

Recent Advances in Functional Carbon Quantum Dots for Antitumour

Rong Cai ¹
Long Xiao ¹
Meixiu Liu ¹
Fengyi Du ²
Zhirong Wang ¹

¹Central Laboratory, Zhangjiagang TCM Hospital Affiliated to Nanjing University of Chinese Medicine, Suzhou, Jiangsu, 215600, People's Republic of China;

²School of Medicine, Zhenjiang, Jiangsu, 212013, People's Republic of China

Abstract: Carbon quantum dots (CQDs) are an emerging class of quasi-zero-dimensional photoluminescent nanomaterials with particle sizes less than 10 nm. Owing to their favourable water dispersion, strong chemical inertia, stable optical performance, and good biocompatibility, CQDs have become prominent in biomedical fields. CQDs can be fabricated by “top-down” and “bottom-up” methods, both of which involve oxidation, carbonization, pyrolysis and polymerization. The functions of CQDs include biological imaging, biosensing, drug delivery, gene carrying, antimicrobial performance, photothermal ablation and so on, which enable them to be utilized in antitumour applications. The purpose of this review is to summarize the research progress of CQDs in antitumour applications from preparation and characterization to application prospects. Furthermore, the challenges and opportunities of CQDs are discussed along with future perspectives for precise individual therapy of tumours.

Keywords: CQDs, antitumour, drug delivery, phototherapy

Introduction

At present, tumours are still one of the main culprits threatening human health. Due to the heterogeneity and metastatic nature of tumours, their diagnosis, treatment and control are not easy. Depending on the different sites and nature of the tumour, a comprehensive analysis of the clinical manifestations and signs of the patient, combined with laboratory examination and imaging and cytopathological examination, usually leads to a definitive diagnosis.¹ However, there is still a lack of ideal and specific methods for early diagnosis, especially for deep tumours. There are many kinds of malignant tumours, which have different properties, involve different tissues and organs, occur during different stages of diseases, and have different responses to various treatments, so most patients need comprehensive treatment.² Surgery is a good way to remove tumours, but it can only eradicate early or earlier solid tumours.³ Drug therapy is chemotherapy that can kill tumours. Since tumour cells differ most from normal cells in their rapid cell division and growth, antitumour drugs usually work by disrupting the mechanisms of cell division, such as DNA replication or chromosomal separation.^{4,5} Nevertheless, most chemotherapeutic drugs are not specific, so they simultaneously kill normal tissue cells that divide, often harming healthy tissues that need to divide to function properly. Radiotherapy uses radiation to kill tumour cells, thereby shrinking the tumours.^{6–8} Radiation can control the growth of tumour cells by damaging their genetic material, thus preventing them from growing or dividing.^{9,10} The disadvantage of radiotherapy is that it is limited to areas of radiation exposure, and it kills tumour cells while affecting normal cells.¹¹ Therefore, the

Correspondence: Zhirong Wang; Fengyi Du
Tel +86 138 6222 4593; +86 159 5290 0513
Email zjgfy_spine_wzr@njucm.edu.cn;
biodfy@ujs.edu.cn



early detection and diagnosis of tumours and the specific elimination of tumour cells have become the direction of antitumour research.

Located in the second period of the IVA group on the periodic table, carbon is a simple, stable and common element that is widely present in nature.¹² Since the last century, carbon has played an important role in the development of nanomaterials.¹³ Although these nanomaterials are mainly composed of the same element, carbon, they have different functions and sizes due to their different structures, which ensures that they have different but extraordinary properties. Carbon nanomaterials have been widely reported and include carbon nanotubes (CNTs),¹⁴ carbon fibres (CFs)¹⁵ and carbon quantum dots (CQDs).¹⁶ CNTs, also known as bucky tubes, are one-dimensional quantum materials with a unique structure of coaxial circular tubes.¹⁷ Research on the nucleation thermodynamics of CNTs is ongoing and has made great progress.¹⁸ CF is a unique fibre with more than 90% carbon content, and it has great application potential in the development of advanced technology.¹⁹ CQDs are carbon-based zero-dimensional nanoparticles with particle sizes less than 10 nm that are composed of dispersed spherical carbon particles.²⁰ Their nuclei are generally formed by sp^2 hybridization, and they have two kinds of structures: lattices and nonlattices.²¹ In 2004, Professor Xiaoyou Xu of the University of South Carolina et al reported for the first time that during the electrophoretic purification of single-walled carbon nanotubes, CQDs that can emit bright fluorescence were found in the products,²² thus introducing CQDs and opening the door for scientific researchers to further explore them. In 2007, Cao et al reported that the two-photon absorption cross sections of prepared CQDs were comparable to the two-photon absorption cross sections of the best semiconductor quantum dots or core shell nanoparticles reported in previous literature and demonstrated that the CQDs could enter breast cancer cells.²³ In 2012, Lin et al mixed CQDs with oxidants such as $KMnO_4$ and Ce^{4+} to stimulate their chemiluminescence, which is the production of light through a chemical reaction.²⁴ In 2017, Yuan et al first fabricated CQDs that had the ability to emit fluorescence from red to blue light species by controlling the fusion and carbonization of citric acid and diamionaphthalene, which provided a new clue for the development of light-emitting diodes.²⁵ In 2019, Guo et al embedded CQDs into alginate gel beads to form composite materials that could absorb rare earth elements, which could be applied as a rare earth element adsorbent for environmental protection.²⁶ In 2021, Tao et al developed CQDs that emitted blue light in water and redshifted in the solid state and reported

that these CQDs could be utilized as fluorescent ink for anti-counterfeiting and for printing high-quality fluorescent images.²⁷ Figure 1 shows the development history of CQDs since their discovery.

With the in-depth exploration of CQDs, researchers have gradually obtained a thorough understanding of them. Generally, the following commonalities can be listed. Due to the influence of precursors, the surface of CQDs is covered with hydrophilic functional groups such as $-COOH$ and $-OH$, and the particle sizes are small, which endows them with the potential for good biocompatibility.^{33–35} Besides, the hydrophilic groups on the surface of CQDs promote the interaction with proteins in vivo, resulting in the formation of protein corona, which changes the original properties of CQDs.^{36,37} Therefore, how to change the surface properties of CQDs to realize the controllable load of proteins and how to regulate the interaction between the two has become one of the hot topics that many scientists compete to study. CQDs also have excellent fluorescence properties, which gives them considerable application prospects in the field of bioluminescence imaging and sensors.³⁸ Due to their small structure, CQDs can be trapped in tumour sites by an enhanced permeability and retention effect (EPR).^{39,40} In normal tissues, the microvascular endothelial space is dense and intact, and CQDs are difficult to penetrate the vascular wall, while in solid tumor tissues, blood vessels are abundant, and the wide vascular wall space, poor structural integrity and missing lymphatic return result in high selective permeability and retention of large CQDs.^{41,42} Therefore, EPR effect promotes the selective distribution of CQDs in tumor tissues, which can increase antitumour efficacy and reduce systemic side effects. Moreover, the additional functions of CQDs can be added by modification and doping. We can utilize the abovementioned characteristics of CQDs to develop and expand their antitumour applications and provide new ideas for the discovery, diagnosis, treatment and monitoring of tumours in the future. This review outlines the features and characterizations of CQDs, introduces the preparation of CQDs, elaborates the application of CQDs in antitumour applications, including drug delivery, phototherapy, monitoring and auto-antitumour applications, and discusses the design of related topics.

Features and Characteristics of CQDs

Preparation of CQDs

The synthesis methods of CQDs include the “top-down” method and the “bottom-up” method.^{43–45} The “top-down”

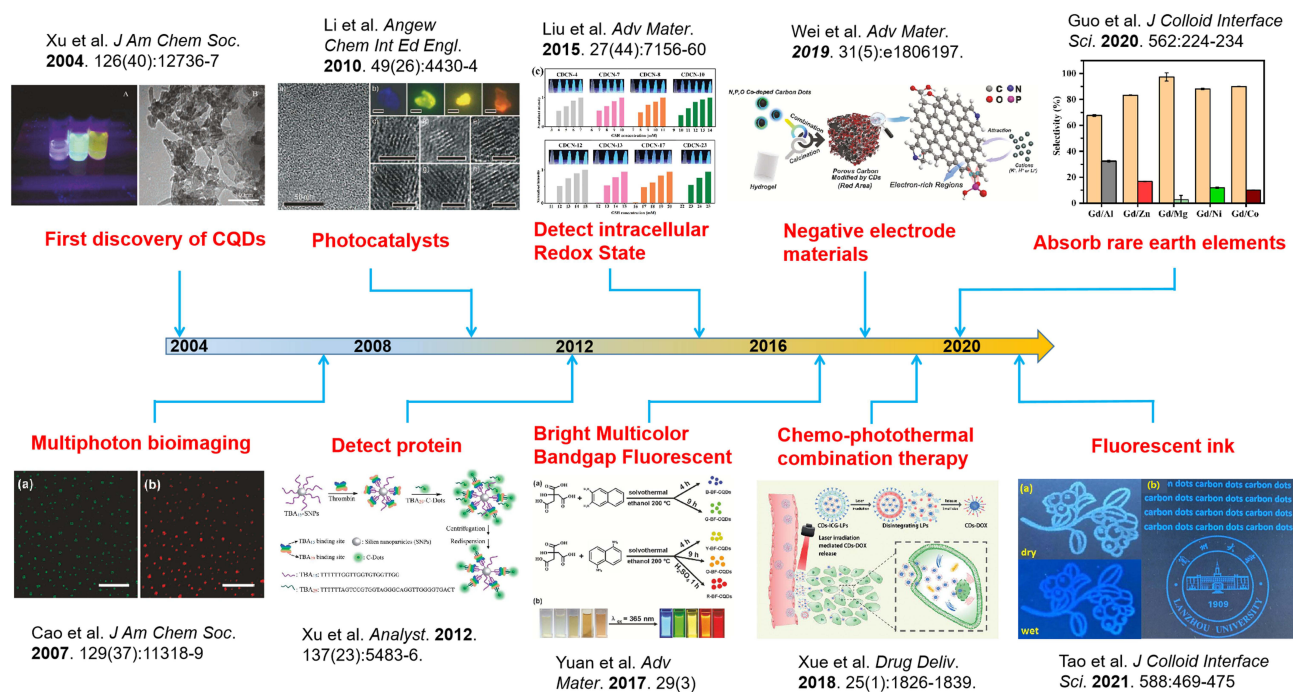


Figure 1 The development history of CQDs.

Notes: Images reprinted with permission from Xu X, Ray R, Gu Y, et al. Electrophoretic analysis and purification of fluorescent single-walled carbon nanotube fragments. *J Am Chem Soc.* 2004;126(40):12736–12737, Copyright (2004) American Chemical Society²²; reprinted with permission from Cao L, Wang X, Meziani MJ, et al. Carbon dots for multiphoton bioimaging. *J Am Chem Soc.* 2007;129(37):11318–11319, Copyright (2007) American Chemical Society²³; reprinted with permission from Yuan F, Wang Z, Li X, et al. Bright Multicolor Bandgap Fluorescent Carbon Quantum Dots for Electroluminescent Light-Emitting Diodes. *Adv Mater.* 2017;29(3). © 2016 WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim²⁵; reprinted with permission from *J Colloid Interface Sci.* 562, Guo Z, Li Q, Li Z, et al. Fabrication of efficient alginate composite beads embedded with N-doped carbon dots and their application for enhanced rare earth elements adsorption from aqueous solutions. 224–234, Copyright 2020, with permission from Elsevier²⁶; reprinted with permission from *J Colloid Interface Sci.* 588, Tao Y, Lin J, Wang D, Wang Y. Na⁺-functionalized carbon dots with aggregation-induced and enhanced cyan emission. 469–475, Copyright (2021), with permission from Elsevier²⁷; reprinted with permission from Li H, He X, Kang Z, et al. Water-soluble fluorescent carbon quantum dots and photocatalyst design. *Angew Chem Int Ed Engl.* 2010;49(26):4430–4434. Copyright © 2010 WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim²⁸, reprinted with permission from Liu Y, Tian Y, Tian Y, et al. Carbon-Dot-Based Nanosensors for the Detection of Intracellular Redox State. *Adv Mater.* 2015;27(44):7156–7160. © 2015 WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim²⁹; reprinted with permission from Xue X, Fang T, Yin L. Multistage delivery of CDs-DOX/ICG-loaded liposome for highly penetration and effective chemo-photothermal combination therapy. *Drug Deliv.* 2018;25(1):1826–1839³⁰; republished with permission of Xu B, Zhao C, Wei W, et al. Aptamer carbon nanodot sandwich used for fluorescent detection of protein. *Analyst.* 2012;137(23):5483–5486, permission conveyed through Copyright Clearance Center, Inc³¹; reprinted with permission from Wei JS, Ding C, Zhang P, et al. Robust Negative Electrode Materials Derived from Carbon Dots and Porous Hydrogels for High-Performance Hybrid Supercapacitors. *Adv Mater.* 2019;31(5):e1806197. © 2018 WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim.³²

method refers to the separation of CQDs with small particle sizes from carbon-based materials (such as CNTs, CF, graphite) by chemical or physical methods, including arc discharge,²² electrochemical methods,⁴⁶ chemical oxidation,⁴⁷ laser ablation⁴⁸ and combustion.⁴⁹ Contrary to the “top-down” method, the “bottom-up” method mainly refers to the formation of CQDs by carbonization and polymerization of a series of small molecules through chemical reactions, including hydrothermal methods,⁵⁰ microwave methods⁵⁵ and template methods.⁶⁰ Table 1 summarizes the advantages and disadvantages of various methods for preparing CQDs.

“Top-Down” Method

Arc discharge is a self-sustaining discharge method that uses gas plasma generated in a sealed reactor to drive

anodic electrodes to decompose CQDs from bulk carbon precursors. To generate high-energy plasma, the heat of the reactor can be as high as tens of thousands of degrees Celsius under the action of the electric current, so arc discharge is the most intense method of gas discharge. The most obvious visual characteristics of arc discharge are bright arc columns and electrode spots, while energy balance is another important law that describes the arc discharge phenomenon. The generation of energy is the JHEAT of the arc, and the divergence of energy occurs through radiation, convection and conduction. Xu et al accidentally harvested three kinds of CQDs with different fluorescence characteristics when purifying single-walled carbon nanotubes by the arc discharge method. These three CQDs can emit blue-green, yellow and orange

Table 1 Various Methods of Preparing CQDs as Well as Their Advantages and Disadvantages

Method	Classification	Advantages	Disadvantages	Reference
Arc discharge	“Top-down” method	Excellent water dispersibility	Particle size with large difference	[22]
Electrochemical method	“Top-down” method	Easy to adjust the particle size and fluorescence properties, high yield	Special equipment required	[46,61]
Chemical oxidation	“Top-down” method	Excellent water dispersion and fluorescence characteristics	Special equipment required	[47,62]
Laser ablation	“Top-down” method	Excellent water dispersion and fluorescence characteristics	Low yield, inhomogeneous particle size and complex operation	[48,63]
Combustion	“Top-down” method	Simple operation	Low yield and inhomogeneous particle size	[49,64]
Hydrothermal method	“Bottom-up” method	Homogeneous particle size and high yield	Long time consuming	[50–54]
Microwave method	“Bottom-up” method	Simple, fast and environmental protection	Low yield and multiple impurities	[55–59]
Template method	“Bottom-up” method	Homogeneous particle size and excellent water dispersibility	Tedious operation	[60]

fluorescence at 365 nm.²² Although this method can synthesize CQDs with good water dispersion, the size of the CQDs formed in the discharge process is different, and they generally have a larger particle size distribution.

