PeerJ

Seasonal trends of the polyp expansion and nutritional condition of *Alcyonium acaule* (Octocorallia, Alcyonacea)

Lucia Rizzo¹, Ida Fiorillo² and Sergio Rossi^{3,4}

ABSTRACT

The ecological physiology of anthozoans, as well as their resistance to stressors, are strongly influenced by environmental factors and the availability of resources. The energy budget of anthozoans can vary seasonally in order to find an equilibrium between the available resources and respiration, polyp activity, growth, and reproduction processes. The variation in the biochemical composition of the animal tissues in these organisms results from a combination of the productivity processes of the water column coupled with the reproductive effort and potential starvation periods of the anthozoans. Here, the seasonal variation in the polyp activity of a slow-growing passive suspension feeder, the octocoral Alcyonium acaule, as well as their carbohydrate, protein and lipid contents, was investigated in a warm temperate environment using *in-situ* observations and biochemical analyses. Polyp activity exhibited a significant variability that was moderately dependent on season, while an aestivation phenomenon in A. acaule (i.e., a resting period in which the anthozoan is not capable of any polyp activity) during the warmer months is clearly observed. Carbohydrate concentrations in the coral species showed a significant increase in the late winter and spring seasons, and the lipid content increased during the spring. A higher abundance of lipids and carbohydrates coincided with a higher primary productivity in the water column, as well as with the octocoral reproduction period. In late autumn, there was a depletion of these biomolecules, with protein levels exhibiting great variability across sampling times. Complex alterations driven by climate change could affect the energy fluxes that depend on the dead or alive particles that are intercepted by marine animal forests. The obtained findings show a food shortage in late summer and autumn of the benthic suspension feeder A. acaule through the integrative descriptors of the ecophysiology of these anthozoans. This research contributes to the knowledge of energy storage capabilities in benthic suspension feeders in general, highlighting the importance of understanding the limits of resistance to starvation periods through these indicators.

Subjects Animal Behavior, Ecology, Marine Biology, ZoologyKeywords Marine animal forest, Habitat-forming species, Anthozoans, Polyp expansion, Organic matter, Octocorals, Seasonal changes

Submitted 28 April 2021 Accepted 30 July 2021 Published 14 October 2021

Corresponding author Lucia Rizzo, lucia.rizzo@szn.it

Academic editor Anastazia Banaszak

Additional Information and Declarations can be found on page 14

DOI 10.7717/peerj.12032

Copyright 2021 Rizzo et al.

Distributed under Creative Commons CC-BY 4.0

OPEN ACCESS

¹ Department of Integrative Marine Ecology, Stazione Zoologica Anton Dohrn, Napoli, Italy

² Institute of Marine Sciences (ICM-CSIC), Barcelona, Spain

³ National Interuniversity Consortium for Marine Sciences (CoNISMa), Roma, Italy

⁴ Department of Biological and Environmental Sciences and Technologies, University of Salento, Lecce, Italy

INTRODUCTION

Coastal benthic ecosystems represent a hotspot of marine biodiversity, forming different habitats and providing shelter to benthic and demersal biota (*Bianchi & Morri, 2000*; *Rossi et al., 2019a*; *Rossi & Rizzo, 2020*). Climate change, acting simultaneously with the growing frequency of anthropic activities, can affect habitat-forming species and alter coastal trophodynamics (*Pusceddu et al., 2016*; *Rizzo et al., 2017, 2020*). These human-driven stressors can influence energy fluxes related to the availability of food quantity and quality, and threaten the fitness of sensitive species (*Rossi et al., 2019a*; *Rossi & Rizzo, 2020*). Seasonal patterns in growth, reproduction, and abundance of marine benthic suspension feeders are well recognized, revealing the strong dependence of coastal ecosystems on large environmental variability (*Clarke, 1988*; *Valiela, 1995*). Understanding the relationship between the temporal changes of environmental factors, water column resources (*e.g.*, nutrients or food availability), and the response of benthic organisms, represents a crucial step in coupling physical and biological processes in the ocean.

Coma et al. (2000) reported a seasonal pattern in the activity in benthic suspension feeders driven by temperature in cold temperate seas, showing an autumnal decrease in activity and in secondary production, a minimum in the winter period, and an increase during spring and summer (Clarke, 1993). Antarctic gorgonians collected in the Eastern Weddell Sea and the Bransfield Strait during primary productivity depletion had a diet based on resuspended material, suggesting a consumption of phytoplankton detritus during the winter (*Elias-Piera et al., 2013*). However, in warm temperate seas, such as the Mediterranean basin, the responses of all benthic suspension feeders are largely heterogeneous (Boero et al., 1986; Boero & Fresi, 1986; Coma et al., 2000). During the summer season, some benthic species face an adverse feeding period and seem to be affected by aestivation (summer dormancy) instead of winter dormancy, including ascidians (Turon, 1992; Turon & Becerro, 1992), bivalves and polychaetes (Ramón, Abelló & Richardson, 1995; Arneri, Giannetti & Antolini, 1998; Sardá, Pinedo & Martin, 1999), and anthozoans (Coma et al., 1998; Garrabou, 1999). Generally, benthic species show the highest reproductive output and growth rate in late winter, spring and early summer (Turon, 1992; Turon & Becerro, 1992; Coma et al., 1998; Garrabou, 1999). Cold-water and warm-water species live in the Mediterranean Sea, showing a coexistence in the array of climatic and hydrologic conditions. The resulting high species richness of this sea is recognized as a hotspot for biodiversity (Bianchi & Morri, 2000).

Different ecological factors and related interactions affect seasonal cycles of benthic species (*Gili & Hughes, 1995; Sardá, Pinedo & Martin, 1999*). Temperature, water movement, and food availability have been identified as crucial environmental factors affecting the dynamics of benthic suspension feeders, such as in *Corallium rubrum (Rossi, Rizzo & Duchêne, 2019), Paramuricea clavata (Coma et al., 1994), Alcyonium acaule* and *Parazoanthus axinellae (Garrabou, 1999)*. In the Mediterranean Sea, changes in temperature, primary productivity, and water stratification create different conditions in food availability. Benthic suspension feeders and their metabolic responses depend on the supply of food (*Coma et al., 1998; Rossi et al., 2006*) associated with the short-time-scale variability of near-bottom seston (*Rossi & Gili, 2007; Rossi & Rizzo, 2021*). In late winter/early spring, the stabilization of surface seawater influences changes in the supply of food, and in autumn, the thermocline breaking mixes the waters and related food (*Estrada, 1996*). During the summer, the strong stratification of the water column leads to a depletion of suspended materials. Polyp expansion (*i.e.*, the opening of polyps in anthozoans, *Rossi, Rizzo & Duchêne, 2019*) can be stimulated by increasing the frequency of food inputs (*Tsounis et al., 2006; Rossi & Gili, 2007; Rossi & Rizzo, 2021*). This triggers a response in passive suspension feeders in front of changes in seston availability, supporting their feeding activity (*Coma et al., 1994; Garrabou, 1999; Rossi, Rizzo & Duchêne, 2019*). However, while climate change and related problems of energy availability have been shown to affect the metabolic rates of these organisms, the underlying basic mechanisms regulating ecophysiological processes in warm, temperate benthic suspension feeders remain poorly investigated.

