti, and cause auditory defects in both mice and humans [13,22-24]. These disruptions have been attributed to both a lack of differentiation of PCs and malformation in PC microtubule development [23]. However, there must also be some regulation of Fgf-receptor expression. One possibility that has been observed in other organ systems is the impact of thyroid hormone regulation of the Fgf-signaling pathway. For example, thyroid hormone stimulates Fgfr expression in undifferentiated cartilage [25,26]. Furthermore, a thyroid hormone response element is present in the promoter region of Fgfr1 [27]. Together, these data suggest that Fgf-signaling could act as an intermediary between thyroid hormone signaling and cytoskeletal development, and motivate further examination of thyroid hormone action specifically in the cochlea.

In this study, we examined the mechanism of thyroid hormone action on inner ear structural development. We found that hypothyroidism led to higher mRNA expression of Fgf-receptors relative to controls, leading to a delay in the down-regulation of Fgfr3. Hypothyroidism also led to delayed OHC and PC differentiation, which may be mediated in part by the aberrant expression of Fgf-receptors. Finally, hypothyroidism disrupted OHC and supporting PC structure, and aberrantly stiffened

embryonic and early postnatal epithelial cells. Here, we show that the hypothyroid-induced cell stiffening may be mediated in part by the disrupted phosphorylation of Cofilin, which has the potential to alter actin dynamics.

Results

Thyroid hormone levels regulate Fgfr expression in the cochlea

To determine if Fgf-signaling is modulated by thyroid hormones, we examined expression of two Fgf-receptors, *Fgfr1* and *Fgfr3*, in cochleae from mice with altered thyroid hormone levels. To create a hyperthyroid condition, cochlear explant cultures were established at embryonic day 14 (E14) and treated with triiodothyronine (T3) for 2 or 4 days beginning after 24 hours *in vitro*. Quantitative RT-PCR (qPCR) data showed a two-fold decrease in *Fgfr1* and *Fgfr3* mRNA levels at both time points (Figure 1A). As endogenous thyroid hormone depletion takes considerably longer than thyroid hormone stimulation in culture [28], we used pharmacological agents to deplete thyroid hormones during development *in vivo*. Pregnant female mice were rendered hypothyroid with a combination of methimazole in the drinking water and low iodine feed, which has previously been shown to induce hypothyroidism in the cochlea *in utero* [29]. To probe for changes in Fgf-receptors, mRNA was extracted from inner ear sensory epithelia. Results from qPCR indicated significant increases in both *Fgfr1* and *Fgfr3* mRNA levels under hypothyroid conditions at both embryonic day 16 (E16) and postnatal day 0 (P0) (Figure 1B). However, by postnatal day 3 (P3), while *Fgfr1* expression was six-fold greater in hypothyroid conditions relative to controls, *Fgfr3* expression was not significantly different from controls (Figure 1B). These data are consistent with negative regulation of Fgfr expression by thyroid hormones within the developing inner ear.

In order to localize the increase in expression of these receptors in the cochlea, we examined Fgfr mRNA expression by *in situ* hybridization in control and hypothyroid cochleae. At P0, *Fgfr1* is localized to cell populations in both the greater epithelial ridge, a collection of cells medial to the sensory epithelium that gives rise to the inner sulcus [30], and the lesser epithelial ridge, which is located lateral to the sensory epithelium and gives rise to the spiral ligament [31] (Figure 1C). In hypothyroid conditions, expression of *Fgfr1* appeared more intense at P0 relative to controls at basal and apical regions of the cochlear duct (Figure 1C). Expansion of the expression domain of *Fgfr1* was not observed. In contrast with *Fgfr1*, *Fgfr3* is normally initially expressed broadly within the sensory epithelium in progenitors that will give rise to both hair cells and supporting cells [32], but by P0 is down-regulated in sensory OHCs of the more mature basal

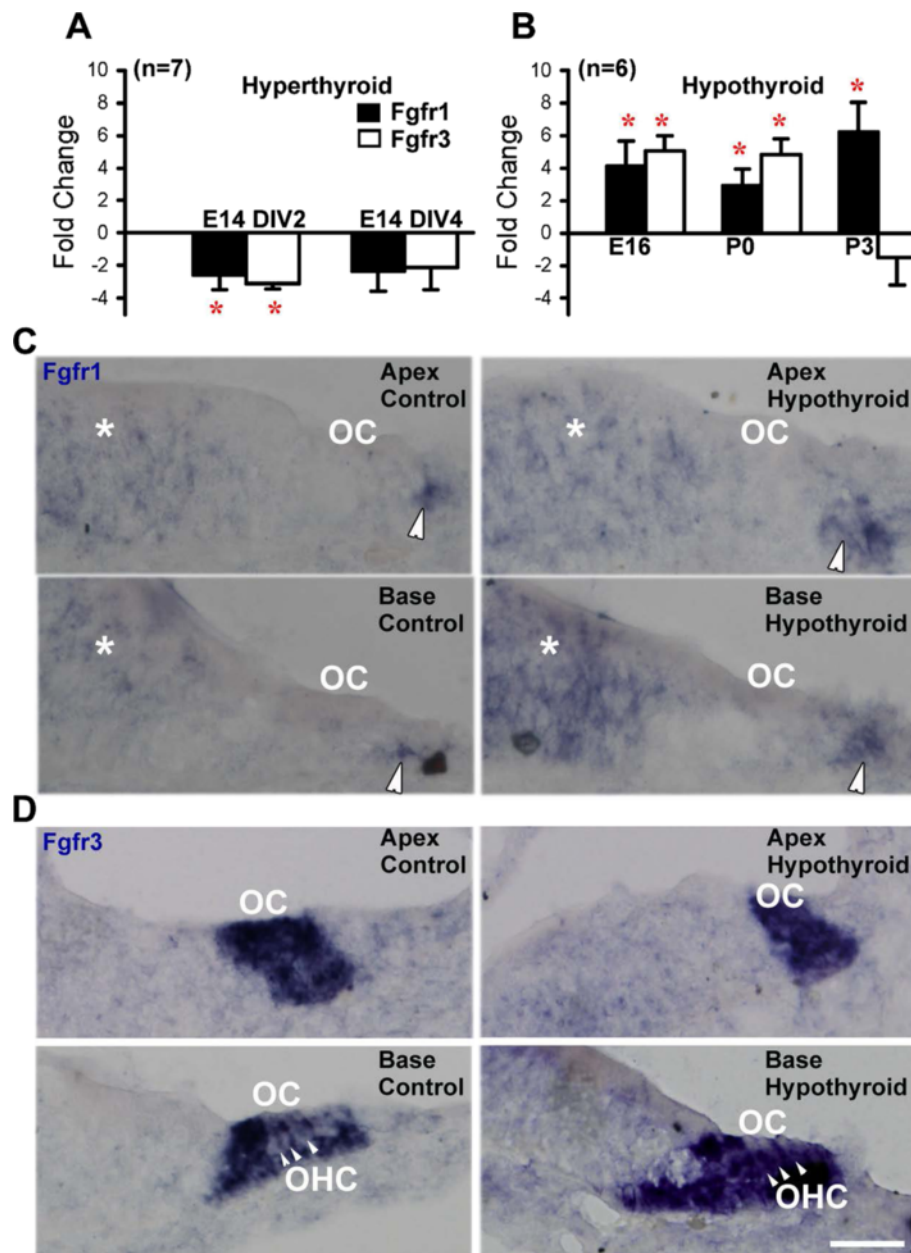


Figure 1 Thyroid hormone levels regulate Fgf-receptor expression in the cochlea. **(A)** QPCR results show significantly decreased *Fgfr1* (black) and *Fgfr3* (white) expression upon treatment with thyroid hormone *in vitro* (*p-value < 0.05). **(B)** QPCR expression of *Fgfr1* and *Fgfr3* is significantly increased when thyroid hormone levels are decreased *in vivo* (*p-value < 0.05). **(C)** *In situ* hybridization for *Fgfr1* mRNA at P0 shows increased expression in the greater (asterisk) and lesser (arrowhead) epithelial ridges in hypothyroid cochleae (right) relative to control (left). **(D)** *Fgfr3* mRNA levels at P0 persist in hypothyroid OHCs (right, arrowheads) relative to controls at the base of the cochlea (left, arrowheads). Scale bar, 20 μ m, applies to all images. OC, organ of Corti; OHC, outer hair cell.

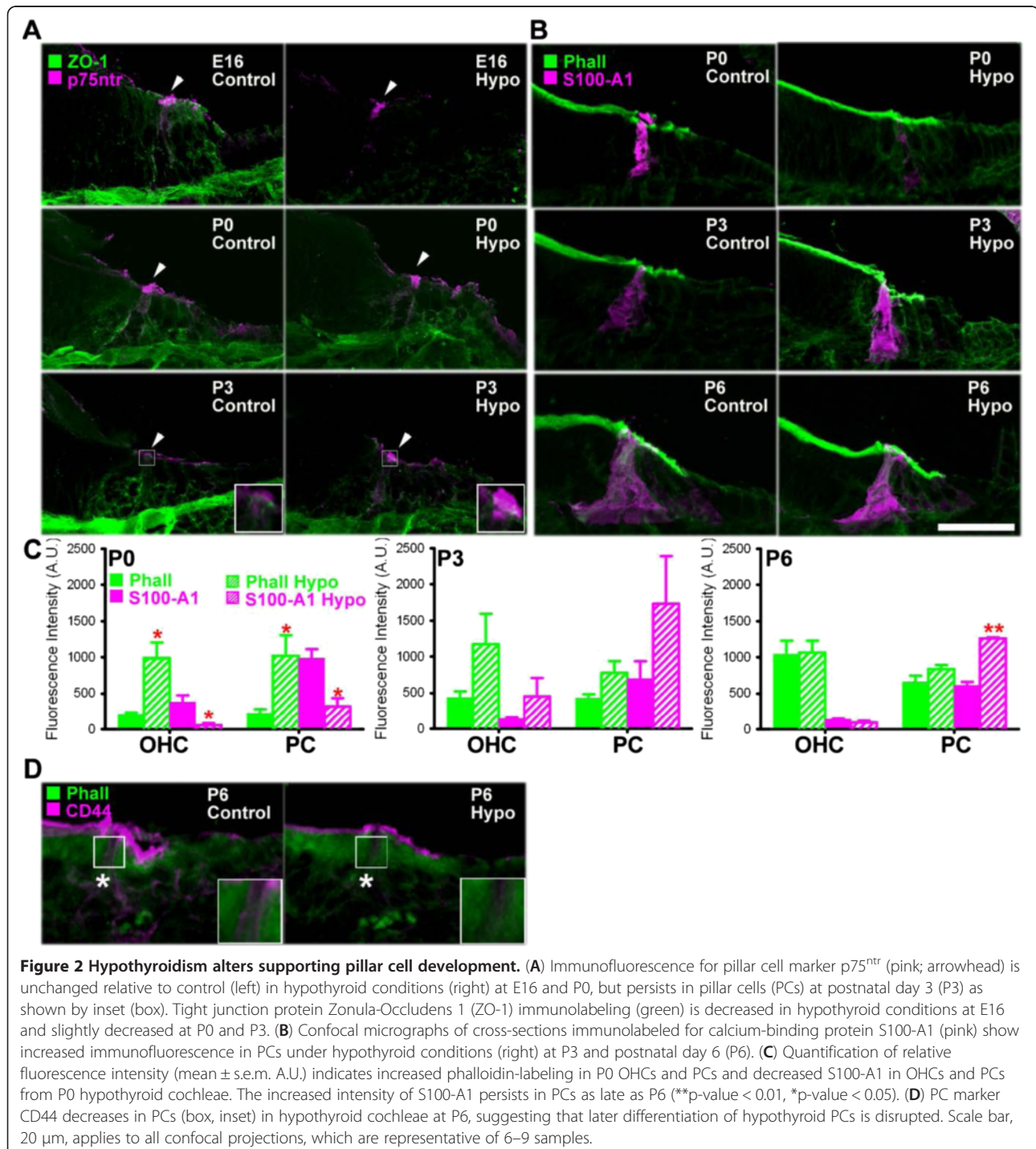
region of the cochlea and maintained in non-sensory supporting pillar and Deiter's cells (Figure 1D). At later developmental time points, *Fgfr3* expression in the apex resembles expression in the base [33]. In hypothyroid conditions, *Fgfr3* expression in the base persisted in OHCs at P0, indicating a delay in down-regulation (Figure 1D). Taken together, these results

show that hypothyroidism leads to a delay in development of the inner ear, and suggest that there may also be a delay in differentiation of both OHCs and PCs at postnatal stages.