Electrochemical methods are simple and convenient preparation methods that can be carried out under normal temperature and pressure. With the function of inhibiting maltose activity, chiral CQDs were fabricated from L- or D-glutamic acid through electrochemical methods to be used as a candidate drug for controlling blood sugar.⁴⁶ Yen et al reported a three-electrode electrochemical method for the direct synthesis of high-quality CQDs in pure water electrolyte.⁶¹ This method provides a step for the preparation of aqueous CQD solutions without further postprocessing, such as filtration, dialysis, centrifugation, column chromatography and gel electrophoresis. The results show that the prepared CQDs have a catalytic effect on H₂O₂ and reduce the charge transfer resistance and ionic diffusion resistance. Although special equipment is needed, the particle size and fluorescence properties of CQDs synthesized by electrochemical methods can be easily adjusted, and the CQDs are formed in high yield.

Chemical oxidation has been widely used to decompose bulk carbon-based materials into CQDs and introduce hydrophilic groups such as -COOH or -OH, which can significantly improve the water dispersion and fluorescence

of CQDs. Qiao et al prepared CQDs by a chemical oxidation method using activated carbon as a raw material in the treatment of nitric acid, which had excellent photostability and cell avirulence and could penetrate living cells, so they had the potential to be utilized as living cell imaging agents.⁴⁷ Hu et al utilized coal tar pitch as a raw material to prepare CQDs-based nanosensors by chemical oxidation for the detection of Cu²⁺, Fe³⁺ and L-ascorbic acid.⁶²

Laser ablation is used to irradiate the surface of carbon-based materials with large particle sizes with a high-energy laser pulse, causing the materials to reach a high temperature and high pressure, quickly heat up and evaporate into the plasma state, and finally crystallize through steam to form CQDs. Yu et al prepared triple valence CQDs in toluene by laser irradiation, which increased the fluorescence lifetime.⁴⁸ Moreover, graphene was observed during production through a real-time detection system. Kang et al transformed Earth's rich resources and low-cost coal into CQDs within 5 minutes through a simple, low-cost, environmentally friendly liquid pulse laser ablation method.⁶³ CQDs prepared by laser ablation have good water dispersibility and strong fluorescence properties, but their wide application is hindered by low yield, uneven particle size and complex operation.

Combustion is a method that involves collecting CQDs from the products of burning organic matter. Zinc oxide and

zinc sulfide CQDs were synthesized by combustion using zinc nitrate and thioacetamide as raw materials and ethylene glycol as fuel, whose water solution exhibited three blue-green emission peaks at 420 nm, 486 nm and 520 nm.⁴⁹ Han et al prepared single- and double-doped CQDs by a combustion method, which could skillfully adjust the doping amount of heteroatoms by changing the precursor concentration.⁶⁴ Although combustion is easy to operate, the yield of CQDs is low, and the particle size is different.

“Bottom-Up” Method

The hydrothermal method is one of the most commonly used methods to synthesize CQDs. The main process is to dissolve small organic molecules and/or polymers in water or organic solvents to form reaction precursors, followed by transferring them to a stainless steel autoclave lined with Teflon and finally forming CQDs at high temperature. Ye et al first prepared two-photon CQDs by a hydrothermal method for detecting intracellular pH, which depended on the disaggregation and aggregation of their physical states, thus contributing to future applications in living systems.⁵⁰ Atchudan et al utilized *P. acidus* and aqueous ammonia as raw materials to form CQDs by a hydrothermal method.⁵¹ The prepared CQDs can be used as fluorescence inks and Fe³⁺ detectors, which provides a new idea for the monitoring of Fe³⁺ in industrial wastewater. Jiang et al prepared room temperature phosphorescence (RTP)-based CQDs from trimellitic acid (TA) by a hydrothermal method; these CQDs were designed to be applied for anti-counterfeiting and information encryption.⁵² Zheng et al utilized amino acids and polymers as raw materials to fabricate CQDs for blood analysis and cell imaging by hydrothermal methods.⁵³ Our group reported a method of preparing iodine-doped CQDs as a high-efficiency CT contrast agent for the first time by hydrothermal carbonization.⁵⁴ Compared with traditional iodine contrast agents, the prepared CQDs not only have unique photoluminescence and X-ray attenuation characteristics but also have the characteristics of a long cycle and passive target CT imaging. The operation of the hydrothermal method is simple, the size of CQDs is almost the same, and the yield is high.

Among the bottom-up methods, the microwave method is the most rapid commercial method. The preparation process involves dissolving the precursor in the solution to form a transparent solution, followed by heating it in a microwave oven. CQDs with PEI modified on the surface were prepared by the microwave method.⁵⁵ Among them, PEI played two important roles: enhanced

fluorescence and gene delivery. Singh et al prepared CQDs with quinolate phosphoribosyl transferase (QPRTase) on the surface by the microwave method, which was used to detect the endogenous neurotoxin quinolinic acid (QA), thus providing a candidate for the treatment of neurological disorders.⁵⁶ In addition, to address the difference in pH between wounds and normal regions, CQDs with the function of monitoring pH were fabricated by the microwave method; these CQDs provided new clues for the identification, treatment and healing of wounds in the future.⁵⁷ In addition, Pierrat et al utilized citric acid and BPEI25K as raw materials to prepare CQDs by the microwave method to transfect nucleic acids in vivo and in vitro, thus providing a new method for drug delivery to the lungs.⁵⁸ Our group also reported a synthesis of nitrogen-doped CQDs that uses citric acid and ammonium acetate as precursors through a microwave method. These CQDs had strong photoluminescence, good water solubility and stable optical properties.⁵⁹ It is worth noting that an inhomogeneous multilayer structure was observed for the first time in nitrogen-doped CQDs. The microwave method is a simple, rapid and environmentally friendly method for the synthesis of CQDs.

The template method is a technique of preparing CQDs with special materials as templates and then removing the template by physical or chemical methods to collect the CQDs. Yang et al synthesized organic templated TiOS under solvent-free conditions, performed heat treatment at 300 °C in vacuum to prepare N-CDs@TiOS, and finally, they transformed the organic template into N-CDs and embedded them into a titanium sulfate matrix to collect the CQDs.⁶⁰ This kind of CQDs can be used as an efficient photosensitizer and has good application potential in visible light-driven photocatalysis, which provides an effective way to prepare CQDs by a template method for visible light catalysis. The CQDs prepared by the template method have good water dispersibility and uniform particle size, but the operation is cumbersome.

Photoluminescence of CQDs

Photoluminescence (PL), a basic characteristic of CQDs, has been widely researched in related fields. PL refers to the electrons of matter jump from the valence band to the conduction band and leave holes in the valence band under the excitation of light.⁶⁵ In the conduction band and valence band, electrons and holes reach the lowest unoccupied excited state by relaxation and enter a quasi-equilibrium state. In this quasi-equilibrium state, electrons and holes

emit light through recombination, forming spectra of intensity or distributing energy across different wavelengths.⁶⁶ Although PL can only be used for qualitative analysis rather than quantitative analysis, it has high resolution.⁶⁷ In recent years, many studies have been performed on the PL mechanism of CQDs, but there has been no unified conclusion.^{68–70} This lack of consensus may be because different raw materials, surface passivators or preparation methods have a great impact on the structure and PL properties of CQDs. It is widely believed that the mechanism of the PL properties of CQDs involves surface defects,⁷¹ lattice defects⁷² and quantum confinement effects.⁷³ The PL analysis of CQDs has the significant advantages of high sensitivity and strong selectivity, including an excitation spectrum and emission spectrum, making it possible to choose a variety of wavelengths for molecular fluorescence analysis.⁷⁴ It is noteworthy that the traditional CQDs are excited by single photon, which leads to photobleaching when they are used for tumour tissue imaging. Two-photon CQDs are excited by two-photon near-infrared light, which can improve the tissue penetration, and thus observe in deep tissues and prolong the observation time.^{75,76} Therefore, their research in tumour tissue imaging is more popular. PL is an important characteristic of CQDs, and PL measurements are helpful for later research. The influence of external pressure on the fluorescence properties of CQDs can be judged by measuring the PL and UV-Vis spectra.⁷⁷ As shown in Figure 2A, Ding et al utilized urea and p-phenylenediamine to fabricate CQDs by a hydrothermal method; the CQDs were filtered through silica column

chromatography into eight different components, each of which emitted a different fluorescence colour.⁷⁸ In addition, the cerium-doped CQDs (Ce-doped CQDs) prepared by our research group also showed excellent fluorescence properties.⁷⁹ As shown in Figure 2B, with increasing excitation wavelength, the fluorescence intensity of Ce-doped CQDs increased gradually, reaching a peak at 360 nm, and then decreased gradually.

UV visible (UV-Vis) spectrophotometry is a method for the determination of the absorbance of the substance in the range of 190–800 nm, which can be used to determine the linear optical absorption behaviour of CQDs.⁸⁰ When the light passes through the solution, the absorption of the substance changes with the wavelength of the light. In the ultraviolet absorption spectrum, electrons undergo four different types of transitions, $\sigma \rightarrow \sigma^* n \rightarrow \sigma^* \pi \rightarrow \pi^*$ and $n \rightarrow \pi^*$ which are arranged according to the decreasing order of energy required by various transition types.⁸¹ Among them, $\sigma \rightarrow \sigma^*$ transitions indicate the presence of C-C bonds,⁸² $n \rightarrow \sigma^*$ transitions demonstrate the presence of -OH and -NH₂ bonds,⁸³ $\pi \rightarrow \pi^*$ transitions occur due to the double bond absorption of aromatic nuclei,⁸⁴ and $n \rightarrow \pi^*$ transitions imply the presence of C=O, C=S, -N=O and -N=N-.⁸⁵ The same concentration of the solution to be measured has different absorbance for different wavelengths of light, while for the same solution to be measured, the higher the concentration, the greater the absorbance, but the wavelength corresponding to the maximum absorption peak (λ_{max}) is the same. Therefore, by measuring the

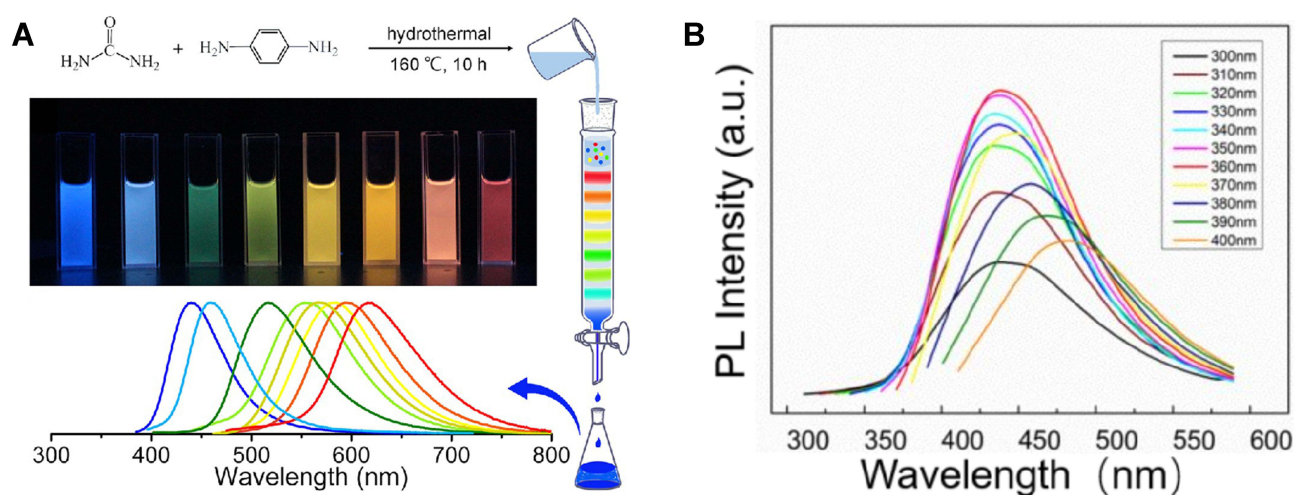


Figure 2 (A) Scheme of CQDs. Reprinted with permission from Ding H, Yu SB, Wei JS, Xiong HM. Full-color light-emitting carbon dots with a surface-state-controlled luminescence mechanism. *ACS Nano*. 2016;10(1):484–491. Copyright (2016) American Chemical Society.⁷⁸ and **(B)** PL spectra of Ce-doped CQDs. Reprinted with permission from Zhang M, Zhao L, Du F, et al. Facile synthesis of cerium-doped carbon quantum dots as a highly efficient antioxidant for free radical scavenging. *Nanotechnology*. 2019;30(32):325101Z.⁷⁹

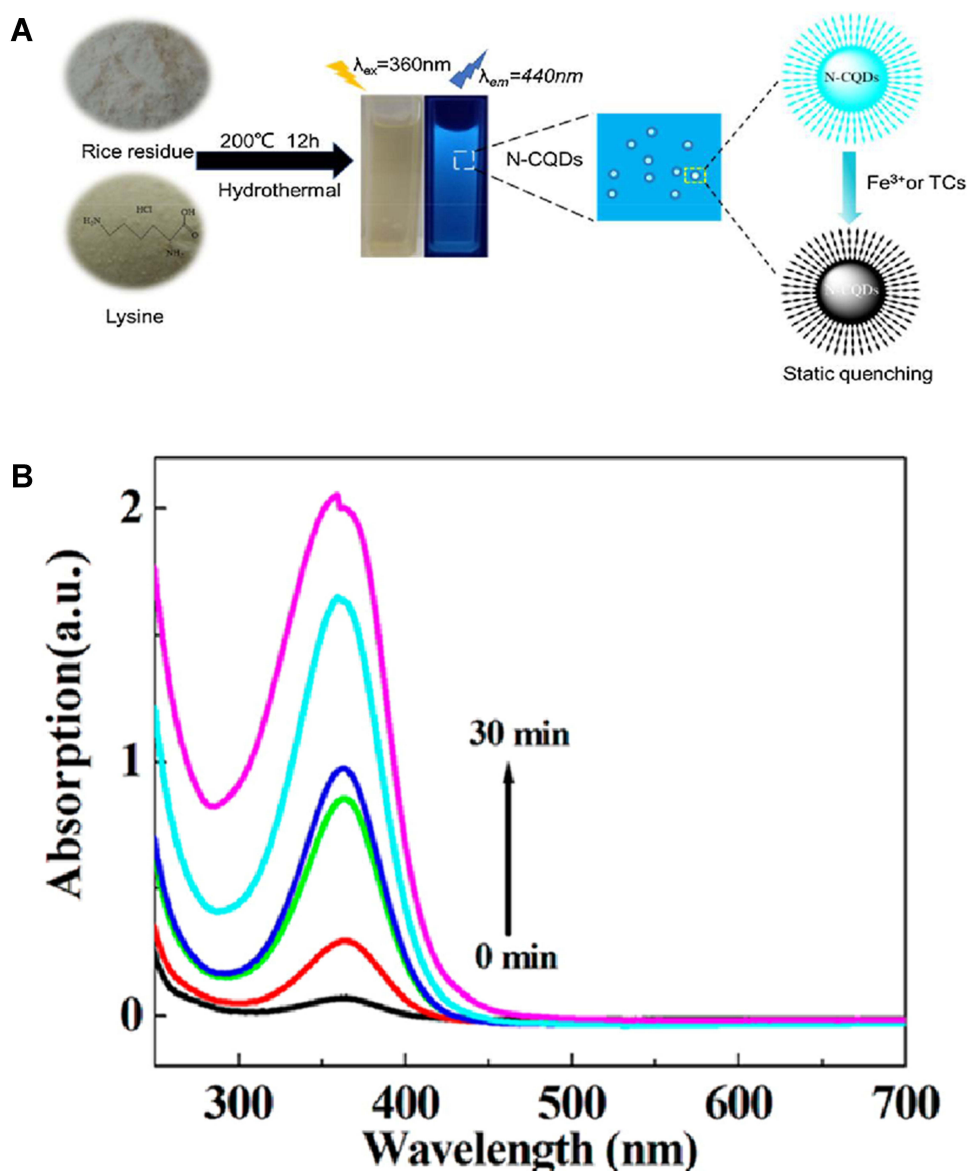


Figure 3 (A) Mechanism of N-CQDs. Reprinted from *J Colloid Interface Sci*, 539, Qi H, Teng M, Liu M, et al. Biomass-derived nitrogen-doped carbon quantum dots: highly selective fluorescent probe for detecting Fe³⁺ ions and tetracyclines. 332–341, Copyright 2019, with permission from Elsevier.⁸⁶ and **(B)** UV-Vis spectra of B-CDs. Reprinted with permission from Liu H, He Z, Jiang LP, Zhu JJ. Microwave-assisted synthesis of wavelength-tunable photoluminescent carbon nanodots and their potential applications. *ACS Appl Mater Interfaces*. 2015;7(8):4913–4920. Copyright (2015) American Chemical Society.⁸⁷
Abbreviations: N-CQDs, nitrogen-doped CQDs; B-CDs, blue luminescent carbon dots.

absorbance of the substance at different wavelengths and drawing the relationship between the absorbance and wavelength, the absorption spectrum of the substance can be obtained. As shown in Figure 3A, Qi et al prepared nitrogen-doped CQDs (N-CQDs), which fluoresced blue under ultraviolet light but were quenched when Fe³⁺ was encountered.⁸⁶ In addition, the prepared aqueous B-CD solution was colourless and transparent under normal environmental conditions.⁸⁷ With the extension of microwave irradiation time, the peak value of the aqueous B-CD solution at 360 nm gradually increased, and its colour gradually

darkened, which indicated that microwave irradiation could shorten the reaction time (Figure 3B).

