# **Cancer Science**

# Unsolicited Review Article

# Long non-coding RNAs and prostate cancer

# Aya Misawa,<sup>1</sup> Ken-ichi Takayama<sup>1,2</sup> and Satoshi Inoue<sup>1,2,3</sup> in

<sup>1</sup>Department of Anti-Aging Medicine, Graduate School of Medicine, The University of Tokyo, Tokyo; <sup>2</sup>Department of Functional Biogerontology, Tokyo Metropolitan Institute of Gerontology, Tokyo; <sup>3</sup>Division of Gene Regulation and Signal Transduction, Research Center for Genomic Medicine, Saitama Medical University, Saitama, Japan

#### Key words

Androgen, androgen receptor, castration resistant prostate cancer, long non-coding RNA, prostate cancer

#### Correspondence

Satoshi Inoue, Department of Functional Biogerontology, Tokyo Metropolitan Institute of Gerontology, 35-2 Sakaecho, Itabashi-ku, Tokyo 173-0015, Japan. Tel: 81-3-3964-3241; Fax: 81-3-3579-4776; E-mail: sinoue@tmig.or.jp

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**N** on-coding RNAs, which are RNA transcripts that do not code for proteins, can be divided into two major groups: small ncRNAs between 18 and 200 nt in length, and lncRNAs, which are larger than 200 nt. miRNAs are evolutionally conserved and single-stranded small non-protein coding transcripts of approximately 18–22 nt that post-transcriptionally regulate gene expression.<sup>(1)</sup> Aberrant expressions of miRNAs have been well documented in most types of cancer.<sup>(2)</sup> Unlike miRNAs, lncRNAs are able to fold into secondary and tertiary structures by which they carry out their function<sup>(3)</sup> Several studies have shown the importance of lncRNAs as modulators of key cellular processes not only in normal physiology<sup>(4,5)</sup> but also in diseases such as cancer, including prostate cancer.<sup>(6–8)</sup> It is assumed that many of these transcripts could serve as future cancer biomarkers.<sup>(9)</sup>

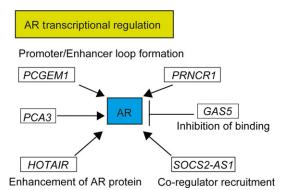
Prostate cancer is one of the leading causes of cancer morbidity and mortality in developed countries. It is currently diagnosed based on the measurement of serum PSA, also known as KLK3, and DRE. Treatment of localized prostate cancer takes into account clinicopathological factors including

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Long non-coding RNAs (IncRNAs) are RNA transcripts larger than 200 nucleotides that do not code for proteins the aberrant expression of which has been documented in various types of cancer, including prostate cancer. Lack of appropriate sensitive and specific biomarkers for prostate cancer has led to overdiagnosis and overtreatment, making IncRNAs promising novel biomarkers as well as therapeutic targets for the disease. The present review attempts to summarize the current knowledge of IncRNA expression patterns and mechanisms in prostate cancer, which contribute to carcinogenesis. In particular, we focused on IncRNAs regulated by androgen receptor and expressed in castration-resistant prostate cancer.

> Gleason score, initial PSA level, patient's age and clinical tumor stage.<sup>(10)</sup> Active surveillance may be best for patients with low-risk disease (i.e. Gleason score of 6 or less), whereas men with high-risk disease (i.e. Gleason score >7, PSA levels >20 ng/mL and clinical tumor stage >pT2c, i.e. tumor involving both prostate lobes) will instead benefit from radical prostatectomy or radiotherapy.<sup>(11)</sup> AR and its downstream signaling are fundamental for the development and progression of both localized and advanced metastatic prostate cancer.<sup>(12)</sup> Therefore, nowadays, the recommended therapy for locally advanced prostate cancer consists of long-term ADT in combination with radiotherapy. ADT decreases circulating testosterone levels to a very low amount (<50 ng/mL), a condition called chemical castration of men. However, some tumors will become hormone refractory following ADT, featured by increasing PSA levels in blood and upregulation of the AR in cancer cells.<sup>(13,14)</sup> Over a period of 12-36 months, a disease state called CRPC evolves in many patients. The ineffectiveness of conventional ADT in these CRPC is a result of activation of AR and its downstream pathways.<sup>(14)</sup> Past studies have



**Fig. 1.** Several long non-coding RNAs such as HOTAIR, PCGEM1 and *PCA3* interact with androgen receptor (AR) signals for castration-resistant prostate cancer progression. *GAS5*, growth arrest-specific-5; HOTAIR, HOX transcript antisense RNA; PCGEM1, prostate cancer gene expression marker 1; PRNCR1, prostate cancer noncoding RNA-1.

revealed that elevated AR expression,<sup>(15)</sup> enhanced AR activity<sup>(16)</sup> and development of AR variants<sup>(17)</sup> are observed in the progression of CRPC. Therefore, identification of AR downstream signals and new molecular mechanisms for AR activation are important to improve the treatment of advanced prostate cancer.

LncRNAs are aberrantly expressed in a variety of human diseases, contributing to pathogenesis or maintaining diseased conditions.<sup>(8)</sup> Aberrantly expressed lncRNAs can be indicative of certain stages of cancer progression, and may predict early progression or efficiently sustain tumor-related signaling pathways.<sup>(8)</sup> Thus, lncRNAs have the potential to be applicable for the diagnosis of prostate cancer, as well as being potential criteria in the choice of therapy and potential new therapeutic targets of CRPC. The present review attempts to summarize the current knowledge of lncRNA expression patterns in prostate cancer and the mechanisms that contribute to prostate carcinogenesis focusing on AR-regulated lncRNAs and lncRNAs expressed in CRPC (Fig. 1).