As mentioned in the introduction, both gain and loss of function mutations in *Fgfr3* lead to deafness [24] and prolonged Fgf-signaling delays PC development [23,32].

Since hypothyroidism leads to a delay in down-regulation of *Fgfr3*, we hypothesized that there may be delays in the maturation of this epithelium. One hallmark of differentiation is the formation and stabilization of junctional complexes, which in the cochlea are composed of adherens and tight junction proteins [34], including Zonula-Occludens 1 (ZO-1). In particular, ZO-1 was recently characterized in this epithelium to be

associated with maturation of OHC apical structure [35]. Therefore, we examined the downstream effect of delayed maturation on immunofluorescence intensity of labeled ZO-1 in control and hypothyroid cochleae. Hypothyroid cochleae showed decreased immunolabeling of ZO-1 relative to control conditions and this was most apparent at E16 (Figure 2A), suggesting both a delay in differentiation and disruption to cell structure.



To look more specifically at supporting cell maturation, expression of neurotrophin receptor p75 (p75^{ntf}), an early marker of developing PCs [36], was examined by immunohistochemistry in cochlear cross-sections. P75^{ntf} is initially expressed in the cochlea at embryonic stages of development [37], and is down-regulated postnatally, which allows for specific labeling of early developing PCs. Confocal images revealed that the onset of p75^{ntf} expression was not delayed at E16 or P0 (Figure 2A). In contrast, at P3, p75^{ntf} expression in the base of the cochlea, which would normally be down-regulated, persisted in hypothyroid samples (Figure 2A), consistent with a delay in maturation of this supporting cell type. Together, these results suggest that cochlear development is delayed in the late embryonic period and PC development is delayed at early postnatal time points.

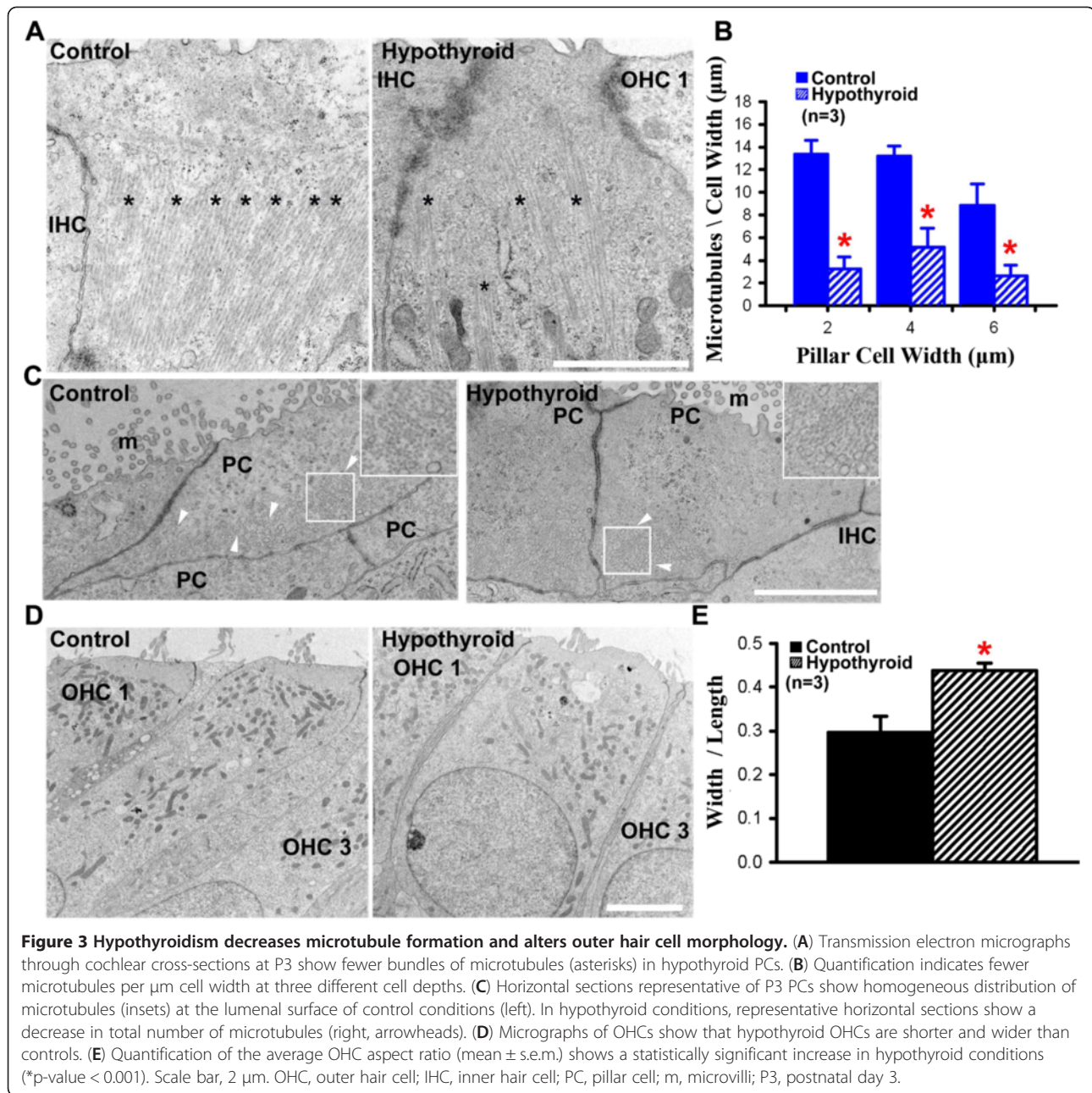
To further assess postnatal PC development, expression of S100-A1 protein was labeled and compared between cochleae in hypothyroid and control conditions. S100 proteins are initially highly enriched throughout the cochlear duct at embryonic stages [38], and are later down-regulated in differentiated PCs after the first postnatal week. While S100-A1 immunofluorescence intensity appeared to be lower in hypothyroid than in control conditions at P0, by P3, S100-A1 immunofluorescence appeared higher in hypothyroid than in control conditions (Figure 2B), which is consistent with the developmental delay observed in hypothyroid cochleae [14]. To quantify this difference, measurements of relative fluorescence intensity (mean \pm s.e.m. A.U.) were calculated and compared between hypothyroid and control conditions. In PCs, S100-A1 intensity decreased from 987 ± 119 A.U. in control conditions to 319 ± 116 A.U. in hypothyroid conditions at P0, as was the case for the OHCs (Figure 2C; p -value < 0.05). Later, at P6, expression of S100-A1 was significantly increased from 609 ± 48 A.U. in controls to 1257 ± 18 A.U. in hypothyroid conditions (Figure 2C; p -value < 0.01) showing that disruptions in these cells persist at later postnatal stages. Additionally, the relative fluorescence intensity of phalloidin in hypothyroid OHCs and PCs showed about a three-fold increase relative to control conditions (Figure 2C; p -value < 0.05).

Finally, to assess late postnatal differentiation of PCs, we examined the relative immunofluorescence of CD44, a protein that is restricted to supporting PCs at P0, increases in intensity through P7 [39] and is reported to be responsive to changes in thyroid hormone levels in the cerebellum [40]. Confocal micrographs of cochlear cross-sections at P6 showed that CD44 fluorescence appeared lower in hypothyroid conditions relative to controls (Figure 2D), suggesting that PCs maintain a sensitivity to thyroid hormone levels during postnatal development that may contribute to cell maturation

beyond the effects of *Fgfr3*. Taken together, these data suggest that hypothyroidism leads to a delay in the timing of PC maturation, as early markers of development that are normally down-regulated persist and late markers of development are delayed under experimental conditions. Additionally, the corresponding delay in down-regulation of *Fgfr* expression also suggests that some of these early effects are mediated through prolonged *Fgfr* expression.

Hypothyroidism disrupts microtubule formation in the cochlea

Previous studies found that hypothyroidism reduces the presence of tubulin in the developing central nervous system [41,42]. Furthermore, we observed a delay in the pattern of S100-A1 expression, which has been implicated in microtubule formation *in vitro* [8,43]. Therefore we asked if hypothyroidism also had consequences for the structural development of PCs and OHCs. To examine cellular structure, transmission electron micrographs were collected from cross-sections of control and hypothyroid cochleae. At P3, control PC cytoplasm contained microtubules that are oriented longitudinally from the luminal surface to the basilar membrane (Figure 3A). In contrast, PCs from hypothyroid cochleae contained fewer microtubules (Figure 3A). Quantitative analyses of the distribution of microtubules after controlling for differences in cell width were calculated and compared between control and hypothyroid conditions. Analysis revealed a significant decrease in the mean number of microtubules (mean \pm s.e.m.) per micron cell width in PCs in hypothyroid conditions (Figure 3B; p -value < 0.05). To confirm this result, horizontal sections from the luminal surface of the PCs were also examined and fewer microtubules were observed in hypothyroid relative to control conditions (Figure 3C). It is of note that the majority of microtubules at the luminal cell surface of PCs in hypothyroid conditions localized to the lateral edge of the PC, whereas there were microtubules observed throughout the cytoplasm of control PCs (Figure 3C). As there was also a significant increase in hypothyroid OHC phalloidin immunofluorescence (Figure 2C), we investigated whether hypothyroid conditions would also lead to structural defects in OHC development. Transmission electron micrographs indicated that hypothyroid OHCs were shorter and wider in comparison to controls at P3 (Figure 3D). To quantify this difference, we measured the average cell width and cell length (mean \pm s.e.m. μ m), and found a significant increase in OHC width: length ratio in hypothyroid cochleae (Figure 3E; p -value < 0.05). Overall, the data suggest that hypothyroidism leads to microtubule-based structural defects in supporting PCs and morphological defects in sensory OHCs.