Characterization and Biocompatibility of CQDs

A requirement for the biological application of CQDs is that they must be harmless to the organism, so analysis of their biocompatibility is the primary task. The excellent biocompatibility of CQDs is due to their small size and surface groups,⁸⁸ as well as biocompatibility of precursors,

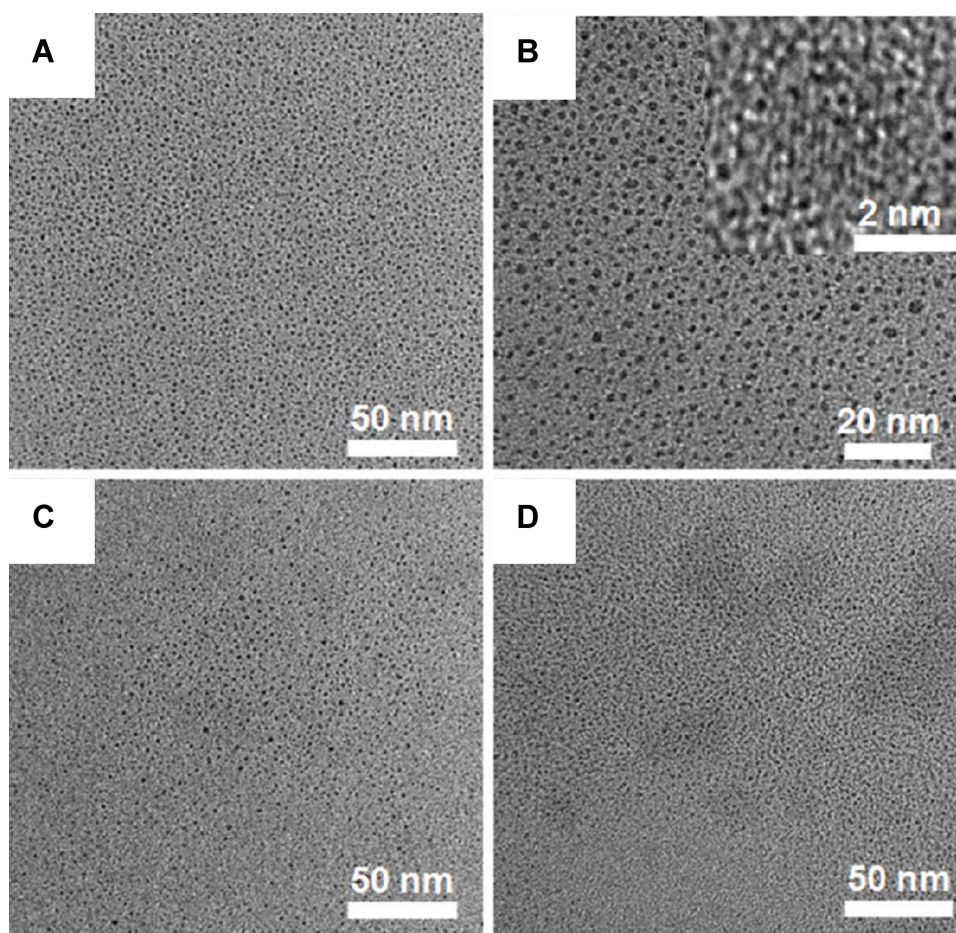


Figure 4 TEM and high-resolution TEM images of (A and B) r-CDs, (C) c-CDs, and (D) l-CDs. Reprinted with permission from Zhang J, Yuan Y, Liang G, Yu SH. Scale-up synthesis of fragrant nitrogen-doped carbon dots from bee pollens for bioimaging and catalysis. *Adv Sci (Weinh)*. 2015;2(4):1500002. © 2015 The Authors. Published by WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim.⁹⁹

Abbreviations: r-CDs, carbon quantum dots from rapeseed flower bee pollen; c-CDs, carbon quantum dots from camellia bee pollen; l-CDs, carbon quantum dots from lotus bee pollen.

distribution of the surface charges and even the structural properties of carbon elements.^{89–91} The particle size of CQDs is generally less than 10 nm, which makes it impossible to observe through ordinary optical microscopy. Moreover, the wavelength of transmission electron microscopy (TEM) is much shorter than that of visible light and ultraviolet light, which enables the resolution of TEM to reach 0.2 nm.^{92–94} The working principle of TEM is to project the accelerated and focused electron beam onto a very thin sample. The direction of the electron beam changes due to the collision between the electron and the atom in the sample, resulting in solid angle reflection.^{95–97} The size of the scattering angle is related to the density and thickness of the sample, so different light and dark images are formed, which are displayed on the imaging device after the image is magnified and focused.⁹⁸

The three kinds of CQDs, including r-CDs (Figure 4A), c-CDs (Figure 4C), and l-CDs (Figure 4D), were distributed uniformly in water, and the particle size was approximately 2 nm.⁹⁹ Furthermore, as shown in Figure 4B, the r-CDs were magnified again, and their crystal structure was observed by high-resolution transmission electron microscopy (HRTEM). In addition, Li et al prepared two CQDs, including ACDs and OCDs.¹⁰⁰ The ACDs had a crystalline structure with a lattice spacing of 0.208 nm, while the OCDs did not, which implied that there were lattice and nonlattice conformations in the CQDs. In addition, TEM showed that the size of the N, P-CDs varied, and the maximum size was approximately 15 nm.¹⁰¹ In addition, the N,P-CDs were magnified again by HRTEM. Different N,P-CDs had different lattice spacings of 0.24 nm, 0.35 nm and 0.51 nm, which implied that the N, P-CDs did not have uniform lattice spacing, which may be related to the plane spacing of the N,P-CDs. Furthermore,

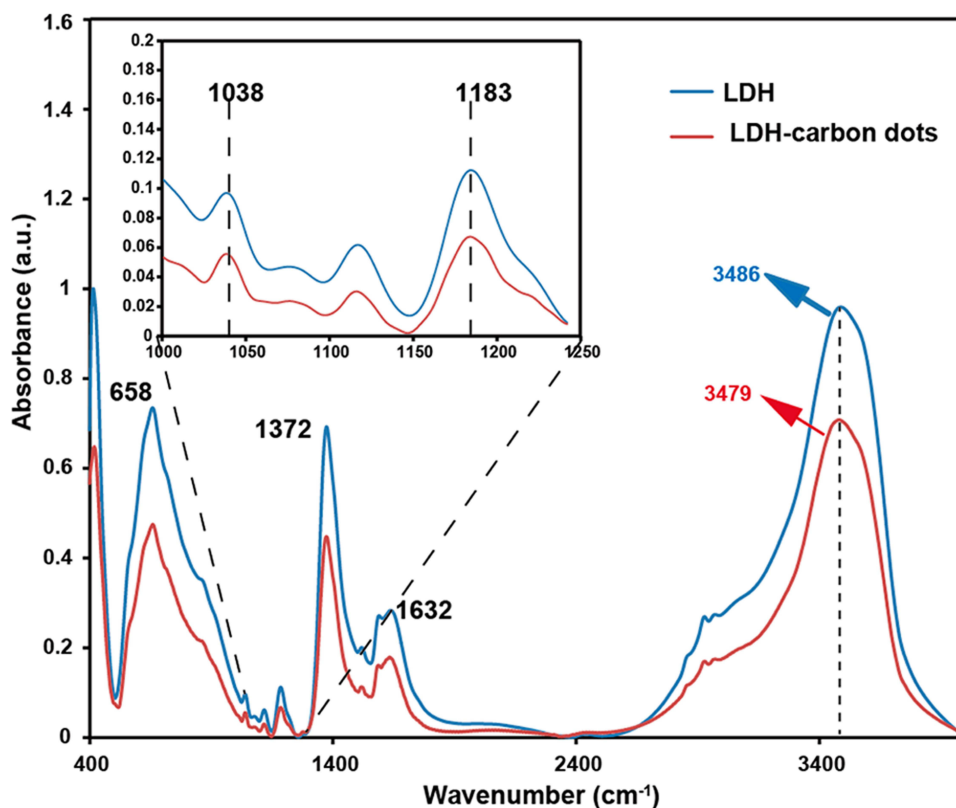


Figure 5 FTIR spectra of LDH and LDH-carbon quantum dots. Reprinted with permission from Zhang M, Yao Q, Lu C, et al. Layered double hydroxide-carbon dot composite: high-performance adsorbent for removal of anionic organic dye. *ACS Appl Mater Interfaces*. 2014;6(22):20225–20233. Copyright (2014) American Chemical Society.¹⁰⁸

Abbreviations: FTIR, Fourier transform infrared spectroscopy; LDH, layered double hydroxide.

energy dispersive X-ray (EDX) spectroscopy is usually used in combination with TEM because it can determine the content of elements in CQDs by comparing the intensities of spectral lines of different elements.

Fourier transform infrared spectroscopy (FTIR), which is mainly composed of a Michelson interferometer and computer, can be used to analyse the chemical structure and surface groups of CQDs.¹⁰² A Michelson interferometer is a precision measuring instrument that uses the interference principle of light to precisely measure the length and tiny changes in length.¹⁰³ The principle is that incident light is reflected back by the corresponding plane mirror after being divided into two beams by the spectroscopy. Because the two beams have the same frequency, the same vibration direction and constant phase difference (ie, the interference conditions are satisfied), interference can occur.¹⁰⁴ The different optical paths of the two beams can be realized by adjusting the length of the interference arm and changing the refractive index of the medium so that different interference patterns can be formed.¹⁰⁵ The intensity distribution of the original

light source can be calculated by Fourier transform of the interference function on a computer.¹⁰⁶ Compared with traditional spectrometers, FTIR has the following advantages: fast scanning speed, high resolution, high sensitivity and wide spectral range.¹⁰⁷ Zhang et al prepared layered double hydroxide-combined CQDs (LDH-carbon quantum dots) capable of adsorbing methyl blue.¹⁰⁸ As shown in Figure 5, compared with the fixed hydrogen bond peak of LDH, the hydrogen bond peak of LDH-carbon quantum dots blueshifted (3490 to 3479 cm^{-1}), suggesting that there was a hydrogen bond between LDH and CQDs. Moreover, the peaks at 1183 cm^{-1} and 1038 cm^{-1} were attributed to the $-\text{SO}_3^-$ of LDH and LDH-carbon quantum dots.

X-ray photoelectron spectroscopy (XPS), an advanced analytical technique for the microanalysis of electronic materials and components, can analyse the chemical elements in CQDs, and it is usually coordinated with auger electron spectroscopy (AES).¹⁰⁹ The principle of XPS is to irradiate the CQDs with X-rays so that the core electrons of the atoms or molecules (also known as photoelectrons)

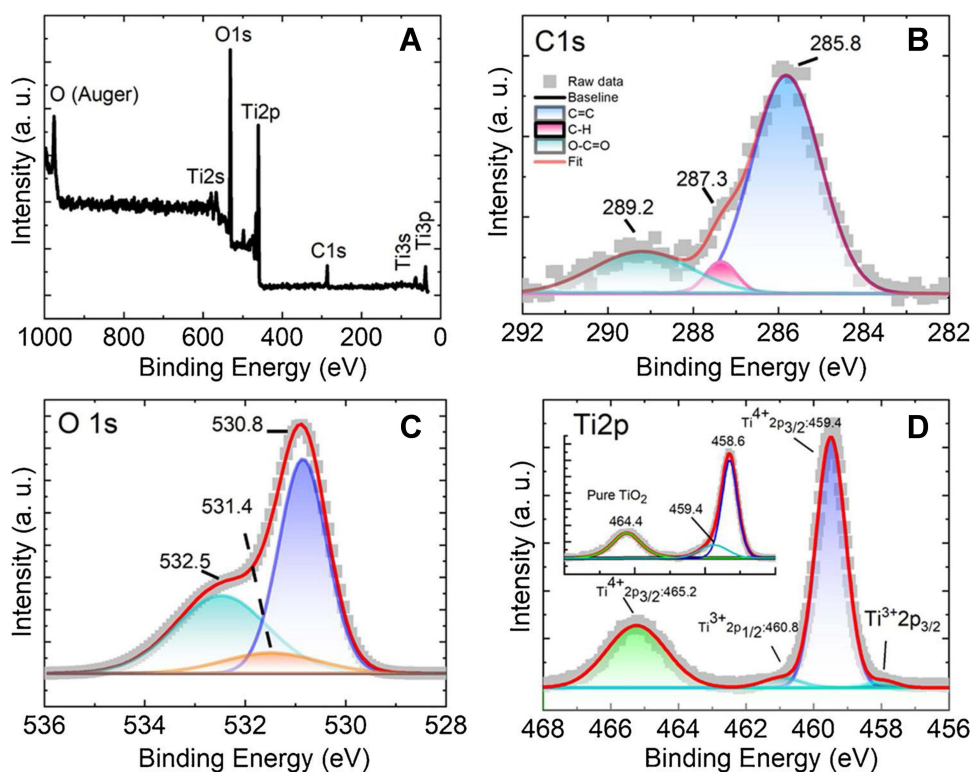


Figure 6 The XPS spectra of (A) survey, (B) C_{1s} , (C) O_{1s} and (D) Ti_{2p} . Reprinted from Mahmood A, Shi G, Wang Z, et al. *J Hazard Mater*, 401, Carbon quantum dots-TiO₂ nanocomposite as an efficient photocatalyst for the photodegradation of aromatic ring-containing mixed VOCs: an experimental and DFT studies of adsorption and electronic structure of the interface. 123402, Copyright 2021, with permission from Elsevier.¹¹³

are excited to emit and then measure the energy of photoelectrons.¹¹⁰ With the kinetic energy/binding energy of photoelectrons as the abscissa and the relative intensity as the ordinate, the photoelectron spectrum can be created to obtain relevant information on the CQDs.¹¹¹ As a modern analytical method, XPS has the following characteristics: the ability to analyse all elements except H and He, strong identification, the ability to observe chemical shifts, quantitative analysis and high sensitivity.¹¹² Figure 6A–C correspond to the survey, C_{1s} and O_{1s} XPS spectra of the obtained CQDs, respectively. As shown in Figure 6D, two peaks at 459.4 eV and 465.2 eV corresponded to the $Ti^{4+}2p_{3/2}$ core level, while the peaks at 457.8 eV and 460.9 eV belonged to $Ti^{3+}2p_{1/2}$. XPS results demonstrated that CQDs were mainly composed of three elements, including carbon, oxygen and titanium.¹¹³

Interestingly, some researchers prepared green CQDs with food as raw material, which fully demonstrated the high biocompatibility of CQDs.^{114,115} Nevertheless, although the biocompatibility of CQDs has been reported in many studies, some researchers have found that there is a “threshold” for the low toxicity of CQDs. Liu et al reported the CQDs prepared at high temperature by mixing

folic acid and $CuCl_2$ with anhydrous ethanol.¹¹⁶ When the CQDs were mixed at different concentrations and added to cells, the cell viability was no less than 90% before 400 $\mu\text{g/mL}$ but gradually decreased from 600 $\mu\text{g/mL}$. Therefore, it can be inferred that the toxicity and side effects of CQDs on cells are related to the type of cells co-cultured, the concentration of CQDs and the co-cultivation time; these results are conducive to the subsequent exploration of the biocompatibility of CQDs and the design of their application in antitumour projects. It is expected that the toxicity assessment of therapeutic CQDs with potential biomedical applications will be carried out through animal models that conform to the norms of international organizations in the future.