# **LncRNAs and Prostate Cancer**

LncRNAs as biomarkers in prostate cancer. PCA3 is one of the most specific prostate cancer biomarkers, which was originally discovered in 1999 by differential display analysis of prostate tissues and cell lines.<sup>(18)</sup> Its expression was shown to be 60-100-fold higher in more than 95% of prostate tumors compared to adjacent non-neoplastic tissues, and is undetectable in other tumor types. PCA3 knockdown inhibits AR signaling, cell growth and viability, suggesting that overexpression of PCA3 may modulate AR signaling in tumor cells. Knockdown of PCA3 leads to partial upregulation of epithelial markers such as E-cadherin, claudin-3 and cytokeratin-18, and downregulation of the mesenchymal marker vimentin.<sup>(19)</sup> PCA3 also regulates the expression of important cancer-related genes involved in apoptosis, angiogenesis, signal transduction, cell adhesion and mitogen-activated kinase kinase 1.<sup>(19)</sup> In addition, a working model of PCA3 has been proposed, in which PCA3 acts as a dominant-negative oncogene that downregulates the unrecognized tumor suppressor Prune Homolog 2 (PRUNE2), a human homolog of the Drosophila prune gene, through a process that involves RNA editing by the formation of *PRUNE2/PCA3* double-stranded RNA.<sup>(20)</sup> Combination of urinary PCA3 and fusion gene TMPRSS2-ERG can increase

specificity in prostate cancer diagnosis compared with serum PSA, and has the potential to substantially reduce unnecessary prostate biopsies. (Functions of lncRNAs in prostate cancer and references are summarized in Table 1 and Fig. 2).

Second chromosome locus-associated with prostate-1 (SChLAP1) is a lncRNA highly expressed in 25% of prostate cancer.<sup>(21)</sup> Its expression is more frequent in CRPC and it is significantly associated with risk of biochemical recurrence, clinical progression, metastasis and prostate cancer-specific mortality.<sup>(21,22)</sup> SChLAP1 interacts with Switch-Sucrose Non-Fermentable (SWI/SNF) complex for chromatin remodeling, counteracting the tumor-suppressor effects of SWI/SNF.<sup>(21)</sup> Analysis of SChLAP1 expression by ISH showed that this lncRNA independently predicts biochemical recurrence after radical prostatectomy.<sup>(23)</sup> Furthermore, SChLAP1 expression also correlated with prostate cancer lethal progression, which makes this lncRNA a useful tissue-based biomarker for identifying PCa patients at higher risk of CRPC progression.<sup>(24)</sup>

*SPRY4 intronic transcript 1 (SPRY4-IT1)* is one of the lncRNAs highly upregulated in PC3 cells and in patient samples compared to non-malignant prostate epithelial cells and matched normal prostate tissues.<sup>(25)</sup> siRNA knockdown of *SPRY4-IT1* inhibited PC3 cellular proliferation and invasion, and increased apoptosis.<sup>(25)</sup> *SPRY4-IT1* was easily detected in all prostate cancer samples with different Gleason scores (6–10) in an RNA chromogenic ISH assay.<sup>(25)</sup> Prostate cancer specificity and easy detection with standard clinical staining procedures of tissue samples makes this lncRNA a useful candidate as a diagnostic biomarker.

Metastasis-associated lung adenocarcinoma transcript 1 (MALAT1) is a lncRNA originally found to be overexpressed in non-small-cell lung cancer tissues with a high risk of metastasis.<sup>(26)</sup> Recent studies showed that MALAT1 was also overexpressed in other human cancers, including breast, pancreas, colon, prostate, and liver cancers.<sup>(27,28)</sup> In prostate cancer, MALATI overexpression was associated with indicators of poor prognosis such as high Gleason score, higher tumor-nodemetastasis stage and serum PSA >20 ng/mL, and its expression was significantly higher in CRPC than in hormone-sensitive prostate cancer.<sup>(29)</sup> In a study comparing the expression of MALAT1 in urinary samples of biopsy-positive and biopsynegative prostate cancer patients, this lncRNA was significantly higher in biopsy-positive samples,<sup>(30)</sup> suggesting that urinary MALAT1 may be a promising diagnostic biomarker. Furthermore, using EZH2 antibody-based RNA immunoprecipitation coupled with high throughput sequencing analysis, it was demonstrated that *MALAT1* binds to EZH2.<sup>(31)</sup> It was indicated that MALAT1 plays a crucial role in EZH2-enhanced migration and invasion in CRPC cell lines, and a positive correlation between MALAT1 and EZH2 has been documented.<sup>(31)</sup>

Transient receptor potential cation channel, subfamily M, member 2-antisense transcript (TRPM2-AS) is an antisense lncRNA transcribed from the opposite strand of the TRPM2 gene that is overexpressed in prostate cancer.<sup>(32)</sup> High TRPM2-AS was associated with poor prognosis and *in vitro* TRPM2-AS knockdown led to prostate cancer cell apoptosis and activation of the TRPM2 gene. Microarray analysis was carried out using PC3 cells which were transfected with an siRNA ablating TRPM2-AS RNA or control siRNA to analyze the mechanisms by which TRPM2-AS maintains cell survival in prostate cancer cells.<sup>(33)</sup> Results showed that TRPM2-AS coordinates the expression of a large number of genes involved in controlling survival, unfolded protein response and cell cycle in prostate cancer cells. Moreover, targets of existing drugs and treatments