In PCs, microtubules are further refined through post-translational modifications such as acetylation [12], which have consequences for both microtubule stability [9] and cell structure [44]. To investigate the effects of hypothyroidism on microtubule stability, immunohistochemistry of acetylated tubulin was performed on whole mount cochleae from control and hypothyroid conditions. Confocal image analysis showed that acetylated tubulin immunofluorescence, a marker for stable microtubules [45,46], was almost undetectable in the PC region at P0 in hypothyroid cochleae (Figure 4A). This was in contrast to control cochleae, which showed

strong immunolabeling of acetylated tubulin in PCs (Figure 4A). Similar results were observed by P3 with lowered overall fluorescence intensity levels of acetylated tubulin in hypothyroid PCs relative to control conditions (Figure 4B). OHCs at P3 also contained a microtubule-based kinocilium [47] that was identified by the enrichment of acetylated tubulin (Figure 4B). However, this structure was also observed under hypothyroid conditions (Figure 4B), suggesting that hypothyroidism does not delay formation or completely disrupt microtubules in OHCs. Finally at postnatal day 5 (P5), there was an absence of acetylated tubulin

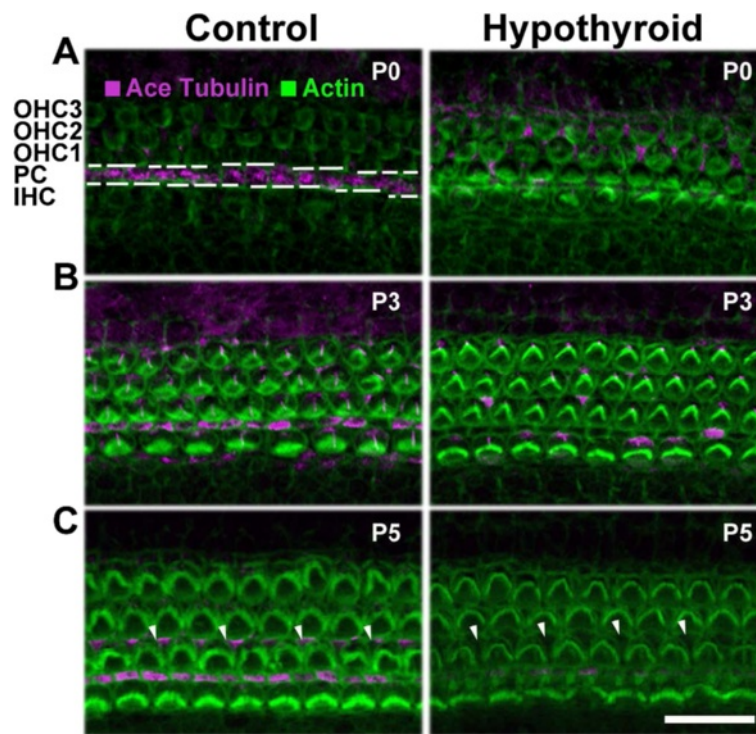


Figure 4 Acetylated tubulin decreases in hypothyroid cochleae. (A) Representative confocal projections of the luminal surface of the cochlear sensory epithelium from the apex of the cochlea at postnatal day 0 (P0). Acetylated tubulin immunofluorescence (pink) is decreased in the PC row (dashed lines) of hypothyroid cochleae. Also, intensity of actin immunolabeling (green) appears increased in hypothyroid conditions. (B) At postnatal day 3 (P3), acetylated tubulin immunolabeling is present in the hypothyroid PC row, but is decreased relative to control. Actin immunofluorescence appears similar between control and hypothyroid conditions. (C) By postnatal day 5 (P5), acetylated tubulin expression appears in supporting pillar and Deiter's cells of control cochleae, but is absent in hypothyroid conditions. Scale bar, 20 μ m, applies to representative confocal projections from 12 samples. OHC, outer hair cell; PC, pillar cell; IHC, inner hair cell.

compared to control in hypothyroid PCs and Deiter's cells (Figure 4C), a type of supporting cell with processes that interdigitate between OHCs. These results are consistent with a decrease in PC microtubule stability in response to hypothyroid conditions at all three time-points tested.

Hypothyroidism stiffens outer hair cells and supporting pillar cells

Previous cell and developmental biology studies suggest that the cytoskeleton plays a role in the control of cell shape [2] and, more recently, in dictating cell mechanical properties [1]. The decreased microtubule formation (Figure 3), and alterations in microtubule stability (Figure 4) of PCs, along with changes in morphology of OHCs (Figure 3) raised the question of whether mechanical properties of these cells might also be altered in hypothyroid conditions. To test this hypothesis, we measured the mechanical properties of these cells in culture with an atomic force microscope (Figure 5A). Average Young's modulus (mean \pm s.e.m kPa), a measure of the cellular resistance to deformation, was calculated and compared between hypothyroid and control

cochleae at late embryonic and early postnatal time points. We hypothesized that decreased cell structural development would lead to a decrease in Young's modulus. However, Young's modulus of OHCs actually showed a trend towards increased stiffness in hypothyroid conditions between E16 and P3 (Figure 5B) with this difference becoming statistically significant at P3 (p-value < 0.05). These results suggest that hypothyroid-related changes in cell morphology stiffen OHCs.

To examine the effects of hypothyroidism on the mechanical properties of developing PCs, we also calculated average Young's modulus for PCs in control and hypothyroid conditions. We found that in controls, PC Young's modulus was approximately five-fold higher between E16 and P5 (Figure 5C). This corresponds to increased microtubule acetylation [48] and suggests that increased cytoskeletal formation leads to cell stiffening. In contrast with controls, hypothyroid PCs were significantly stiffer at E16 and P0 (Figure 5C; p-value < 0.05). However by P5, hypothyroid PCs showed significantly reduced Young's modulus relative to controls (Figure 5C; p-value < 0.05). In summary, these results indicate that hypothyroidism leads to a

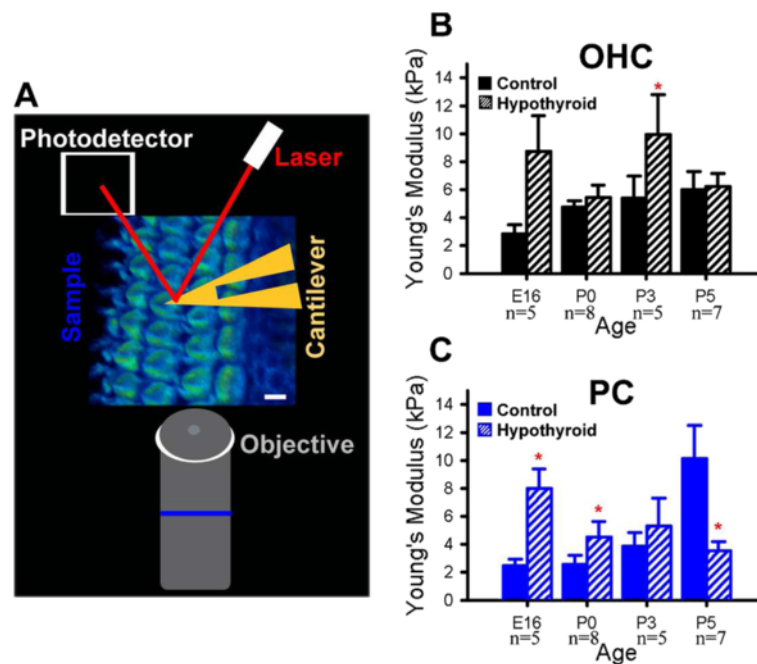


Figure 5 Hypothyroidism stiffens outer hair cells and supporting pillar cells. (A) Schematic of Atomic Force Microscopy (AFM) experiments. Organ of Corti explants were probed to determine surface mechanical properties of outer hair cells (OHCs) and supporting pillar cells (PCs) with an AFM that measures the relative movement of a laser reflected from a cantilever probe onto a photodiode detector. (B) Average Young's modulus (mean \pm s.e.m. kPa) was calculated for control and hypothyroid OHCs. OHCs in hypothyroid cochleae show a trend towards greater stiffness that is significant relative to control conditions at P3 (*p-value < 0.05). (C) Average Young's modulus is significantly increased in hypothyroid PCs at E16 and P0. However, hypothyroid PCs have significantly reduced Young's modulus by P5 relative to control conditions.

stiffening of OHCs and PCs early in development, and that at later time points hypothyroid PCs fail to develop mature mechanical properties.

Actin is responsible for the aberrant increase in pillar cell and outer hair cell stiffness

The decreased stiffness of hypothyroid PCs at P5 is consistent with the observed decrease in microtubules (Figure 3) and lack of microtubule stability (Figure 4). However, the hypothyroid PC stiffening at E16 and P0 and OHC stiffening at P3 were unexpected. One possible explanation for this effect could be the increase in filamentous actin in response to decreased thyroid hormone signaling, as observed by the increased phalloidin fluorescence intensity (Figure 2C). If actin mediates cell mechanical properties, then disruptions to the actin cytoskeleton should lead to decreased cell stiffness in hypothyroid conditions. To test this hypothesis, cochlear explants from control and hypothyroid animals were treated with Latrunculin A, a cell-permeant toxin [49] that sequesters actin monomers [50] and depolymerizes actin filaments [49]. Consistent with this mechanism, cochleae treated at P0 with Latrunculin A showed decreased phalloidin

immunofluorescence (Figure 6A). In controls, a 30 minute incubation with Latrunculin A resulted in a significant decrease in the average Young's modulus (mean \pm s.e.m. kPa) of OHCs from 6.06 ± 0.64 kPa to 4.20 ± 0.31 kPa, while PCs in the same samples were not significantly different after treatment (Figure 6B; p-value < 0.01). This suggests that the disruption to OHC actin and reduced OHC stiffness does not also significantly disrupt PC Young's modulus. However, PCs maintained in hypothyroid conditions and treated with Latrunculin A showed a significant decrease in Young's modulus at both P0, from 4.93 ± 1.11 kPa untreated to 1.07 ± 0.08 kPa after treatment, and at P3, from 9.83 ± 1.02 kPa untreated to 2.48 ± 0.50 kPa after treatment (Figure 6D; p-value < 0.001). OHCs maintained in hypothyroid conditions also showed a decreased Young's modulus in response to Latrunculin A administration, which is similar to results observed in euthyroid conditions. However, the magnitude of the decreased OHC Young's modulus was much greater than in controls (Figure 6C; p-value < 0.05), suggesting a greater sensitivity to actin disruptions in hypothyroid conditions. These results suggest that PC surface mechanical properties in hypothyroid cochleae at E16 and P0 are mediated through changes in actin.