Soft corals (no skeletal axis) belong to the order Alcyonacea, a main structural group of benthic colonial invertebrates found across all latitudes and depths, from shallow waters to the deep sea (*Dinesen, 1983; Lasker, Gottfried & Coffroth, 1983; Spalding et al., 2001; Mortensen & Buhl-Mortensen, 2004; Roberts et al., 2009). Alcyonium acaule* Marion, 1878 (Cnidaria: Octocorallia: Alcyonacea) is colonial with a sessile soft coral thick-bodied and finger-like lobes. This species grows to form a cluster up to 20 cm in height. It is a slow growth organism with a life span of >20 years (*Teixidó, Garrabou & Harmelin, 2011*). This soft coral is abundant in the Mediterranean Sea, in the pre-coralligenous and coralligenous bioconcretions, *i.e.*, outcrops consist of coralline algal frameworks that grow in dim light conditions (*Fiorillo et al., 2013; Ambroso et al., 2013; Martin et al., 2014*). In certain cases, this species forms dense aggregations, which reinforces its role in community organization (*Garrabou, 1999; Ambroso et al., 2013*).

The aim of this study was to evaluate seasonal differences in polyp expansion and tissue concentrations of proteins, carbohydrates, and lipids of the benthic invertebrate *A. acaule* in a temperate sea (*i.e.*, Mediterranean Sea) in relation to temporal coupling with environmental variables. The variation in the biochemical composition of the tissue of the soft coral *A. acaule* may depend on productivity fluctuations of the water column, and seasonal fluctuations in seston quantity and quality (*Rossi et al., 2003; Rossi & Gili, 2005*).

MATERIALS & METHODS

Study area and sampling

One hundred and twenty *A. acaule* colonies were observed and sampled in the Marine Protected Area of the Medes Islands (NW Mediterranean Sea, Catalonia, Spain), in Tascons Grossos (40°02′55″'N, 3°13′30″E, Fig. 1). Sampling was carried out at 11–19 m depth in a channel of rocky blocks by SCUBA diving from June 2002 to June 2003. Dense *Alcyonium acaule* colonies live on rocky bottom at 10–50 m depth in the study area, together with other benthic suspension feeders and coralline algae (*Gili & Ros, 1985*). The channel is characterized by alternating currents (north/south and south/north) that can reach high speeds (*Rossi, 2002; Rossi & Gili, 2007; Rossi, Rizzo & Duchêne, 2019*).



Figure 1 Map of the sampling location indicated with a star (Tascons Grossos, Marine Protected Area of the Medes Islands) in the Mediterranean Sea. Full-size DOI: 10.7717/peerj.12032/fig-1

Environmental variables

Sea water temperatures were recorded three times each month at two different depths (5 and 20 m depth), ~100 m away from the sampling area (*Rossi & Gili, 2005*), using an inverted thermometer according to *Cebrián, Duarte & Pascual (1996*). Water transparency and seston abundance were evaluated applying the Secchi disk protocol (*Preisendorfer, 1986*). A vertical Secchi disk (VSD) was observed each 10–12 days from a boat 20–30 m from the sampling point. Observations were always made between 10:00 a.m. and 12:00 p.m. to avoid interference from incident light (*Preisendorfer, 1986*).

Seasonal polyp expansion

Polyp expansion was observed in *Alcyonium acaule* over the entire year. Colonies with expanded polyps or completely contracted polyps were observed visually and registered monthly three times a season (100 healthy colonies grouped in sets of 10 were observed each time randomly) by SCUBA diving. Only colonies showing polyps with 100% expansion were assigned as expanded polyps, while colonies with closed polyps were recorded as contracted polyps. Colonies in between these stages were not counted. This alcyonarian also exhibits dormant colonies that were also recorded (*i.e.*, those animals with contracted polyps and a thin film covering the surface, *Garrabou*, *1999*).

Biochemical analyses

During the sampling, apical branches (no more than one cm length) were cut and transported in a cooler (4 °C). Samples were frozen in liquid nitrogen within 30–40 min of sampling. The samples were held at -80 °C, freeze-dried at -110 °C and a pressure of 100 mbar, and then stored frozen at -20 °C pending biochemical analyses.

Results have been reported in μ g Protein-Carbohydrate-Lipid mg⁻¹ OM (Organic Matter). Organic matter was obtained by taking 10–15 mg of the tissue each month (over 1 year), heating the tissue at 80 °C for 48 h and weighing the sample to obtain dry weight. The same tissue was ashed for 4 h at 500 °C. The difference between dry weight and ash weight was considered organic matter (*Rossi et al., 2006*).

All analyses were performed spectrophotometrically (colorimetrically) following the protocol of *Rossi et al.* (2006). For proteins, a 10–15 mg piece of tissue was homogenized in 6 ml 1 N NaOH, and quantified using the method of *Lowry et al.* (1951), using bovine serum albumin as the standard (Sigma-Aldrich, St. Louis, MO, USA). For carbohydrates, a 15–20 mg piece of each branch was weighed in a microbalance (precision: ±0.01 mg), and the tissue was homogenized in 6 ml of double distilled water. Carbohydrates were quantified using the method reported by *DuBois et al.* (1956), using (D)-glucose as standard (Sigma-Aldrich, St. Louis, MO, USA). For lipids, a 15–20 mg piece of dry tissue was homogenized in 6 ml of Chloroform-Methanol (2:1 v:v); lipids were quantified according to the method of *Barnes & Blackstock* (1973), using cholesterol as standard (Sigma-Aldrich, St. Louis, MO, USA).

Statistical analyses

The differences in (i) polyp expansion and (ii) protein, carbohydrate, lipid concentrations and OM composition of *A. acaule* among several seasons were assessed across sampling times by multivariate analyses.