Applications of CQDs in Antitumour Diagnosis

At present, clinical cancer is often found in the middle or even late stage, making later treatment much more difficult. When it is not discovered until the cancer develops to the late stage, the cancer is difficult to handle and shortens the life

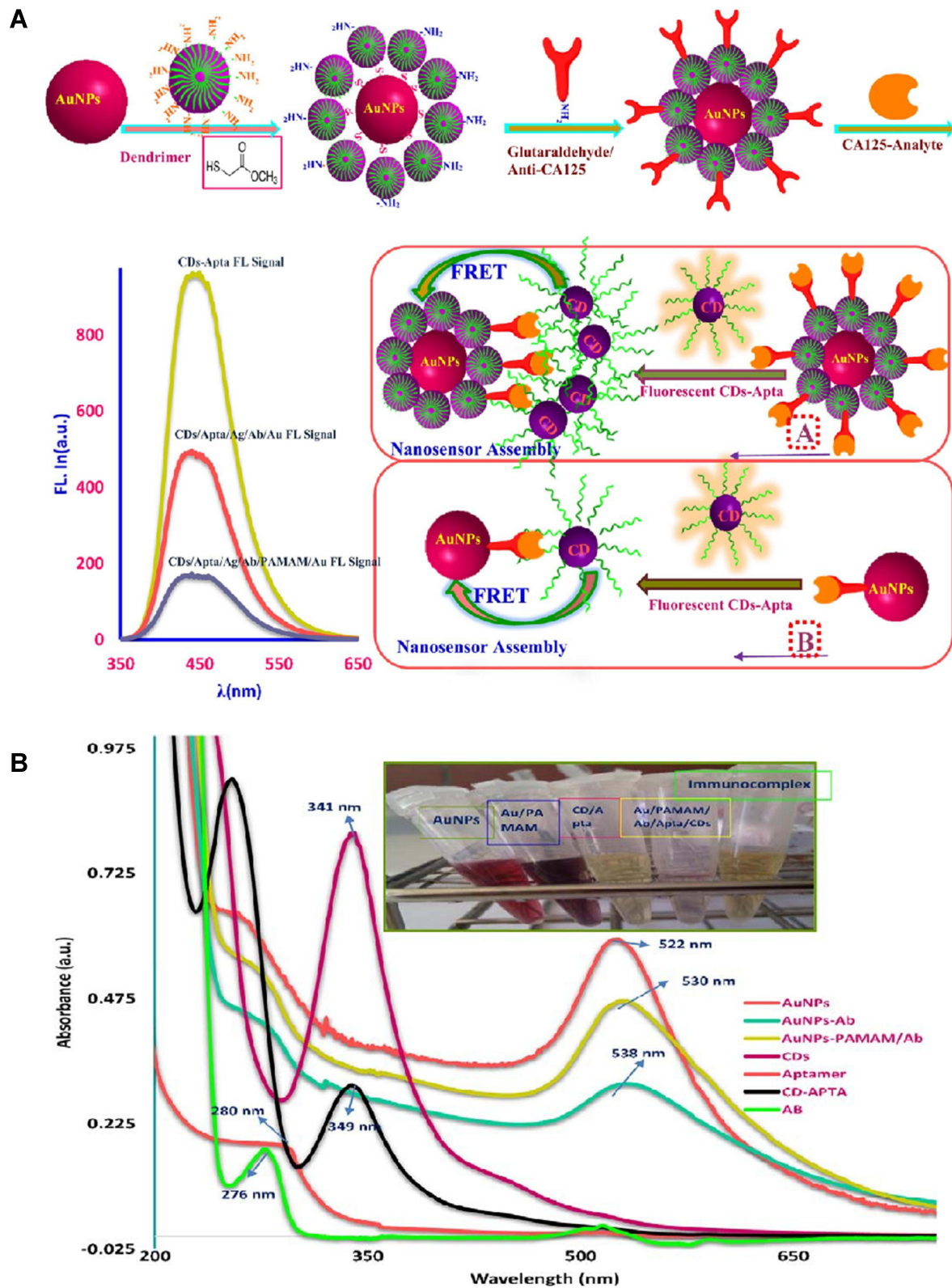


Figure 7 (A) The action mechanism diagram and **(B)** UV-vis absorption spectrum of AuNPs-PAMAM/Ab. Reprinted from Hamd-Ghadareh S, Salimi A, Fathi F, Bahrami S. Biosens Bioelectron, 96, An amplified comparative fluorescence resonance energy transfer immunosensing of CA125 tumor marker and ovarian cancer cells using green and economic carbon dots for bio-applications in labeling, imaging and sensing. 308–316, Copyright 2017, with permission from Elsevier.¹³⁵
Abbreviation: FRET, fluorescence resonance energy transfer.

span.¹¹⁷ The treatment of advanced cancer can only rely on drugs,¹¹⁸ chemotherapy¹¹⁹ or radiotherapy¹²⁰ to kill other free tumour cells, but these methods cannot completely kill all tumour cells in vivo. In addition, advanced cancer easily metastasizes and spreads, leading to the invasion of tumour cells in other normal parts of the body, which makes it impossible to control the tumours simply by removing the original area.¹²¹ Therefore, the development of tumour cell probes for early cancer screening is vital,¹²² and CQDs have become candidates for probes of tumour cells due to their excellent fluorescence properties and high sensitivity.

Because they are fluorescent, CQDs become excited after receiving electromagnetic radiation, thus emitting the same or different emission spectrum as the excitation spectrum and showing fluorescence.¹²³ Due to this characteristic, CQDs have been widely used in fluorescence imaging and sensing.^{124–126} When two fluorescent groups meet, if the emission spectrum of one fluorescent group is consistent with the excitation spectrum of the other, the fluorescence signal of the former weakens or even disappears, which is called fluorescence resonance energy transfer (FRET).¹²⁷ The conditions for effective FRET between the donor and the acceptor are specific.¹²⁸ First, the emission spectrum of the donor and the absorption spectrum of the acceptor must overlap.¹²⁹ Second, FRET requires that the donor and the energy acceptor be close enough, generally 7–10 nm apart, and the FRET decreases significantly with distance.¹³⁰ Finally, the fluorescent chromophores of the donor and the acceptor must be arranged correctly.¹³¹ Due to this principle, many researchers have combined FRET with CQDs for biological

analysis.^{132–134} Using CQDs as donors and AuNPs as acceptors, Somayeh et al assembled a FRET-based sensor that could detect CA125 on the surface of ovarian cancer cells. Due to the $-NH_2$ groups, anti-CA125 could be modified on the surface of AuNPs.¹³⁵ The sandwich structure of the AuNPs-CA125-CDs could be formed when the CA125-trapped AuNPs met the DNA-coated CQDs, which promoted the occurrence of FRET and led to the fluorescence quenching of the CQDs. As shown in Figure 7A, by $-S-$ wrapping PAMAM-Dendrimer with $-NH_2$ on the surface of AuNPs, the $-NH_2$ on the surface of AuNPs could be amplified, which made the fluorescence quenching signal of CQDs more intense, thus greatly improving the sensitivity of the device to CA125. Moreover, the UV-Vis absorption spectrum showed that the absorbance of AuNPs-PAMAM/Ab was higher than that of AuNPs-Ab at any wavelength, which implied that the adjunction of PAMAM-Dendrimer could significantly enhance the fluorescence quenching signal (Figure 7B).

In recent years, as a combination of chemiluminescence and electrochemical methods, electrochemiluminescence (ECL) has attracted great interest due to its high sensitivity and selectivity.¹³⁶ ECL is an electricity generation phenomenon in which certain substances are generated by electrochemical methods, and then these substances or biomass and other substances react further.¹³⁷ It retains the advantages of chemiluminescence methods, such as high sensitivity, wide linear range and convenient observation, and has many incomparable advantages, including good reproducibility, easy control and partial repeatability.¹³⁸ ECL not only greatly promotes biochemistry and molecular biology research but

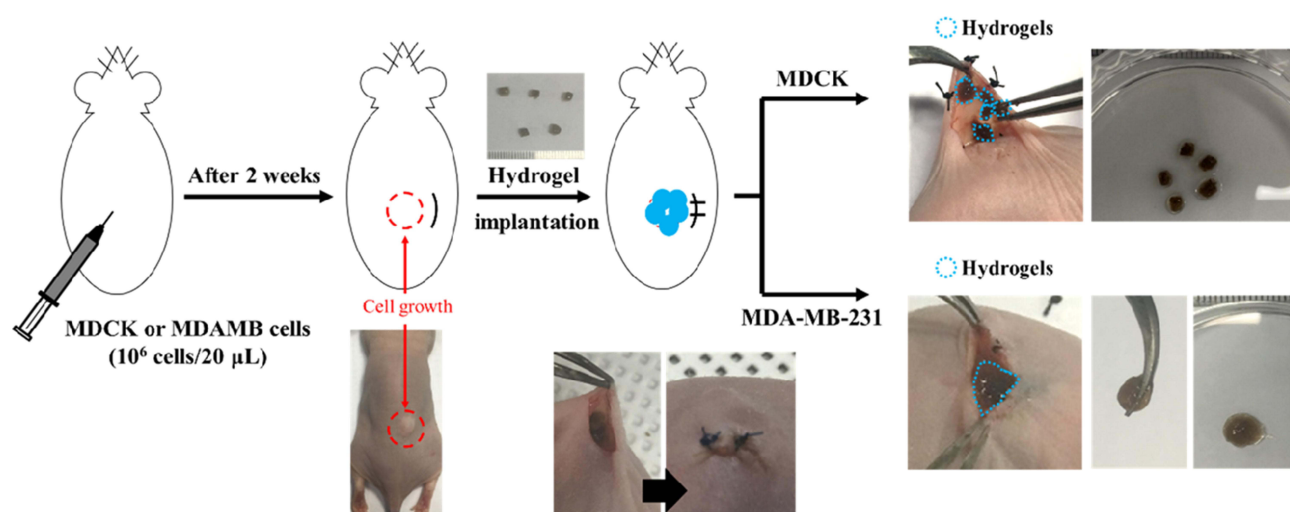


Figure 8 Photographic images of Gel-UPy/dsCQD hydrogel under normal and tumour environment. Reprinted with permission from Won HJ, Ryplida B, Kim SG, et al. Diselenide-bridged 1710 carbon-dot-mediated self-healing, conductive, and adhesive wireless hydrogel sensors for label-free breast cancer detection. *ACS Nano*. 2020;14(7):8409–8420. Copyright (2020) American Chemical Society.¹⁴⁵

also brings a technological revolution in clinical diagnosis, in which ECL probes play an important role.¹³⁹ Although CQDs have been developed as ECL probes,^{140,141} they are not widely used because of their low ECL yield. Therefore, the modification of CQDs to amplify the ECL signal is conducive to the application of ECL technology in biological analysis. Qiu et al designed an ECL system by combining CQDs with magnetic beads.¹⁴² MCF-7 cells and MDA-MB-231 cells were captured on an ITO electrode by folic acid, and then HA-solid-state ZnCQDs could bind to CD44 on the surface of tumour cells through HA, resulting in an enhanced ECL effect. It can be utilized for monitoring tumour cells and evaluating the heterogeneity of CD44 expression, which is conducive to the early diagnosis and screening of breast

cancer. Moreover, the greater amounts of H₂O₂ secreted by tumour cells can be used in active metabolism to induce stronger ECL emission from NHCDs combined with Au NPs, which provides another method of cancer monitoring.¹⁴³ Nevertheless, without electrode plates, DNA or RNAs as aptamers that can identify HER2 on the surface of MCF-7 cells are applied to amplify the ECL signal through CQDs or synergistic metal-organic frameworks.¹⁴⁴

In addition to FRET and ECL, CQDs can also be utilized to monitor the tumour environment by structural modification, spatial conformation or properties. Won et al combined gelatine and ureidopyriminone (Upy) into hydrogels (Gel-UPy) by performing isocyanate-amine group reactions, followed by mixing CQDs into the Gel-UPy to prepare a self-

Table 2 A Summary of Applications for Antitumour CQDs in Diagnosis

Theory	Raw Materials	Model	Function	Reference
FRET	Citric acid, ethylenediamine	MCF-7 cells	Diagnosis of breast cancer	[148]
FRET	Carbon powders	OVCAR3 cells	Monitor the presence of tumour cells	[149]
FRET	Citric acid, ethylenediamine	MCF-7 and HeLa cells	Diagnose as well as killing of tumour cells	[150]
FRET	Glucose, PAAS	HeLa/HepG2 cells	Determining prognosis of cancer and investigation of cancer metastasis	[151]
ECL	Zinc acetate, PEI	MCF-7 and MDA-MB-231	Monitor the presence of tumour cells	[142]
ECL	Citric acid, DMF	Hela and MCF-7 cells	Monitor the presence of tumour cells	[143]
ECL	–	MCF-7 cells	Diagnosis of HER2 and HER2-overexpressed living tumour cells	[144]
ECL	Candle soot, nitric acid	Hela and MCF-7 cells	Highly sensitive and selective tumor cells detection, DNA and protein analysis	[152]
Coagulation and dissolution of hydrogels	DOPA-Br, selenium ions	MDA-MB-231 cells	Breast cancer detection	[145]
Character dependent fluorescence properties	Citric acid, ethylenediamine	–	Diagnosis of early lung cancer	[146]
Detect GSH	GSH, HAuCl ₄ , glucose	HIBEpiC cells	Monitor the presence of tumour cells	[153]
Targeting C6 glioma cells	D-glucose, L-aspartic acid	C6 glioma cells	Diagnostic, targeting and therapeutic of brain cancer cells	[154]
Change of emission wavelength	Alizarin carmine	SMMC-7721 cells	Distinguish between tumour cells and normal cells	[147]
Detect Carcinoembryonic antigen	Citric acid, urea	Liver cancer mice beared HepG2 cells	Detect tumour cells and drug delivery	[155]

Abbreviations: PAAS, poly(acrylate sodium); PEI, polyethylenimine; DMF, N, N-Dimethylformamidot estimable; DOPA, dihydroxyphenylalanine; GSH, glutathione.

healing Gel-UPy/dsCD hydrogel.¹⁴⁵ The self-healing properties of the Gel-UPy/dsCD hydrogel depended on GSH and H₂O₂, which are produced by tumour cells rather than normal cells. As shown in Figure 8, after planting 5 pieces of Gel-UPy/dsCQD hydrogel into the cell inoculation site of mice, one hour later, the Gel-UPy/dsCQD hydrogel of the MDCK group was still a single slice, while the MDA-MB-231 group converged into a whole gel structure. This result indicates that the Gel-UPy/dsCQD hydrogel can self-heal at the tumour site but not in normal cells, so it can be used to monitor tumour cells through its self-healing function. In addition, Kalytchuk et al prepared CQDs with ICPA (a molecular fluorophore from the family of pyridines) on the surface, which caused fluorescence quenching when the CQDs were cooled to a solid and returned to the liquid state when heated, recovering their fluorescence.¹⁴⁶ However, when the CQDs were in a cool solid-state, the addition of alcohol prevented fluorescence quenching. Therefore, they can be used to measure alcohol, which is conducive to the early diagnosis of lung cancer. In addition, CQDs can monitor tumours by using the fluorescence change of CQDs as the detection signal. Li et al fabricated CQDs that could produce 430 nm and 642 nm emission under 360 nm laser irradiation.¹⁴⁷ When they encountered GSH, they only produced 430 nm excitation light, which made their fluorescence change from blue green to dark blue. The GSH content in tumour sites is much greater than that in normal tissues, so we can distinguish tumour cells from normal cells by the change in fluorescence colour of CQDs.