#### Table 1. LncRNAs implicated in PCa

| Expression<br>in PCa | LncRNAs   | Role                | Implications in PCa   | References  |
|----------------------|-----------|---------------------|---|-------------|
| ↑                    | PCA3      | Biomarker           | Enhances AR signaling, cell growth and viability. Regulates the expression of important cancer-related genes  | 18–20       |
| ↑                    | SChLAP1   |                     | Overexpression is associated with risk of biochemical recurrence, clinical progression, metastasis and PCa-specific mortality   | 21,24       |
| $\uparrow$           | SPRY4-IT1 |                     | siRNA knockdown inhibits cell proliferation and invasion, and increases apoptosis   | 25          |
| 1                    | MALAT1    |                     | Overexpression is associated with indicators of poor prognosis. Binds to EZH2 to enhance migration and invasion   | 26–31       |
| ↑                    | TRPM2-AS  |                     | Transcribed from the antisense strand of <i>TRPM2</i> , which is activated with <i>TMPR2-AS</i> knockdown. Overexpression associated with poor prognosis                                    | 32,33       |
| ↑                    | NEAT1     |                     | Expression associated with PCa cell progression. Alters the epigenetic status of target genes to drive oncogenic growth   | 34          |
| $\uparrow$           | PCGEM1    | AR-related          | Promotes cell proliferation by regulating c-Myc   | 35,37,46,47 |
| ↑                    | PCAT1     |                     | Upregulated in high-grade localized and metastatic PCa. Promotes cell proliferation by regulating <i>c-Myc</i> . Represses <i>BRCA2</i>   | 38,39       |
| $\uparrow$           | PCAT6     |                     | Predictive of tumor progression by AR signaling. Overexpressed in primary and metastatic  | 8,40,41     |
| ↑                    | PCAT7     |                     | PCa. siRNA-mediated knockdown reduces cell growth   |             |
| ↑                    | PCAT18    |                     | Predictive of tumor progression by AR signaling. Metastatic PCa specific. Induced by AR. siRNA-mediated knockdown reduces cell growth   | 8,42        |
| ↑                    | PRNCR1    |                     | Associated with prostate cancer susceptibility. siRNA knockdown attenuates cell viability and AR activity. Could be involved in prostate carcinogenesis through AR                          | 44,45       |
| ↑                    | CTBP1-AS  |                     | Transcribed from the antisense strand of <i>CTBP1</i> . Promotes castration-resistant prostate tumor growth by regulating epigenetically cancer-associated genes                            | 46,49       |
| 1                    | HOTAIR    |                     | Repressed by androgen and upregulated in CRPC after deprivation therapies. Binds to AR to prevent its degradation. Overexpression increases cell growth and invasion                        | 58          |
| 1                    | SOCS2-AS1 |                     | Transcribed from the antisense strand of SOCS2. Promotes cell proliferation, migration and anti-apoptosis.  | 59          |
| ↑                    | POTEF-AS1 |                     | Transcribed from the antisense strand of <i>POTEF</i> . Promotes cell proliferation and anti-<br>apoptosis  | 60          |
| $\downarrow$         | GAS5      | Tumor<br>suppressor | Represses AR action and promotes apoptosis. Downregulated in CRPC. Reciprocal regulation of <i>GAS5</i> levels and mTOR inhibitor action  | 62–64       |
| $\downarrow$         | H19       | ••                  | Upregulation of <i>H19</i> represses cell migration. H19-derived miR-675 targets TGF $\beta$ 1 to repress cell migration  | 69          |
| Ļ                    | PCAT29    |                     | First AR-repressed IncRNA that functions as a tumor suppressor. Low <i>PCAT29</i> expression correlated with poor prognostic outcomes. Overexpression suppresses cell growth and metastasis | 43          |

AR, androgen receptor; BRCA2, breast cancer susceptibility gene 2; CRPC, castration-resistant prostate cancer; CTBP1, C-terminal binding protein 1; EZH2, enhancer of zeste homolog 2; GAS5, growth arrest-specific 5; IncRNAs, long non-coding RNAs; mTOR, mammalian target of rapamycin; PCa, prostate cancer; POTEF, prostate, ovary, testis expressed protein family member-F gene; SOCS2, suppressor of cytokine signaling 2; TGF-β1, transforming growth factor beta 1; TRPM2-AS, transient receptor potential cation channel, subfamily M, member 2-antisense transcript.

were found to be consistently regulated by *TRPM2-AS* knockdown. Thus, the essential role of *TRPM2-AS* as a key regulator of survival in prostate cancer makes this lncRNA a suitable therapeutic target for further clinical studies.

Estrogen receptor alpha (ER $\alpha$ ) is expressed in prostate cancer, independent of AR status and, in CRPC, ER $\alpha$  signaling constitutes an effective mechanism to bypass the androgen/ AR axis. In a study combining ChIP- and RNA-seq data in prostate cancer cells, an ER $\alpha$ -specific non-coding transcriptome signature was identified, where the lncRNA, nuclear enriched abundant 1 (NEAT1), was the most significantly overexpressed lncRNA in prostate cancer.<sup>(34)</sup> NEAT1 expression was associated with prostate cancer progression, and prostate cancer cells expressing high levels of this lncRNA were resistant to androgen deprivation or AR antagonists. NEAT1 is recruited to the promoter regions of target genes and alters their epigenetic status to favor transcription and drive oncogenic growth. NEAT1 levels increase after longterm androgen-deprivation therapies, and its expression is significantly higher in CRPC compared to hormone-sensitive prostate cancer, which indicates that  $ER\alpha$ -*NEAT1* interaction may promote castration resistance. Elevated *NEAT1* levels are associated with early biochemical recurrence and metastatic spread, making this lncRNA both a potential therapeutic target and a reliable prognostic biomarker.

**LncRNAs related with AR signaling.** Prostate cancer gene expression marker 1 (PCGEM1) and prostate cancer-associated ncRNA transcript 1 (PCAT1) are highly prostate-specific lncRNAs and considered attractive biomarkers. PCGEM1 was one of the oncogenic lncRNAs identified in prostate cancer and regulated by androgen.<sup>(35)</sup> PCGEM1 is expressed in at least half of prostate tumors and functional analysis revealed oncogenic properties such as promotion of cell proliferation and colony formation.<sup>(36)</sup> It was demonstrated that PCGEM1 promotes cancer cell proliferation by regulating *c-Myc*.<sup>(37)</sup> PCAT1 is a lncRNA which is expressed highly specifically in prostate and upregulated in high-grade localized (Gleason score >7) and metastatic prostate cancer. PCAT1 induces cell

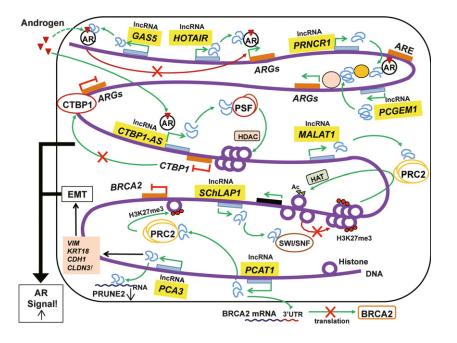


Fig. 2. Epigenetic mechanisms of long non-coding RNAs (IncRNAs) in prostate cancer. Summary of functional roles of IncRNAs in prostate cancer is shown. ARE, androgen response element; ARGs, androgen responsive genes; BRCA2, breast cancer susceptibility gene 2; CDH1, E-cadherin; CLDN3, claudin-3; CTBP1-AS, C-terminal binding protein 1 transcript; EMT, epithelial antisense to mesenchymal transition; HAT, histone acetyl transferase; HDAC, histone deacetylase; KRT18, cytokeratin-18; MALAT1, metastasis-associated lung adenocarcinoma transcript 1; PCAT1, prostate cancer-associated ncRNA transcript 1; PRC2, polycomb repressive complex 2; SChLAP1, second chromosome locus-associated with prostate-1; SWI/ non-fermentable; SNF. switch-sucrose VIM. vimentin.