The design consisted of two factors: Season (S, as fixed factor with four levels) and Sampling Time (T(S)), as random factor with three levels, nested in Season), with n = 3 for each combination of factors. Multivariate and univariate permutational analysis of variance (PERMANOVA, McArdle & Anderson, 2001) was based on Euclidean distances of untransformed data of polyp expansion and previously normalized data of proteins, carbohydrates, lipids and OM composition, using 9,999 random permutations of the appropriate units (Anderson & Braak, 2003). When significant differences were encountered (p < 0.05), post-hoc pairwise tests for the fixed factor were carried out, to ascertain the consistency of the differences among seasons. Because of the restricted number of unique permutations in the pairwise tests, p values were obtained from Monte Carlo samplings. For illustrating differences in the composition of OM, significant terms were plotted using canonical analysis of principal coordinates (CAP) (Anderson & Willis, 2003) for the factor Season. To determine whether the environmental features (*i.e.*, seawater temperature at -5 m, seawater temperature at -20 m, water transparency) influenced polyp expansion and organic matter composition of A. acaule, multivariate multiple regression analyses and distance-based Redundancy Analyses (dbRDA, Stepwise regression) were performed (DistLM, McArdle & Anderson, 2001). The analyses were performed using the software PRIMER v. 6 (Clarke & Gorley, 2006).





RESULTS

Environmental variables

The tested environmental variables are shown in Fig. 2. The temperatures at -5 m ranged from 11 to 23 °C, whilst at -20 m varied from 11 to 21 °C (Fig. 2). The summer was the warmest season, with a mean of 21 °C at -5 m and 20 °C at -20 m depth, followed by spring (17 and 16 °C, respectively), autumn (16 °C at both depths) and winter (13 and 12 °C, respectively). Water transparency was assessed by Secchi disk which ranged from 5 to 29 m (Fig. 2). It was higher during the summer (approx. 22 m), and decreased in the winter (~11 m, Fig. 2).

Seasonal polyp expansion

The polyp expansion of *A. acaule* through the entire year is shown in Fig. 2 in relation to the environmental variables investigated. Although associated with a wide variability, a low percentage of polyp expansion occurred in summer together with the phenomenon of dormant stage that was observed in some colonies. In a low number of colonies, dormancy was also present during the late spring and early autumn.

The PERMANOVA revealed that the polyp expansion of *A. acaule* polyps varied significantly among sampling times within different seasons, while significant seasonal variations had not been observed (Table 1). The results of the DistLM analyses revealed that the temperature at -20 m significantly explained variations in polyp expansion (p < 0.05, 14%). In particular, the three considered environmental variables explained cumulatively 20% of polyp expansion variation (Table 2, Fig. 3).

Biochemical composition of A. acaule organic matter

The highest carbohydrate contents were found in winter $(32.69 \pm 1.50 \ \mu g \ mg^{-1} \ OM)$, followed by the concentrations detected in spring $(28.89 \pm 2.03 \ \mu g \ mg^{-1} \ OM)$ and summer $(24.52 \pm 1.59 \ \mu g \ mg^{-1} \ OM)$; the lowest concentrations were reported in autumn with an average of $21.94 \pm 1.51 \ \mu g \ mg^{-1} \ OM$ (Fig. 4). Carbohydrate concentration in winter was significantly higher than autumn and winter, while in autumn it was

1	8	1 8		
Source	df	MS	Pseudo-F	P (perm)
S	3	110.79	1.37	
T(S)	8	80.57	3.96	***
Res	108	20.33		
Total	119			

Table 1 Results of multivariate permutational analyses of variance (PERMANOVA) in polyp expansion of A. *acaule* among seasons (S) and sampling times (T(S)).

Notes:

df, degree of freedom; MS, mean sum of squares; Pseudo-F, Pseudo-F statistic; *P* (perm), permutational level of probability.

*** P < 0.001.

 Table 2 DistLM analyses estimating the proportion of polyp expansion of A. acaule explained singularly (marginal tests) and cumulatively (sequential tests) by environmental features.

Marginal tests			
Variable	Р	Prop.	
Temperature at −5 m	*	0.12	
Temperature at -20 m	*	0.14	
Transparency	*	0.11	
Sequential tests			
Variable	Р	Prop.	Cumul.
Temperature at −20 m	*	0.14	0.14
Transparency	ns	3.25 E-02	0.17
Temperature at -5 m	ns	3.61 E-02	0.20

Notes:

p < 0.05.
 ns, not significant; Prop., proportion of explained variance; Cumul., cumulative results.



Figure 3 DbRDA ordination after DistLM analysis, showing the relationships among the environmental features and polyp expansion of *A. acaule*.

Full-size DOI: 10.7717/peerj.12032/fig-3



Figure 4 Seasonal carbohydrate concentrations in the tissue of *A. acaule*. Different letters indicate statistically significant differences among seasons (p > 0.05).

Full-size 🖾 DOI: 10.7717/peerj.12032/fig-4

Table 3 Results of permutational analyses of variance (PERMANOVA) in carbohydrate, protein and lipid concentrations of *A. acaule* among seasons (S) and sampling times (T(S)).

		Carboh	ydrates		Proteir	18		Lipids		
Source	df	MS	Pseudo-F	P (perm)	MS	Pseudo-F	P (perm)	MS	Pseudo-F	P (perm)
S	3	5.02	9.40	**	2.10	1.17		4.58	6.32	*
T(S)	8	0.53	0.82	ns	1.79	2.98	*	0.72	1.13	ns
Res	24	0.65			0.60			0.64		
Total	35									

Notes:

df, degree of freedom; MS, mean sum of squares; Pseudo-F, Pseudo-F statistic; P (perm), permutational level of probability; ns, not significant. * P < 0.05.

** P < 0.01.



Figure 5 Temporal variation in carbohydrate concentrations of *A. acaule*. Different letters indicate statistically significant differences among seasons (p > 0.05).

Full-size 🖾 DOI: 10.7717/peerj.12032/fig-5



Figure 6 Seasonal lipid concentrations in the tissue of A. acaule. Different letters indicate statisticallysignificant differences among seasons (p > 0.05).Full-size \square DOI: 10.7717/peerj.12032/fig-6

Table 4 Results of multivariate permutational analyses of variance (PERMANOVA) in organic matter composition of A. *acaule* among seasons (S) and sampling times (T(S)).