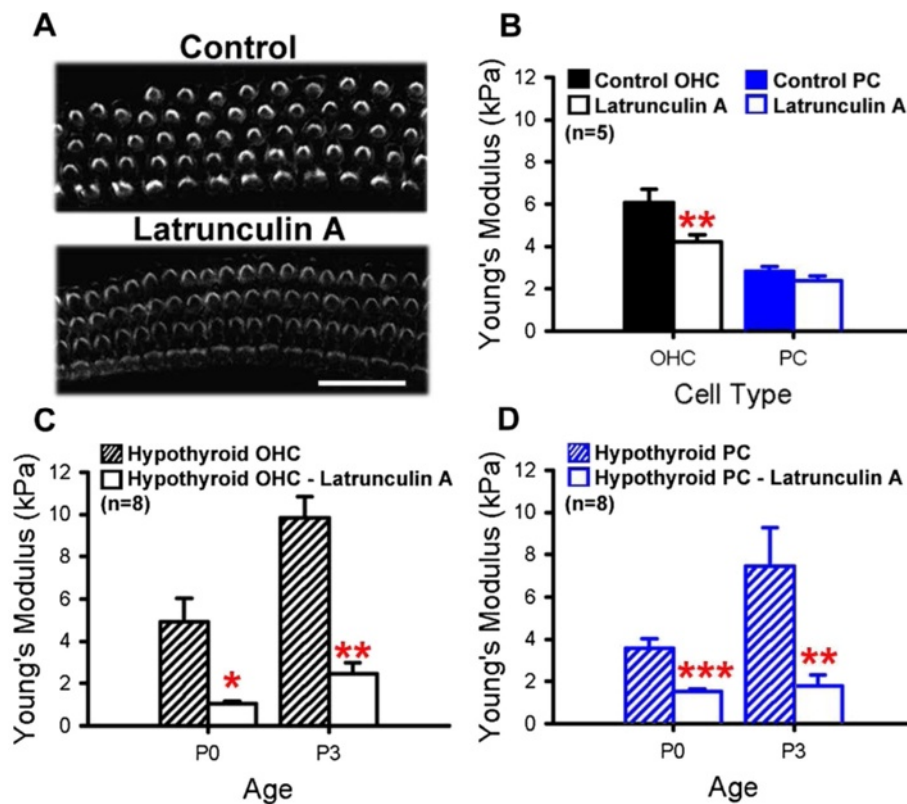


Figure 6 Hypothyroidism increases sensitivity of supporting cells to actin disruption. (A) Representative confocal z-projections of whole mount explants treated at P0 with Latrunculin A show decreased phalloidin immunofluorescence relative to vehicle control. (B) In controls, average Young's modulus (mean \pm s.e.m. kPa) of outer hair cells (OHCs, black) is significantly decreased with Latrunculin A treatment, while pillar cells (PCs, blue) are not significantly different from Latrunculin A treatment at P0 (**p-value < 0.01). (C) Hypothyroid OHCs also have a decreased Young's modulus in treated conditions relative to vehicle control (*p-value < 0.05). (D) In contrast with controls, in which Latrunculin A treatment did not affect PC Young's modulus, hypothyroid PCs have a significant decrease in stiffness when treated with Latrunculin A (***p-value < 0.001).

Cofilin activity mediates changes in actin dynamics under hypothyroid conditions

The results presented above suggested a relationship in which thyroid hormone levels might be acting through Fgfr-signaling to induce actin-based stiffening of hypothyroid OHCs and PCs. Therefore, we explored possible direct regulation of actin-modulating proteins in response to activation of the Fgf-signaling pathway. In particular, receptor tyrosine kinases have the potential to mediate actin dynamics by controlling the phosphorylation of Cofilin [51]. Cofilin, when not phosphorylated by phosphorylated-Lim-domain kinase (p-LIMK), can increase the turnover of actin leading to increased polymerization of actin filaments [52-54]. To examine the potential role of this pathway, total protein was isolated from control and hypothyroid cochleae and then probed with antibodies against phosphorylated and total Cofilin protein. At E16, hypothyroid cochleae appeared to have lower levels of p-Cofilin relative to control (Figure 7A). To quantify this difference, the average relative density (mean \pm s.e.m. A.U.) of labeled protein was calculated and compared between control

and hypothyroid conditions. Total Cofilin was not significantly different between control and hypothyroid conditions (Figure 7A). A similar pattern of p-Cofilin at E16 was also observed at P0, with p-Cofilin decreased from 2.38 ± 0.23 A.U. in control to 0.68 ± 0.15 A.U. in hypothyroid conditions (Figure 7B, p-value < 0.05). As was the case for E16, total protein levels of Cofilin were not significantly different between control and hypothyroid cochleae at P0 and P3. By postnatal day 3 (P3), the relative intensity and average relative density of p-Cofilin labeled proteins were now similar between control and treated conditions (Figure 7C). These results are consistent with the hypothesis that hypothyroidism leads to prolonged Fgfr expression and activation, which may have the effect of decreasing phosphorylation of Cofilin and, as a result, to increasing actin stability.

While the static localization of actin filaments has long been understood to maintain cell shape and membrane tension [55,56], the dynamics of actin treadmilling for cell maturation and mechanics are just beginning to be realized. In particular, the dynamic actin cytoskeleton is

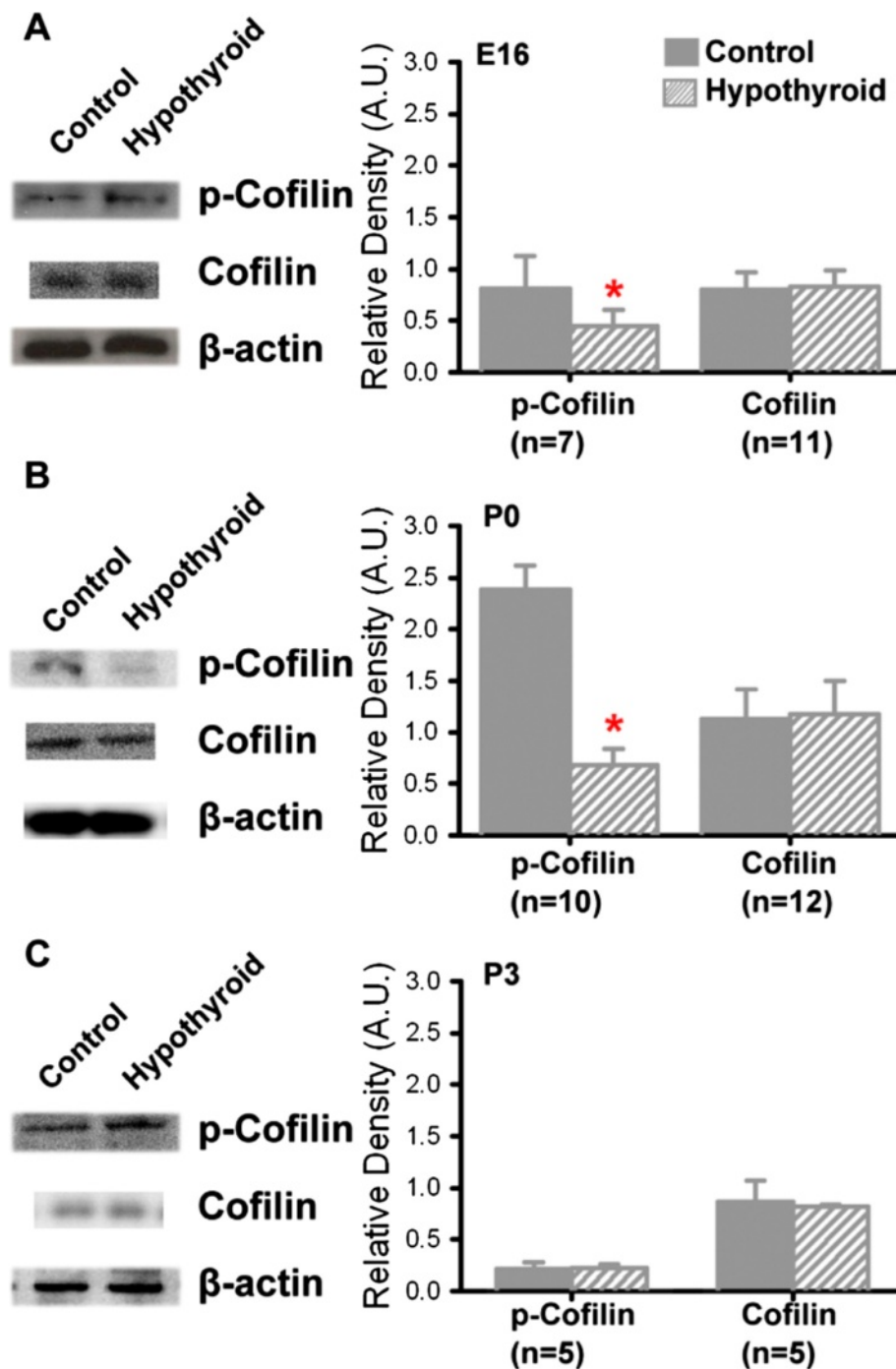


Figure 7 Hypothyroidism leads to decreased phosphorylated Cofilin. (A) Representative western blots for protein isolated from embryonic day 16 (E16) cochleae show decreased intensity of p-Cofilin in hypothyroid conditions relative to control. Total Cofilin appears to be unchanged. Average relative density (mean \pm s.e.m.) of phosphorylated, labeled proteins indicates decreased expression of p-Cofilin in hypothyroid cochleae. Average relative density of Cofilin is not significantly different between control and hypothyroid. (B) At postnatal day 0 (P0), intensity of p-Cofilin in hypothyroid conditions appears to be decreased relative to control conditions. Average relative density of p-Cofilin is significantly decreased in hypothyroid relative to control conditions (*p-value < 0.05). (C) By postnatal day 3 (P3), the relative intensity and average relative density of p-Cofilin labeled protein is unchanged between control and treated conditions. As for E16, total protein level of Cofilin is not significantly different between control and hypothyroid cochleae at P0 and P3.

primarily controlled by actin-binding proteins that regulate nucleation, polymerization and disassembly [57]. Specifically, the actin-depolymerizing factor, Cofilin, has been shown to be necessary for actin filament assembly [58,59]. The results presented above demonstrated decreased Cofilin phosphorylation during hypothyroidism. However, changes in specific cell types could not be determined. Therefore, in order to localize the observed changes in p-Cofilin in hypothyroid cochleae, the expression of p-Cofilin by immunofluorescence in cochlear cross-sections at E16, P0, and P3 was examined. At E16, expression of p-Cofilin was observed only in the nuclei of OHCs and cells of the greater epithelial ridge, but was present in both the cytoplasm and nuclei of supporting pillar cells and cells of the lesser epithelial ridge (Figure 8). This pattern of expression persisted in both basal and apical sections of control conditions through P3 (Figure 8). In hypothyroid conditions, p-Cofilin immunofluorescence appeared to be reduced in the sensory epithelium relative to controls at E16, P0, and P3 (Figure 8). In particular, a marked reduction in p-Cofilin was observed in PC cytoplasm and in OHC nuclei at P0 and P3 (Figure 8). These data support a role for lower Cofilin phosphorylation in the stabilization of actin in PCs under hypothyroid conditions. The localization of p-Cofilin in OHCs indicates that phosphorylation takes place predominantly in the nucleus, which suggests that Cofilin is active primarily in the dense actin meshwork of sensory OHCs, and may

explain in part the differences in OHC and PC responses to hypothyroidism.