proliferation in vitro and has repressive effects on a broad range of genes, including the tumor suppressor gene, *breast* cancer susceptibility gene 2 (BRCA2).<sup>(38)</sup> PCAT1 also promotes cell proliferation by interaction with c-Myc by acting as a decoy for *c-Myc*-targeting miRNAs.<sup>(39)</sup> Other PCAT-family members, *PCAT6*, *PCAT 7*, and *PCAT 18* were predictive of tumor progression by modulation of AR signaling.<sup>(8,40)</sup> Expressions of PCAT6 and PCAT7 were higher in primary and metastatic prostate cancer, and siRNA-mediated knockdown of either of them reduced cell growth and soft agar colony formation in both androgen-dependent LNCaP and -independent LNCaP sublines.<sup>(41)</sup> PCAT18 is highly prostate-specific and expressed especially in metastatic prostate cancer tissues. Expression level of PCAT18 is regulated by AR signaling.<sup>(42)</sup> siRNA-mediated knockdown of PCAT18 also reduced cell growth. *PCAT29* was identified as an AR-repressed lncRNA that functions as a tumor suppressor.<sup>(43)</sup> *PCAT29* was suppressed by androgen and upregulated by castration in a prostate cancer xenograft model. PCAT29 knockdown significantly increased proliferation and migration of prostate cancer cells, whereas *PCAT29* overexpression conferred the opposite effect. In prostate cancer patient specimens, low PCAT29 expression correlated with poor prognostic outcomes, suggesting that decrease of this lncRNA may identify a subset of patients at higher risk for disease recurrence.

*Prostate cancer noncoding RNA-1 (PRNCR1)* was initially identified as a novel lncRNA transcribed from 8q24, the genomic locus most significantly associated with prostate cancer susceptibility found in a genome-wide association study.<sup>(44)</sup> Knockdown of *PRNCR1* by siRNA attenuated prostate cancer cell viability and AR transactivation activity, indicating that *PRNCR1* could be involved in prostate carcinogenesis possibly through AR. In another study, *PRNCR1* and *PCGEM1* were found to successively bind to AR and strongly enhance AR-mediated gene activation programs and proliferation in prostate cancer cells.<sup>(45)</sup> Later, it was shown that *PCGEM1* and *PRNCR1* do not interact with AR and that neither gene was a prognostic factor for prostate cancer.<sup>(46)</sup> A study using AR+/ androgen-dependent prostate cancer xenograft models revealed

that *PRNCR1* was scarcely expressed and that *PCGEM1* expression was lower in metastatic prostate cancer compared to primary tumors.<sup>(47)</sup> Thus, the roles of these lncRNAs in prostate cancer are still being debated, and further studies are necessary to elucidate the relation of *PCGEM1* and AR, and its clinical significance.

Our study analyzing a global AR transcriptional network by mapping genome-wide androgen-regulated TTS using CAGE and ARBS using ChIP-seq has revealed comprehensive AR-regulated transcripts from intergenic or AS regions of genes in prostate cancer cells.<sup>(48)</sup> Based on this study, a lncRNA located at the AS region of the *C-Terminal Binding Protein 1* (*CTBP1*) gene, *CTBP1-AS*, was found to promote tumor growth of both hormone-sensitive prostate cancer and CRPC models by regulating cancer-associated genes epigenetically.<sup>(49)</sup> These studies revealed the importance of androgen-regulated lncRNAs in prostate cancer progression.

HOX Transcript Antisense RNA (HOTAIR) is a well-known IncRNA found to play important roles in several tumors.<sup>(50–57)</sup> In prostate cancer, HOTAIR was reported as a IncRNA repressed by androgen.<sup>(58)</sup> It was markedly upregulated in CRPC after androgen deprivation therapies. HOTAIR was found to bind to AR protein to block its interaction with E3 ubiquitin ligase Mouse double minute 2 homolog (MDM2), preventing AR ubiquitination and degradation. HOTAIR overexpression increased prostate cancer cell growth and invasion. Consequently, HOTAIR expression was sufficient to induce androgen-independent AR activation and drive an ARmediated transcriptional program in the absence of androgen. HOTAIR may drive androgen-independent AR activity and CRPC progression, being a potential therapeutic target.

Our comprehensive analysis of AR-regulated lncRNAs in two prostate cancer cell lines and CRPC model cells by directional RNA-seq analysis identified an androgen-regulated lncRNA, *suppressor of cytokine signaling 2-antisense transcript 1 (SOCS2-AS1)*, located at the antisense strand of *SOCS2*. Expressions of *SOCS2-AS1* and *SOCS2* expression were androgen-dependent and their expression was suppressed by AR knockdown as well as by an anti-androgen,

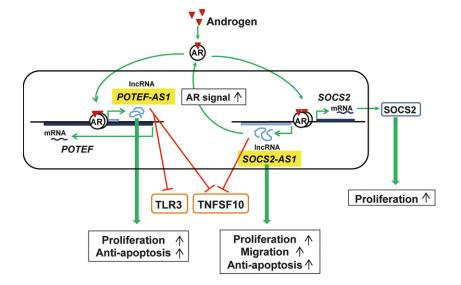


Fig. 3. Androgen-regulated lncRNAs, SOCS2-AS1 and POTEF-AS1 promote prostate cancer growth. Androgen-induced SOCS2-AS1 and SOCS2 promote cell growth. Furthermore, SOCS2-AS1 activates androgen signaling to enhance cell growth, migration and anti-apoptosis signals, repressing genes related to the apoptosis pathway. POTEF-AS1, represses genes related to Toll-like receptor (TLR) signaling and apoptosis pathways, promoting cell growth and inhibiting apoptosis.

bicalutamide. *SOCS2-AS1*, which is highly expressed in CRPC model cells, promoted castration-resistant and androgen-dependent cell growth and inhibited apoptosis. Furthermore, *SOCS2-AS1* promoted androgen signaling by modulating the epigenetic control for AR target genes including *Tumor Necrosis Factor Superfamily Member 10 (TNFSF10)*, suggesting that *SOCS2-AS1* plays an important role in the development of CRPC by repressing apoptosis.<sup>(59)</sup> (Fig. 3.)

We recently identified another AR-dependent lncRNA transcribed from the AS strand of *prostate, ovary, testis expressed protein family member-F gene (POTEF)*, which was named *POTEF-Antisense transcript-1 (POTEF-ASI)*.<sup>(60)</sup> POTEF is one of the proteins encoded by the POTE family genes, which are preferentially expressed in prostate, ovary, testis and placenta.<sup>(61)</sup> *POTEF-AS1* promoted cell growth, inhibited apoptosis and repressed genes related to TLR signaling and apoptosis pathways in LNCaP cells, suggesting that *POTEF-AS1* would play a key role in the progression of prostate cancer by repressing TLR signaling.<sup>(60)</sup> (Fig. 3.)

