

Full-Genome Sequence Analysis of a Reassortant Strain of *Bluetongue virus* Serotype 16 from Southern India

Lalit Kumar,^a Kanisht Batra,^a Deepika Chaudhary,^a Akhil Kumar Gupta,^a Anita Dalal,^a Brindha Kalyanaraman,^b Ganesan P. Irulappan,^b Vinay Kumar,^a Sushila Maan^a

College of Veterinary Sciences, LLR University of Veterinary and Animal Sciences, Hisar, Haryana, India^a; Centre for Animal Health Studies, Tamil Nadu Veterinary and Animal Sciences University, Madhavaram Milk Colony, Chennai, India^b

The complete genome sequence of a reassortant field strain (IND2014/01) of *Bluetongue virus* (BTV) serotype 16, isolated from sheep from southern India in 2014, was sequenced. The total genome size was 19,186 bp. Sequence comparisons of all genome segments, except segment 5 (Seg-5), showed that IND2014/01 belonged to the major eastern toptotype of BTV.

Received 14 June 2016 Accepted 24 June 2016 Published 18 August 2016

Citation Kumar L, Batra K, Chaudhary D, Gupta AK, Dalal A, Kalyanaraman B, Irulappan GP, Kumar V, Maan S. 2016. Full-genome sequence analysis of a reassortant strain of *Bluetongue virus* serotype 16 from southern India. *Genome Announc* 4(4):e00783-16. doi:10.1128/genomeA.00783-16.

Copyright © 2016 Kumar et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Sushila Maan, sushilamaan105@gmail.com.

Bluetongue virus (BTV) is the type species of the genus *Orbivirus* within the family *Reoviridae* (1), which causes economically important bluetongue disease in domestic and wild ruminants. BTV is a nonenveloped, icosahedral, and double-stranded RNA virus containing three concentric protein layers, transmitted biologically by certain species of *Culicoides* biting midges (2–4). The linear double-stranded RNA (dsRNA) consists of 10 segments encoding seven structural (VP1 to VP7) and five nonstructural (NS1 to NS5) proteins (5, 6). Twenty-seven serotypes of BTV have been reported so far, and there is evidence of two additional putative serotypes (5). There are also characteristic regional variants (topotypes) of each genome segment, which have developed due to separate evolution of BTV strains from different continents by acquiring multiple point mutations, insertion/deletion, and reassortment events. Complete genome data become crucial in order to explicate the emergence and molecular epidemiology of these viruses with segmented genome.

In India, 11 BTV serotypes (BTV-1, BTV-2, BTV-3, BTV-5, BTV-9, BTV-10, BTV-12, BTV-16, BTV-21, BTV-23, and BTV-24) have been isolated since 2001 (6). BTV-16 has been reported earlier from the southern states of India, and at least four complete genomes of BTV-16 have been published from India, China, Australia, and the South African reference (7–10).

We report here the whole-genome sequence analysis of an Indian isolate (IND2014/01) of BTV-16 that was isolated from sheep blood in the Karur district of Tamil Nadu. The virus was isolated using KC cells and then grown in bulk in BHK-21 cells. The viral dsRNA was purified using TRIzol reagent (Life Technologies), which was then used to synthesize cDNA by full-length amplification of cDNA (FLAC) method (11), followed by sequencing on a capillary sequencer using gene-specific primers (11).

Segments 1 to 10 (Seg-1 to Seg-10) of IND2014/01 are 3,945, 2,931, 2,772, 1,981, 1,765, 1,637, 1,156, 1,125, 1,052, and 822 bp, respectively. Phylogenetic analysis showed that IND2014/01 contains genome segments derived mainly from eastern lineages, as segments 1, 3, 7, 8, 9, and 10 have grouped within the eastern

topotype along with viruses from Australia, China, and Far East with very high level of nucleotide and amino acid sequence identities. Analysis of Seg-5 showed its grouping within the western toptotype cluster, a phenomenon that is repeatedly being seen in isolates collected post-1982. Possibly, the western NS1 contributes to enhanced transmission of the virus. Analyses of serotype-determining segments (Seg-2 and Seg-6) have grouped IND2014/01 within serotype 16, with high level of sequence identity (99%) to the previous BTV-16 isolates within the eastern toptotype, confirming its serotype.

Multiple BTV serotypes are currently circulating in the Indian subcontinent (5, 12, 13), thus potentially creating opportunities for the generation and circulation of novel reassortant viruses with unique characteristics. Hence, full-genome constellation analysis and sharing of genomic data are warranted to timely identify the newly emergent viruses.

Accession number(s). The complete genome sequence of BTV isolate IND2014/01 was deposited in GenBank under the accession numbers [KX302634](https://www.ncbi.nlm.nih.gov/nuccore/KX302634) to [KX302643](https://www.ncbi.nlm.nih.gov/nuccore/KX302643).

ACKNOWLEDGMENTS

We acknowledge funding support from BBSRC-DBT (BT/IN/Indo-UK/FADH/46/SM/2013) and DBT Bio-CARE grant (BT/Bio-CARE/04/261/2011-12). Sushila Maan is a DBT Bio-CARE scientist.