Discussion

Effects of thyroid hormone signaling in the inner ear may be regulated through prolonged Fgf-signaling

Our data support previous findings that thyroid hormone levels coordinate the timing of development in many target tissues, including the skeletal [60] and central nervous systems [61,62]. However, the specific molecular pathways that mediate the effects of thyroid hormone are largely still unknown. Thyroid hormones primarily regulate transcriptional activity through thyroid hormone receptors (reviewed in [63]). Based on similarity in phenotypes and existing data from other systems, we examined Fibroblast growth factor receptors after altering thyroid hormone levels. In the organ of Corti, decreased levels of thyroid hormone were shown to result in prolonged expression of *Fgfr1* and *Fgfr3*, respectively (Figure 1), suggesting that thyroid hormone receptors negatively regulate Fgf-receptor expression in the developing cochlea. Interestingly, *Fgfr1* has been shown to accelerate differentiation when inactivated in differentiating osteoblasts [64] and a thyroid hormone response element has been identified in the promoter region of *Fgfr1* [27]. In the organ of Corti, it seems that *Fgfr3* regulates the timing of hair cell and supporting cell development as shown by experiments in which ectopic activation of *Fgfr3* delays

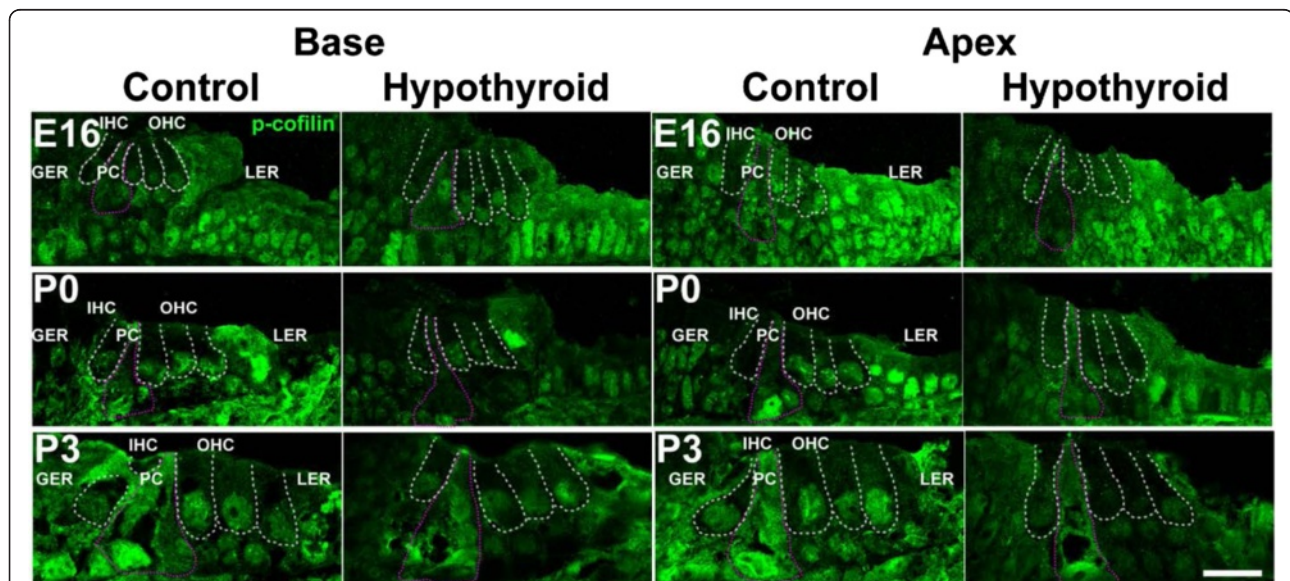


Figure 8 Phosphorylated Cofilin localizes to the cochlear sensory epithelium and is decreased in hypothyroid conditions. Representative cross-sections through the organ of Corti at the indicated time points. P-Cofilin is indicated by immunofluorescence in 12 μ m projections. Immunofluorescence is present in the nuclei of cells in the greater epithelial ridge (GER) and outer hair cells (OHCs, white dashed lines), as well as in the nuclei and cytoplasm of pillar cells (PCs, pink dotted lines) and cells of the lesser epithelial ridge (LER). P-Cofilin is decreased in OHCs and PCs of hypothyroid cochleae relative to control. Scale bar, 20 μ m, applies to all images. GER, greater epithelial ridge; IHC, inner hair cell; PC, pillar cell; OHC, outer hair cell; LER, lesser epithelial ridge.

differentiation of supporting cells [23,32]. The observation of similar delays in the down-regulation of p75^{NTR} and S100 proteins (Figure 2) in differentiating cells in hypothyroid conditions is consistent with the idea that the maintenance of Fgfr3 signaling mediates at least some of the effects observed in hypothyroidism of the cochlea. However, these data do not rule out an additional layer of regulation by thyroid hormones on Fgf-signaling. Indeed, thyroid hormone has been shown to play a role in heparin sulfate expression in the developing growth plate [65], which suggests an additional pathway, through which thyroid hormone could enhance Fgf-signaling. Overall, these findings suggest that the timing of down regulation of Fgf-signaling is important for the developing inner ear. However, the persistent expression of both Fgfr3 and of Fgf8, the most likely cognate ligand for Fgfr3 in the organ of Corti, into adult stages, suggests that Fgf-signaling may also have a continuous role in inner ear tissue morphology. While it seems reasonable to suggest that the mechanism to decrease Fgfr3-signaling would be to decrease the presence of ligand [66,67], we propose that thyroid hormones might also mediate Fgf-signaling at this stage of inner ear development by regulating the level of Fgf-receptor expression in the organ of Corti.

Using the drug methimazole, hypothyroidism was induced in utero to examine the cellular response to a lack of thyroid hormone in the organ of Corti. Thyroid hormone levels *in utero* are thought to be primarily from a maternal source and transmitted through the placenta [68,69]. Methimazole transmission has also been shown to cross the placenta [70] and to reduce the serum levels of thyroid hormone [71]. While the effectiveness of this drug has led to widespread use as a treatment in clinical cases of hyperthyroidism for adults and, in particular, during pregnancy [72], methimazole is not without side effects. In less than 1% of human subjects, agranulocytosis, a lowered white blood cell count, and liver toxicity resulting in jaundice were reported [73,74]. However, considering the relatively limited time course of methimazole treatment in these studies, it seems unlikely that the treatment led to these side effects.

Thyroid hormone may disrupt cytoskeletal formation through prolonged Fgf-signaling

A key observation of the experiments described here was the alteration in cytoskeletal formation in response to hypothyroidism. While disruption of the cytoskeleton in cases of hypothyroidism has long been known [3,7,42], the molecular basis for these effects is not yet fully understood. The results presented here suggest that Fgf signaling may act as an intermediary, with prolonged Fgf-signaling leading to disruptions in microtubule formation. In the case of microtubule structure,

we extend existing research by providing two complementary mechanisms for this hypothyroid-induced structural defect. First, deletion of *Fgfr3* leads to decreased β -tubulin [23], further supporting the relationship between Fgf-signaling and microtubule number. Second, the increase in S100-A1 in PCs is representative of elevated intracellular calcium levels [75,76], which could directly impair microtubule formation and is observed in *Fgfr3*-deficient mice [77].

In addition to the microtubule disruptions in PCs (Figures 3 and 4), there also appeared to be a higher fluorescence intensity of phalloidin-labeled actin in hypothyroid conditions relative to controls. Phalloidin, a member of the phallotoxin group of F-actin binding peptides, has been used to quantify the amount of F-actin in cells [78,79]. This probe has a diameter of 12–15 Å [80], which has the advantages of low steric hindrance to bind close to a 1:1 ratio [80] and therefore does not compete with actin binding proteins [81]. This probe also has a similar affinity for both large and small filaments [82], but does not bind monomeric G-actin. This is a limitation of the technique as it is not possible with only one probe to examine whether or not the ratio of F-actin-to-G-actin has been altered in hypothyroid relative to control conditions. However, the advantages of this probe support our conclusion that stronger phalloidin fluorescence intensity, combined with electron micrographs showing no apparent difference in cuticular plate thickness, suggests that there is a higher density of actin, which may cause in part measured stiffening at the luminal surface of the cochlea.

There is increasing evidence that Fgf-signaling mediates actin dynamics through signaling [83], which may be the key link between hypothyroidism and the observed cell structural defects. Indeed, Fgf-signaling is necessary for the dynamic changes in actin remodeling that lead to invagination of the developing otocyst [84]. In the hypothyroid cochlea, Fgf-receptors might mediate actin dynamics through activation of p21-activated kinases (PAKs) or ROCK [85,86] that can then phosphorylate LIMK to inhibit Cofilin activity via phosphorylation [87,88]. The prolonged Fgfr-expression shown in hypothyroid conditions coincides with a decrease in phosphorylation of Cofilin (Figure 7), which is consistent with the role for this protein in the rate of actin filament turnover. Since both increased actin filaments and decreased Cofilin phosphorylation were observed in hypothyroid cochleae, these results are consistent with this proposed mode of regulation. This might in part explain the disruption to OHC morphology (Figure 3), which is interesting given that recently it has been proposed in other developing tissues that the rearrangement of actin filaments generally would produce a disruption in cell morphology [89]. Additionally, disrupted

p75^{ntf} expression has also been shown to mediate actin dynamics through down-regulation of Rho-GTP signaling [90]. Since p75^{ntf} expression appeared to persist in hypothyroid conditions at P3 (Figure 2), the contribution of Egf-signaling may also contribute indirectly through a p75^{ntf}-dependent mechanism.

Alterations in cytoskeletal dynamics may affect cell mechanical properties

Ultimately, impaired functional ability of supporting PCs to resist deformation at P5 (Figure 5) is not exactly concomitant with the decrease in microtubule number observed in electron micrographs of hypothyroid pillar cells relative to controls. However, it is possible that the lack of acetylation observed at P5 (Figure 4), and not absolute microtubule number, contributes to the formation of the tunnel of Corti by the pillar cells. It is also possible that a lack of acetylation has other effects on microtubule length and polymerization [91,92]. Future studies directly disrupting acetylation, while also controlling for microtubule length and protofilament number, will add important *in vitro* data that can be brought back into microtubule-based cell structural assays to further support what has been shown here.

In the case of actin structure, the findings that hypothyroid OHCs and PCs are aberrantly stiffer than controls (Figure 5) and are increasingly sensitive to Latrunculin A (Figure 6) implicate the actin cytoskeleton, rather than microtubule development, in hypothyroid-induced cell stiffening observed from E16 to P3. In this study, atomic force microscopy (AFM) has been used to probe cytoskeletal structures in living cells without membrane disruption or fixation. While the cell membrane is an integral part of stabilizing the cytoskeleton, previous results have shown that membrane stiffness is negligible in eukaryotic cells relative to cytoskeletal stiffness [93,94]. Furthermore, with increasing indentation depth—in this study indentation was 1.5 μm —the bulk of Young's modulus has been shown to be the result of cytoskeletal stiffness [48], and in the case of the developing cochlea, reflects the stiffness of the actin mesh at the cell apex. Indeed, by combining AFM with live-cell fluorescence imaging techniques, it is possible to get single cell specificity. For example, after treatment with the cytoskeletal disrupting agent Latrunculin A, the OHC but not the neighboring PC Young's modulus was decreased in cochlear explant cultures. Overall, these AFM data suggest that the calculated Young's modulus reflects cell surface mechanical properties, which are dominated by cytoskeletal components.

Cell-specific responses to thyroid hormone may lend insight into cell structural development

While data presented here were only able to measure the consequences of disrupted actin dynamics in hair

cells and supporting PCs, disruptions to actin dynamics could have broad consequences for many supporting cell types in the developing organ of Corti. For example, the greater epithelial ridge is composed of columnar epithelial cells that undergo structural changes leading to the formation of the fluid filled space known as the inner sulcus [95]. While we know that these cells support synapse formation of sensory inner hair cells in the organ of Corti [96,97], the aberrant cell cytoplasm in this region may explain part of the pathology of hypothyroidism. By localizing phosphorylated Cofilin expression in control and hypothyroid cochlear cross-sections (Figure 8), we observed not only the aberrant defect in the sensory epithelium, but also the cell-specific differences in phosphorylated cofilin expression throughout the cochlear duct. Cofilin phosphorylation has also been observed to mediate actin dynamics in a number of developing systems [98,99]. The relationship between hypothyroidism and cell-structural defects in the developing cochlea might prove to be the basis for additional studies of morphogenesis in developing systems that rely on coordination by thyroid hormones [60,62]. Future studies also examining actin dynamics under reduced thyroid hormone levels could lend insight into a mechanism by which developmental malformations to the structure and function of the organ of Corti may be prevented.