**LncRNAs as tumor suppressors.** Growth arrest-specific 5 (GAS5) is a tumor suppressive lncRNA that promotes apoptosis and represses AR action by sequestering androgen/AR complex and preventing binding of the complex to target genes.<sup>(62)</sup> Its expression level was shown to decline when prostate cancer cells acquire castrate resistance.<sup>(63)</sup> Recently, regulation of GAS5 expression by mTOR inhibitor was found in prostate cancer cells. Several mTOR inhibitors increased cellular GAS5 expression levels and inhibited cell growth in AR-positive LNCaP and 22Rv1 cells, whereas silencing of GAS5 in these cells decreased sensitivity to mTOR inhibitors may be used to increase GAS5 levels to enhance cellular apoptosis.

Genomic imprinting is a form of epigenetic gene regulation that results in expression from a single allele in a parent-of-origin-dependent manner. One of the most extensively studied imprinting-associated lncRNAs in cancer is *Long Intergenic Non-Protein Coding RNA 8 (H19).*<sup>(65)</sup> *H19* is located in an imprinted region of chromosome 11 near the *insulin-like growth factor 2 (IGF2)* gene, the former expressed only from the maternally inherited chromosome, whereas the latter is expressed only from the paternally inherited chromosome.<sup>(66)</sup> *H19* was found to be a lncRNA which plays a critical role in choriocarcinoma<sup>(67)</sup> and bladder cancer.<sup>(68)</sup> However, in metastatic prostate cancer, *H19* has a tumor suppressive role by repressing the effects of *transforming growth factor beta 1 (TGF \beta1)*. *H19* and *H19*-derived miR-675 were both significantly downregulated in metastatic prostate cancer cells compared to non-metastatic prostate cancer cells.<sup>(69)</sup> Upregulating *H19* increased miR-675 levels and repressed cell migration. In addition, miR-675 repressed TGF\beta1 translation by directly binding to the 3'UTR.

# **Clinical Use of IncRNAs and Limitations**

LncRNAs still remain uncharacterized RNA molecules as a result of their low expression levels, poor conservation, and high tissue/cell specificity.<sup>(70)</sup> In prostate cancer diagnosis, the main priority is identifying novel biomarkers to reliably distinguish between low-risk and high-risk patients who need definitive treatment.<sup>(71)</sup> Although the majority of studies have focused on the intracellular roles of lncRNAs, there is an increasing interest in the potential roles of extracellular circulating lncRNAs. Urinary PCA3 is now widely used as a biomarker for detection of cancer and has been approved by the United States Food and Drug Administration.<sup>(72)</sup> Measuring PCA3 was shown to be superior to serum PSA test and digital rectal examination in the sensitivity and specificity for prostate cancer diagnosis.<sup>(73,74)</sup> However, its utility as a first-line test or to detect high-grade prostate cancer remains under investigation.<sup>(75)</sup> Therefore, other methods for prostate cancer diagnosis using lncRNAs have been developed, such as MALAT1-derived (MD) mini-RNA (MD-mini-RNA).<sup>(76)</sup> Plasma MD-mini-RNA was found to improve diagnostic accuracy for predicting the results of prostate biopsy in patients with high PSA levels (>4 ng/mL). These circulating lncRNAs have shown promise as non-invasive biomarkers across a wide spectrum of diseases and altered physiological states.<sup>(77,78)</sup> Release of cellular RNA species into circulation may reflect disease-specific tissue injury and/or remodeling, or potential intercellular signaling. However, the biological functions and the mechanisms of circulating lncRNAs have not been elucidated. Uniform normalization protocol of sample collection, lncRNAs extraction, endogenous control selection, quality assessment, and quantitative data analysis has not been established. To adopt circulating lncRNA into

clinical practice, the following aspects should be investigated: normalization of sample preparation such as temperature, volume and reagents used to extract and store lncRNAs, selection of reliable endogenous controls, method to assess the quality of lncRNA and reduction of bias by increasing the size of the research cohort.

# FANTOM Projects to Investigate the Functional Role of IncRNAs

Recent advances in transcriptomics have improved coverage as well as detection accuracy of profiled RNA molecules. The FANTOM project, starting in early 2000, is one of the longest running collaborative projects in genomics that aims to functionally characterize mammalian genomes.<sup>(79,80)</sup> FANTOM1 and FANTOM2 generated more than 100 000 mouse fulllength cDNAs that showed that the portion of the genome encoding proteins is small, whereas the majority of it is involved in producing non-coding RNAs. FANTOM3 uncovered the promoter landscape of the mammalian genome, allowing precise identification of TSS, and the existence of antisense transcription, by combining CAGE and full-length cDNA isolation.<sup>(80)</sup> FANTOM4 adopted CAGE and 454 Life Sciences sequencing combined with Illumina microarrays to study a model of differentiation in human THP-1 (myeloid leukemia) cells, and to define the transcriptional regulatory network based on TSS activity that explained such a timely process.<sup>(79)</sup> Within the FANTOM5 project, the consortium profiled nearly 2000 human and 1000 mouse samples, representative of the majority of cell types and tissues, using CAGE followed by next-generation single molecule sequencing (HeliScope).<sup>(81,82)</sup> A current limitation of FANTOM5, besides the coverage of species, lies in the approaches taken to explore RNAs. As the CAGE protocol is designed to capture only the 5'-end of capped long RNA molecules, the internal structure of long RNAs and small regulatory RNAs remains unexplored. To complement the CAGE profiles, CAGEscan, RNA-seq and small RNA sequencing data are being analyzed to be added to the FANTOM web resource, and existing databases and interfaces are being upgraded. The consortium is already developing the sixth FANTOM project which aims to uncover the function of long non-coding RNAs by high-throughput screening coupled with CAGE.

