

Review Article

The Brazilian Journal of INFECTIOUS DISEASES

www.elsevier.com/locate/bjid



Insights into the epidemiology of community-associated methicillin-resistant *Staphylococcus aureus* in special populations and at the community-healthcare interface



Letícia Calixto Romero), Maria de Lourdes Ribeiro de Souza da Cunha 🗅 *

Universidade Estadual Paulista (UNESP), Institute of Biosciences, Department of Chemical and Biological Sciences, Sector of Microbiology and Immunology, Botucatu, SP, Brazil

ARTICLE INFO

Article history: Received 29 June 2021 Accepted 15 September 2021 Available online 19 October 2021

Keywords: Molecular typing Nasal carriage CA-MRSA LA-MRSA MRSA evolution SCCmec

ABSTRACT

The current epidemic proportions of infections caused by Staphylococcus aureus strains and especially by methicillin-resistant S. aureus (MRSA) are one of today's many threats to global public health, particularly in underdeveloped countries where significant gaps on the subject exist. The rapid spread and diversification of pandemic clones that exhibit remarkably increasing virulence and antimicrobial resistance pose a risk to the effective prevention and treatment of a wide range of infections. Undoubtedly, the remarkable versatility involving the pathogenesis and resistance of these bacteria is perpetuated through geographic and temporal factors inherent to clonal evolution and is reflected in the dramatic epidemiological changes of MRSA which, after decades prevailing in healthcare settings, have emerged in the community. Denominated community-associated [CA]-MRSA, these strains are particularly prevalent in some population groups, facilitating the spread of successful clones that are potentially capable of triggering severe community-acquired infections. Therefore, a broad approach to local epidemiological aspects in less studied regions, but nonetheless at latent risk of endemic spread that may reach global proportions, is necessary. In Brazil, despite limited molecular epidemiology data, CA-MRSA strains predominantly characterized as SCCmec IV, often classified as CC30-ST30, CC5-ST5 and CC8-ST8, seem to be spreading across different population groups in different regions of the country. Another important fact addressed in this review is the identification of the ST398-MRSA-IV/V clone and methicillin-susceptible S. aureus (MSSA) in healthy individuals from the community. Although susceptible to methicillin, the ST398 clone is associated with severe infections in humans and animals, denominated livestock-associated MRSA. It is therefore important to encourage assertive actions by all government sectors and by society, with a reassessment of current public health measures in light of the new perspectives arising from the scientific and epidemiological data on MRSA.

© 2021 Sociedade Brasileira de Infectologia. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)

* Corresponding author at.

E-mail address: mlrs.cunha@unesp.br

(M.d.L.R. de Souza da Cunha). https://doi.org/10.1016/j.bjid.2021.101636

^{1413-8670/© 2021} Sociedade Brasileira de Infectologia. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)

The epidemiology of Staphylococcus aureus has undergone a conceptual revolution in recent decades. This phenomenon was due in part to important changes in the epidemiological behavior of these bacteria, but also to a reassessment of old concepts in view of new knowledge arising from clinical and experimental research. Two key factors support the contemporary perspective of *S. aureus*, virulence and antimicrobial resistance. Therefore, a broad and simultaneous approach to these phenomena and the evaluation of their impact on the population are necessary, especially in developing countries such as Brazil where available data are often limited.

Severe infections caused by bacteria that are resistant to the antibiotics commonly used in clinical practice have emerged as a global health problem in the 21st century. Nowadays, antimicrobial resistance is a complex and multifactorial problem that affects society as a whole. Population characteristics, living conditions, agglomerations, injectable drug use, misuse of antibiotics, and underlying diseases all contribute to the selection and perpetuation of resistant bacteria. The current scenario of antimicrobial resistance is a matter of concern and tends to progress to what the World Health Organization calls the "post-antibiotic era", which may be the near future.¹

We continue to fail to contain the spread of resistance genes and we now have clinically important and potentially problematic organisms circulating in the community. Particularly methicillin-resistant *S. aureus* (MRSA) represent one of today's many threats to global public health because of the rapid spread and diversification of pandemic clones that exhibit remarkably increasing virulence and antimicrobial resistance.

Emergence of community-associated Staphylococcus aureus and virulence and resistance determinants

The isolation of MRSA was described for the first time in England in 1961.² In subsequent decades, the prevalence and epidemiology of MRSA have undergone dramatic changes, particularly in the 1990s, with an increase in the number of reports of infections associated with genetically distinct strains originating in the community.³ Thus, MRSA, which until then exclusive agents of healthcare-associated infections (HAIs), started to be recognized as the causal agents of severe disease acquired in the community and were later denominated community-associated [CA]-MRSA.⁴

For a long time, the clinical definition of CA-MRSA infections has been based on the absence of risk factors for hospital-associated MRSA (HA-MRSA) infection, such as a history of recent hospitalization and medical procedures such as dialysis, surgery or catheter use, and many studies continue to use these criteria.⁵ Furthermore, different genetic markers are strongly associated with strains typically defined as HA-MRSA or CA-MRSA. These markers will be addressed below.

The genetic determinant of methicillin resistance in MRSA, the *mecA* gene (which encodes a penicillin binding protein with low affinity, PBP2a), is transported by a mobile genetic element called staphylococcal cassette chromosome *mec* (SCCmec) which, by inference, confers resistance to all other β -lactam antibiotics, except for ceftaroline and ceftobiprole. The latter are fifth-generation cephalosporins indicated for the treatment of infections caused by MRSA.⁶ The acquisition via horizontal gene transfer and insertion of SCCmec into the chromosome of susceptible strains lead to the emergence of resistant staphylococcal strains. The high diversity in the structural organization and genetic content of these elements allows to trace the evolutionary origin of MRSA clones.^{7,8}

Characteristically, the largest SCC*mec* types (I, II and III) are associated with HA-MRSA isolates, while SCC*mec* types IV and V, which are smaller, are frequently carried by typical isolates of CA-MRSA.^{9,10} However, with the rising prevalence and global spread of MRSA in the community over the past three decades, the epidemiological and molecular distinction of the origin of these strains has become blurred, with both CA-MRSA and HA-MRSA strains circulating in the community and concomitantly acting as nosocomial pathogens.^{7,11}