In contrast to normal cells, tumour cells have a unique acidic environment, tumour markers and related genes; these factors can be considered in the design of new treatments. Table 2 summarizes the application of CQDs for monitoring tumours or antitumour drugs in related research. At present, the optical properties of CQDs are mostly used in tumour monitoring research, in which FRET and ECL are the most widely used, while other properties are rarely involved. However, whether the other properties of CQDs, such as changes in surface functional groups, size and properties, could be utilized to monitor tumours remains to be explored in the future. In addition to the early diagnosis of tumours and the release of antitumour drugs, monitoring the heterogeneity, mutation, metastasis and recurrence of tumours is a possible application of CQD probes.

Drug Delivery

Traditional antitumour drugs have low bioavailability due to their nontargeting and metabolic dynamics, and they can

damage normal cells.¹⁵⁶ Therefore, it is urgent to improve the bioavailability of antitumour drugs. The natural, ultra-small structure endows the CQDs with EPR, which actively causes them to gather in the tumour site.¹⁵⁷ Using CQDs as carriers to deliver antitumour drugs to the body not only solves the drug targeting problem but also increases the time of drug aggregation in the tumour site.¹⁵⁸

Doxorubicin (DOX), as a broad-spectrum antitumour drug, can inhibit the synthesis of RNA and DNA, inducing the death of tumour cells in various growth cycles.^{159–161} DOX can restrain DNA superhelix topoisomerase II by embedding and inhibiting the interaction between polymer biosynthesis and DNA, thus destroying the replication of the DNA strand.^{162,163} It can also act on chromatin with transcriptional activity, which leads to histone changes. In the clinic, DOX has been widely used in the treatment of various diseases, including breast cancer,¹⁶⁴ lung cancer,¹⁶⁵ ovarian cancer,¹⁶⁶ leukaemia,¹⁶⁷ and gastric cancer.¹⁶⁸ Therefore, loading CQDs with DOX and incorporating them into the body for tumour treatment has become possible. Nitrogen- and phosphorus-doped CQDs (PNHCs) loaded with DOX were prepared by spontaneously heating glucose to boiling without additional heating operation.¹⁶⁹ The tumour volume of tumour-bearing mice treated with PNHCs-DOX was larger than that of tumour-bearing mice treated with free DOX. Moreover, the weight of tumour-bearing mice treated with free DOX gradually decreased, while that of the mice treated with PNHCs-DOX showed an upward trend, which indicated that PNHCs-DOX could not only inhibit tumour growth but also improve the tumour targeting of DOX, exhibiting the advantage of the EPR of CQDs. In addition, compared to free DOX, DOX-loaded and transferrin-modified C-Dot-Trans-Dox showed better therapeutic effects for brain cancer.¹⁷⁰ Since both tumour cells and the blood brain barrier (BBB) have transferrin receptors, it is possible for C-Dot-Trans-Dox to cross the BBB and gather in brain tumour sites, which is promising for the treatment of brain tumour-related diseases in the future. Moreover, due to the nitrogen functional groups on the surface of the CSCNP-R-CQDs, they could enter not only the tumour cell nucleus but also the cancer stem cell nucleus, which provides new possibilities for tumour eradication.¹⁷¹ As a pH-sensitive drug, DOX is more soluble under acidic conditions than under neutral conditions. Tumour tissues and cells can provide an appropriate pH environment for DOX release, which also leads to the tumour targeting of DOX. With the rapid development of CQDs technology, researchers are committed to combining it with other chemical compounds, biomaterials and even technology to play

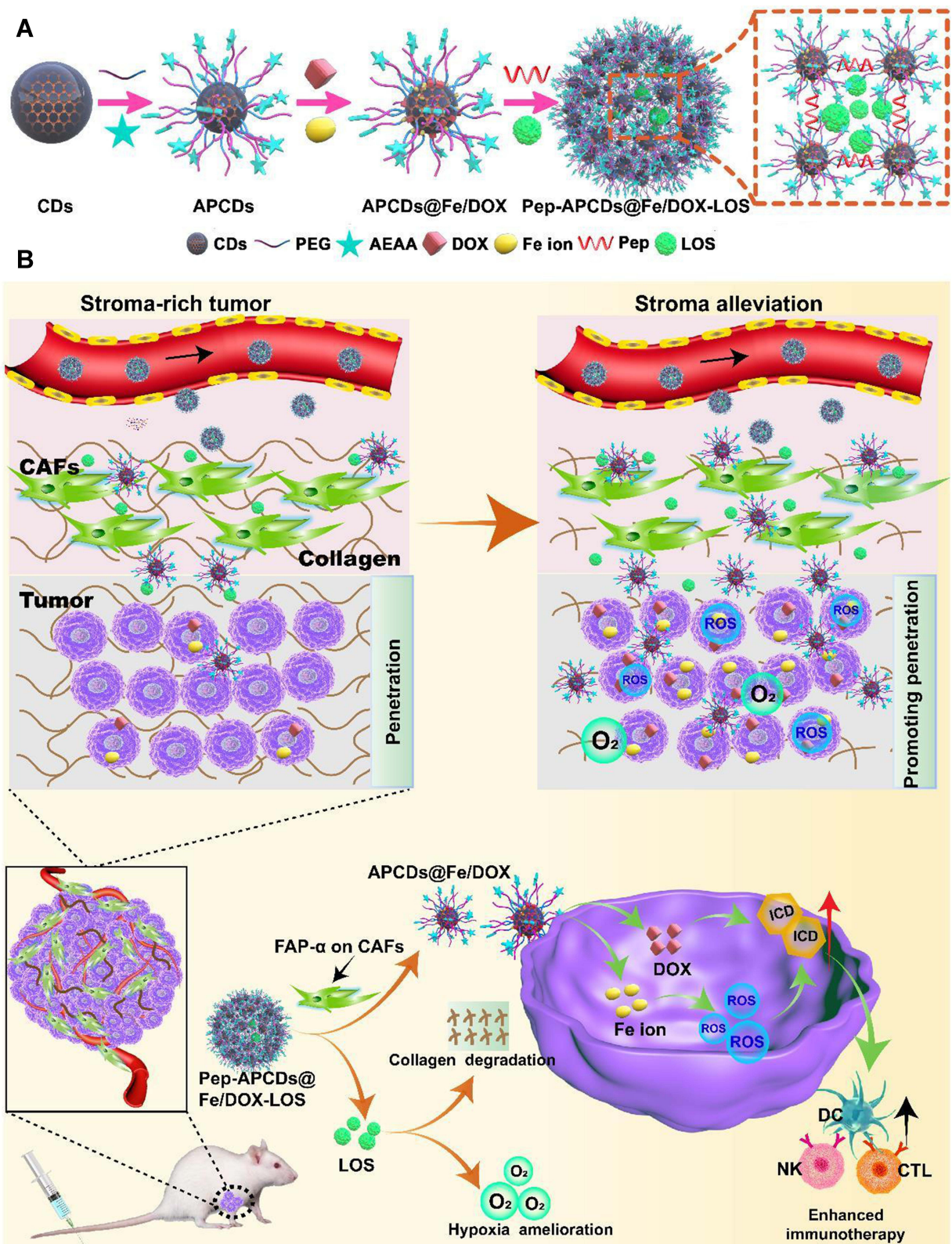


Figure 9 The (A) formation process and (B) mechanism of Pep-APCDs@Fe/DOX-LOS. Reprinted from Hou L, Chen D, Wang R, et al. Transformable honeycomb-like nanoassemblies of carbon dots for regulated multisite delivery and enhanced antitumor chemoimmunotherapy. *Angew Chem Int Ed Engl.* 2021;60(12):6581–6592. © 2020 Wiley-VCH GmbH.¹⁷²

Abbreviations: PEG, polyethylene glycol; AEAA, aminoethyl anisamide; Pep, peptides; LOS, losartan; CAFs, cancer-associated fibroblasts.

enhanced antitumour roles. Hou et al modified aminoethyl anisamide (AEAA) onto CQDs loaded with DOX and Fe ions to form APCDs@Fe/DOX. Then, APCDs@Fe/DOX were connected with each other by Asp-Ala-thl-gly-pro-Ala peptides (Pep) to form losartan (Los) wrapped in a mesoporous structure, thus finally forming Pep-APCDs@Fe/DOX-LOS.¹⁷² Pep was sensitive to the fibroblast-activating protein- α (FAP- α), while FAP- α was overexpressed in cancer-associated fibroblasts (CAFs). When Pep-APCDs@Fe/DOX-LOS flowed through the CAFs, Pep was degraded, while APCDs@Fe/DOX and Los were released. Los, as a modifier of the tumour microenvironment, could deplete the tumour matrix. Along with Fe ions and DOX, Los had the ability to accelerate the infiltration of T cells and NK cells, promote the release of proinflammatory cytokines, and down-regulate the recruitment of immunosuppressive cells (Figure 9). Kang et al coated silicon dioxide on the surface of a nanotube template and increased the temperature, forming CQDs on the surface of silicon dioxide.¹⁷³ The template simultaneously dissolved to form a hollow structure (C-hMOS), and then the C-hMOS was filled with DOX by soaking. Moreover, after heat treatment, the DOX-delivering C-hMOS showed more significant fluorescence intensity, which endowed them with the ability to observe the

distribution of DOX in vivo. In addition, Türk et al fabricated pH-sensitive hydrogels by self-crosslinking hydroxyapatite, NCQDs and DOX through Schiff bases, hydrogen bonds and ion interactions for the first time.¹⁷⁴ After reaching the tumour site, the Schiff base broke away and DOX was released.

Similar to other platinum drugs, oxaliplatin (II) has an antitumour mechanism that targets DNA, and platinum atoms cross-bind with DNA to prevent its replication and transcription.^{175–177} Nevertheless, due to drug resistance and side effects, it is possible to use Pt(IV) complexes instead of oxaliplatin (II).¹⁷⁸ Pt(IV) complexes are prodrugs of oxaliplatin(II) that can be transformed into oxaliplatin(II) by a reduction reaction, and tumour cells can provide an inherent reducing environment. Based on this principle, CQDs were prepared by modified thermal pyrolysis with citric acid as the carbon source and polyene polyamine as the passivator, following which a Pt(IV) complex (oxa(IV)-COOH) was combined with CQDs to form CD-Oxa by dehydration condensation between the amino carboxyl groups.¹⁷⁸ Under excitation light at 405 nm, 488 nm and 555 nm, CD-Oxa could produce bright polychromatic fluorescence, which was conducive to its effective monitoring of tumour cells (Figure 10A–E). Moreover, with increasing drug concentration, the viability

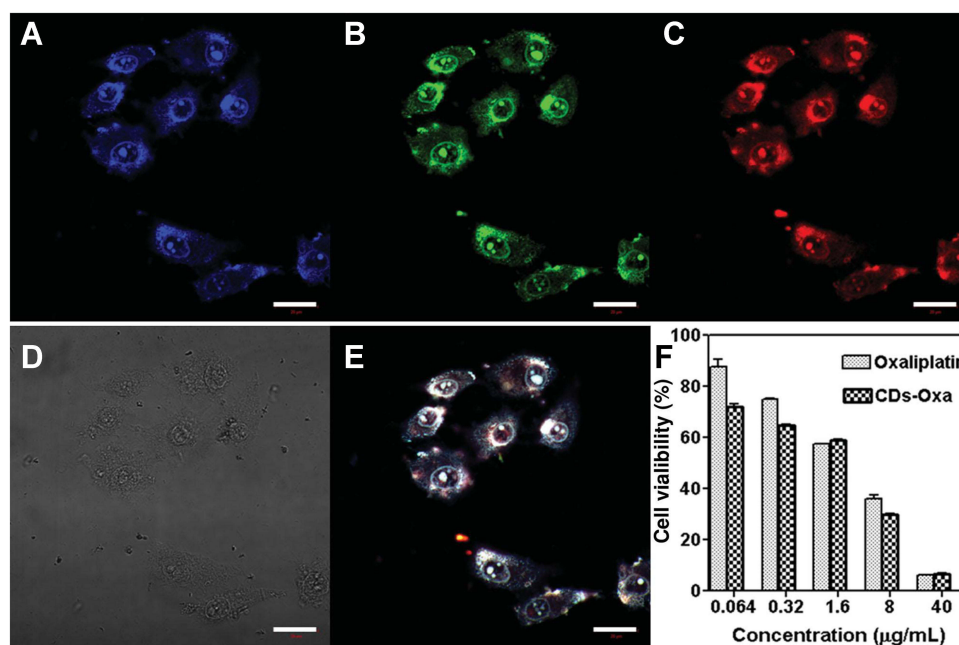


Figure 10 Confocal fluorescence images of HeLa cells treated with CD-Oxa under (A) 405 nm, (B) 488 nm, (C) 555 nm, (D) bright field and (E) overlay. (F) Cell viability of HepG2 cells treated with oxaliplatin (II) or CD-Oxa. Reprinted from Zheng M, Liu S, Li J, et al. Integrating oxaliplatin with highly luminescent carbon dots: an unprecedented theranostic agent for personalized medicine. *Adv Mater.* 2014;26(21):3554–3560. © 2020 Wiley-VCH GmbH.¹⁷⁸

Abbreviation: Oxa, oxaliplatin.