FANTOM projects revealed that the human genome is widely transcribed, producing thousands of lncRNAs. Most lncRNAs lack typical signatures of selective constraints. Given

### References

- 1 Ambros V. The functions of animal microRNAs. *Nature* 2004; 431: 350-5.
- 2 Garzon R, Calin GA, Croce CM. MicroRNAs in cancer. Annu Rev Med 2009; 60: 167–79.
- 3 Gibb J, Hayley S, Poulter MO, Anisman H. Effects of stressors and immune activating agents on peripheral and central cytokines in mouse strains that differ in stressor responsivity. *Brain Behav Immun* 2011; **25**: 468–82.
- 4 Feng S, Yao J, Chen Y et al. Expression and functional role of reprogramming-related long noncoding RNA (lincRNA-ROR) in glioma. J Mol Neurosci 2015; 56: 623–30.
- 5 Hu L, Wu Y, Tan D et al. Up-regulation of long noncoding RNA MALAT1 contributes to proliferation and metastasis in esophageal squamous cell carcinoma. J Exp Clin Cancer Res 2015; 34: 7.
- 6 Kandoth C, McLellan MD, Vandin F et al. Mutational landscape and significance across 12 major cancer types. *Nature* 2013; 502: 333–9.
- 7 Ciriello G, Miller ML, Aksoy BA, Senbabaoglu Y, Schultz N, Sander C. Emerging landscape of oncogenic signatures across human cancers. *Nat Genet* 2013; 45: 1127–33.

their diversity in biogenesis and their low expression and conservation levels, the functional relevance of most lncRNAs remains unclear. Future studies will enable understanding of lncRNA functions and to identify relevant lncRNA in diseases including prostate cancer.

### Conclusion

The main priority in clinical prostate cancer research is identifying novel biomarkers to reliably distinguish between low-risk patients and high-risk patients who need definitive treatment. Strong evidence has been presented about the significance of lncRNAs in prostate cancer, and the challenge is now to determine lncRNAs functionally relevant with oncogenic changes. Although recent studies showed that measuring these transcripts in clinical samples could improve the prediction of prognosis, identification of more molecular mechanisms that lead to metastatic prostate cancer would enable us to determine treatment modalities at the time of diagnosis.

### **Disclosure Statement**

Authors declare no conflicts of interest for this article.

#### Abbreviations

| ADT      | androgen deprivation therapy             |
|----------|--|
| AR       | androgen receptor                        |
| ARBS     | androgen receptor binding site           |
| ARE      | androgen responsive element              |
| AS       | antisense                                |
| CAGE     | cap analysis of gene expression          |
| ChIP-seq | chromatin immunoprecipitation sequencing |
| CRPC     | castration-resistant prostate cancer     |
| DRE      | digital rectal examination               |
| EZH2     | enhancer of zeste homolog 2              |
| ISH      | in situ hybridization                    |
| KLK3     | kallikrein-related peptidase 3           |
| IncRNAs  | long non-coding RNAs                     |
| miRNAs   | microRNAs                                |
| ncRNAs   | non-coding RNAs                          |
| nt       | nucleotide                               |
| PCA3     | prostate cancer antigen 3                |
| PSA      | prostate-specific antigen                |
| RNA-seq  | RNA sequencing                           |
| TLR      | Toll-like receptor                       |
| TTS      | transcriptional stat site                |

- 8 Prensner JR, Chinnaiyan AM. The emergence of lncRNAs in cancer biology. *Cancer Discov* 2011; 1: 391–407.
- 9 Iyer MK, Niknafs YS, Malik R *et al.* The landscape of long noncoding RNAs in the human transcriptome. *Nat Genet* 2015; **47**: 199–208.
- 10 Attard G, Parker C, Eeles RA et al. Prostate cancer. Lancet 2016; 387: 70-82.
- 11 Bill-Axelson A, Holmberg L, Garmo H et al. Radical prostatectomy or watchful waiting in early prostate cancer. N Engl J Med 2014; 370: 932–42.
- 12 Scher HI, Sawyers CL. Biology of progressive, castration-resistant prostate cancer: directed therapies targeting the androgen-receptor signaling axis. J Clin Oncol 2005; 23: 8253–61.
- 13 Smith MR, Cook R, Lee KA, Nelson JB. Disease and host characteristics as predictors of time to first bone metastasis and death in men with progressive castration-resistant nonmetastatic prostate cancer. *Cancer* 2011; **117**: 2077–85.
- 14 Chen CD, Welsbie DS, Tran C et al. Molecular determinants of resistance to antiandrogen therapy. Nat Med 2004; 10: 33–9.
- 15 Chen G, Shukeir N, Potti A *et al.* Up-regulation of Wnt-1 and beta-catenin production in patients with advanced metastatic prostate carcinoma: potential pathogenetic and prognostic implications. *Cancer* 2004; **101**: 1345–56.