The virulence and pathogenesis of CA-MRSA is somewhat complex and several factors probably contribute to the fitness of these strains during colonization and spread in the population. Studies suggest that the increased virulence of these strains may be associated with their capacity to evade death by neutrophils, the increased production of phenol-soluble modulins, high activity of the accessory gene regulator (Aqr) system, the presence of SCCmec of low fitness cost, and a specific repertoire of toxins.^{4,7} In addition, the production of Panton-Valentine leucocidin (PVL), a cytotoxin that causes tissue necrosis and leukocytes destruction, was originally considered an important determinant of CA-MRSA virulence, playing a key role in the pathogenesis of necrotizing pneumonia.^{7,8,12} However, it is believed that its importance has been overestimated since PVL exerts its aggravating potential only in specific types and scenarios of infection, with the prevalence varying between strains and geographic areas, a fact suggesting that other virulence factors contribute to this process.^{4,7,12}

Molecular epidemiology of CA-MRSA in Brazil

Since their emergence and isolation, CA-MRSA strains have evolved differently in different geographic areas, a fact that has rendered the global epidemiology of these pathogens remarkably heterogeneous, with successful clones predominating in certain regions. This versatility is not only geographical but also temporal, with epidemic clones being replaced with emerging lineages following a pattern of clonal evolution over time.⁷

Certainly, the current situation related to these resistant strains has taken notorious proportions that require the development and improvement of tools capable of distinguishing isolated strains and of outlining the epidemiological panorama of *S. aureus* in the world and over time. In addition to SCC*mec* typing, some methods are useful to identify and to screen for different MRSA clones, particularly multilocus sequence typing (MLST). This method permits to infer clonal relationships between *S. aureus* isolates described in different parts of the world based on allele profiles classified as sequence types (ST), which can be grouped into clusters, the so-called clonal complexes (CC). In Brazil, the first descriptions of CA-MRSA infections date back to the early 2000s and were reported in outpatients without a history of hospitalization or surgery from Porto Alegre, southern region of the country. These isolates belonged to the Oceania Southwest Pacific (OSPC) clone, ST-30-MRSA- IV.¹³ The same clone was isolated in Rio de Janeiro between 2004 and 2006, together with the USA400 clone (ST-1-MRSA-IV); the latter is associated with HAIs.¹⁴ The pandemic clone USA300 (ST-8-MRSA-IV), which caused respiratory infection in a patient from Porto Alegre, was also detected in the same study.¹⁴ Both studies identified isolates carrying the *lukF* gene, which encodes PVL.

Until the rise of CA-MRSA, the multidrug-resistant clone ST239-SCC*me*cIIIA, known as the Brazilian epidemic clone (BEC), and its variants predominated as nosocomial pathogens throughout the country.^{15–17} In the early 2000s, the ST5-SCC*me*cIV clone, known as the pediatric clone, which was not multidrug-resistant at first, was identified in Brazilian hospitals.^{18,19} Subsequently, this lineage seems to have developed multidrug resistance, as well as important virulence factors such as biofilm formation and enterotoxin production.^{15,20}

Nowadays, in Latin America, SCC*mec* type IV is frequently detected in clones circulating in the community and in healthcare settings.²¹ In Brazil, infections with CA-MRSA have been reported in the cities of Porto Alegre, Rio de Janeiro, São Paulo, Botucatu, Recife, and Salvador, which ranged from skin and soft tissue infections to necrotizing pneumonia and severe sepsis. These infections were associated with clones harboring SCC*mec* type IV, similar to the OSPC clone (ST-30-MRSA-IV), and frequently carrying PVL-coding genes.^{13,14,22–28}

Our research group also described a case of systemic CA-MRSA infection in an adolescent living in a small town in the interior of São Paulo (Bofete/SP), who had no history of healthcare exposure or recent travel.²⁷ The isolate was characterized as ST5-MRSA-IV, carrying the genes encoding PVL and enterotoxin, and belonged to the same clonal complex as a lineage described in Argentina²⁹ that is genetically related to the pediatric clone. These findings alert to the presence of CA-MRSA in small towns in rural Brazil and reinforce the need for comprehensive studies that contribute to elucidating the prevalence of these clones. Certainly, the spread of successful clones, especially in reservoirs of the community, facilitates the transport of endemic strains into households and puts the population at increased risk of infection.³⁰

Community prevalence of Staphylococcus aureus and CA-MRSA

Infections with CA-MRSA are particularly prevalent in some population groups. These groups include sportsmen, men who have sex with men, prison inmates, injection drug users, military personnel, children, people in correctional facilities or shelters, people living in overcrowded or low socioeconomic conditions, HIV-infected patients, and members of remote populations (such as Australian Pygmies and Aborigines), as well as American Indians, Alaskan Natives, and Pacific Islanders.³¹ However, it should be noted that all of these data are derived from sparse studies conducted in different countries. The studies are diverse in terms of objectives and methods and do not allow to draw a complete and coherent picture of the global epidemiology of CA-MRSA. In addition, there are significant gaps, with limited information for regions in Africa and South America, and the imminent endemic spread of CA- MRSA in underdeveloped countries has raised concern regarding probable devastating global consequences.⁷

Studies on the prevalence of asymptomatic *S. aureus* carriage are important since colonization is a possible precursor stage of invasive disease.³² Furthermore, a model developed by Macal et al.³³ to study the dynamics of transmission of and infection with CA-MRSA in Chicago demonstrated that the majority of transmission events of these strains originated from asymptomatic carriers of MRSA and, less frequently, from individuals with active infection. This fact is interesting, considering that colonization is more common and usually lasts longer compared to infection, and should encourage the reassessment of public measures focused only on the control of clinically apparent infections.³³

In 2011, our research group conducted a population-based survey that identified a prevalence of asymptomatic nasal S. *aureus* carriers of 32.7%, with six out of 686 urban residents of Botucatu, São Paulo, Brazil, being colonized with CA-MRSA (prevalence of 0.9%).³⁴ All isolates carried SCC*mec* type IV and belonged to CC5. The study also estimated more than 1,000 MRSA carriers in the city.