Source	df	MS	Pseudo-F	P (perm)	
S	3	11.70	3.84	**	
T(S)	8	3.05	1.61	ns	
Res	24	1.90			
Total	35				
Groups	t	<i>P</i> (MC)	Groups	t	P (MC)
WI vs SP	2.05	ns	SP vs SU	1.30	ns
WI vs SU	1.69	ns	SP vs AU	2.88	*
WI vs AU	3.86	**	SU vs AU	0.80	ns

Notes:

df, degree of freedom; MS, mean sum of squares; Pseudo-F, Pseudo-F statistic; *P* (perm), permutational level of probability; *P* (MC), probability level after Monte Carlo simulations; t, pairwise tests; ns, not significant; WI, winter, SP, spring, SU, summer and AU, autumn.

P < 0.05.

** P < 0.01.

significantly lower than winter and spring (Table 3). PERMANOVA results revealed that protein levels varied significantly across sampling times of several seasons, while no differences have been observed among seasons (Table 3, Fig. 5). The highest lipid contents were found in spring, followed by the concentrations detected in summer and winter; the lowest concentrations were reported for autumn (Fig. 6). Differences in lipid contents did not vary among sampling times, while significant seasonal differences were found (Table 3). In particular, the lipid concentrations in spring were significantly higher than autumn and winter.

The multivariate PERMANOVA tests revealed that the biochemical composition varied significantly among seasons (Table 4). The pairwise comparisons showed significant differences in OM composition between winter and autumn and between autumn and spring. The CAP plot showed a segregation between seasons (Fig. 7), which is mostly explained by the increased contents of proteins and carbohydrates during the winter and an increment of lipids during the spring.



Figure 7 CAP plot of organic matter composition in tissue of *A. acaule* in different seasons. Vectors represent the correlations of each concentrations of carbohydrates, proteins and lipids. Full-size DOI: 10.7717/peerj.12032/fig-7

 Table 5 DistLM analyses estimating the proportion of organic matter composition in A. acaule

 explained singularly (marginal tests) and cumulatively (sequential tests) by environmental features.

Marginal tests			
Variable	Р	Prop.	
Temperature at -5 m	ns	7.85E-02	
Temperature at -20 m	*	9.35E-02	
Transparency	ns	7.58E-02	
Sequential tests			
Sequential tests Variable	 P	Prop.	Cumul.
Sequential tests Variable Temperature at -20 m	P *	Prop. 9.35E–02	Cumul. 9.35E–02
Sequential tests Variable Temperature at -20 m Temperature at -5 m	P * *	Prop. 9.35E–02 0.1	Cumul. 9.35E–02 0.20

Notes: * *p* < 0.05.

ns, not significant; Prop., proportion of explained variance; Cumul., cumulative results.

The results of the DistLM analyses revealed that the temperature at -20 m explained variations in organic matter composition significantly (p < 0.05, 9.35%), followed by temperature at -5 m and water transparency, which altogether explained cumulatively 26% of organic matter composition variation (Table 5). This was shown by the dbRDA analysis where these environmental variables cumulatively explained about 21% of organic matter composition along the first two axes (Fig. 8).





DISCUSSION

The findings of this study couples seasonal polyp activity and nutritional condition with key environmental fluctuations in a slow-growing, passive suspension feeder in warm temperate waters. Polyp activity in the field highlighted a high variability that was loosely dependent on different seasons, suggesting an inter-individual variability in the response to water column changes (*Duchêne et al., 2000; Duchêne, 2017*). The complex dynamics of polyp expansion and the trophic status of *A. acaule* have revealed that, among the environmental variables investigated, water temperature greatly influenced polyp activity and organic matter composition of the octocoral.

Similar to previous studies on anthozoan polyp activity (*Garrabou, 1999; Rossi, 2002; Previati et al., 2010; Rossi, Rizzo & Duchêne, 2019*), the current study found a significant negative relationship between temperature and polyp activity. *Previati et al. (2010)* showed that respiration is the main factor conditioning polyp contraction at high temperatures, a result previously suggested by *Coma et al. (2002)*. The high variability detected in the polyp expansion can be explained by the sensitivity of polyps to water temperature, and especially seston abundance and quality (*Rossi, Rizzo & Duchêne, 2019*).

Seston availability affects Mediterranean gorgonians (Cnidaria, Octocorallia) differently according to species-specific trophic strategies (*i.e.*, mixotrophic and heterotrophic) and population density (*Rossi & Rizzo*, 2021). Likewise, complex hydrodynamic conditions, including water temperatures varying in time and space, and the water current regime, act simultaneously to shape polyp activity (*Rossi, Rizzo & Duchêne, 2019*). Water movement may increase plankton concentrations (*Sebens & DeRiemer, 1977; Palardy, Grottoli & Matthews, 2006; Rossi & Rizzo, 2021*), affecting the polyp expansion. The cumulative effect of seston availability thus appears to be a major factor controlling spawning behaviour as described below.

In the present study, we observed significant differences in carbohydrate and lipid concentrations among seasons, while proteins exhibited great variability across sampling times. Protein levels seem to be lower than values found in Paramuricea clavata (Rossi et al., 2006) and Leptogorgia sarmentosa (Rossi, 2002); it may be due to the proteinic skeleton of these gorgonians. However, the lipid and carbohydrate concentrations have been suggested as a proxy of resource mobilization (Clarke, 1977; Costello, 1998), and along with Heat Shock Protein (HSP) expression, can be used as biomarkers of environmental changes and food availability for benthic suspension feeders (Rossi, Snyder & Gili, 2006). Our results show carbohydrate concentrations in line with the data reported in the literature for other anthozoans (Ben-David-Zaslow & Benayahu, 1999; Rossi, 2002; Rossi et al., 2006; Rossi & Tsounis, 2007). Lipid concentrations were lower than values found in Corallium rubrum by Rossi & Tsounis (2007) and in Eunicella singularis (Gori et al., 2007). The highest lipid concentrations in A. acaule were found during the spring. Before summer aestivation, organisms reproduce and need a significant quantity of energy to compensate for the gonadal release and the reduced food availability (Coma et al., 2000; Rossi et al., 2006). Indeed, A. acaule is gonochoristic and a surface brooder that retains eggs on the external surface of female colonies with mucous strings (Garrabou, 1999; Fiorillo et al., 2013; Teixidó et al., 2016), spawning once a year in late spring-early summer (*Fiorillo et al., 2013*) in coincidence with its higher lipid contents. Rapidly increasing solar irradiance during the spring has been suggested as the trigger for oocyte maturation and vitellogenesis, nevertheless other underlying mechanisms could be playing a key role (*Fiorillo et al., 2013*). Summer is characterized by transparent and warm waters, with low plankton and seston concentration (Rossi & Gili, 2005). These conditions impose high basal metabolic energy consumption on benthic suspension feeders (Coma et al., 2000; Rossi et al., 2006; Previati et al., 2010). After the summer, the stratified water column breakdowns as a consequence of autumn storms, mixing the available food and influencing activities and behaviours of benthic organisms in the complex coastal hydrodynamics (Estrada, 1996; Rossi & Gili, 2005; Rossi et al., 2019a). After spawning, the energy invested (Rossi et al., 2006; Rossi & Tsounis, 2007; Viladrich et al., 2017) can be partially recovered in the autumnal peak of primary production (Estrada, 1996; Rossi & Gili, 2005; Rossi et al., 2019b). However, in late autumn, benthic suspension feeders face a period of food scarcity due to the high resuspension of low-quality organic matter (*Rossi et al.*, 2019b). Carbohydrates and lipids reach minimum levels in autumn in many octocorals (Rossi, 2002; Rossi et al., 2006; Rossi & Tsounis, 2007; Gori et al., 2012). This feeding stress is also observed in the HSP 70 and 90 expression, which is especially related to carbohydrate mobilization (Rossi, Snyder & Gili, 2006).