- 16 Waltering KK, Helenius MA, Sahu B *et al.* Increased expression of androgen receptor sensitizes prostate cancer cells to low levels of androgens. *Cancer Res* 2009; **69**: 8141–9.
- 17 Sun S, Sprenger CC, Vessella RL *et al.* Castration resistance in human prostate cancer is conferred by a frequently occurring androgen receptor splice variant. *J Clin Invest* 2010; **120**: 2715–30.
- 18 Bussemakers MJ, van Bokhoven A, Verhaegh GW *et al.* DD3: a new prostate-specific gene, highly overexpressed in prostate cancer. *Cancer Res* 1999; 59: 5975–9.
- 19 Lemos AE, Ferreira LB, Batoreu NM, de Freitas PP, Bonamino MH, Gimba ER. PCA3 long noncoding RNA modulates the expression of key cancerrelated genes in LNCaP prostate cancer cells. *Tumour Biol* 2016; 37: 11339–48.
- 20 Salameh A, Lee AK, Cardó-Vila M et al. PRUNE2 is a human prostate cancer suppressor regulated by the intronic long noncoding RNA PCA3. Proc Natl Acad Sci USA 2015; 112: 8403–8.
- 21 Prensner JR, Iyer MK, Sahu A *et al.* The long noncoding RNA SChLAP1 promotes aggressive prostate cancer and antagonizes the SWI/SNF complex. *Nat Genet* 2013; **45**: 1392–8.
- 22 Prensner JR, Zhao S, Erho N *et al.* RNA biomarkers associated with metastatic progression in prostate cancer: a multi-institutional high-throughput analysis of SChLAP1. *Lancet Oncol* 2014; 15: 1469–80.
- 23 Mehra R, Shi Y, Udager AM *et al.* A novel RNA in situ hybridization assay for the long noncoding RNA SChLAP1 predicts poor clinical outcome after radical prostatectomy in clinically localized prostate cancer. *Neoplasia* 2014; 16: 1121–7.
- 24 Mehra R, Udager AM, Ahearn TU *et al.* Overexpression of the long noncoding RNA SChLAP1 independently predicts lethal prostate cancer. *Eur Urol* 2016; **70**: 549–52.
- 25 Lee B, Mazar J, Aftab MN *et al.* Long noncoding RNAs as putative biomarkers for prostate cancer detection. *J Mol Diagn* 2014; 16: 615–26.
- 26 Ji P, Diederichs S, Wang W et al. MALAT-1, a novel noncoding RNA, and thymosin beta4 predict metastasis and survival in early stage non-small cell lung cancer. Oncogene 2003; 22: 8031–41.
- 27 Lin R, Maeda S, Liu C *et al.* A large noncoding RNA is a marker for murine hepatocellular carcinomas and a spectrum of human carcinomas. *Oncogene* 2007; 26: 851–8.
- 28 Konishi H, Ichikawa D, Yamamoto Y et al. Plasma level of metastasis-associated lung adenocarcinoma transcript 1 is associated with liver damage and predicts development of hepatocellular carcinoma. Cancer Sci 2016; 107: 149–54.
- 29 Ren S, Liu Y, Xu W *et al.* Long noncoding RNA MALAT-1 is a new potential therapeutic target for castration resistant prostate cancer. *J Urol* 2013; 190: 2278–87.
- 30 Wang F, Ren S, Chen R *et al.* Development and prospective multicenter evaluation of the long noncoding RNA MALAT-1 as a diagnostic urinary biomarker for prostate cancer. *Oncotarget* 2014; **5**: 11091–102.
- 31 Wang D, Ding L, Wang L et al. LncRNA MALAT1 enhances oncogenic activities of EZH2 in castration-resistant prostate cancer. Oncotarget 2015; 6: 41045–55.
- 32 Orfanelli U, Jachetti E, Chiacchiera F *et al.* Antisense transcription at the TRPM2 locus as a novel prognostic marker and therapeutic target in prostate cancer. *Oncogene* 2015; **34**: 2094–102.
- 33 Lavorgna G, Chiacchiera F, Briganti A, Montorsi F, Pasini D, Salonia A. Expression-profiling of apoptosis induced by ablation of the long ncRNA TRPM2-AS in prostate cancer cell. *Genom Data* 2014; 3: 4–5.
- 34 Chakravarty D, Sboner A, Nair SS et al. The oestrogen receptor alpha-regulated lncRNA NEAT1 is a critical modulator of prostate cancer. Nat Commun 2014; 5: 5383.
- 35 Srikantan V, Zou Z, Petrovics G et al. PCGEM1, a prostate-specific gene, is overexpressed in prostate cancer. Proc Natl Acad Sci USA 2000; 97: 12216– 21.
- 36 Petrovics G, Zhang W, Makarem M et al. Elevated expression of PCGEM1, a prostate-specific gene with cell growth-promoting function, is associated with high-risk prostate cancer patients. Oncogene 2004; 23: 605–11.
- 37 Hung CL, Wang LY, Yu YL et al. A long noncoding RNA connects c-Myc to tumor metabolism. Proc Natl Acad Sci USA 2014; 111: 18697–702.
- 38 Prensner JR, Chen W, Iyer MK et al. PCAT-1, a long noncoding RNA, regulates BRCA2 and controls homologous recombination in cancer. Cancer Res 2014; 74: 1651–60.
- 39 Prensner JR, Chen W, Han S et al. The long non-coding RNA PCAT-1 promotes prostate cancer cell proliferation through cMyc. Neoplasia 2014; 16: 900–8.
- 40 Rönnau CG, Verhaegh GW, Luna-Velez MV, Schalken JA. Noncoding RNAs as novel biomarkers in prostate cancer. *Biomed Res Int* 2014; 2014: 591703.
- 41 Du Z, Fei T, Verhaak RG *et al.* Integrative genomic analyses reveal clinically relevant long noncoding RNAs in human cancer. *Nat Struct Mol Biol* 2013; **20**: 908–13.