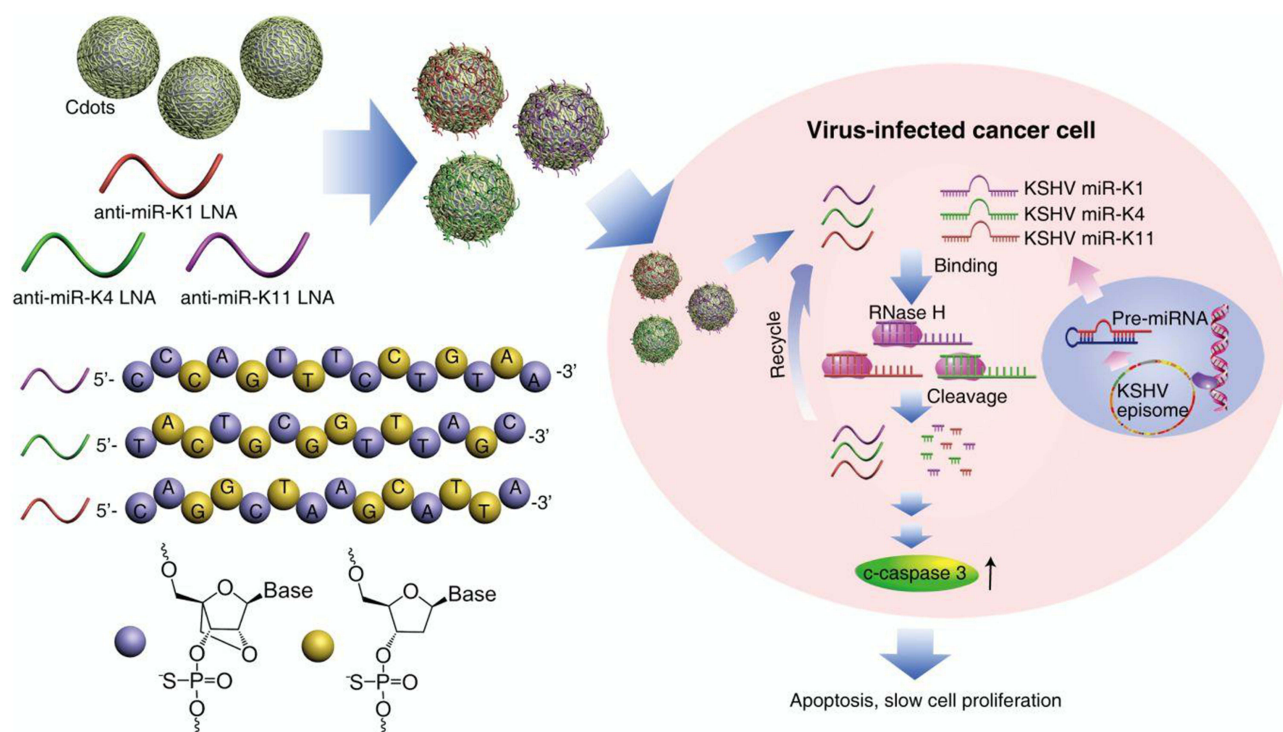


Figure 11 The schematic diagram of Cdots/LNA inducing apoptosis and slowing down proliferation. Reprinted with permission from Ju E, Li T, Liu Z, et al. Specific inhibition of viral MicroRNAs by carbon dots-mediated delivery of locked nucleic acids for therapy of virus-induced cancer. *ACS Nano*. 2020;14(1):476–487. Copyright (2020) American Chemical Society.¹⁸⁷

Abbreviations: LNA, locked nucleic acid; KSHV, Kaposi's sarcoma-associated herpesvirus.

of HepG2 human tumour cells decreased gradually, and the decrease in the CD-Oxa group was greater than that of the free oxaliplatin group, which indicated that the curative effect of CD-Oxa was better than that of free oxaliplatin (Figure 10F). In addition, Feng et al mounted PEG-(PAH/DMMA) on the surface of CQDs containing Pt(IV) (CDs-Pt(IV)). Among them, PEG-(PAH/DMMA) was equivalent to a “cap”, which was not hydrolysed under normal conditions but hydrolyzed under the acidic conditions of tumour tissues, causing CDs-Pt(IV) to enter the cells. Finally, the Pt(IV) prodrug was reduced to oxaliplatin(II), which binds to the nucleic acid of tumour cells.¹⁷⁹ Similarly, PEG-RGD was modified on CDs-Pt(IV), in which PEG and RGD were linked by benzimidazole bonds.¹⁸⁰ RGD could target the RGD receptor on the surface of tumour cells, but the RGD receptor was not specific to tumour cells. Therefore, PEG was designed to block RGD on the surface of CDs-Pt(IV). When CDs-RGD-Pt(IV)-PEG reached the tumour cells, the benzimidazole bond was hydrolysed under acidic conditions to expose RGD to achieve the targeting of tumour cells.

Gene therapy refers to the transfer of exogenous “repair genes” into target cells to treat related diseases at the gene level.^{181–183} The free gene is easily degraded by lysosomes after entering the cells, so it needs to be protected by the vector and then sent to the target cells to solve the problem.¹⁸⁴ Although single CQDs do not have the ability to carry genes, if gene carrying substances are modified on their surface, they can carry genes into target cells for gene therapy.^{185,186} Three kinds of LNA-based oligonucleotides (anti-miR-K1, anti-miR-K4, and anti-miR-K11) were loaded onto CQDs by PEI modification.¹⁸⁷ As shown in Figure 11, after entering lymphoma cells, Cdots/LNA could knock down miR-K1, miR-K4 and miR-K11 of Kaposi's sarcoma virus (KSHV), thus leading to apoptosis and growth inhibition of KSHV-positive primary active lymphoma cells through the caspase-3 pathway. Moreover, a novel DNA self-assembly nanostructure based on nitrogen-doped CQDs known as NPNCd was reported.¹⁸⁸ The conjugation of NPNCd with KRAS siRNA showed a superior gene knockdown effect, which provided inspiration for the clinical treatment of non-small-cell lung cancer.

In addition to DOX, oxaliplatin and “therapeutic” genes, other forms of antitumour drugs have also been widely

Table 3 A Summary of Applications for Antitumour CQDs in Drug Delivery

Drug	Raw Materials	Model	Administration	Function	Reference
DOX	Glucose, EDA, H ₃ PO ₄	Hepatoma mice beared HepG ₂ cells	I.V.	Nuclear targeting of tumor cells	[169]
DOX	Carbon nanopowder	SJGBM2 cells	–	Cross the BBB	[170]
DOX	4-amino-Phenol, potassium periodate	Cervical carcinoma mice beared Hela cells	I.V.	Enter the cancer stem cell nucleus	[171]
DOX	Tyrosine-derived melanin	Breast cancer mice beared 4T1 cells	I.V.	Synergetic Fenton reaction and immunotherapy	[172]
DOX	SiO ₂	Breast cancer mice beared MCF-7 cells	I.T.	pH response of DOX release	[173]
DOX	1,4,5,8-tetraminoanthraquinone, citric acid	Brain glioma mice beared U87 cells, cervical carcinoma mice beared Hela cells	I.V.	Mimicking large amino acids, cross the BBB	[189]
DOX	PEI, H ₂ O ₂ , anhydrous ethanol	A549 cells	–	Deliver nucleic acid and hydrophobic drug cargos	[193]
DOX	Graphite, sugarcane waste	Hela cells	–	Synergetic mesoporous silica nanoparticles	[194]
DOX	Glycerol, PEI	Liver cancer mice beared MHCC-97L cells	I.V.	Treatment synergetic monitoring	[195]
DOX	Gelatin	MDA-MB-231 cells	–	Through mitochondria enhanced apoptosis and autophagy pathway	[196]
DOX	Citric acid	MCF-7 cells	–	Overcoming drug resistance	[197]
DOX	Glucose, PEG	Breast cancer mice beared 4T1 cells	I.V.	Encapsulated into nanoparticles, detachable at tumor site.	[198]
Pt(IV)	Citric acid, polyene polyamine	Hepatoma mice beared H22 cells	I.T.	Multicolor imaging	[178]
Pt(IV)	Citric acid, diethylenetriamine	Cervical carcinoma mice beared UI4 cells	I.V.	PEG controlled drug release	[179]
Epirubicin and temozolomide	Carbon nano powders	U87 cells	–	Epirubicin, temozolomi and transferrin triple conjugated	[199]
LNA	PEI, citric acid	Primary active lymphoma mice beared BCBLI-Luc cells	I.P.	Inhibiting irus-induced cancer	[187]
siRNA	EDTA, citric acid	KRAS-mutated NSCLC cells	–	Induced DNA polymerization	[188]
TAAAs	Mannose, EDTA, phosphoric acid	Hepatoma mice beared HepaI–6 cells	I.T.	Synergetic immunotherapy, amplifying microwave ablation	[190]

(Continued)

Table 3 (Continued).

Drug	Raw Materials	Model	Administration	Function	Reference
AgNPs	Lemon slices	MCF 7 cells	–	Producing ROS for killing tumor cells	[191]
Amygdalin	Golden acid	Hep3B cells	–	Natural targeting tumor cells	[192]
Dextran	Acrylic acid and ethylenediamine	SKOV3 cells	–	Synergistic nanogels release drug in controlled manner	[200]

Abbreviations: EDA, 1,2-ethylenediamine; PEI, polyethylenimine; PEG, polyethylene glycol; EDTA, ethylenediamine tetraacetic acid; TAAs, tumor-associated antigens; ROS, reactive oxygen species.

researched. Large neutral amino acid transporter 1 (LAT1) is highly expressed in various tumour cells but is rarely expressed in normal cells. LAT1 transports amino acids into tumour cells by binding with amino and carboxyl groups on the surface of amino acids. Li et al utilized 1,4,5,8-tetraaminoanthraquinone (TAAQ) and citric acid as raw materials to prepare CQDs based on simulated large amino acids (LAAM TC-CQDs).¹⁸⁹ There were many paired amino and carboxyl groups on the surface of the LAAM TC-CQDs, which made the tumour cells mistakenly believe that they were amino acids; thus, the drug-loaded LAAM TC-CQDs could be transported into tumour cells through LAT1. It is exciting that LAT1 is expressed not only in tumour cells but also in the BBB, which suggests that LAAM TC-CQDs can be utilized to treat brain cancer across the BBB. In addition, microwave ablation (MWA) uses a special needle to puncture the lesion, transmits RF microwaves to the needle tip, generates a large quantity of heat at this position, and destroys the focus, thus treating the disease. Tumour cells killed by MWA can release tumour-associated antigens (TAAs). Some CQDs can capture TAAs, while some carbohydrate modifiers can target dendritic cells (DCs), and mannose-based CQDs (Man-CDs) combine the two.¹⁹⁰ Man-CDSs can deliver captured TAAs to DCs, activate the immune system and amplify MWA, thus skillfully combining drug delivery with immunotherapy. Silver nanoparticles are natural nanodrugs that kill tumour cells. Ghosal et al utilized fresh lemon slices as raw materials to prepare green CQDs by a hydrothermal method and then added solid silver nitrate to CQDs aqueous solution to synthesize CQDs loaded with silver nanoparticles (CQD@AgNPs) in situ.¹⁹¹ CQD@AgNPs kill tumour cells by producing ROS without any other external roles. Amygdalin, a natural chemotherapeutic drug, is nontoxic, but when it is metabolized by β -glucosidase, it produces toxic hydrocyanic acid, which can

be catalysed into nontoxic thiocyanate by thiocyanase. Kalaiyarasan et al used a gold acid as a raw material to prepare CQDs with many carboxylic acids on the surface and then functionalized them with almond protein by dehydration or esterification to form Amy@CQDs.¹⁹² Malignant cells contain more β -glucosidase than normal cells, while normal cells contain more thiocyanase than tumour cells, so Amy@CQDs can cleverly achieve selective killing of tumour cells after entering the body.

Table 3 summarizes the application of CQDs as drug carriers in antitumour research. To achieve good clinical antitumour effects, the selection of CQDs and drugs is very important. The choice of antitumour drugs can be made according to the different manifestations of drugs on normal tissues and tumour tissues. Tumour tissues have more acidic substances, H_2O_2 , glutathione and tumour markers than normal tissues. If the antitumour drugs are nucleic acids, it is necessary to prepare CQDs with gene carrying materials such as PEI and chitosan or modify them on the surface of CQDs. Moreover, encapsulation of CQDs in macromolecules such as nanoparticles and hydrogels is also a possible strategy. In addition, CQDs can be reformed by doping, grafting and modification so that they can combine with antitumour drugs to target tumour sites to expand the therapeutic effect against cancer. In summary, the primary purpose of these methods is to improve the targeting of drug delivery and prolong the residence time of drugs in tumour sites, thus achieving better antitumour efficacy than that of free drugs.

Monitoring the Release of Antitumour Drugs

Antitumour drugs need to be released in time after reaching the tumour sites to exert antitumour effects, and CQDs can monitor their release in real time. P-CQDs were CQDs

covered with PEI on the surface, which could emit blue fluorescence under the activation of the excitation signal under normal circumstances.²⁰¹ When hyaluronic acid-conjugated doxorubicin (HA-Dox) was coated on the surface of P-CQDs, the fluorescence of P-CQDs was quenched by FRET, in which P-CQDs and HA-Dox were the donor and the acceptor, respectively. When P-CQDs/HA-Dox reached

and entered tumour cells, the HA on the surface of P-CQDs/HA-Dox was degraded into small fragments by intracellular hyaluronidase; thus, DOX was released, and the P-CQDs were exposed. Additionally, the fluorescence recovery of P-CQDs could be used to monitor drug release. Similarly, under the effect of FRET, CQDs-Pt(IV) emitted blue, green and red fluorescence, but the blue fluorescence was limited

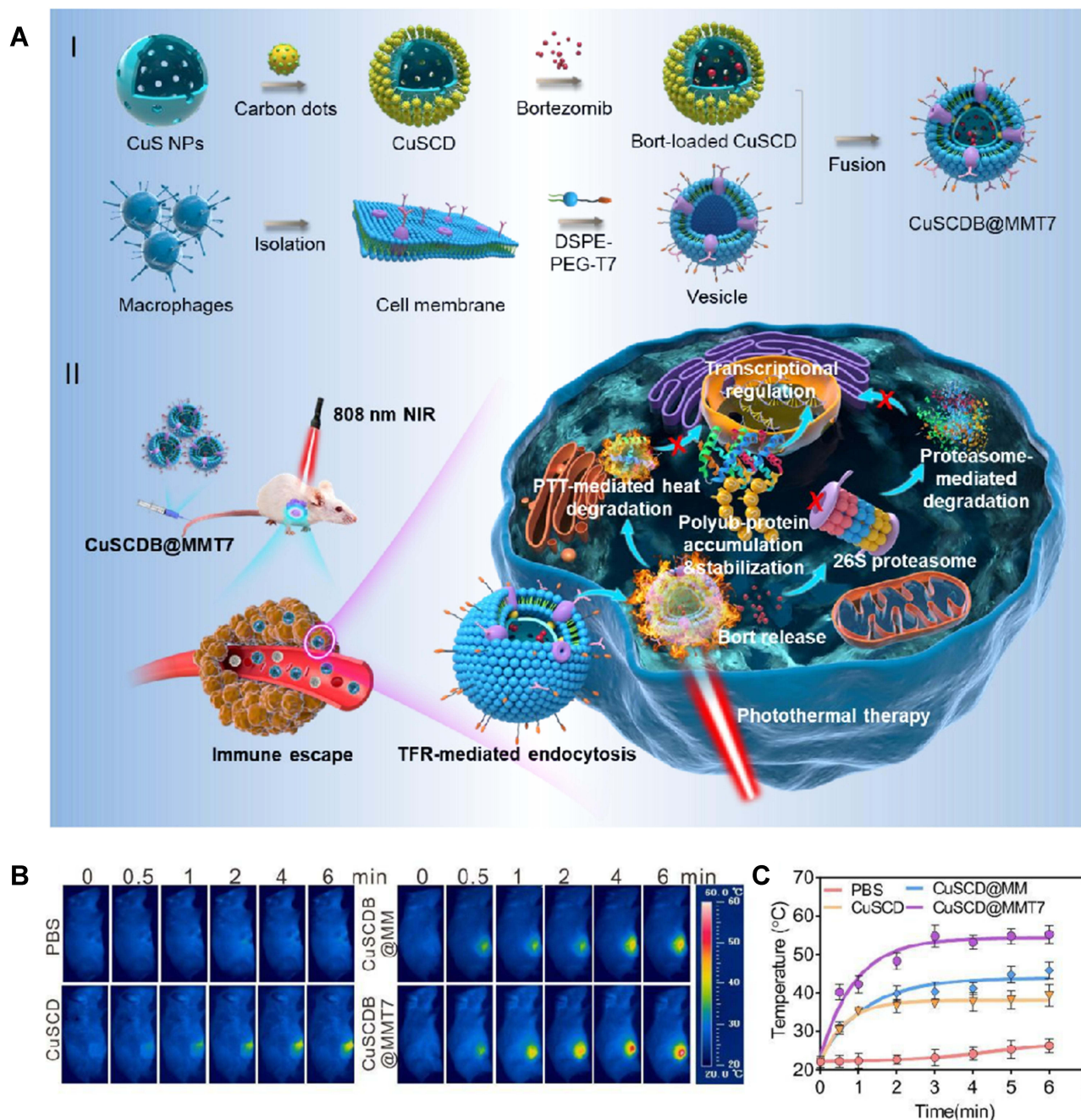


Figure 12 (A) Preparation and mechanism of CuSCDB@MMT7, **(B)** thermography images and **(C)** temperature curves of breast cancer mice bearing 4T1 cells treated with PBS, CuSCD, CuSCDB and CuSCDB@MMT7. Reprinted with permission from Yu Y, Song M, Chen C, et al. Bortezomib-encapsulated CuS/carbon dot nanocomposites for enhanced photothermal therapy via stabilization of polyubiquitinated substrates in the proteasomal degradation pathway. *ACS Nano*. 2020;14(8):10688–10703. Copyright (2020) American Chemical Society.²¹⁵

Abbreviations: DSPE, distearoyl phosphoethanolamine; PEG, polyethylene glycol.

by the quenching agent dabsyl.²⁰² Under the condition of an intracellular reduction reaction, the three substances disintegrated and released Pt(IV), and the blue fluorescence of the CQDs was restored, which was used to monitor the activation of antitumour prodrugs in a complex biological micro-environment. Li et al encapsulated the CQDs and DOX into mesoporous silica nanoparticles to form DOX-CQDs/PNVCL@MSNs.²⁰³ PNVCL was folded on the surface of the DOX-CQD/PNVCL@MSNs by hydrophobic phase transition, which blocked the orifices and hindered DOX release and concealed the fluorescence of the CQDs. Due to the acidic conditions of the tumour, the Schiff base linkage between MSNs and CQDs/PNVCL was broken, and then DOX and CQDs were released, so the release of antitumour drugs can be monitored by the fluorescence of CQDs.