Conclusion

We find that hypothyroidism leads to a delay in the down-regulation of Egf-receptors and to a decrease in microtubule formation and acetylation at early postnatal stages. While the lack of microtubules ultimately reduce supporting pillar cell stiffness, we find that hypothyroidism actually stiffens outer hair cells at P3 and pillar cells at E16 and P0 as a result of increased F-actin. Our data also show that the increased dynamics of actin in these cells might be the result of hypothyroid-induced Fgf signaling and a decreased phosphorylation of Cofilin. Together, these data implicate the sensitivity of cell structural development to thyroid hormones, and suggest that thyroid hormone signaling might coordinate the time-course of tissue morphogenesis.

Methods

Cochlear explant cultures and pharmacological treatments

Cochleae of Institute for Cancer Research (ICR) mice (Charles River Laboratories, Frederick, Maryland, USA) were cultured at specific stages between E16 and P5 as previously described [76] and plated on No.1 glass coverslips (Corning, New York, USA). To increase thyroid hormone signaling, explants were treated with either 5 μM triiodothyronine or 5 μM reverse triiodothyronine

(Sigma, St. Louis, Missouri, USA), an inactive form of thyroid hormone, as a control, in DMSO. To decrease thyroid hormone signaling, animals were treated with 0.02% methimazole (Sigma) and 10% sucrose in drinking water and low iodine feed administered *ad libitum*. All animal care and procedures were approved by the Animal Care and Use Committee at NIH and complied with the NIH guidelines for the care and use of animals.

Transmission electron microscopy

Cochleae from 3 control and 3 hypothyroid animals at P3 were isolated and placed immediately in 0.1 M phosphate buffer (pH 7.4) containing 4% paraformaldehyde and 2% glutaraldehyde for 30 minutes at room temperature followed by 2 hours at 4°C. The inner ears were then washed in phosphate buffer and cacodylate buffer, post-fixed with 1% osmium tetroxide, and dehydrated through a graded alcohol series before being embedded in Poly/BED 812 resin (Polysciences Inc., Warrington, Pennsylvania, USA) as previously described [48]. Thin sections of about 75 nm were cut using a Leica Reichert ultramicrotome, stained with lead citrate, mounted on 200-Cu mesh grids, and examined at room temperature using a JEOL transmission electron microscope (Akishima, Tokyo, Japan) at 80 kV with 15,000× magnification. Images were acquired using a Hamamatsu Camera (Hamamatsu Photonics K.K., Japan) and Advanced Microscopy Techniques Camera System software version 534.4 (Woburn, Massachusetts, USA). Images were cropped in Adobe Photoshop CS4 (Adobe). To quantify morphological changes in OHCs and PCs, transverse sections from three animals for each condition were analyzed with ImageJ [100] analysis software for the presence of microtubules at distances 2, 4, and 6 μm from the luminal surface of PCs. Measurements of OHC length and width were also made. Analysis of statistical significance was determined using student's T-test.

Immunohistochemistry and confocal image analysis

Samples of the same time point were harvested on the same day under the same conditions, fixed in 4% paraformaldehyde for 4 hours, rinsed, passed through an increasing sucrose gradient, and embedded in OCT (Sakura, New York, USA) in parallel. All sections to be examined from the same time point for comparison were cryosectioned at 12 μm thickness on the same day. Sections were processed in parallel with the same stock solutions, which included permeabilization with 0.2% Tween-20 in PBS, blocking with 10% normal horse serum and incubation overnight in primary antibody (p75^{ntr}, Covance, Princeton, New Jersey, USA, 1:1000; ZO-1, Millipore, 1:1000, S100-A1, Neomarkers, Kalamazoo, Michigan, USA, 1:500; CD44, BD Pharmingen, Franklin Lakes, New Jersey, USA, 1:200; p-Cofilin, Santa Cruz Biotechnology, Santa Cruz, California, USA, 1:500) at

4°C. Primary antibodies were detected using either Alexa Fluor 488 or 546 (Invitrogen, 1:1000) conjugated secondary antibodies. Directly conjugated Phalloidin 633 (Invitrogen, 1:5000) was applied to all samples. Cochleae prepared for whole mount immunohistochemistry were fixed for 2 hours and were labeled with a primary antibody against acetylated tubulin (Sigma, 1:750). Samples were mounted in Fluoromount-G (Southern Biotech, Birmingham, Alabama, USA), using No. 1½ glass coverslips (Corning, New York, USA) adhered with nail polish. All fluorescence images were acquired at room temperature with LSM 510 acquisition software as 12 μm Z-stacks with 1 μm optical sectioning using a Zeiss 510 LSM Confocal Microscope with 40X oil objective [1.3 numerical aperture (NA); Plan-Neofluar]. Using line scan analysis, fluorescence intensity was measured from projected Z-stacks with ImageJ [100] analysis software in 2 μm areas at the luminal surfaces of both PCs and OHCs, as this region was previously observed with Transmission Electron Microscopy to maintain a homogeneous cell cytoplasm from E16 through P5 [48]. Average fluorescence intensity (mean ± s.e.m. Arbitrary Units) was calculated from 6 samples, and compared using student's T-test.

In situ hybridization

Cochleae from 6 animals per condition—hypothyroid or control—were harvested on the same day at the same time point and fixed in a stock solution of 4% paraformaldehyde in 1X PBS overnight. Samples were then rinsed, passed through an increasing sucrose gradient, and embedded in OCT (Sakura, New York, USA) in parallel using the same solutions under the same conditions. Samples were cryosectioned in parallel at 12 μm thickness. All steps of the in situ hybridization were carried out in parallel on 6 sections from 6 animals per condition as previously described [93] with probes specific to *Fgfr1* and *Fgfr3*. All samples had the same exposure to the same reaction mixture, as previously described [101] for the same length of time.

Quantitative real-time polymerase chain reaction (qPCR)

Cochleae were removed from the temporal bone, and the surrounding mesenchyme, scala vestibuli and scala tympani were removed. 6–8 cochleae were pooled and total RNA was isolated using RNAqueous (Ambion, Grand Island, New York, USA) reagents. cDNA was synthesized from 500 ng total RNA for each condition using a Superscript III first strand synthesis kit (Invitrogen). Amplification was performed with SYBR Green (Applied Biosystems, Foster City, California, USA). Amplification of all mRNA was performed on an ABI Prism 7000 (Applied Biosystems) with the following cycling conditions: 40 cycles of 95°C for 15 seconds, and 60°C for 1 minute.

To calculate fold change, gene expression was normalized to glyceraldehyde 3-phosphate dehydrogenase (GAPDH) and statistical significance was confirmed with student's T-test. Primer sequences used are as follows: Fgfr1, F 3'-ATGGTTGACCGTTCTGGAAG-5'; Fgfr1, R 3'-AGAAAAGGGTACGCAGCAGA-5'; Fgfr3, F 3'-GAGACTTGGCTGCCAGAAAC-5'; Fgfr3, R 3'-GGGCTCACATTTGTGGTCTT-5', GAPDH, F 3'-ATCCTGTAGGCCAGGTCATG-5'; GAPDH, R 3'-TATGCCCGAGGACAATAAGG-5'.

Western blot analysis

Cochlear explants were freshly isolated before total protein extraction. Proteins were extracted from 8–10 cochleae in RIPA buffer containing complete mini protease inhibitor cocktail (Roche), complete phosphatase inhibitor cocktail (Roche), 1 mM Na₃VO₄, and 500 mM NaF. Protein was quantified using the Dc protein assay kit (Bio Rad) and Lowry method using a ND-1000 Spectrophotometer (Nanodrop, Wilmington, Delaware, USA). 25 µg total protein per condition was loaded onto 4-12% SDS-PAGE gels (Invitrogen) run for 2 hours at 120 V, transferred to nitrocellulose membrane (Invitrogen), and run for 3 hours at 80 V at 4°C. Membranes were blocked in 0.05% TBS-T containing BLOTTO (Rockland, Gilbertsville, Pennsylvania, USA) and 1% BSA (Sigma). Primary antibodies were incubated in blocking solution at the following concentrations: p-Cofilin (1:500, Abcam); Cofilin (1:5000, Abcam). Primary antibodies were conjugated to horseradish peroxidase anti-rabbit secondary antibody (1:5000, Amersham) and detected using ECL Detection Reagents (Amersham). Membranes were visualized using Image Station 4000R (KODAK) and Carestream Molecular Imaging Software (New Haven, Connecticut, USA). Image analysis was performed with ImageJ [100] using the Gel Analysis plug-in method to calculate relative density. Relative density of phosphorylated protein signal was normalized to β-actin (Sigma, 1:5000) signal for the same tissue and under the same experimental conditions. Statistical significance was determined with Welch's T-test [102].

Atomic force microscopy and live cell imaging

Experiments were performed using a Bioscope II and Bioscope Catalyst AFM (Bruker, Santa Barbara, California, USA) head mounted onto a Zeiss Axiovert 200 inverted microscope and controlled via a Nanoscope V controller. Pyramidal shaped, gold-coated, silicon nitride cantilever probes with 0.03 N/m spring constant (Bruker) were used for all measurements. Cells were identified after being loaded with 500 nM Calcein AM vital dye (Invitrogen) in Leibovitz's media (Invitrogen) for 30 minutes and rinsed with fresh Leibovitz's media. Contact mode AFM was applied to all samples in Leibovitz's

media at 1.5 µm maximum indentation using 3 µm ramps at 1 Hz continuous force ramping to collect 3 force-distance curves for the center of each identified hair cell or supporting cell. All force-distance curves were analyzed with custom analysis software in MATLAB (Mathworks, Natick, Massachusetts, USA) and fit to the Sneddon model [103] to measure Young's modulus, which is a material property of the cellular resistance to deformation and was calculated with the formula $F = (2/\pi \tan \alpha)(E/(1-\nu^2))\delta^2$; where F is applied force, ν is Poisson's ratio and assumed to be 0.5, δ is cantilever indentation, and α is cantilever tip angle. Average Young's modulus (mean ± s.e.m. kPa) was calculated from the average of sample measurements of 10 cells within a given region of interest such as the base or the apex of the cochlea. Statistical analyses were performed using Welch's T-test [102].

Abbreviations

OHCs: Outer hair cells; PCs: Pillar cells; Fgf: Fibroblast growth factor; Fgfr3: Fibroblast growth factor receptor 3; P75^{NTR}: Neurotrophin receptor p75; E16: Embryonic day 16; P0: Postnatal day 0; kPa: Kilo-Pascal; LIMK: Lim-domain-kinase; ROCK: Rho-associated protein kinase.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

KBS designed the experiments and acquired the data, KBS and RSP performed EM experiments, NG wrote software to analyze AFM data. KBS analyzed the data and performed statistical analyses. KBS, RSP, NG, RSC, and MWK contributed to interpretation of data. KBS drafted the manuscript and RSP, NG, RSC and MWK revised it critically for important intellectual content. All authors read and approved the final manuscript.

Acknowledgements

We thank Ya-Xian Wang for help with tissue preparation for TEM, and S. Raft, D.S. Sharlin, and A. Fridberger for comments on earlier versions of this manuscript. This research was supported by the National Institute on Deafness and Other Communication Disorders (NIDCD) Intramural Research Program [DC000059 to M.W.K., DC0003333 to R.S.C.] and in part with an Intramural Fellowship to Promote Diversity, from the NIH Office of the Director to K.B.S.