Our findings agree with the results of the National Health and Nutrition Examination Survey (NHANES) which, in 2001-2002, estimated a 30.8% prevalence of *S. aureus* in the community and a 0.8% prevalence of colonization with MRSA in the United States.³⁵ In subsequent years, more precisely in 2003-2004, the estimated prevalence of *S. aureus* exhibited a statistically significant reduction, dropping to 27.1%. On the other hand, MRSA colonization rates almost doubled during the same period (from 0.8% to 1.5%).³⁶ It should be noted that, up to 2001, the most prevalent CA- MRSA clone in the United States was USA400 (ST1-SCC*mec* IV), which was largely replaced by the USA300 clone (ST8-SCC*mec* IVa), one of the most successful clones of all time,⁸ although it is currently declining.³⁷

Other studies were conducted in Brazil to investigate asymptomatic MRSA colonization in different population groups. Some of those studies provided data on clonal characterization, which are summarized in Table 1. The majority of studies are concentrated in the state of São Paulo and generally indicate a rate of S. aureus colonization very similar to that described in the population-based study conducted in Botucatu, with the observation of higher rates among users of primary healthcare services, children attending public day care centers, and indigenous people. On the other hand, despite the correlated predominance of SCCmec IV, the prevalence of MRSA carriage was higher in the other studies, except for one involving prison inmates and indigenous people. These results are somewhat intriguing and disagree with data previously described for prisoners, American Indians and Australian Aborigines.^{36,38}

Table 1 – Prevalence of MRSA colonization outside hospital settings reported in Brazil.								
Population (n)	Source of isolation	Prevalence of colonization		SCCmec type		Year	Reference	
		S. aureus	MRSA		(city/state)			
Patients with insulin- dependent diabetes (312)	N, O	30.4	4.8	IV, I, II	Botucatu/SP	2015/18	44	
Nursing home resi- dents (226)	N, O, A	33.6	8	IV, I, II	Botucatu/SP	2019	46	
Nursing home resi- dents (300)	Ν	17.7	3.7	II, IV	Bauru/SP	2017	45	
Illicit drug users and alcoholics (138)	N, O	28.3	2.9	IV, I	Botucatu/SP	2020	50	
Psychiatric patients (82)	N, O	24.3	7.3	IV, I, II	Botucatu/SP	2020	50	
Prison inmates (302)	Ν	16.5	0.7	IV	Avaré/SP	2009/10	76	
Patients with wounds attending basic health units (171)	N, W	51.5	8.7	IV, II	Botucatu/SP	2010/13	52	
Healthcare workers of Family Health Strat- egy Units (63)	Ν	74.6	53.9	IV, III, I	Cidade do Oeste Pau- lista/SP	2017	56	
Indigenous people (400)	N, O	47.6	0.7	IV	Indigenous communi- ties in northern and southeastern Brazil *	2017	77	
Urban population (686)	Ν	32.7	0.9	IV	Botucatu/SP	2011	34	
Children (≤ 5 years) attending day care centers (1,192)	Ν	31.1	1.2	III, IV	Goiânia/GO	2005	55	
Healthy children (1 to 6 years) attending public day care cen- ters (148)	Ν	47.3	7.4	IV, V	Vitória da Conquista/BA	2019	53	
People living with HIV/ AIDS (368)	N, O	26.0	2.7	IV	Botucatu and region/SP	2018	66	
People living with HIV/ AIDS (500)	Ν	31.4	4.4	II, V	Recife/PE	2018	78	

N: nasal; O: oropharynx; A: anal; W: wounds; n: number of subjects; SP: São Paulo (southeastern region); GO: Goiás (midwest region); BA: Bahia (northeastern region).

* Kopenoti and Tereguá indigenous villages located in the municipality of Avaí, São Paulo, and Kaxinawá, Ashaninka, Shanenawa and Poyanawas indigenous villages located in the municipality of Cruzeiro do Sul, Acre.

Epidemiology of Staphylococcus aureus and CA-MRSA in special populations

An important facet of the epidemiology of S. aureus is the fact that these bacteria infect "special populations". This term refers to population strata that can be differentiated based on ecological pressures and/or specific conditions of morbidity, such as the elderly, bedridden patients and patients with chronic diseases, particularly diabetes mellitus. Studies suggest that the rate of nasal colonization with S. aureus and MRSA may be higher in diabetic patients than in non-diabetic subjects.^{35,39,40} Within this context, an influence of glycemic control on the S. aureus carriage rate has been demonstrated, considering that hyperglycemia reduces the activation of macrophages.^{39,40} These patients are also more susceptible to persistent infections, particularly skin infections such as those caused by S. aureus, and are more likely to develop severe MRSA pneumonia due to increased blood glucose levels and suppression of the immune response.^{41,42} A prospective cohort study showed that patients with both type 1 and

type 2 diabetes mellitus had an increased risk of bacterial skin infections; a higher risk of recurrence is observed in patients with type 1 diabetes, which can be explained by the higher frequency of nasal carriage observed among insulin-dependent diabetic patients.⁴³ A recent study conducted by our group evaluated nasal and oropharyngeal MRSA carriage in patients with insulin-dependent diabetes from Botucatu and found a 30.4% prevalence of colonization with S. *aureus* and a 4.8% prevalence of colonization with MRSA.⁴⁴ The S. *aureus* colonization rate was very similar to the overall prevalence found in the population of the same city in a previous study.³⁴ However, MRSA isolates were more frequent among diabetic patients (4.8% vs 0.9%).

The emergence of CA-MRSA also poses a special risk to known vulnerable populations such as the elderly, especially residents of long-term care facilities. Within this context, the prevalence of colonization with S. *aureus* and MRSA among elderly residents of nursing homes and bedridden patients from Botucatu and Bauru (interior of São Paulo) was investigated by two different studies. In Bauru, the prevalence of S. *aureus* and MRSA nasal carriage was 17.7% and 3.7%, respectively.⁴⁵ On the other hand, a higher colonization rate was observed in Botucatu, which reached 33.6% for S. *aureus* and 8% for MRSA, probably because the analysis included extra-nasal sites (oropharynx and anal).⁴⁶

Long-term care facilities are intermediate spaces between the community and the hospital and may represent a possible reservoir of multidrug-resistant microorganisms.⁴⁷ Furthermore, the social and interactive characteristics of nursing homes, such as shared meals, recreation and therapeutic facilities, certainly facilitate the spread of communicable diseases.48 However, little is known about the prevalence of MRSA among older adults, with rates ranging from 0.7 to 2.0% in studies conducted around the world that are in contrast to the above-mentioned findings.⁴⁹ In particular, a recent study investigating nasopharyngeal colonization in 776 older outpatients attending the largest geriatric clinic in the city of São Paulo found a prevalence of S. aureus and MRSA carriage of 15.9% and 2.3%, respectively.⁴⁹ Here it is important to note that nursing homes in Brazil can be public, private or philanthropic, and the quality of care varies widely, a fact that influences the spread of pathogens.45