Photothermal Therapy

In recent years, photothermal therapy (PTT) has attracted extensive research, which aims to use a photothermal conversion agent to absorb near-infrared light (650–900 nm) and convert the absorbed light energy into heat energy to further achieve the deep penetration of tissues and the minimum heating of nontarget tissues.^{204–206} Therefore, the choice of photothermal agent is the key factor in determining the success of photothermal therapy. First, to obtain effective treatment results, near-infrared photothermal agent-mediated tumour treatment usually requires a near-infrared laser irradiation power density significantly less than the maximum allowable skin exposure.²⁰⁷ Second, photothermal agents are applied for photothermal treatment of the tumour, and they should have good biocompatibility with surrounding tissues.²⁰⁸ Third, only when the size of the photothermal agent is small enough can it enter the tissue cells for photothermal therapy, so it must have superior tissue targeting.²⁰⁹ At present, widely used photothermal materials include noble metal nanoparticles such as Au, Ag and Pt,^{210,211} carbon materials such as graphene and carbon nanorods,²¹² metal and nonmetal compounds such as CuS and ZnS,²¹³ and organic dyes such as indocyanine green and Prussian blue,²¹⁴ while few studies have mentioned CQDs.

CuS nanoparticles carrying CQDs and bortezomib were synthesized, coated with macrophage membranes and modified with the T7 peptide to form CuSCDB@MMT7.²¹⁵ After CuSCDB@MMT7 enters the body, the immune system does not attack them because they are protected by the macrophage membrane, but instead, it transports them to tumour cells through the T7 peptide. CQDs are used for photothermal ablation under 808 nm NIR, while bortezomib is a proteasome inhibitor

that can protect tumour suppressor proteins from degradation, thus uniting in opposition to tumours (Figure 12A). As shown in Figures 12B and C, the photothermal conversion efficiency of the composites increased with increasing irradiation time. Compared to the PBS, CuSCD and CuSCD@MM groups, the CuSCD@MMT7 group could reach 50 °C in 3 minutes, indicating its superior photothermal conversion rate, thus demonstrating the feasibility of thermal ablation therapy for tumour cells. Wang et al prepared a self-crosslinked chitosan hydrogel (CCHN) coated with DOX and CQDs with an average particle size of 65 nm to endow injectability.²¹⁶ When CCHNs reached the tumour site, chitosan hydrolysed and released CQDs and DOX. CQDs kill tumour cells through PTT; however, DOX is an antitumour drug, so this system achieves a dual antitumour effect of PTT synergistic drug delivery. In addition, Qian et al developed a CQDs-based mesoporous silica scaffold (CQD@MSN) for photothermal synergistic immunotherapy.²¹⁷ CQD@MSNs targeted tumour sites and participated in photothermal ablation, while biodegradable CQD@MSN fragments could obtain TAAs from photothermally lethal tumour cells and then carry them away from necrotic tissue, initiating the immune response by stimulating the proliferation and activation of NK cells and macrophages.

Photodynamic Therapy

Photodynamic therapy (PDT) is a new method for treating tumour diseases with photosensitive drugs and laser activation, whose mechanism of action is to produce ROS by the interaction between a photosensitizer and oxygen to kill tumour cells.^{218–220} The photosensitizer absorbed by the tissue is stimulated by laser irradiation of a specific wavelength, and the excited photosensitizer transfers energy to the surrounding oxygen to generate highly active singlet oxygen.²²¹ Singlet oxygen reacts with adjacent biological macromolecules to induce cytotoxicity, which leads to cell damage and even death.²²² In the treatment of oesophageal cancer,²²³ lung cancer,²²⁴ skin cancer,²²⁵ breast cancer²²⁶ and other diseases, PDT is a new tumour treatment technology that has risen and developed in recent years. As a cold photochemical reaction, the basic elements of PDT are oxygen, photosensitizer and visible light (commonly applied with a laser). Only by choosing an appropriate photosensitizer can PDT achieve the purpose of killing tumours. First, photosensitizers have a certain selectivity and affinity for tumour tissues and remain in tumour sites for a long time so that the

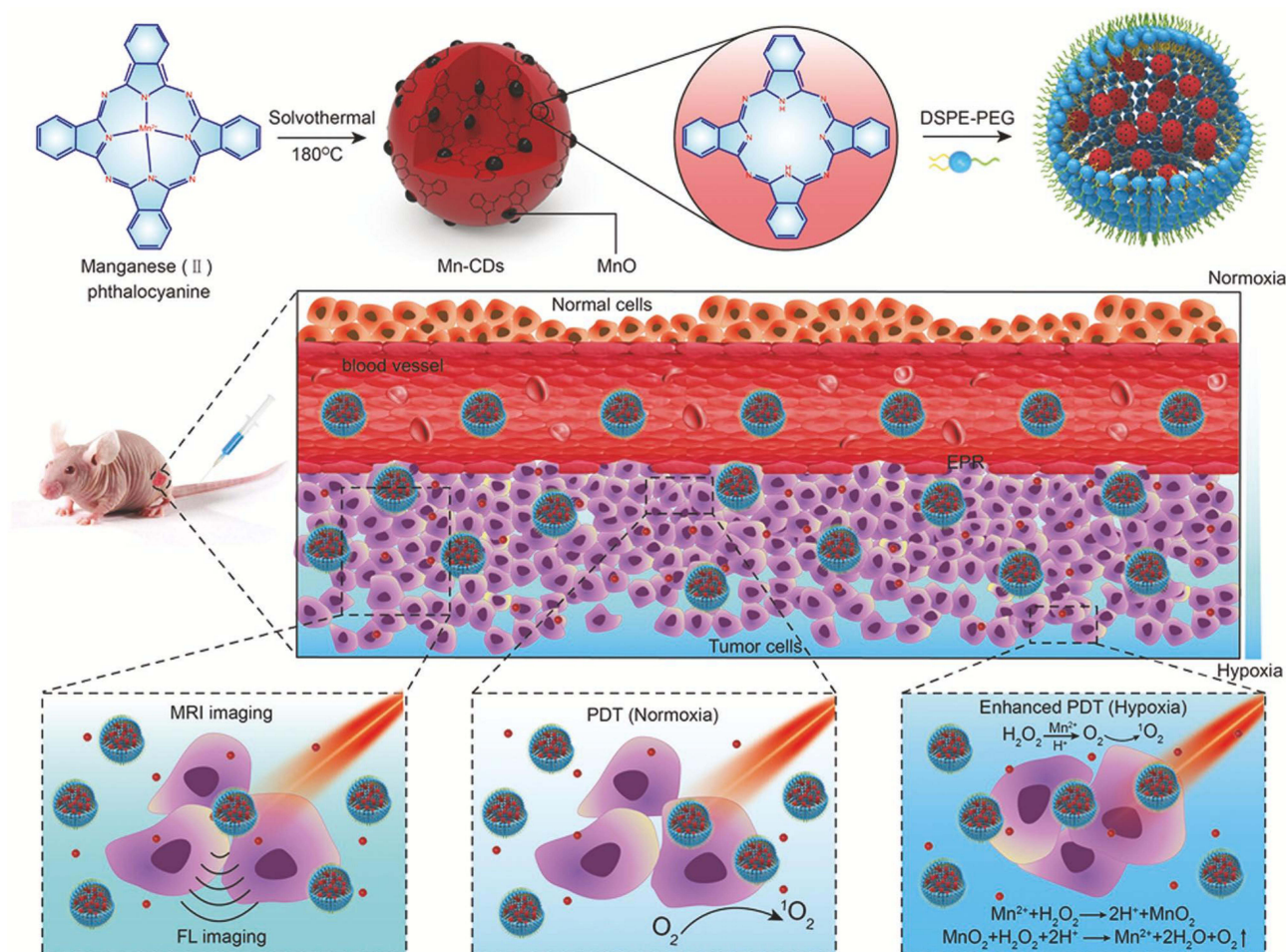


Figure 13 Preparation and mechanism of Mn-CDs. Reprinted from Jia Q, Ge J, Liu W, et al. A magneto fluorescent carbon dot assembly as an acidic H₂O₂-driven oxygenerator to regulate tumor hypoxia for simultaneous bimodal imaging and enhanced photodynamic therapy. *Adv Mater.* 2018;30(13):e1706090. © 2018 WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim.²³⁰

Abbreviations: DSPE, distearoyl phosphoethanolamine; PEG, polyethylene glycol.

concentration difference between tumour sites and normal tissues is maximized.²²⁷ Second, a photosensitizer can produce singlet oxygen in tumour tissue by illumination.²²⁸ Last, the photosensitizer is excited by light of an appropriate wavelength.²²⁹

Jia et al utilized manganese phthalocyanine (Mn-PC) as a raw material to prepare hydrophobic magnetic fluorescent CQDs (Mn-CDs) by a high-temperature method, followed by inserting PEG on the surface to improve their hydrophilicity.²³⁰ Mn²⁺ in Mn-CDs could be used as a catalyst to decompose a large amount of H₂O₂ at the tumour sites into oxygen, and ¹O₂ could be converted into ROS under irradiation with NIR light (635 nm), killing tumour cells (Figure 13). The lack of O₂ in tumour tissues limits the efficacy of PDT in antitumour therapy. Therefore, increasing O₂ content in tumour sites becomes the primary premise of PDT phototherapy. In addition,

although C₃N₄, a water splitting material, can decompose water into O₂ by PDT, the decomposition efficiency is not high. To solve this problem, Zheng et al embedded CQDs and protoporphyrin IV (PpIV) on C₃N₄ to form PCCN.²³¹ Among them, CQDs were used to enhance the water decomposition of C₃N₄, thereby increasing the production of O₂, followed by transforming O₂ into ROS by PpIV under light and killing the tumour cells. Nevertheless, traditional PDT applies ultraviolet and visible light as the excitation light source, which cannot penetrate the tissues sufficiently, so PDT can only be used for shallow tumours. It is not effective for deep tumours, which greatly limits the scope of PDT in clinical applications. At present, there are studies that combine upconversion nanophosphors (UCNPs) with PDT; such methods use low-energy near-infrared light as the excitation light source and upconvert

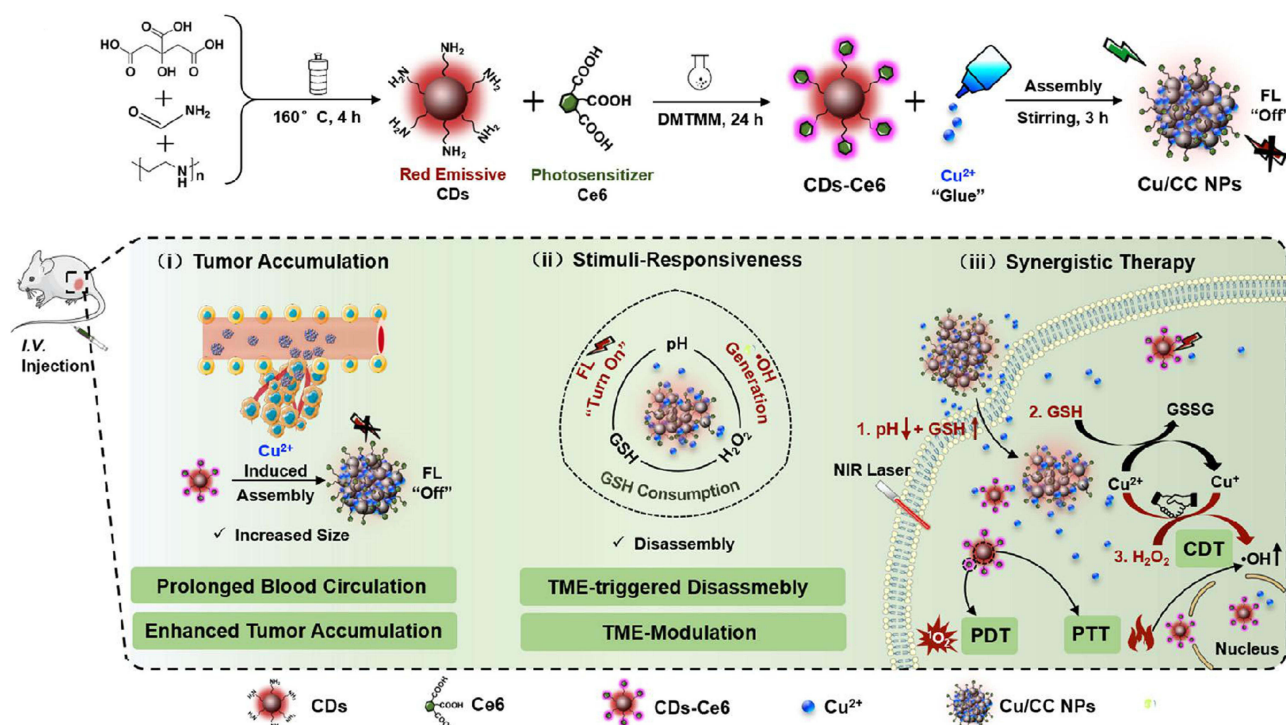


Figure 14 Preparation and mechanism of Cu/CC NPs. Reprinted from Sun S, Chen Q, Tang Z, et al. Tumor microenvironment stimuli-responsive fluorescence imaging and synergistic cancer therapy by Carbon-Dot-Cu²⁺ nanoassemblies. *Angew Chem Int Ed Engl.* 2020;59(47):21041–21048. © 2020 Wiley-VCH GmbH.²³⁹

Abbreviations: Ce6, Chlorin e6; CDs-Ce6, Carbon quantum dots modified with Ce6; Cu/CC NPs, Nanoparticles loaded with Cu²⁺ and CDs-Ce6; DMTMM, 4-(4,6-Dimethoxy[1,3,5]triazin-2-yl)-4-methylmorpholinium chloride hydrate.

it into high-energy light to achieve the required spectrum of PDT photosensitizers.^{232,233} Near-infrared light has a deeper light transmission depth, which allows PDT to be utilized for deeper tissue treatment. Chan et al combined UCNP with CQDs for the treatment of tumours through PDT.²³⁴ Among them, UCNP were used as light converters to transform 808 nm near-infrared light into ultraviolet light for CQDs to produce ROS and then kill tumour cells.