Author details

¹Section on Developmental Neuroscience, Laboratory of Cochlear Development, National Institute on Deafness and other Communication Disorders, National Institutes of Health, Bethesda, MD, USA. ²Section on Auditory Mechanics, Laboratory of Cellular Biology, National Institute on Deafness and other Communication Disorders, National Institutes of Health, Bethesda, MD, USA. ³Center for Hearing and Communication Research and Department of Clinical Science, Intervention, and Technology, Karolinska Institutet, Stockholm, Sweden. ⁴Advanced Imaging Core, National Institute on Deafness and other Communication Disorders, National Institutes of Health, Bethesda, MD, USA. ⁵Current address: Laboratory of Joseph Opferman, Department of Biochemistry, St. Jude Children's Research Hospital, 262 Danny Thomas Place, Memphis, TN 38105-3678, USA. ⁶Current address: Drittes Physikalisches Institut, Georg-August-Universität, Göttingen, Germany.

Received: 22 August 2012 Accepted: 29 January 2013

Published: 9 February 2013

References

1. Huang S, Ingber DE: The structural and mechanical complexity of cell-growth control. *Nat Cell Biol* 1999, **1**:E131–E138.

2. Mammoto A, Ingber DE: **Cytoskeletal control of growth and cell fate switching.** *Curr Opin Cell Biol* 2009, **21**:864–870.
3. Bernal J: **Thyroid hormones and brain development.** *Vitam Horm* 2005, **71**:95–122.
4. Koibuchi N, Jingu H, Iwasaki T, Chin WW: **Current perspectives on the role of thyroid hormone in growth and development of the cerebellum.** *Cerebellum* 2003, **2**:279–289.
5. Shepard AR, Eberhardt NL: **Molecular mechanisms of thyroid hormone action.** *Clin Lab Med* 1993, **13**:531–541.
6. Rosenfeld MG, Glass CK: **Coregulator codes of transcriptional regulation by nuclear receptors.** *J Biol Chem* 2001, **276**:36865–36868.
7. Thompson CC, Potter GB: **Thyroid hormone action in neural development.** *Cereb Cortex* 2000, **10**:939–945.
8. Donato R: **Calcium-independent, pH-regulated effects of S-100 proteins on assembly-disassembly of brain microtubule protein in vitro.** *J Biol Chem* 1988, **5**:106–110.
9. Gelfand VI, Bershadsky AD: **Microtubule dynamics: mechanism, regulation, and function.** *Annu Rev Cell Biol* 1991, **7**:93–116.
10. Ito M, Spicer SS, Schulte BA: **Cytological changes related to maturation of the organ of Corti and opening of Corti's tunnel.** *Hear Res* 1995, **88**:107–123.
11. Raphael Y, Athey BD, Wang Y, Lee MK, Altshuler RA: **F-actin, tubulin, and spectrin in the organ of Corti: comparative distribution in different cell types and mammalian species.** *Hear Res* 1994, **1**:173–187.
12. Tannenbaum J, Slepecky NB: **Localization of microtubules containing posttranslationally modified tubulin in cochlear epithelial cells during development.** *Cell Motil Cytoskeleton* 1997, **38**:146–162.
13. Colvin JS, Bohne BA, Harding GW, McEwen DG, Ornitz DM: **Skeletal overgrowth and deafness in mice lacking fibroblast growth factor receptor 3.** *Nat Genet* 1996, **12**:390–397.
14. Rusch A, Ng L, Goodyear R, Oliver D, Lisoukov I, Vennstrom B, Richardson G, Kelley MW, Forrest D: **Retardation of cochlear maturation and impaired hair cell function caused by deletion of all known thyroid hormone receptors.** *J Neurosci* 2001, **21**:9792–9800.
15. Inoshita A, Iizuka T, Okamura HO, Minekawa A, Kojima K, Furukawa M, Kusunoki T, Ikeda K: **Postnatal development of the organ of Corti in dominant-negative Gjb2 transgenic mice.** *Neuroscience* 2008, **156**:1039–1047.
16. Bradley DJ, Towle HC, Young WS 3rd: **Alpha and beta thyroid hormone receptor (TR) gene expression during auditory neurogenesis: evidence for TR isoform-specific transcriptional regulation in vivo.** *Proc Natl Acad Sci USA* 1994, **18**:439–443.
17. Lautermann J, ten Cate WJ: **Postnatal expression of the alpha-thyroid hormone receptor in rat cochlea.** *Hear Res* 1997, **107**:23–28.
18. Ng L, Goodyear RJ, Woods CA, Schneider MJ, Diamond E, Richardson GP, Kelley MW, Germain DL, Galton VA, Forrest D: **Hearing loss and retarded cochlear development in mice lacking type 2 iodothyronine deiodinase.** *Proc Natl Acad Sci USA* 2004, **101**:3474–3479.
19. Ng L, Hernandez A, He W, Ren T, Srinivas M, Ma M, Galton VA, St Germain DL, Forrest D: **A protective role for type 3 deiodinase, a thyroid hormone-inactivating enzyme, in cochlear development and auditory function.** *Endocrinology* 2009, **150**:1952–1960.
20. Sharlin DS, Visser TJ, Forrest D: **Developmental and cell-specific expression of thyroid hormone transporters in the mouse cochlea.** *Endocrinology* 2011, **152**:5053–5064.
21. Tucker JB, Paton CC, Richardson GP, Mogensen MM, Russell IJ: **A cell surface-associated centrosomal layer of microtubule-organizing material in the inner pillar cell of the mouse cochlea.** *J Cell Sci* 1992, **102**:215–226.
22. Hayashi T, Cunningham D, Bermingham-McDonogh O: **Loss of Fgfr3 leads to excess hair cell development in the mouse organ of Corti.** *Dev Dyn* 2007, **236**:525–533.
23. Puligilla C, Feng F, Ishikawa K, Bertuzzi S, Dabdoub A, Griffith AJ, Fritzsche B, Kelley MW: **Disruption of fibroblast growth factor receptor 3 signaling results in defects in cellular differentiation, neuronal patterning, and hearing impairment.** *Dev Dyn* 2007, **236**:1905–1917.
24. Mansour SL, Twigg SR, Freeland RM, Wall SA, Li C, Wilkie AO: **Hearing loss in a mouse model of Muenke syndrome.** *Hum Mol Genet* 2009, **18**:43–50.
25. Stevens DA, Harvey CB, Scott AJ, O'Shea PJ, Barnard JC, Williams AJ, Brady G, Samarut J, Chassande O, Williams GR: **Thyroid hormone activates fibroblast growth factor receptor-1 in bone.** *Mol Endocrinol* 2003, **17**:1751–1766.
26. Barnard JC, Williams AJ, Rabier B, Chassande O, Samarut J, Cheng SY, Bassett JH, Williams GR: **Thyroid hormones regulate fibroblast growth factor receptor signaling during chondrogenesis.** *Endocrinology* 2005, **146**:5568–5580.
27. O'Shea PJ, Guigon CJ, Williams GR, Cheng SY: **Regulation of fibroblast growth factor receptor-1 (FGFR1) by thyroid hormone: identification of thyroid hormone response element in the murine Fgfr1 promoter.** *Endocrinology* 2007, **148**:5966–5976.
28. Dezonne RS, Stipursky J, Gomes FC: **Effect of thyroid hormone depletion on cultured murine cerebral cortex astrocytes.** *Neurosci Lett* 2009, **467**:58–62.
29. Knipper M, Zinn C, Maier H, Praetorius M, Rohbock K, Kopschall I, Zimmermann U: **Thyroid hormone deficiency before the onset of hearing causes irreversible damage to peripheral and central auditory systems.** *J Neurophysiol* 2000, **83**:3101–3112.
30. Hinojosa R: **A note on development of Corti's organ.** *Acta Otolaryngol* 1977, **84**:238–251.
31. Chole RA, Tinling SP: **Bone lining cells of the mammalian cochlea.** *Hear Res* 1994, **75**:233–243.
32. Mueller KL, Jacques BE, Kelley MW: **Fibroblast growth factor signaling regulates pillar cell development in the organ of Corti.** *J Neurosci* 2002, **22**:9368–9377.
33. Hayashi T, Ray CA, Younkins C, Bermingham-McDonogh O: **Expression patterns of FGF receptors in the developing mammalian cochlea.** *Dev Dyn* 2010, **239**:1019–1026.
34. Nunes FD, Lopez LN, Lin HW, Davies C, Azevedo RB, Gow A, Kachar B: **Distinct subdomain organization and molecular composition of a tight junction with adherens junction features.** *J Cell Sci* 2006, **119**:4819–4827.
35. Etournay R, Lepelletier L, Boutet de Monvel J, Michel V, Cayet N, Leibovici M, Weil D, Foucher I, Hardelin JP, Petit C: **Cochlear outer hair cells undergo an apical circumference remodeling constrained by the hair bundle shape.** *Development* 2010, **137**:1373–1383.
36. Sano H, Mukai J, Monoo K, Close LG, Sato TA: **Expression of p75NTR and its associated protein NADE in the rat cochlea.** *Laryngoscope* 2001, **111**:535–538.
37. Schecterson LC, Bothwell M: **Neurotrophin and neurotrophin receptor mRNA expression in the developing inner ear.** *Hear Res* 1994, **73**:92–100.
38. Buckiova D, Syka J: **Calbindin and S100 protein expression in the developing inner ear in mice.** *J Comp Neurol* 2009, **513**:469–482.
39. Hertzano R, Puligilla C, Chan SL, Timothy C, Depireux DA, Ahmed Z, Wolf J, Eisenman DJ, Friedman TB, Riazuddin S, et al: **CD44 is a marker for the outer pillar cells in the early postnatal mouse inner ear.** *J Assoc Res Otolaryngol* 2010, **11**:407–418.
40. Dong H, Yauk CL, Rowan-Carroll A, You SH, Zoeller RT, Lambert I, Wade MG: **Identification of thyroid hormone receptor binding sites and target genes using ChIP-on-chip in developing mouse cerebellum.** *PLoS One* 2009, **4**:e4610.
41. Aniello F, Couchie D, Grippo D, Nunez J: **Regulation of five tubulin isoforms by thyroid hormone during brain development.** *J Neurochem* 1991, **57**:1781–1786.
42. Lakshmy R, Srinivasarao P: **Effect of thiocyanate on microtubule assembly in rat brain during postnatal development.** *Int J Dev Neurosci* 1997, **15**:87–94.
43. Sorci G, Agneletti AL, Donato R: **Effects of S100A1 and S100B on microtubule stability. An in vitro study using triton-cytoskeletons from astrocyte and myoblast cell lines.** *Neuroscience* 2000, **99**:773–783.
44. Wloga D, Gaertig J: **Post-translational modifications of microtubules.** *J Cell Sci* 2010, **123**:3447–3455.
45. Gundersen GG, Khawaja S, Bulinski JC: **Generation of a stable, posttranslationally modified microtubule array is an early event in myogenic differentiation.** *J Cell Biol* 1989, **109**:2275–2288.
46. Takemura R, Okabe S, Umeyama T, Kanai Y, Cowan NJ, Hirokawa N: **Increased microtubule stability and alpha tubulin acetylation in cells transfected with microtubule-associated proteins MAP1B, MAP2, or tau.** *J Cell Sci* 1992, **103**:953–964.
47. Vago P, Ripoll C, Tournebize R, Lenoir M: **Distribution of actin and tubulin in outer hair cells isolated from developing rat cochlea: a quantitative study.** *Eur J Cell Biol* 1996, **69**:308–315.
48. Szarama KB, Gavara N, Petralia RS, Kelley MW, Chadwick RS: **Cytoskeletal changes in actin and microtubules underlie the developing surface mechanical properties of sensory and supporting cells in the mouse cochlea.** *Development* 2012, **139**:2187–2197.