As reported for diabetic patients in the aforementioned study,⁴⁴ SCC*mec* characterization of MRSA isolated from older adults living in the same region showed a predominance of SCC*mec* types I, II and IV,^{45, 46} with SCC*mec* I and II occasionally being risk factors for HA-MRSA. Similar SCC*mec* typing results were obtained for MRSA isolated from patients of a psychiatric hospital and from outpatients with skin infections treated at a dermatology clinic in the same city.^{50,51}

Another interesting study drew attention to the circulation of strains that were resistant to different classes of antimicrobial agents in the community, demonstrating a high prevalence of *S. aureus* (51.5%) and MRSA (8.7%) isolated from patients with skin wounds treated at primary healthcare centers in Botucatu, with no reports of risk factors for HA-MRSA. The strains carried SCCmec II and IV and a positive association was found between nasal carriage of *S. aureus* and MRSA and their respective presence in the wound.⁵² However, the gene encoding PVL was not detected, a finding that is consistent with the low prevalence of this gene in MRSA-IV reported in Brazil.⁵²

Clonal diversity of CA-MRSA and epidemiological changes

Studies investigating the clonal profile of CA-MRSA isolates throughout Brazil identified strains predominantly characterized as SCC*mec* IV, including those that cause staphylococcal infections; special attention must be paid to clone ST5-IV whose frequent identification suggests its spread, particularly in Botucatu and region. These and other studies on the clonal characterization of MRSA isolated from the community in Brazil are described in Table 2, which provides an overview of CA-MRSA clones in Brazil reported in the literature.

Reports of isolates carrying SCCmec type V are less common in Brazil, and SCCmec III isolates are usually multidrug-resistant and associated with HAIs.^{53,54} However, both types were found colonizing the nares of healthy children attending day care centers.^{53,55} In fact, strains carrying SCCmec III seem to be adapting to the community and are no longer restricted to the hospital.⁵⁵ SCCmec III MRSA isolates have also been described colonizing health professionals working in primary healthcare centers in the inland of the country, including community health agents who perform tasks that require a greater level of contact with the population.⁵⁶ Like SCCmec III, SCCmec types I and II are commonly found circulating in health services, including primary healthcare settings, as reported by Pereira-Franchi et al.^{52,57} and Goes et al.⁵⁶; a history of hospitalization is frequently associated with this finding.

Interesting findings from a study conducted in a hospital in the interior of São Paulo indicate polyclonal endemicity, with hegemony of SCCmec III but unrelated to the Brazilian endemic clone, and suggest wide dissemination of MRSA in Brazilian hospitals, in which SCCmec IV isolates are acquired in the healthcare settings.⁵⁸ In fact, several epidemiological studies have reported blurring of boundaries between CA-MRSA and HA-MRSA, indicating a significant overlap between the two groups.⁵⁹ Nevertheless, SCCmec typing continues to be an important tool to elucidate the evolution of MRSA, as well as to understand and monitor constant epidemiological changes and to guide therapeutic decisions. The concept of

Clonal complex	Sequence type	SCCmec type	Study location (city)	References	
CC5	ST5	IV	Botucatu, Avaré, Bofete, São Paulo, Vitória da Conquista, Pardinho	27, 34, 44, 53,57, 66, 76, 77	
		Ι	Botucatu	44	
		II	Botucatu	57	
	ST2594	IV	Botucatu	34	
	ST1176	IV	Botucatu, Fartura	34, 57, 66	
	ST6	IV	Botucatu	66	
CC8	ST8	IV	Botucatu, Porto Alegre, Paranapanema	44, 66, 57, 14,25, 66	
	ST239	III	Goiânia	55	
CC45	ST1120	V	Goiânia	55	
	ST45	IV	Porto Alegre, Vitória da Conquista	25, 53	
	ST2228	IV	Vitória da Conquista	53	
CC1	ST1	IV	Rio de Janeiro, Porto Alegre	14, 20	
CC30	ST30	IV	Porto Alegre, Rio de Janeiro, Goiânia, Botucatu	13, 14, 79, 55,80, 25, 66	
CC121	ST121	IV	Goiânia	55	
Singleton	ST12	IIIa	Goiânia	55	

CC: Clonal complex; ST: Sequence type.

SCC*mec* elements should be revised considering their notorious ability to carry other genes that are essential for the increased survival of staphylococci in different environments. 11,59

A study conducted in Bahia identified a high prevalence of nasal colonization with CA-MRSA (47.3%) in healthy children attending public day care centers, with the description of lineages belonging to CC5 (ST5), CC45 (ST45 and ST2228) and CC398 (ST389) that carry SCCmec IV and V.⁵³ Interestingly, the ST398 clone, which is traditionally associated with livestock, has emerged among human patients, including in Brazil, and will be discussed below. Complexes CC5 and CC45 belong to clonal groups involved in a global pandemic caused by MRSA; however, CC5 lineages are notably the most frequently detected in Brazil, especially ST5. Other clones included in CC5 have also been reported in Brazil, such as ST2594, ST1176 and ST6. The ST1176 clone was described for the first time in S. aureus SCCmec IV of patients admitted to a hospital in the city of São Paulo and was identified in Botucatu (140 miles away from São Paulo) by two community studies; one of these studies also identified a new clone, ST2594, circulating in the city.^{34,57,60}

Reports of CA-MRSA are less common in European countries where infections with MRSA are typically healthcare associated.⁶¹ Characteristically, there is an extensive clonal diversity of CA-MRSA in Europe, with a high degree of geographic segregation of clones, associated with a considerably low but rising prevalence in some countries.⁶² In general, the European clone ST80-MRSA-IV (lukSF-PV positive) predominates, although the USA300 clone (ST8-MRSA-IV) has been reported throughout the United Kingdom and Europe.⁶² Greece in particular is the country with the highest incidence of CA-MRSA in Europe.⁶³ On the other hand, CA-MRSA infections have emerged in Nordic countries and in The Netherlands, where rates of HA-MRSA are very low.⁶¹ An important epidemiological aspect in The Netherlands is the emergence of clone ST398-V, which is responsible for about 20% of cases of MRSA infection in the country.^{61,62}