- 42 Crea F, Watahiki A, Quagliata L *et al.* Identification of a long non-coding RNA as a novel biomarker and potential therapeutic target for metastatic prostate cancer. *Oncotarget* 2014; **5**: 764–74.
- 43 Malik R, Patel L, Prensner JR et al. The IncRNA PCAT29 inhibits oncogenic phenotypes in prostate cancer. Mol Cancer Res 2014; 12: 1081–7.
- 44 Chung S, Nakagawa H, Uemura M et al. Association of a novel long noncoding RNA in 8q24 with prostate cancer susceptibility. *Cancer Sci* 2011; 102: 245–52.
- 45 Yang L, Lin C, Jin C et al. lncRNA-dependent mechanisms of androgenreceptor-regulated gene activation programs. *Nature* 2013; 500: 598–602.
- 46 Prensner JR, Sahu A, Iyer MK *et al.* The IncRNAs PCGEM1 and PRNCR1 are not implicated in castration resistant prostate cancer. *Oncotarget* 2014; **5**: 1434–8.
- 47 Parolia A, Crea F, Xue H et al. The long non-coding RNA PCGEM1 is regulated by androgen receptor activity in vivo. *Mol Cancer* 2015; **14**: 46.
- 48 Takayama K, Tsutsumi S, Katayama S *et al.* Integration of cap analysis of gene expression and chromatin immunoprecipitation analysis on array reveals genome-wide androgen receptor signaling in prostate cancer cells. *Oncogene* 2011; **30**: 619–30.
- 49 Takayama K, Horie-Inoue K, Katayama S et al. Androgen-responsive long noncoding RNA CTBP1-AS promotes prostate cancer. EMBO J 2013; 32: 1665–80.
- 50 Gupta RA, Shah N, Wang KC *et al.* Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis. *Nature* 2010; 464: 1071–6.
- 51 Kogo R, Shimamura T, Mimori K *et al.* Long noncoding RNA HOTAIR regulates polycomb-dependent chromatin modification and is associated with poor prognosis in colorectal cancers. *Cancer Res* 2011; **71**: 6320–6.
- 52 Geng YJ, Xie SL, Li Q, Ma J, Wang GY. Large intervening non-coding RNA HOTAIR is associated with hepatocellular carcinoma progression. J Int Med Res 2011; 39: 2119–28.
- 53 Niinuma T, Suzuki H, Nojima M *et al.* Upregulation of miR-196a and HOTAIR drive malignant character in gastrointestinal stromal tumors. *Cancer Res* 2012; **72**: 1126–36.
- 54 Li D, Feng J, Wu T *et al.* Long intergenic noncoding RNA HOTAIR is overexpressed and regulates PTEN methylation in laryngeal squamous cell carcinoma. *Am J Pathol* 2013; **182**: 64–70.
- 55 Kim K, Jutooru I, Chadalapaka G et al. HOTAIR is a negative prognostic factor and exhibits pro-oncogenic activity in pancreatic cancer. Oncogene 2013; 32: 1616–25.
- 56 Nie Y1, Liu X, Qu S et al. Long non-coding RNA HOTAIR is an independent prognostic marker for nasopharyngeal carcinoma progression and survival. Cancer Sci 2013; 104: 458–64.
- 57 Ge XS, Ma HJ, Zheng XH *et al.* HOTAIR, a prognostic factor in esophageal squamous cell carcinoma, inhibits WIF-1 expression and activates Wnt pathway. *Cancer Sci* 2013; **104**: 1675–82.
- 58 Zhang A, Zhao JC, Kim J *et al.* LncRNA HOTAIR enhances the androgenreceptor-mediated transcriptional program and drives castration-resistant prostate cancer. *Cell Rep* 2015; **13**: 209–21.
- 59 Misawa A, Takayama K, Urano T, Inoue S. Androgen-induced long noncoding RNA (lncRNA) SOCS2-AS1 promotes cell growth and inhibits apoptosis in prostate cancer cells. *J Biol Chem* 2016; **291**: 17861–80.
- 60 Misawa A, Takayama K, Suzuki Y, Fujimura T, Homma Y, Inoue S. Androgen-induced lncRNA POTEF-AS1 regulates apoptosis related pathway to facilitate cell survival in prostate cancer cells. *Cancer Sci* 2016; **108**: 373–9.
- 61 Bera TK, Saint Fleur A, Lee Y et al. POTE paralogs are induced and differentially expressed in many cancers. Cancer Res 2006; 66: 52–6.
- 62 Kino T, Hurt DE, Ichijo T, Nader N, Chrousos GP. Noncoding RNA gas5 is a growth arrest- and starvation-associated repressor of the glucocorticoid receptor. *Sci Signal* 2010; 3: ra8.
- 63 Pickard MR, Mourtada-Maarabouni M, Williams GT. Long non-coding RNA GAS5 regulates apoptosis in prostate cancer cell lines. *Biochim Biophys Acta* 2013; 1832: 1613–23.
- 64 Pickard MR, Mourtada-Maarabouni M, Williams GT. Reciprocal regulation of GAS5 lncRNA levels and mTOR inhibitor action in prostate cancer cells. *Prostate* 2015; 75: 693–705.
- 65 Bartolomei MS, Zemel S, Tilghman SM. Parental imprinting of the mouse H19 gene. *Nature* 1991; **351**: 153–5.
- 66 Zemel S, Bartolomei MS, Tilghman SM. Physical linkage of two mammalian imprinted genes, H19 and insulin-like growth factor 2. *Nat Genet* 1992; **2**: 61–5.
- 67 Lustig-Yariv O, Schulze E, Komitowski D *et al.* The expression of the imprinted genes H19 and IGF-2 in choriocarcinoma cell lines. Is H19 a tumor suppressor gene? *Oncogene* 1997; **15**: 169–77.
- 68 Ariel I, Sughayer M, Fellig Y et al. The imprinted H19 gene is a marker of early recurrence in human bladder carcinoma. Mol Pathol 2000; 53: 320–3.
- 69 Zhu M, Chen Q, Liu X *et al.* IncRNA H19/miR-675 axis represses prostate cancer metastasis by targeting TGFB1. *FEBS J* 2014; **281**: 3766–75.
- 70 Ponting CP, Oliver PL, Reik W. Evolution and functions of long noncoding RNAs. Cell 2009; 136: 629–41.

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- 71 Kung JT, Colognori D, Lee JT. Long noncoding RNAs: past, present, and future. *Genetics* 2013; **193**: 651–69.
- 72 Groskopf J, Aubin SM, Deras IL *et al.* APTIMA PCA3 molecular urine test: development of a method to aid in the diagnosis of prostate cancer. *Clin Chem* 2006; **52**: 1089–95.
- 73 Ouyang B, Bracken B, Burke B *et al.* A duplex quantitative polymerase chain reaction assay based on quantification of alpha-methylacyl-CoA racemase transcripts and prostate cancer antigen 3 in urine sediments improved diagnostic accuracy for prostate cancer. *J Urol* 2009; **181**: 2508–13.
- 74 Heidenreich A, Bastian PJ, Bellmunt J et al. EAU guidelines on prostate cancer. Part 1: screening, diagnosis, and local treatment with curative intentupdate 2013. Eur Urol 2014; 65: 124–37.
- 75 Roobol MJ, Schroder FH, van Leeuwen P *et al.* Performance of the prostate cancer antigen 3 (PCA3) gene and prostate-specific antigen in prescreened men: exploring the value of PCA3 for a first-line diagnostic test. *Eur Urol* 2010; **58**: 475–81.
- 76 Ren S, Wang F, Shen J et al. Long non-coding RNA metastasis associated in lung adenocarcinoma transcript 1 derived miniRNA as a novel plasma-

based biomarker for diagnosing prostate cancer. Eur J Cancer 2013; 49: 2949–59.

- 77 Tong YS, Wang XW, Zhou XL *et al.* Identification of the long non-coding RNA POU3F3 in plasma as a novel biomarker for diagnosis of esophageal squamous cell carcinoma. *Mol Cancer* 2015; **14**: 3.
- 78 Zhou X, Yin C, Dang Y et al. Identification of the long non-coding RNA H19 in plasma as a novel biomarker for diagnosis of gastric cancer. Sci Rep 2015; 5: 11516.
- 79 FANTOM Consortium. The transcriptional network that controls growth arrest and differentiation in a human myeloid leukemia cell line. *Nat Genet* 2009; **41**: 553–62.
- 80 Carninci P, Kasukawa T, Katayama S et al. The transcriptional landscape of the mammalian genome. Science 2005; **309**: 1559–63.
- 81 Lizio M, Harshbarger J, Abugessaisa I *et al.* Update of the FANTOM web resource: high resolution transcriptome of diverse cell types in mammals. *Nucleic Acids Res* 2017; 4; 45(D1): D737–43.
- 82 Thompson JF, Steinmann KE. Single molecule sequencing with a HeliScope genetic analysis system. *Curr Protoc Mol Biol* 2010; https://doi.org/10.1002/ 0471142727.mb0710s92.