FUNDING INFORMATION

This work, including the efforts of Sushila Maan, was funded by Biotechnology and Biological Sciences Research Council (BBSRC [UK]) and Department of Biotechnology, Ministry of Science and Technology (DBT [India]) (BT/IN/Indo-UK/FADH/46/SM/2013 and BT/Bio-CARE/04/261/2011-12) (jointly funded project).

The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

REFERENCES

- Attoui H, Maan S, Anthony SJ, Mertens PPC. 2009. Bluetongue virus, other orbiviruses and other reoviruses: their relationships and taxonomy,

- p 23–46. In Mellor PS, Baylis M, Mertens PPC (ed), Bluetongue monograph, 1st ed. Elsevier/Academic Press, London, United Kingdom.
2. Maan S, Maan NS, Nomikou K, Batten C, Antony F, Belaganahalli MN, Samy AM, Reda AA, Al-Rashid SA, El Batel M, Oura CA, Mertens PP. 2011. Novel bluetongue virus serotype from Kuwait. *Emerg Infect Dis* 17:886–889. <http://dx.doi.org/10.3201/eid1705.101742>.
 3. Mellor PS, Boorman J, Baylis M. 2000. *Culicoides* biting midges: their role as arbovirus vectors. *Annu Rev Entomol* 45:307–340. <http://dx.doi.org/10.1146/annurev.ento.45.1.307>.
 4. Tabachnick WJ. 2004. *Culicoides* and the global epidemiology of bluetongue virus infection. *Vet Ital* 40:144–150.
 5. Maan S, Maan NS, Belaganahalli MN, Rao PP, Singh KP, Hemadri D, Putty K, Kumar A, Batra K, Krishnajyothi Y, Chandel BS, Reddy GH, Nomikou K, Reddy YN, Attoui H, Hegde NR, Mertens PP. 2015. Full-genome sequencing as a basis for molecular epidemiology studies of bluetongue virus in India. *PLoS One* 10:e0131257. <http://dx.doi.org/10.1371/journal.pone.0131257>.
 6. Maan S, Maan NS, Batra K, Kumar A, Gupta A, Rao PP, Hemadri D, Reddy YN, Guimera M, Belaganahalli MN, Mertens PP. 2016. Reverse transcription loop-mediated isothermal amplification assays for rapid identification of eastern and western strains of bluetongue virus in India. *J Virol Methods* 234:65–74. <http://dx.doi.org/10.1016/j.jviromet.2016.04.002>.
 7. Maan S, Maan NS, Singh KP, Belaganahalli MN, Guimera M, Pullinger G, Nomikou K, Mertens PP. 2012. Complete genome sequence analysis of a reference strain of bluetongue virus serotype 16. *J Virol* 86:10255–10256. <http://dx.doi.org/10.1128/JVI.01672-12>.
 8. Minakshi P, Singh R, Ranjan K, Kumar P, Joshi CG, Reddy YK, Prasad G. 2012. Complete genome sequence of bluetongue virus serotype 16 of goat origin from India. *J Virol* 86:8337–8338. <http://dx.doi.org/10.1128/JVI.01128-12>.
 9. Boyle DB, Bulach DM, Amos-Ritchie R, Adams MM, Walker PJ, Weir R. 2012. Genomic sequences of Australian bluetongue virus prototype serotypes reveal global relationships and possible routes of entry into Australia. *J Virol* 86:6724–6731. <http://dx.doi.org/10.1128/JVI.00182-12>.
 10. Yang T, Liu N, Xu Q, Sun E, Qin Y, Zhao J, Wu D. 2011. Complete genomic sequence of bluetongue virus serotype 16 from China. *J Virol* 85:13472. <http://dx.doi.org/10.1128/JVI.06402-11>.
 11. Maan S, Rao S, Maan NS, Anthony SJ, Attoui H, Samuel AR, Mertens PP. 2007. Rapid cDNA synthesis and sequencing techniques for the genetic study of bluetongue and other dsRNA viruses. *J Virol Methods* 143:132–139. <http://dx.doi.org/10.1016/j.jviromet.2007.02.016>.
 12. Krishnajyothi Y, Maan S, Kandimalla K, Maan NS, Tutika RB, Reddy YV, Kumar A, Mrunalini N, Reddy GH, Putty K, Ahmed SM, Reddy YN, Hemadri D, Singh KP, Mertens PP, Hegde NR, Rao PP. 2016. Isolation of bluetongue virus 24 from India—an exotic serotype to Australasia. *Transbound Emerg Dis* 63:360–364. <http://dx.doi.org/10.1111/tbed.12512>.
 13. Rao PP, Hegde NR, Reddy YN, Krishnajyothi Y, Reddy YV, Susmitha B, Gollapalli SR, Putty K, Reddy GH. 2016. Epidemiology of bluetongue in India. *Transbound Emerg Dis* 63:e151–e164.