Clone ST398

Described for the first time in France, the CC398-ST398 lineage originated in animals and is one of the most important genotypes of livestock-associated S. *aureus* (LA-MRSA). First documented in countries with intensive animal farming (cattle, horses, pigs), these microorganisms are increasingly spreading among humans not exposed to these animals.⁸ Human infections with the ST398 clone have sporadically been reported in various geographic regions, including Europe, the Americas and Asia, with a higher prevalence in Europe and China.⁶⁴

Cases of human colonization and infection with methicillin-susceptible S. *aureus* (MSSA)-CC398 appear to be more common, whereas MRSA-ST398 strains are mainly detected in animals due to clonal adaptation.⁶⁵ These strains are generally not associated with the production of PVL or enterotoxins, but exhibit a remarkable diversity of resistance genes, including resistance to trimethoprim, tetracycline, macrolides, lincosamides, gentamicin, ciprofloxacin and trimethoprim-sulfamethoxazole, as well as to antibiotics used in livestock farming.^{7,8} Other important features of this emerging clone include its ability to acquire virulence genes, low host specificity, and skilled mobility and spread, characteristics that render this lineage a real threat.⁷

In Brazil, asymptomatic colonization with the ST398 clone was identified in MSSA and MRSA isolated from diabetic patients (MSSA/MRSA-ST398-IV)⁴⁴ and people living with HIV/AIDS (MSSA-ST398)⁶⁶ in Botucatu/SP, healthy children in Vitória da Conquista/BA (MRSA-ST398-V),⁵³ and children attending day care centers and public hospitals in Niterói/RJ (MRSA-ST398-IV/V). The genes encoding PVL were detected in these isolates.⁶⁷ These findings indicate that the ST398 clone might be emerging in the community in different parts of the country and in different population groups, with this occurrence being underreported, as suggested by Neto et al.⁶⁷

The ST398 clone has also occasionally been reported to cause severe infections in Brazil. A prospective study conducted at the Hospital of the Botucatu Medical School revealed a considerable number of MSSA isolates that belonged to the ST398 clone and that were associated with cases of pneumonia and a higher mortality rate.⁶⁸ For example, Gales et al.⁶⁹ reported a case of fatal pneumonia caused by MSSA-ST398 in a cancer patient from São Paulo.⁶⁹

MRSA and COVID-19

The current circumstances of the COVID-19 pandemic, as well as of previous pandemics, raise important issues that need to be investigated. Studies show that about half of the patients hospitalized with COVID-19 who die are co-infected with fungi and bacteria.^{70,71} The widespread use of antibiotics in the treatment of these bacterial co-infections highlights the importance of considering possible effects on the global prevalence of antibiotic-resistant bacteria.⁷²

During an outbreak of the so-called severe acute respiratory syndrome (SARS-CoV-1) in 2003, an increased frequency of MRSA transmission and infection was reported among intensive care unit patients, which was strongly linked to ventilator-associated pneumonia.⁷³ Furthermore, bacterial pathogens, including *S. aureus*, have been recognized to be involved in infections secondary to influenza and are a common cause of post-influenza bacterial pneumonia. Within this context, the emergence of CA-MRSA strains in recent decades has changed the clinical scenario of these infections.⁷⁴

Certainly, the current pandemic scenario will lead to significant changes in the pattern of endemic pathogens such as *S. aureus.* Studies highlight the complex and multifaceted interrelationships and interdependence of antimicrobial resistance determinants in past, current and future pandemics. Potential interventions to support the reduction of antimicrobial prescriptions during the COVID-19 pandemic require urgent consideration.⁷⁵

Conclusion

We understand that a broad and simultaneous approach to the special populations described above, combining epidemiological strategies and the genetic characterization of staphylococci, will provide valuable information. We also believe that the evaluation of different groups will provide insights into the pathogenesis and spread of MRSA strains.

Cases of CA-MRSA colonization have been described in all regions of Brazil; however, the use of molecular typing techniques that provide more refined data on the epidemiology of these pathogens in Brazil is still limited. In addition, available data on CA-MRSA infections in Brazil are mainly derived from case reports. It is nevertheless possible to catch a glimpse of aspects of the epidemiology of these pathogens in the country, recognizing possible reservoirs in the community, especially children attending day care centers, institutionalized older adults, and diabetic patients.

On the other hand, some studies have reported the lack of an epidemiological link between the type of SCC*mec* and a history of hospital admission, corroborating the current global scenario of the widespread transmission of MRSA isolates between the community and hospitals. All of these points, together with the lack of epidemiological surveillance of *S. aureus* and MRSA in Brazil, highlight the importance of the adoption of assertive strategies by all government sectors and by society, with a reassessment of current public policies in light of the new perspectives arising from epidemiological findings.

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

We thank the National Council for Technological and Scientific Development (CNPq; Grant 303603/2020-8) and PROPG Edital 19/2021.

REFERENCES

- 1. Hwang AY, Gums JG. The emergence and evolution of antimicrobial resistance: impact on a global scale. Bioorg Med Chem. 2016;24:6440–5.
- Jevons MP. "Celbenin" resistant Staphylococci. Br Med J. 1961;1:124–5.
- Chambers HF, DeLeo FR. Waves of resistance: Staphylococcus aureus in the antibiotic era. Nat Rev Microbiol. 2009;7:629–41.
- 4. Otto M. Community-associated MRSA: what makes them special? Int J Med Microbiol. 2013;303:324–30.
- Cataldo MA, Taglietti F, Petrosillo N. Methicillin-resistant Staphylococcus aureus: a community health threat. Postgrad Med. 2010;122:16–23.
- CLSI. Performance Standards for Antimicrobial Susceptibility Testing. CLSI supplement M100. 28th ed. Wayne, PA: Clinical and Laboratory Standards Institute; 2018. p. 258.
- Lakhundi S, Zhang K. Methicillin-resistant Staphylococcus aureus: molecular characterization, evolution, and epidemiology. Clin Microbiol Rev. 2018;31. e00020-18.
- 8. Monaco M, Pimentel de Araujo F, Cruciani M, Coccia EM, Pantosti A. Worldwide epidemiology and antibiotic resistance

of Staphylococcus aureus. Curr Top Microbiol Immunol. 2017;409:21–56.