The aestivation process may lead to dormancy states (*Coma et al., 2000*). We could observe the aestivation phenomenon in *A. acaule* during the summer and, to a minor extent, in the late spring and early autumn. Among benthic fauna, periods of activity that alternate to dormancy are common (*Boero & Fresi, 1986*; *Boero et al., 1986*; *Turon, 1992*; *Turon & Becerro, 1992*). Contrary to what is observed in warm temperate water, seasonality in the dynamics of marine benthic suspension feeders is characterized by winter dormancy in cold temperate seas (*Coma et al., 2000*). Generally, the Mediterranean

aestivation is characterized by a summer energy shortage (reduced polyp activity, colony dormancy, low oxygen consumption, depletion of energy storage) and a decrease of energy investment in secondary production (*i.e.*, growth and reproduction) during the summer period, however the aestivation processes can differ widely among taxa (*Coma et al., 2000; Coma et al., 2002*).

Studies on octocorals showed that at high temperatures the metabolic rate decreases in a dormancy-like stage (*Coma et al., 2000; Previati et al., 2010*). The increase in temperature reflects high respiration rates, reducing the available energy storage of benthic suspension feeders (*Rossi et al., 2006*). Anthozoans, indeed, are characterized by a decrease in investment in secondary production and in feeding activity during the summer period (*Coma et al., 1998*), with some species exhibiting non-feeding periods (*Garrabou, 1999*). The role of resting stages has been recognized as a key factor in life cycle dynamics and in the shaping of marine assemblages in some studies that focus on temporal fluctuations, not only in benthic invertebrates but also in phyto-and zooplankton communities (*Boero et al., 1996; Marcus & Boero, 1998; Montresor et al., 2013*).

The importance of discerning between seasonal energy invested or stored during periods of insufficient food (or energy and nutrient) availability has been discussed (*Clarke, 1977; Epp, Bricelj & Malouf, 1988; Alonzo, Mayzaud & Razouls, 2000*). Metabolic adjustments aimed to reduce energy losses could be decisive for avoiding mass mortality events of benthic suspension feeders that have been reported in the summer (*Cerrano et al., 2000; Garrabou et al., 2009*). Understanding the capability to store energy, coupled with benthic suspension feeding activity, could be key factors to help understand the mass mortalities detected in the Mediterranean Sea. It has been suggested that feeding capabilities will be constrained by temperature shifts in the near future (*Galli et al., 2016*) under the scenario of warm temperate seas, combined with water stratification in shallow areas and a depletion of primary productivity (*Galli, Solidoro & Lovato, 2017*).

CONCLUSIONS

Our findings demonstrate a food shortage in late summer and late autumn for the benthic suspension feeder *A. acaule.* Polyp activity highlighted a high variability that was only slightly dependent on seasons, while the aestivation phenomenon, a resting period in which the Anthozoan is not capable of any polyp activity, during the warm months has been quantified. This research supports the hypothesis that some Mediterranean benthic suspension feeders exhibit a seasonal pattern of trophic status, characterized by aestivation. Further research will elucidate the complex alterations driven by climate change in marine animal forests and in their processes of carbon immobilization (*Coppari, Zanella & Rossi, 2019; Rossi & Rizzo, 2020*), as food quantity and quality will be central to face fast environmental changes and anthropogenic stressors (*Rossi, Rizzo & Duchêne, 2019*). The knowledge of energy fluxes in these benthic organisms related to changes in environmental variables is important to promote conservation measures addressed to the mitigation of human-induced and climate-driven changes.

ACKNOWLEDGEMENTS

We want to thank Rafel Coma, Cristina Linares, David Diaz and Bernat Hereu for field assistance, Josep-Maria Gili for several key discussions about some ideas shown in the paper, and Roberta Johnson and Paride Papadia for the English review. The authors are especially grateful to Josep Pasqual for the environmental data collection.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This research was funded by the Spanish project MEDGORG, CICYT (REN2000-0633-C03-01/MAR). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors: Spanish project MEDGORG, CICYT: REN2000-0633-C03-01/MAR.

Competing Interests

The authors declare that they have no competing interests.

Author Contributions

- Lucia Rizzo analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Ida Fiorillo performed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Sergio Rossi conceived and designed the experiments, performed the experiments, authored or reviewed drafts of the paper, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:

The raw measurements are available in the Supplemental File.

Supplemental Information

Supplemental information for this article can be found online at http://dx.doi.org/10.7717/ peerj.12032#supplemental-information.

REFERENCES

- Alonzo F, Mayzaud P, Razouls S. 2000. Egg production, population structure and biochemical composition of the subantarctic copepod *Paraeuchaeta antarctica* in the Kerguelen Archipelago. *Marine Ecology Progress Series* 205:207–217 DOI 10.3354/meps205207.
- Ambroso S, Gori A, Dominguez-Carrió C, Gili J-M, Berganzo E, Teixidó N, Greenacre M, Rossi S. 2013. Spatial distribution patterns of the soft corals *Alcyonium acaule* and *Alcyonium palmatum* in coastal bottoms (Cap de Creus, northwestern Mediterranean Sea). *Marine Biology* 160(12):3059–3070 DOI 10.1007/s00227-013-2295-4.