49. Spector I, Shochet NR, Kashman Y, Groweiss A: **Latrunculins: novel marine toxins that disrupt microfilament organization in cultured cells.** *Science* 1983, **219**:493–495.
50. Coue M, Brenner SL, Spector I, Korn ED: **Inhibition of actin polymerization by latrunculin A.** *FEBS Lett* 1987, **213**:316–318.
51. Marcoux N, Vuori K: **EGF receptor activity is essential for adhesion-induced stress fiber formation and cofilin phosphorylation.** *Cell Signal* 2005, **17**:1449–1455.
52. Carlier MF, Laurent V, Santolini J, Melki R, Didry D, Xia GX, Hong Y, Chua NH, Pantaloni D: **Actin depolymerizing factor (ADF/cofilin) enhances the rate of filament turnover: implication in actin-based motility.** *J Cell Biol* 1997, **136**:1307–1322.
53. Lappalainen P, Drubin DG: **Cofilin promotes rapid actin filament turnover in vivo.** *Nature* 1997, **388**:78–82.
54. Ichetovkin I, Grant W, Condeelis J: **Cofilin produces newly polymerized actin filaments that are preferred for dendritic nucleation by the Arp2/3 complex.** *Curr Biol* 2002, **12**:79–84.
55. Ingber DE: **Cellular tensegrity: defining new rules of biological design that govern the cytoskeleton.** *J Cell Sci* 1993, **104**:613–627.
56. Luo L: **Actin cytoskeleton regulation in neuronal morphogenesis and structural plasticity.** *Annu Rev Cell Dev Biol* 2002, **18**:601–635.
57. Pak CW, Flynn KC, Bamberg JR: **Actin-binding proteins take the reins in growth cones.** *Nat Rev Neurosci* 2008, **9**:136–147.
58. Chen Q, Pollard TD: **Actin filament severing by cofilin is more important for assembly than constriction of the cytokinetic contractile ring.** *J Cell Biol* 2011, **195**:485–498.
59. Wiggan O, Shaw AE, DeLuca JG, Bamberg JR: **ADF/cofilin regulates actomyosin assembly through competitive inhibition of myosin II binding to F-actin.** *Dev Cell* 2012, **22**:530–543.
60. Bassett JH, Williams GR: **The molecular actions of thyroid hormone in bone.** *Trends Endocrinol Metab* 2003, **14**:356–364.
61. Leonard JL, Farwell AP: **Thyroid hormone-regulated actin polymerization in brain.** *Thyroid* 1997, **7**:147–151.
62. Forrest D, Reh TA, Rusch A: **Neurodevelopmental control by thyroid hormone receptors.** *Curr Opin Neurobiol* 2002, **12**:49–56.
63. Harvey CB, Williams GR: **Mechanism of thyroid hormone action.** *Thyroid* 2002, **12**:441–446.
64. Jacob AL, Smith C, Partanen J, Ornitz DM: **Fibroblast growth factor receptor 1 signaling in the osteo-chondrogenic cell lineage regulates sequential steps of osteoblast maturation.** *Dev Biol* 2006, **15**:315–328.
65. Bassett JH, Swinhoe R, Chassande O, Samarut J, Williams GR: **Thyroid hormone regulates heparan sulfate proteoglycan expression in the growth plate.** *Endocrinology* 2006, **147**:295–305.
66. Pivola U, Ylikoski J, Trokovic R, Hebert JM, McConnell SK, Partanen J: **FGFR1 is required for the development of the auditory sensory epithelium.** *Neuron* 2002, **15**:671–680.
67. Shim K, Minowada G, Coling DE, Martin GR: **Sprouty2, a mouse deafness gene, regulates cell fate decisions in the auditory sensory epithelium by antagonizing FGF signaling.** *Dev Cell* 2005, **8**:553–564.
68. Obregon MJ, Mallol J, Pastor R, Morreale de Escobar G, Escobar del Rey F: **L-thyroxine and 3,5,3'-triiodo-L-thyronine in rat embryos before onset of fetal thyroid function.** *Endocrinology* 1984, **114**:305–307.
69. Morreale de Escobar G, Pastor R, Obregon MJ, Escobar del Rey F: **Effects of maternal hypothyroidism on the weight and thyroid hormone content of rat embryonic tissues, before and after onset of fetal thyroid function.** *Endocrinology* 1985, **117**:1890–1900.
70. Mortimer RH, Cannell GR, Addison RS, Johnson LP, Roberts MS, Bernus I: **Methimazole and propylthiouracil equally cross the perfused human term placental lobule.** *J Clin Endocrinol Metab* 1997, **82**:3099–3102.
71. Calvo R, Obregon MJ, Escobar del Rey F, Morreale de Escobar G: **The rat placenta and the transfer of thyroid hormones from the mother to the fetus. Effects of maternal thyroid status.** *Endocrinology* 1992, **131**:357–365.
72. Azizi F: **The safety and efficacy of antithyroid drugs.** *Expert Opin Drug Saf* 2006, **5**:107–116.
73. Arab DM, Malatjalian DA, Rittmaster RS: **Severe cholestatic jaundice in uncomplicated hyperthyroidism treated with methimazole.** *J Clin Endocrinol Metab* 1995, **80**:1083–1085.
74. Tajiri J, Noguchi S: **Antithyroid drug-induced agranulocytosis: special reference to normal white blood cell count agranulocytosis.** *Thyroid* 2004, **14**:459–462.
75. Weisenberg RC: **Microtubule formation in vitro in solutions containing low calcium concentrations.** *Science* 1972, **177**:1104–1105.
76. Keith C, DiPaola M, Maxfield FR, Shelanski ML: **Microinjection of Ca⁺⁺ – calmodulin causes a localized depolymerization of microtubules.** *J Cell Biol* 1983, **97**:1918–1924.
77. Szarama KB, Stepanyan R, Petralia RS, Gavara N, Frolenkov GI, Kelley MW, Chadwick RS: **Fibroblast growth factor receptor 3 regulates microtubule formation and cell surface mechanical properties in the developing organ of Corti.** *BioArchitecture* 2012, **2**(6):214–219. doi:10.4161/bioa.22332.
78. Hilmo A, Howard TH: **F-actin content of neonate and adult neutrophils.** *Blood* 1987, **69**:945–949.
79. Pender N, McCulloch CA: **Quantitation of actin polymerization in two human fibroblast sub-types responding to mechanical stretching.** *J Cell Sci* 1991, **100**:187–193.
80. Wulf E, Deboen A, Bautz FA, Faulstich H, Wieland T: **Fluorescent phalloxin, a tool for the visualization of cellular actin.** *Proc Natl Acad Sci USA* 1979, **76**:4498–4502.
81. Huang ZJ, Haugland RP, You WM, Haugland RP: **Phalloxin and actin binding assay by fluorescence enhancement.** *Anal Biochem* 1992, **200**:199–204.
82. Verderame M, Alcorta D, Egnor M, Smith K, Pollack R: **Cytoskeletal F-actin patterns quantitated with fluorescein isothiocyanate-phalloidin in normal and transformed cells.** *Proc Natl Acad Sci USA* 1980, **77**:6624–6628.
83. Ribeiro C, Ebner A, Affolter M: **In vivo imaging reveals different cellular functions for FGF and Dpp signaling in tracheal branching morphogenesis.** *Dev Cell* 2002, **2**:677–683.
84. Sai X, Ladher RK: **FGF signaling regulates cytoskeletal remodeling during epithelial morphogenesis.** *Curr Biol* 2008, **18**:976–981.
85. Kouhara H, Hadari YR, Spivak-Kroizman T, Schilling J, Bar-Sagi D, Lax I, Schlessinger J: **A lipid-anchored Grb2-binding protein that links FGF-receptor activation to the Ras/MAPK signaling pathway.** *Cell* 1997, **89**:693–702.
86. Shin EY, Woo KN, Lee CS, Koo SH, Kim YG, Kim WJ, Bae CD, Chang SI, Kim EG: **Basic fibroblast growth factor stimulates activation of Rac1 through a p85 betaPIX phosphorylation-dependent pathway.** *J Biol Chem* 2004, **279**:1994–2004.
87. Bamberg JR: **Proteins of the ADF/cofilin family: essential regulators of actin dynamics.** *Annu Rev Cell Dev Biol* 1999, **15**:185–230.
88. Ohashi K, Hosoya T, Takahashi K, Hing H, Mizuno K: **A Drosophila homolog of LIM-kinase phosphorylates cofilin and induces actin cytoskeletal reorganization.** *Biochem Biophys Res Commun* 2000, **276**:1178–1185.
89. Davidson LA: **Embryo mechanics: balancing force production with elastic resistance during morphogenesis.** *Curr Top Dev Biol* 2011, **95**:215–241.
90. Deponti D, Buono R, Catanzaro G, De Palma C, Longhi R, Meneverri R, Bresolin N, Bassi MT, Cossu G, Clementi E, et al: **The low-affinity receptor for neurotrophins p75NTR plays a key role for satellite cell function in muscle repair actin via RhoA.** *Mol Biol Cell* 2009, **20**:3620–3627.
91. Brown A, Li Y, Slaughter T, Black MM: **Composite microtubules of the axon: quantitative analysis of tyrosinated and acetylated tubulin along individual axonal microtubules.** *J Cell Sci* 1993, **104**:339–352.
92. Cueva JG, Hsin J, Huang KC, Goodman MB: **Posttranslational acetylation of a-tubulin constrains protofilament number in native microtubules.** *Curr Biol* 2012, **22**:1066–1074.
93. Rotsch C, Radmacher M: **Drug-induced changes of cytoskeletal structure and mechanics in fibroblasts: an atomic force microscopy study.** *Biophys J* 2000, **78**:520–535.
94. Radmacher M: **Studying the mechanics of cellular processes by atomic force microscopy.** *Methods Cell Biol* 2007, **83**:347–372.
95. Lim DJ, Anniko M: **Developmental morphology of the mouse inner ear. A scanning electron microscopic observation.** *Acta Otolaryngol. Suppl.* 1985, **422**:1–69.
96. Whitlon DS, Zhang X: **Polysialic acid in the cochlea of the developing mouse.** *Int J Dev Neurosci* 1997, **15**:657–669.
97. Tritsch NX, Yi E, Gale JE, Glowatzki E, Bergles DE: **The origin of spontaneous activity in the developing auditory system.** *Nature* 2007, **450**:50–55.
98. Gurniak CB, Perlas E, Witke W: **The actin depolymerizing factor n-cofilin is essential for neural tube morphogenesis and neural crest cell migration.** *Dev Biol* 2005, **278**:231–241.
99. Pham H, Yu H, Laski FA: **Cofilin/ADF is required for retinal elongation and morphogenesis of the Drosophila rhabdome.** *Dev Biol* 2008, **318**:82–91.

100. Rasband WS: *ImageJ*. Bethesda, Maryland, USA: US National Institutes of Health; (1997–2011) <http://imagej.nih.gov/ij>.
101. Wu DK, Oh SH: Sensory organ generation in the chick inner ear. *J Neurosci* 1996, **16**:6454–6462.
102. Welch BL: The generalization of “Student’s” problem when several different population variances are involved. *Biometrika* 1947, **34**:28–35.
103. Sneddon IN: The relation between load and penetration in the axisymmetric Boussinesq problem for a punch of arbitrary profile. *Int J Eng Sci* 1965, **3**:47–57.

doi:10.1186/1471-213X-13-6

Cite this article as: Szarama *et al.*: Thyroid hormone increases fibroblast growth factor receptor expression and disrupts cell mechanics in the developing organ of corti. *BMC Developmental Biology* 2013 **13**:6.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