- 9. Ito T, Ma XX, Takeuchi F, Okuma K, Yuzawa H, Hiramatsu K. Novel Type V staphylococcal cassette chromosome *mec* driven by a novel cassette chromosome recombinase, *ccrC*. Antimicrob Agents Chemother. 2004;48:2637–51.
- Ma XX, Ito T, Tiensasitorn C, et al. Novel type of staphylococcal cassette chromosome mec identified in community-acquired methicillin- resistant Staphylococcus aureus strains. Antimicrob Agents Chemother. 2002;46:1147– 52.
- David MZ, Glikman D, Crawford SE, et al. What is communityassociated methicillin-resistant Staphylococcus aureus? J Infect Dis. 2008;197:1235–43.
- 12. Lee AS, de Lencastre H, Garau J, et al. Methicillin- resistant Staphylococcus aureus. Nat Rev Dis Primer. 2018;4:1–23.
- Ribeiro A, Dias C, Silva-Carvalho MC, et al. First report of infection with community-acquired methicillin-resistant Staphylococcus aureus in South America. J Clin Microbiol. 2005;43:1985–8.
- 14. Ribeiro A, Coronado AZ, Silva-Carvalho MC, et al. Detection and characterization of international community-acquired infections by methicillin- resistant Staphylococcus aureus clones in Rio de Janeiro and Porto Alegre cities causing both community- and hospital-associated diseases. Diagn Microbiol Infect Dis. 2007;59:339–45.
- Andrade MM, Luiz WB, da Silva Oliveira Souza R, Amorim JH. The history of methicillin- resistant *Staphylococcus aureus* in Brazil. Can J Infect Dis Med Microbiol. 2020;2020: 1–18.
- 16. Sader HS, Pignatari AC, Hollis RJ, Jones RN. Evaluation of interhospital spread of methicillin- resistant Staphylococcus aureus in Sao Paulo, Brazil, using pulsed-field gel electrophoresis of chromosomal DNA. Infect Control Hosp Epidemiol. 1994;15:320–3.
- Teixeira LA, Resende CA, Ormonde LR, et al. Geographic spread of epidemic multiresistant Staphylococcus aureus clone in Brazil. J Clin Microbiol. 1995;33:2400–4.
- de Miranda OP, Silva-Carvalho MC, Ribeiro A, et al. Emergence in Brazil of methicillin-resistant Staphylococcus aureus isolates carrying SCCmecIV that are related genetically to the USA800 clone. Clin Microbiol Infect. 2007;13:1165–72.
- Oliveira GA, Faria JB, Levy CE, Mamizuka EM. Characterization of the Brazilian endemic clone of methicillin-resistant Staphylococcus aureus (MRSA) from hospitals throughout Brazil. Braz J Infect Dis. 2001;5:163–70.
- 20. Silva-Carvalho MC, Bonelli RR, Souza RR, et al. Emergence of multiresistant variants of the community-acquired methicillin-resistant *Staphylococcus aureus* lineage ST1-SCCmecIV in 2 hospitals in Rio de Janeiro, Brazil. Diagn Microbiol Infect Dis. 2009;65:300–5.
- Rodríguez-Noriega E, Seas C, Guzmán-Blanco M, et al. Evolution of methicillin-resistant Staphylococcus aureus clones in Latin America. Int J Infect Dis. 2010;14:e560–6.
- 22. Caraciolo FB, Maciel MAV, Santos JB dos, Rabelo MA, Magalhães V. Antimicrobial resistance profile of Staphylococcus aureus isolates obtained from skin and soft tissue infections of outpatients from a university hospital in Recife -PE, Brazil. An Bras Dermatol. 2012;87:857–61.
- 23. Fortes CQ, Espanha CA, Bustorff FP, et al. First reported case of infective endocarditis caused by community-acquired methicillin-resistant *Staphylococcus aureus* not associated with healthcare contact in Brazil. Braz J Infect Dis. 2008;12:541–3.
- 24. d'Azevedo PA, Inoue FM, Andrade SS, Tranchesi R, Pignatari ACC. Pneumonia necrotizante por Staphylococcus aureus resistente à meticilina. Rev Soc Bras Med Trop. 2009;42:461–2.
- 25. Gelatti LC, Bonamigo RR, Inoue FM, et al. Communityacquired methicillin-resistant Staphylococcus aureus carrying

SCCmec type IV in southern Brazil. Rev Soc Bras Med Trop. 2013;46:34–8.