- Anderson M, Braak CT. 2003. Permutation tests for multi-factorial analysis of variance. *Journal of Statistical Computation and Simulation* 73(2):85–113 DOI 10.1080/00949650215733.
- Anderson MJ, Willis TJ. 2003. Canonical analysis of principal coordinates: a useful method of constrained ordination for ecology. *Ecology* 84:511–525 DOI 10.1890/0012-9658(2003)084[0511:.
- Arneri E, Giannetti G, Antolini B. 1998. Age determination and growth of *Venus verrucosa* L. (Bivalvia: Veneridae) in the southern Adriatic and the Aegean Sea. *Fisheries Research* 38(2):193–198 DOI 10.1016/s0165-7836(98)00146-5.
- Barnes H, Blackstock J. 1973. Estimation of lipids in marine animals and tissues: detailed investigation of the sulphophosphovanilun method for 'total' lipids. *Journal of Experimental Marine Biology and Ecology* 12(1):103–118 DOI 10.1016/0022-0981(73)90040-3.
- **Ben-David-Zaslow R, Benayahu Y. 1999.** Temporal variation in lipid, protein and carbohydrate content in the Red Sea soft coral *Heteroxenia fuscescens*. *Journal of the Marine Biological Association of the United Kingdom* **79(6)**:1001–1006 DOI 10.1017/s002531549900123x.
- **Bianchi CN, Morri C. 2000.** Marine biodiversity of the Mediterranean Sea: situation, problems and prospects for future research. *Marine pollution bulletin* **40(5)**:367–376 DOI 10.1016/S0025-326X(00)00027-8.
- Boero F, Balduzzi A, Bavestrello G, Caffa B, Vietti RC. 1986. Population dynamics of *Eudendrium glomeratum* (Cnidaria: Anthomedusae) on the Portofino Promontory (Ligurian Sea). *Marine Biology* 92(1):81–85 DOI 10.1007/bf00392749.
- Boero F, Belmonte G, Fanelli G, Piraino S, Rubino F. 1996. The continuity of living matter and the discontinuities of its constituents: do plankton and benthos really exist? *Trends in Ecology & Evolution* 11(4):177–180 DOI 10.1016/0169-5347(96)20007-2.
- Boero F, Fresi E. 1986. Zonation and evolution of a rocky bottom hydroid community. *Marine Ecology* 7(2):123–150 DOI 10.1111/j.1439-0485.1986.tb00152.x.
- **Cebrián J, Duarte CM, Pascual J. 1996.** Marine climate on the Costa Brava (northwestern Mediterranean) littoral. *Publicaciones especiales Instituto Español de Oceanografía* **22**:9–21.
- Cerrano C, Bavestrello G, Bianchi CN, Cattaneo-vietti R, Bava S, Morganti C, Morri C, Picco P, Sara G, Schiaparelli S, Siccardi A, Sponga F. 2000. A catastrophic mass-mortality episode of gorgonians and other organisms in the Ligurian Sea (North-western Mediterranean). *Ecology Letters* 3(4):284–293 DOI 10.1046/j.1461-0248.2000.00152.x.
- Clarke A. 1977. Seasonal variations in the total lipid content of *Chorismus antarcticus* (Pfeffer) (Crustacea: Decapoda) at South Georgia. *Journal of Experimental Marine Biology and Ecology* 27(1):93–106 DOI 10.1016/0022-0981(77)90056-9.
- Clarke A. 1988. Seasonality in the antarctic marine environment. *Comparative Biochemistry and Physiology Part B: Comparative Biochemistry* **90(3)**:461–473 DOI 10.1016/0305-0491(88)90285-4.
- Clarke A. 1993. Temperature and extinction in the sea: a physiologist's view. *Paleobiology* 19(4):499–518 DOI 10.1017/S0094837300014111.
- Clarke KR, Gorley RN. 2006. PRIMER V6: user manual/tutorial. Plymouth: Plymouth Marine Laboratory.
- Coma R, Gili J-M, Zabala M, Riera T. 1994. Feeding and prey capture cycles in the aposymbiontic gorgonian *Paramuricea clavata*. *Marine Ecology Progress Series* 115:257–270 DOI 10.3354/meps115257.
- Coma R, Ribes M, Gili JM, Zabala M. 1998. An energetic approach to the study of life-history traits of two modular colonial benthic invertebrates. *Marine Ecology Progress Series* 162:89–103 DOI 10.3354/meps162089.

- Coma R, Ribes M, Gili J-M, Zabala M. 2000. Seasonality in coastal benthic ecosystems. *Trends in Ecology & Evolution* 15(11):448–453 DOI 10.1016/s0169-5347(00)01970-4.
- Coma R, Ribes M, Gili JM, Zabala M. 2002. Seasonality of in situ respiration rate in three temperate benthic suspension feeders. *Limnology and Oceanography* 47(1):324–331 DOI 10.4319/lo.2002.47.1.0324.
- Coppari M, Zanella C, Rossi S. 2019. The importance of coastal gorgonians in the blue carbon budget. *Scientific Reports* 9(1):13550 DOI 10.1038/s41598-019-49797-4.
- **Costello J. 1998.** Physiological response of the hydromedusa *Cladonema californicum* Hyman (Anthomedusa: Cladonemidae) to starvation and renewed feeding. *Journal of Experimental Marine Biology and Ecology* **225(1)**:13–28 DOI 10.1016/s0022-0981(97)00204-9.
- **Dinesen ZD. 1983.** Patterns in the distribution of soft corals across the central Great Barrier Reef. *Coral Reefs* 1(4):229–236 DOI 10.1007/bf00304420.
- **DuBois M, Gilles KA, Hamilton JK, Rebers PA, Smith F. 1956.** Colorimetric method for determination of sugars and related substances. *Analytical Chemistry* **28(3)**:350–356 DOI 10.1021/ac60111a017.
- **Duchêne J-C. 2017.** Activity rhythm measurement in suspension feeders. In: *Marine Animal Forests*. Berlin: Springer, 761–785.
- **Duchêne J-C, Jordana E, Charles F, Grémare A, Amouroux J-M. 2000.** Experimental study of filtration activity in *Ditrupa arietina* (Annelida Polychaeta) using an automated image analysis system. *Oceanologica Acta* **23**(7):805–817 DOI 10.1016/s0399-1784(00)01124-5.
- Elias-Piera F, Rossi S, Gili JM, Orejas C. 2013. Trophic ecology of seven Antarctic gorgonian species. *Marine Ecology Progress Series* 477:93–106 DOI 10.3354/meps10152.
- **Epp J, Bricelj VM, Malouf RE. 1988.** Seasonal partitioning and utilization of energy reserves in two age classes of the bay scallop *Argopecten irradians irradians* (Lamarck). *Journal of Experimental Marine Biology and Ecology* **121**(2):113–136 DOI 10.1016/0022-0981(88)90250-x.
- Estrada M. 1996. Primary production in the northwestern Mediterranean. *Scientia Marina* 60:55–64.
- Fiorillo I, Rossi S, Alva V, Gili JM, López-González PJ. 2013. Seasonal cycle of sexual reproduction of the Mediterranean soft coral *Alcyonium acaule* (Anthozoa, Octocorallia). *Marine Biology* 160(3):719–728 DOI 10.1007/s00227-012-2126-z.
- Galli G, Bramanti L, Priori C, Rossi S, Santangelo G, Tsounis G, Solidoro C. 2016. Modelling red coral (*Corallium rubrum*) growth in response to temperature and nutrition. *Ecological Modelling* 337:137–148 DOI 10.1016/j.ecolmodel.2016.06.010.
- Galli G, Solidoro C, Lovato T. 2017. Marine heat waves hazard 3D maps and the risk for low motility organisms in a warming Mediterranean Sea. *Frontiers in Marine Science* 4:136 DOI 10.3389/fmars.2017.00136.
- **Garrabou J. 1999.** Life-history traits of *Alcyonium acaule* and *Parazoanthus axinellae* (Cnidaria, Anthozoa), with emphasis on growth. *Marine Ecology Progress Series* **178**:193–204 DOI 10.3354/meps178193.
- Garrabou J, Coma R, Bensoussan N, Bally M, Chevaldonné P, Cigliano M, Diaz D, Harmelin JG, Gambi MC, Kersting DK, Ledoux JB, Lejeusne C, Linares C, Marschal C, Pérez T, Ribes M, Romano JC, Serrano E, Teixido N, Torrents O, Zabala M, Zuberer F, Cerrano C. 2009. Mass mortality in Northwestern Mediterranean rocky benthic communities: effects of the 2003 heat wave. *Global Change Biology* 15(5):1090–1103 DOI 10.1111/j.1365-2486.2008.01823.x.
- Gili JM, Hughes RG. 1995. The ecology of marine benthic hydroids. *Oceanography and Marine Biology: An Annual Review* 33:351–426.