- 26. Gelatti LC, Sukiennik T, Becker AP, et al. Sepse por Staphylococus aureus resistente à meticilina adquirida na comunidade no sul do Brasil. Rev Soc Bras Med Trop. 2009;42:458–60.
- 27. Camargo CH, da Cunha M de LR de S, Bonesso MF, da Cunha FP, Barbosa AN, Fortaleza CMCB. Systemic CA-MRSA infection following trauma during soccer match in inner Brazil: clinical and molecular characterization. Diagn Microbiol Infect Dis. 2013;76:372–4.
- 28. Gomes RT, Lyra TG, Alves NN, Caldas RM, Barberino M-G, Nascimento-Carvalho CM. Methicillin-resistant and methicillin-susceptible community-acquired Staphylococcus aureus infection among children. Braz J Infect Dis. 2013;17:573–8.
- 29. Sola C, Paganini H, Egea AL, et al. Spread of epidemic MRSA-ST5-IV clone encoding PVL as a major cause of community onset staphylococcal infections in Argentinean children. PLoS ONE. 2012;7:e30487.
- **30.** Knox J, Uhlemann A-C, Lowy FD. Staphylococcus aureus infections: transmission within households and the community. Trends Microbiol. 2015;23:437–44.
- **31.** Hansra NK, Shinkai K. Cutaneous community-acquired and hospital-acquired methicillin- resistant *Staphylococcus aureus*: cutaneous MRSA. Dermatol Ther. 2011;24:263–72.
- Jarvis WR. The epidemiology of colonization. Infect Control Hosp Epidemiol. 1996;17:47–52.
- 33. Macal CM, North MJ, Collier N, et al. Modeling the transmission of community-associated methicillin-resistant Staphylococcus aureus: a dynamic agent-based simulation. J Transl Med. 2014;12:124.
- 34. Pires FV, da Cunha M de LR de S, Abraão LM, Martins PYF, Camargo CH, Fortaleza CMCB. Nasal carriage of Staphylococcus aureus in Botucatu, Brazil: a population-based survey. PloS One. 2014;9:e92537.
- Graham PL, Lin SX, Larson EL. A U.S. population-based survey of Staphylococcus aureus colonization. Ann Intern Med. 2006;144:318–25.
- Malcolm B. The Rise of Methicillin-Resistant Staphylococcus aureus in U.S. Correctional Populations. J Correct Health Care. 2011;17:254–65.
- Glaser P, Martins-Simões P, Villain A, et al. Demography and intercontinental spread of the USA300 community-acquired methicillin-resistant Staphylococcus aureus Lineage. mBio. 2016;7. e02183-15.
- 38. Tong SYC, McDonald MI, Holt DC, Currie BJ. Global Implications of the Emergence of Community-Associated Methicillin-Resistant Staphylococcus aureus in Indigenous Populations. Clin Infect Dis. 2008;46:1871–8.
- Lipsky BA, Pecoraro RE, Chen MS, Koepsell TD. Factors affecting staphylococcal colonization among NIDDM outpatients. Diabetes Care. 1987;10:483–6.
- 40. Wertheim HFL, Melles DC, Vos MC, van Leeuwen W, van Belkum A, Verbrugh HA, et al. The role of nasal carriage in Staphylococcus aureus infections. Lancet Infect Dis. 2005;5:751–62.
- Trivedi U, Parameswaran S, Armstrong A, et al. Prevalence of multiple antibiotic resistant infections in diabetic versus nondiabetic wounds. J Pathog. 2014;2014:1–6.
- Zhang Q-R, Chen H, Liu B, Zhou M. Methicillin-resistant Staphylococcus aureus pneumonia in diabetics: a single-center, retrospective analysis. Chin Med J (Engl). 2019;132(12):1429–34.
- **43.** Muller LM aJ, Gorter KJ, Hak E, et al. Increased risk of common infections in patients with type 1 and type 2 diabetes mellitus. Clin Infect Dis. 2005;41:281–8.
- 44. Teixeira NB, Castelo Branco Fortaleza CM, de Souza MC, Monteiro Pereira TA, de Camargo Colenci BP, Ribeiro de Souza da Cunha M de L. Molecular characterization of

methicillin- resistant Staphylococcus aureus among insulindependent diabetic individuals in Brazil. Ann Clin Microbiol Antimicrob. 2021;20:1–12.

- **45.** da Silveira M, da Cunha M de LR de S, de Souza CSM, Correa AAF, Fortaleza CMCB. Nasal colonization with methicillin-resistant *Staphylococcus aureus* among elderly living in nursing homes in Brazil: risk factors and molecular epidemiology. Ann Clin Microbiol Antimicrob. 2018;17:18.
- 46. Silva LP. Epidemiologia Molecular de Staphylococcus aureus em Pacientes Acamados em Domicílio ou Vivendo em Instituições de Longa Permanência Para Idosos no Município de Botucatu, SP. Botucatu: Universidade Estadual Paulista Júlio de Mesquita Filho, Instituto de Biociências; 2019. [dissertation on the Internet][cited 2021 Jun 5]Available from: https://repositorio. unesp.br/handle/11449/183106.
- **47**. Jans B, Schoevaerdts D, Huang T-D, et al. Epidemiology of multidrug-resistant microorganisms among nursing home residents in Belgium. PloS One. 2013;8:e64908.
- 48. Dumyati G, Stone ND, Nace DA, Crnich CJ, Jump RLP. Challenges and strategies for prevention of multidrugresistant organism transmission in nursing homes. Curr Infect Dis Rep. 2017;19:18.
- **49**. Zanella RC, Brandileone MC de C, Almeida SCG, et al. Nasopharyngeal carriage of Streptococcus pneumoniae, Haemophilus influenzae, and Staphylococcus aureus in a Brazilian elderly cohort. PLoS ONE. 2019;14:e0221525.
- 50. Silvestre MAD de O, Barbosa M, Teixeira NB, et al. Illicit drug users, alcoholics, and psychiatric patients: Staphylococcus aureus and methicillin-resistant Staphylococcus aureus colonization on the border between community and healthcare settings. Infect Control Hosp Epidemiol. 2020: 1–3.
- 51. Bonesso MF, Marques SA, Camargo CH, Fortaleza CMCB, Cunha MLRS. Community- associated methicillin-resistant Staphylococcus aureus in non-outbreak skin infections. Braz J Microbiol. 2014;45:1401–7.
- 52. Pereira-Franchi EPL, Barreira MRN, Costa NSLM, Fortaleza CMCB, Cunha MLRS. Prevalence of and risk factors associated with the presence of *Staphylococcus aureus* in the chronic wounds of patients treated in primary health care settings in Brazil. Rev Soc Bras Med Trop. 2017;50:833–8.
- 53. de Carvalho SP, de Almeida JB, Andrade YMFS, et al. Molecular characteristics of methicillin-resistant *Staphylococcus aureus* isolates from hospital and community environments in northeastern Brazil. Braz J Infect Dis. 2019;23:134–8.
- 54. Caboclo RMF, Cavalcante FS, Iorio NLP, et al. Methicillinresistant Staphylococcus aureus in Rio de Janeiro hospitals: Dissemination of the USA400/ST1 and USA800/ST5 SCCmec type IV and USA100/ST5 SCCmec type II lineages in a public institution and polyclonal presence in a private one. Am J Infect Control. 2013;41:e21–6.
- 55. Lamaro-Cardoso J, de Lencastre H, Kipnis A, et al. Molecular epidemiology and risk factors for nasal carriage of Staphylococcus aureus and Methicillin-Resistant S. aureus in infants attending day care centers in Brazil. J Clin Microbiol. 2009;47:3991–7.
- 56. Goes ICR dos S, Romero LC, Turra AJ, et al. Prevalence of nasal carriers of methicillin-resistant Staphylococcus aureus in primary health care units in Brazil. Rev Inst Med Trop São Paulo. 2021;63:e14.
- 57. Pereira-Franchi EPL, Barreira MRN, da Costa N de SLM, et al. Molecular epidemiology of methicillin-resistant Staphylococcus aureus in the Brazilian primary health care system. Trop Med Int Health. 2019;24:339–47.
- Rodrigues MVP, Fortaleza CMCB, Riboli DFM, Rocha RS, Rocha C, Cunha M de LR de S d. Molecular epidemiology of methicillin-resistant Staphylococcus aureus in a burn unit from Brazil. Burns. 2013;39:1242–9.

- Nagasundaram N, Sistla S. Existence of multiple SCCmec elements in clinical isolates of methicillin-resistant Staphylococcus aureus. J Med Microbiol. 2019;68:720–7.
- Carmo MS, Inoue F, Andrade SS, et al. New multilocus sequence typing of MRSA in São Paulo, Brazil. Braz J Med Biol Res. 2011;44:1013–7.
- **61**. Otter JA, French GL. Molecular epidemiology of communityassociated meticillin-resistant *Staphylococcus aureus* in Europe. Lancet Infect Dis. 2010;10:227–39.
- 62. Chua K, Laurent F, Coombs G, Grayson ML, Howden BP. Not Community-Associated Methicillin-Resistant Staphylococcus aureus (CA-MRSA)! A Clinician's Guide to Community MRSA -Its Evolving Antimicrobial Resistance and Implications for Therapy. Clin Infect Dis. 2011;52:99–114.
- Mediavilla JR, Chen L, Mathema B, Kreiswirth BN. Global epidemiology of community- associated methicillin resistant Staphylococcus aureus (CA-MRSA). Curr Opin Microbiol. 2012;15:588–95.
- 64. Gonçalves da Silva A, Baines SL, Carter GP, et al. A phylogenomic framework for assessing the global emergence and evolution of clonal complex 398 methicillin-resistant Staphylococcus aureus. Microb Genomics. 2017;3:e000105.
- 65. Chroboczek T, Boisset S, Rasigade J-P, Tristan A, Bes M, Meugnier H, et al. Clonal complex 398 Methicillin Susceptible Staphylococcus aureus: a frequent unspecialized human pathogen with specific phenotypic and genotypic characteristics. PLoS ONE. 2013;8:e68462.
- 66. Souza CSM de [UNESP. Determinação da relação clonal e virulência de Staphylococcus aureus isolados de pacientes vivendo com HIV/AIDS e seus familiares [Thesis on the Internet]. Botucatu: Universidade Estadual Paulista Júlio de Mesquita Filho, Faculdade de Medicina de Botucatu; 2018. [cited 2021 Jun 6]; Available from: https://repositorio.unesp.br/handle/11449/154171
- 67. André Neto ED, Pereira RFA, Snyder RE, et al. Emergence of methicillin-resistant Staphylococcus aureus from clonal complex 398 with no livestock association in Brazil. Mem Inst Oswaldo Cruz. 2017;112:647–9.
- 68. Bonesso MF, Yeh AJ, Villaruz AE, et al. Key role of α-Toxin in fatal pneumonia caused by Staphylococcus aureus sequence type 398. Am J Respir Crit Care Med. 2016;193:217–20.
- 69. Gales AC, Deshpande LM, de Souza AG, Pignatari ACC, Mendes RE. MSSA ST398/t034 carrying a plasmid-mediated Cfr and Erm(B) in Brazil. J Antimicrob Chemother. 2015;70:303–5.
- 70. Chen N, Zhou M, Dong X, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus

pneumonia in Wuhan, China: a descriptive study. Lancet Lond Engl. 2020;395:507–13.

- 71. Zhou F, Yu T, Du R, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. Lancet Lond Engl. 2020;395:1054–62.
- 72. Vaillancourt M, Jorth P. The unrecognized threat of secondary bacterial infections with COVID-19. mBio. 2020;11(4).
- 73. Yap FHY, Gomersall CD, Fung KSC, et al. Increase in Methicillin- Resistant Staphylococcus aureus acquisition rate and change in pathogen pattern associated with an outbreak of severe acute respiratory syndrome. Clin Infect Dis. 2004;39:511–6.
- 74. Chung DR, Huh K. Novel pandemic influenza A (H1N1) and community-associated methicillin- resistant *Staphylococcus aureus* pneumonia. Expert Rev Anti Infect Ther. 2015;13:197–207.
- **75.** Rawson TM, Moore LSP, Zhu N, et al. Bacterial and fungal coinfection in individuals with coronavirus: a rapid review to support COVID- 19 antimicrobial prescribing. Clin Infect Dis. 2020;71:2459–68.
- 76. Witzel C, Castelo Branco Fortaleza C, Sena Martins de Souza C, Moraes Riboli D, Ribeiro de Souza da Cunha M de L. Nasopharyngeal carriage of Staphylococcus aureus among imprisoned males from Brazil without exposure to healthcare: risk factors and molecular characterization. Ann Clin Microbiol Antimicrob. 2014;13:25.
- 77. Abraão LM. Carreamento nasal/oral de Staphylococcus aureus em populações indígenas do norte e sudeste do brasil: resistência antimicrobiana, virulência, fatores de risco e epidemiologia molecular [Thesis on the Internet]. Botucatu: Universidade Estadual Paulista Júlio de Mesquita Filho, Faculdade de Medicina de Botucatu; 2017. [cited 2021 Jun 4]; Available from: https://repositorio.unesp.br/handle/11449/ 151310
- 78. Regina Pedrosa Soares C, de Lira CR, Cunha MAH, et al. Prevalence of nasal colonization by methicillin-resistant Staphylococcus aureus in outpatients living with HIV/AIDS in a Referential Hospital of the Northeast of Brazil. BMC Res Notes. 2018;11:794.
- 79. de Araújo BES, Borchert JM, Manhães PG, et al. A rare case of pyomyositis complicated by compartment syndrome caused by ST30–staphylococcal cassette chromosome *mec* type IV methicillin-resistant *Staphylococcus aureus*. Am J Emerg Med. 2010;28:537.e3–6.
- **80.** Razera F, Stefani SD, Bonamigo RR, Olm GS, Dias CAG, Narvaez GA. CA-MRSA em furunculose: relato de caso do sul do Brasil. An Bras Dermatol. 2009;84:515–8.