- Gili JM, Ros J. 1985. Study and cartography of the Benthic communities of Medes Islands (NE Spain). *Marine Ecology* 6(3):219–238 DOI 10.1111/j.1439-0485.1985.tb00323.x.
- Gori A, Linares C, Rossi S, Coma R, Gili J-M. 2007. Spatial variability in reproductive cycle of the gorgonians *Paramuricea clavata* and *Eunicella singularis* (Anthozoa, Octocorallia) in the Western Mediterranean Sea. *Marine Biology* 151(4):1571–1584 DOI 10.1007/s00227-006-0595-7.
- Gori A, Viladrich N, Gili JM, Kotta M, Cucio C, Magni L, Bramanti L, Rossi S. 2012.
 Reproductive cycle and trophic ecology in deep versus shallow populations of the Mediterranean gorgonian *Eunicella singularis* (Cap de Creus, northwestern Mediterranean Sea). *Coral Reefs* 31(3):823–837 DOI 10.1007/s00338-012-0904-1.
- Lasker HR, Gottfried MD, Coffroth MA. 1983. Effects of depth on the feeding capabilities of two octocorals. *Marine Biology* 73(1):73–78 DOI 10.1007/bf00396287.
- Lowry O, Rosebrough N, Farr AL, Randall R. 1951. Protein measurement with the folin phenol reagent. *Journal of Biological Chemistry* 193(1):265–275 DOI 10.1016/s0021-9258(19)52451-6.
- Marcus NH, Boero F. 1998. Minireview: the importance of benthic-pelagic coupling and the forgotten role of life cycles in coastal aquatic systems. *Limnology and Oceanography* 43(5):763–768 DOI 10.4319/lo.1998.43.5.0763.
- Martin CS, Giannoulaki M, De Leo F, Scardi M, Salomidi M, Knittweis L, Pace ML, Garofalo G, Gristina M, Ballesteros E, Bavestrello G, Belluscio A, Cebrian E, Gerakaris V, Pergent G, Pergent-Martini C, Schembri PJ, Terribile K, Rizzo L, Ben Souissi J, Bonacorsi M, Guarnieri G, Krzelj M, Macic V, Punzo E, Valavanis V, Fraschetti S. 2014. Coralligenous and maërl habitats: predictive modelling to identify their spatial distributions across the Mediterranean Sea. Scientific Reports 4(1):193 DOI 10.1038/srep05073.
- McArdle BH, Anderson MJ. 2001. Fitting multivariate models to community data: a comment on distance-based redundancy analysis. *Ecology* 82(1):290–297 DOI 10.1890/0012-9658(2001)082[0290:FMMTCD]2.0.CO;2.
- Montresor M, Di Prisco C, Sarno D, Margiotta F, Zingone A. 2013. Diversity and germination patterns of diatom resting stages at a coastal Mediterranean site. *Marine Ecology Progress Series* **484**:79–95 DOI 10.3354/meps10236.
- Mortensen PB, Buhl-Mortensen L. 2004. Distribution of deep-water gorgonian corals in relation to benthic habitat features in the Northeast Channel (Atlantic Canada). *Marine Biology* 144(6):1223–1238 DOI 10.1007/s00227-003-1280-8.
- Palardy JE, Grottoli AG, Matthews KA. 2006. Effect of naturally changing zooplankton concentrations on feeding rates of two coral species in the Eastern Pacific. *Journal of Experimental Marine Biology and Ecology* 331(1):99–107 DOI 10.1016/j.jembe.2005.10.001.
- Preisendorfer RW. 1986. Secchi disk science: visual optics of natural waters1. *Limnology and Oceanography* 31(5):909–926 DOI 10.4319/lo.1986.31.5.0909.
- Previati M, Scinto A, Cerrano C, Osinga R. 2010. Oxygen consumption in Mediterranean octocorals under different temperatures. *Journal of Experimental Marine Biology and Ecology* 390(1):39–48 DOI 10.1016/j.jembe.2010.04.025.
- Pusceddu A, Fraschetti S, Scopa M, Rizzo L, Danovaro R. 2016. Meiofauna communities, nematode diversity and C degradation rates in seagrass (*Posidonia oceanica* L.) and unvegetated sediments invaded by the algae *Caulerpa cylindracea* (Sonder). *Marine Environmental Research* 119:88–99 DOI 10.1016/j.marenvres.2016.05.015.
- Ramón M, Abelló P, Richardson CA. 1995. Population structure and growth of *Donax trunculus* (Bivalvia: Donacidae) in the western Mediterranean. *Marine Biology* 121(4):665–671 DOI 10.1007/bf00349302.

- Rizzo L, Pusceddu A, Bianchelli S, Fraschetti S. 2020. Potentially combined effect of the invasive seaweed *Caulerpa cylindracea* (Sonder) and sediment deposition rates on organic matter and meiofaunal assemblages. *Marine Environmental Research* 159(2):104966 DOI 10.1016/j.marenvres.2020.104966.
- Rizzo L, Pusceddu A, Stabili L, Alifano P, Fraschetti S. 2017. Potential effects of an invasive seaweed (*Caulerpa cylindracea*, Sonder) on sedimentary organic matter and microbial metabolic activities. *Scientific reports* 7(1):12113 DOI 10.1038/s41598-017-12556-4.
- Roberts JM, Wheeler A, Freiwald A, Cairns S. 2009. *Cold-Water Corals*. Cambridge: Cambridge University Press.
- **Rossi S. 2002.** Environmental factors affecting the trophic ecology of benthic suspension feeders. PhD Thesis, University of Barcelona.
- Rossi S, Gili J-M. 2005. Composition and temporal variation of the near-bottom seston in a Mediterranean coastal area. *Estuarine, Coastal and Shelf Science* **65(3)**:385–395 DOI 10.1016/j.ecss.2005.05.024.
- Rossi S, Gili J-M. 2007. Short-time-scale variability of near-bottom seston composition during spring in a warm temperate sea. *Hydrobiologia* 575(1):373–388 DOI 10.1007/s10750-006-0390-y.
- Rossi S, Gili J-M, Coma R, Linares C, Gori A, Vert N. 2006. Temporal variation in protein, carbohydrate, and lipid concentrations in *Paramuricea clavata* (Anthozoa, Octocorallia): evidence for summer autumn feeding constraints. *Marine Biology* 149(3):643–651 DOI 10.1007/s00227-005-0229-5.
- Rossi S, Gravili C, Milisenda G, Bosch-Belmar M, De Vito D, Piraino S. 2019b. Effects of global warming on reproduction and potential dispersal of Mediterranean Cnidarians. *The European Zoological Journal* 86(1):255–271 DOI 10.1080/24750263.2019.1631893.
- Rossi S, Grémare A, Gili J-M, Amouroux J-M, Jordana E, Vétion G. 2003. Biochemical characteristics of settling particulate organic matter at two north-western Mediterranean sites: a seasonal comparison. *Estuarine, Coastal and Shelf Science* **58(3)**:423–434 DOI 10.1016/S0272-7714(03)00108-2.
- Rossi S, Isla E, Bosch-Belmar M, Galli G, Gori A, Gristina M, Ingrosso G, Milisenda G,
 Piraino S, Rizzo L, Schubert N, Soares M, Solidoro C, Thurstan RH, Viladrich N, Willis TJ,
 Ziveri P. 2019a. Changes of energy fluxes in marine animal forests of the Anthropocene: factors shaping the future seascape. *ICES Journal of Marine Science* 76(7):2008–2019
 DOI 10.1093/icesjms/fsz147.
- **Rossi S, Rizzo L. 2020.** Marine animal forests as carbon immobilizers or why we should preserve these three-dimensional alive structures. In: *Perspectives on the Marine Animal Forests of the World*. Berlin: Springer International Publishing, 333–400.
- Rossi S, Rizzo L. 2021. The importance of food pulses in Benthic–Pelagic coupling processes of passive suspension feeders. *Water* 13(7):997 DOI 10.3390/w13070997.
- Rossi S, Rizzo L, Duchêne J-C. 2019. Polyp expansion of passive suspension feeders: a red coral case study. *PeerJ* 7(3):e7076 DOI 10.7717/peerj.7076.
- Rossi S, Snyder MJ, Gili J-M. 2006. Protein, carbohydrate, lipid concentrations and HSP 70 HSP 90 (stress protein) expression over an annual cycle: useful tools to detect feeding constraints in a benthic suspension feeder. *Helgoland Marine Research* 60(1):7–17 DOI 10.1007/s10152-005-0009-0.
- Rossi S, Tsounis G. 2007. Temporal and spatial variation in protein, carbohydrate, and lipid levels in Corallium rubrum (Anthozoa, Octocorallia). *Marine Biology* **152(2)**:429–439 DOI 10.1007/s00227-007-0702-4.

- Sardá R, Pinedo S, Martin D. 1999. Seasonal dynamics of macroinfaunal key species inhabiting shallow soft-bottoms in the Bay of Blanes (NW Mediterranean). *Acta Oecologica* 20(4):315–326 DOI 10.1016/s1146-609x(99)00135-6.
- Sebens KP, DeRiemer K. 1977. Diel cycles of expansion and contraction in coral reef anthozoans. *Marine Biology* 43(3):247–256 DOI 10.1007/bf00402317.
- Spalding M, Spalding MD, Ravilious C, Green EP. 2001. World atlas of coral reefs. University of California Press Marine Pollution Bulletin 44(4):350 DOI 10.1016/S0025-326X(01)00310-1.
- Teixidó N, Bensoussan N, Gori A, Fiorillo I, Viladrich N. 2016. Sexual reproduction and early life-history traits of the Mediterranean soft coral *Alcyonium acaule*. *Marine Ecology* 37(1):134–144 DOI 10.1111/maec.12255.
- Teixidó N, Garrabou J, Harmelin J-G. 2011. Low dynamics, high longevity and persistence of sessile structural species dwelling on Mediterranean Coralligenous Outcrops. *PLOS ONE* 6(8):e23744 DOI 10.1371/journal.pone.0023744.
- Tsounis G, Rossi S, Laudien J, Bramanti L, Fernández N, Gili J-M, Arntz W. 2006. Diet and seasonal prey capture rates in the Mediterranean red coral (*Corallium rubrum L.*). *Marine Biology* 149(2):313–325 DOI 10.1007/s00227-005-0220-1.
- Turon X. 1992. Periods of non-feeding in *Polysyncraton lacazei* (Ascidiacea: Didemnidae): a rejuvenative process? *Marine Biology* 112(4):647–655 DOI 10.1007/bf00346183.
- Turon X, Becerro MA. 1992. Growth and survival of several asoidian species from Ihe northwestern Mediterranean. *Marine Ecology Progress Series* 82:235–247 DOI 10.3354/meps082235.
- Valiela I. 1995. Marine ecological processes. New York: Springer.
- Viladrich N, Bramanti L, Tsounis G, Martinez-Quitana A, Ferrier-Pagès C, Rossi S. 2017. Variation of lipid and free fatty acid contents during larval release in two temperate octocorals according to their trophic strategy. *Marine Ecology Progress Series* 573:117–128 DOI 10.3354/meps12141.