Photothermal Therapy and Synergetic Photodynamic Therapy

PTT or PDT alone cannot completely eliminate tumours, so the combination of these two methods improves the curative effects.^{235–237} Based on the upconversion luminescence (UCL) principle described above, the combination of PDT and PTT for tumour phototherapy provides a new opportunity to apply CQDs in tumour treatment. Using ZnPc as a photosensitizer and CQDs as thermal sensitizers, Lv et al prepared a nanomaterial with CQDs and UCNP, in which the CQDs could gradually heat and ablate tumour cells while the UCNP could transfer energy to ZnPc to generate ROS to

kill the tumour cells under 980 nm near-infrared light irradiation; this method was expected to yield twice the result with half the effort.²³⁸ The CQDs grafted with the photosensitizer Ce6 were mixed with Cu²⁺ to form nanoparticles (Cu/CC NPs).²³⁹ As shown in Figure 14, Under the low pH, glutathione (GSH) overexpression and H₂O₂ surplus of the tumour environment, the Cu/CC NPs disintegrated and released CDs-Ce6 and Cu²⁺. GSH can eliminate ROS, which seriously hinders the PDT pathway of killing tumour cells. Therefore, the addition of Cu²⁺ can initiate Fenton-like reactions to eliminate H₂O₂ and produce O₂ for PDT while also removing GSH and strengthening PDT. Moreover, due to FRET, the assembled Cu/CC NPs hide the fluorescence of the CQDs, but they disintegrate and release CQDs when entering tumour cells, thus exposing the fluorescence of the CQDs and providing the possibility for monitoring their entrance into tumour cells. Hua et al grafted HPPH into CQDs doped with Cu²⁺ and Gd³⁺ to form BCCGH.²⁴⁰ CQDs, as thermal sensitizers, enabled PTT under 808 nm laser irradiation, while HPPH was a photosensitizer that produced ROS to kill tumour cells under 671 nm laser irradiation. In addition, Zhang et al combined PTT, PDT,

Table 4 A Summary of Applications for Antitumour CQDs in Phototherapy

Phototherapy	Raw Materials	Model	Administration	Thermal Sensitizer / Photosensitizer	Therapeutic Efficiency	Function	Reference
PTT	Saccharomyces, ethylenediamine	Breast cancer mice beared 4T1 cells	I.V.	CQDs	39.7%	Synergetic protease system, the encapsulation of macrophage membrane prevents nanoparticles from being attacked by the immune system	[215]
PTT	D(t)-Glucose, EDTA	Breast cancer mice beared 4T1 cells	I.V.	CQDs	57.4%	Synergetic chitosan hydrogel, CDs and DOX were released by hydrolysis of chitosan	[216]
PTT	Purchased from Xianfeng Nano (China)	Breast cancer mice beared 4T1 cells	I.V.	CQDs	–	Synergetic immunotherapy	[217]
PTT	Citric acid, glycine	Osteosarcoma mice beared UMR-106 cells	I.T.	CQDs	–	Synergetic bone regeneration and bacterial eradication	[242]
PTT	TNP, BPEI ₁₈₀₀	Breast cancer mice beared 4T1 cells	I.V.	CQDs	–	Enhancement of photothermal conversion efficiency by doping MoS ₂	[243]
PTT	TNP, BPEI ₁₈₀₀	Breast cancer mice beared 4T1 cells	I.V.	CQDs	28.4%	Enhancement of photothermal conversion efficiency by doping black phosphorus	[244]
PTT	Watermelon juice	Cervical carcinoma mice beared Hela cells	I.V.	CQDs	–	Killing tumor cells by photothermal ablation	[245]
PTT	Hyaluronic acid, citric acid, Ethylenediamine	Breast cancer mice beared 4T1 cells	I.V.	CQDs	27.4%	Synergetic drug delivery	[246]
PDT	L-cysteine, m-phenylenediamine	Cervical carcinoma mice beared U14 cells	I.V.	Protoporphyrin IX (PpIX)	–	Targeting tumour nuclei	[35]
PDT	Chitosan, ethylenediamine, mercaptosuccinic acid	MCF-7 cells	–	Rose bengal (RB)	–	Natural targeting mitochondria without additional modification of mitochondrial targeting groups	[247]

PDT	Manganese(II) phthalocyanine	Breast cancer mice beared 4T1 cells	I.V.		Mn-CDs	-	Mn ²⁺ promoted Fenton-like reaction	[230]
PDT	Citric acid, urea	Breast cancer mice beared 4T1 cells	I.V.		Protoporphyrin IX	-	C ₃ N ₄ products O ₂ for amplifying PDT	[231]
PDT	Lychee exocarp	Liver cancer mice beared Bel-7404 cells	I.V.		Ce6	-	Detect tumour cells	[247]
PDT	Citric acid monohydrate, L-cysteine	Hela cells	-		TMPyP	-	FRET mediated PDT production	[248]
PDT	Urea, sodium citrate	Oral carcinoma mice beared CAL 27 cell	I.T.		CNQDs	25%	Utilizing 808 nm near-infrared light to generate UV light band for realizing PDT	[234]
PTT&PDT	Citric acid, PEI	Breast cancer mice beared 4T1 cells	I.V.		CQDs/Ce6	37%	CQDs as the thermal sensitizer while Ce6 as the photosensitizer	[239]
PTT&PDT	L-cysteine, o-phenylenediamine	Cervical carcinoma mice beared U14 cells	I.V.		CQDs/HPPH	68.4%	Synergetic MR imaging	[240]
PTT&PDT	FAC, polylysine acid	Cervical carcinoma mice beared Hela cells	I.V.		Fe ₃ O ₄ -CDs /BPQDs	36.2%	Synergetic MR imaging	[249]
PTT&PDT	Polydopamine, folic acid	LNCaP cells	-		CQDs/polydopamine	-	Photosensitizer polydopamine was used as raw material	[250]
PTT&PDT	DME, Hypocrella bambusae	Breast cancer mice beared 4T1 cells	I.V.		CQDs/hypocrella bambusae	-	Synergetic PA imaging	[251]
PTT&PDT	Carbon nanotubes	Liver cancer mice beared H22 cells	I.T.		CQDs/ZnPc	-	Doping UCL	[238]

(Continued)

Table 4 (Continued).

Phototherapy	Raw Materials	Model	Administration	Thermal Sensitizer / Photosensitizer	Therapeutic Efficiency	Function	Reference
PTT&PDT	MnO ₂ and Cu(II)	Breast cancer mice beared 4T1 cells	I.V.	Mn,Cu-CDs	-	Four combination therapy of PTT/PDT/starving-like /immunotherapy	[241]
PTT&PDT	-	B16-F0 tumor-bearing nude mice	I.V.	GNRs/CDs	-	Synergetic FL and PA imaging	[252]

Abbreviations: EDTA, ethylenediamine tetraacetic acid; TNP, 1,3,5-trinitrotyrene; BPEI, Branched polyethyleneimine; PEI, polyethyleneimine; FAC, cyclophosphamide; HPPH, hydroxyphenyl-phenyl-hydantoin; DMF, N,N-Dimethylformamide.

starvation-like therapy and immunotherapy to fight tumours, in which Mn,Cu-CDs were utilized as both photosensitizers and thermal sensitizers, while the glucose-metabolic reaction agent glucose oxidase (GOx) snatched glucose from the tumour cells, causing them to starve.²⁴¹ Moreover, the tumour-associated antigens released by the above three ways of killing tumour cells stimulated cytotoxic T lymphocytes to penetrate distant tumours through the PD-1/PD-L1 pathway, which not only killed the primary tumours but also blocked the growth of distant tumours.

Compared with that of other nanoparticles, research on the application of CQDs in phototherapy is very limited. Table 4 summarizes the application of CQDs as thermal sensitizers/photosensitizers in antitumour research. Moreover, most CQD-mediated PTT systems are designed to encapsulate CQDs into nanoparticles. PTT requires that the thermal sensitizer has a superior photothermal conversion rate, and CQDs are candidates for thermal sensitizers owing to their naturally dark colour. The efficiency of photothermal conversion can be enhanced by screening raw materials and reforming synthetic CQDs. To design materials for CQD-mediated PDT, we need to bind a photosensitizer to the CQDs, and because tumour sites lack enough oxygen to produce ROS, so we also need to add specific materials to produce O₂. Among them, some transition metals, such as Co²⁺, Cd²⁺, Cu²⁺, Ag⁺, Mn²⁺ and Ni²⁺, can accelerate or replace Fe²⁺ to promote a Fenton-like reaction, thus becoming oxygen suppliers. Moreover, due to the low tumour eradication rates of PTT and PDT alone, the combination of PTT and PDT or with other therapies, including immunotherapy, radiotherapy, chemotherapy, hormone therapy and precision medicine, to achieve a higher tumour eradication rate needs further research.

Tumour-Targeting Lethal CQDs

Tumour-targeting lethal CQDs have antitumour effects without any other synergism and depend on the production of ROS after they are absorbed by tumour cells. Such CQDs only kill tumours, while normal cells can escape them. Using polyethylene glycol (PEG) as a carbon source, imidazole as a nitrogen source and phosphoric acid as a phosphorus source, Bajpai et al prepared N- and P-doped CQDs (NPCDs) by a heating method that could induce cell cycle arrest, autophagy and apoptosis in B16F10 melanoma cells.²⁵³ Yao et al utilized ginsenoside as a raw material, adding appropriate amounts of citric acid and EDA to improve the fluorescence performance to prepare

antitumour Re-CQDs,²⁵⁴ while Emam and Ahmed used NaOH to dissolve carrageenan and pullulan and then fabricated two kinds of CQDs by a hydrothermal method, which were used for antitumour and antiviral activities.²⁵⁵

Summary and Outlook

In this review, the preparation methods, features and characterizations of CQDs were briefly introduced, and the application prospects of CQDs in the antitumour field were also summarized. The preparation methods of CQDs include “top-down” methods and “bottom-up” methods. The difference is that the former separates the CQDs from matter with a large molecular weight, while the latter produces CQDs by carbonization of small molecules. CQDs have excellent biocompatibility due to their small structure and hydrophilic groups on the surface. Moreover, although there is no consensus on the luminescence mechanism of CQDs, it is known to be related to the vacancies of surface groups and epitopes.

The applications of CQDs in antitumour therapy mainly include the detection and diagnosis of early tumours, delivery of antitumour drugs as carriers, monitoring the release of antitumour drugs, phototherapy as photosensitizers or thermosensitizers, and targeted killing of tumour cells by autonomic production of ROS. Early-stage tumours have no obvious clinical manifestations, and the changes in tumour markers are very small, which makes them difficult to detect, diagnose and treat early. CQDs have excellent specificity and sensitivity, and they can be used to effectively monitor early-stage tumours by means of FRET, ECL, and structural modifications to enable the detection of tumour markers or the tumour microenvironment and to release antitumour drugs. CQDs are most widely utilized in antitumour therapy for drug delivery, including traditional drugs and gene therapy drugs. The small size of CQDs increases their specific surface area, which is conducive to the improvement of the encapsulation and loading efficiency of a series of antitumour drug molecules. In addition, the high biocompatibility, low relative cytotoxicity and good cell uptake ability of CQDs also make them excellent materials for drug delivery. CQDs can combine with drugs by covalent bonds, noncovalent bonds, or both, so they can create different types of connections to exhibit high performance while maintaining their original properties. Moreover, because tumour cells are more sensitive to the thermal environment than normal cells, CQDs can be utilized as thermal sensitizers to achieve the targeted killing of tumour cells. Moreover, a photosensitizer can be doped into CQDs or encapsulated in nanoparticles with CQDs to achieve the antitumour effect of synergistic PTT and PDT. The mechanism of

PDT is to produce ROS to damage tumour cells, which is consistent with the mechanism of killing tumour cells by tumour-targeting lethal CQDs.

Although CQDs have made significant contributions to the discovery, diagnosis and treatment of tumours, some challenges remain to be further explored. First, the combination of CQDs with other therapies, such as immunotherapy, gene therapy, endocrine therapy, cryotherapy and radiotherapy, needs to be further studied and promoted. Second, the biocompatibility of CQDs is concentration-dependent. Once the concentration threshold is exceeded, CQDs become toxic. The ameliorative concentration threshold for improving the biocompatibility without affecting the properties of CQDs remains to be further studied. Finally, PTT therapy with CQDs is only suitable for photothermal ablation of superficial tumours, so its treatment of deep tumours is a direction of future research.

Acknowledgments

We greatly acknowledge the financial support from the National Nature Science Foundation of China (82074473), the Natural Science Foundation of Jiangsu province (BK20191201, BE2020666), the Suzhou Health Personnel Training Project (GSWS2019074 and GSWS2020103), the Zhangjiagang Health Personnel Training Project (ZJGWSRC2020002), the Zhangjiagang Health System Youth Science and Technology Project (ZYYQ2005).

Disclosure

The authors report no conflicts of interest in this work.

References

1. Chaput F, Amer R, Baglivo E, et al. Intraocular T-cell lymphoma: clinical presentation, diagnosis, treatment, and outcome. *Ocul Immunol Inflamm*. 2017;25(5):639–648. doi:10.3109/09273948.2016.1139733
2. Wang SY, Chen XX, Li Y, Zhang YY. Application of multimodality imaging fusion technology in diagnosis and treatment of malignant tumors under the precision medicine plan. *Chin Med J (Engl)*. 2016;129(24):2991–2997. doi:10.4103/0366-6999.195467
3. Fisher B. Biological research in the evolution of cancer surgery: a personal perspective. *Cancer Res*. 2008;68(24):10007–10020. doi:10.1158/0008-5472.CAN-08-0186
4. Epstein RJ. Drug-induced DNA damage and tumor chemosensitivity. *J Clin Oncol*. 1990;8(12):2062–2084. doi:10.1200/JCO.1990.8.12.2062
5. Wang J, Wang H, Wang H, et al. Nonviolent self-catabolic DNase nanosponges for smart anticancer drug delivery. *ACS Nano*. 2019;13(5):5852–5863. doi:10.1021/acsnano.9b01589
6. Laperriere NJ, Bernstein M. Radiotherapy for brain tumors. *CA Cancer J Clin*. 1994;44(2):96–108. doi:10.3322/canjclin.44.2.96
7. Wang X, Hu C, Eisbruch A. Organ-sparing radiation therapy for head and neck cancer. *Nat Rev Clin Oncol*. 2011;8(11):639–648. doi:10.1038/nrclinonc.2011.106

8. de Dios NR, Murcia-Mejía M. Current and future strategies in radiotherapy for small-cell lung cancer. *J Clin Transl Res.* 2020;6(4):97–108.
9. Ianzini F, Kosmacek EA, Nelson ES, et al. Activation of meiosis-specific genes is associated with depolyploidization of human tumor cells following radiation-induced mitotic catastrophe. *Cancer Res.* 2009;69(6):2296–2304. doi:10.1158/0008-5472.CAN-08-3364
10. Mendonca MS, Howard KL, Farrington DL, et al. Delayed apoptotic responses associated with radiation-induced neoplastic transformation of human hybrid cells. *Cancer Res.* 1999;59(16):3972–3979.
11. Negi H, Merugu SB, Mangukiya HB, et al. Anterior gradient-2 monoclonal antibody inhibits lung cancer growth and metastasis by upregulating p53 pathway and without exerting any toxicological effects: a Preclinical Study. *Cancer Lett.* 2019;449:125–134. doi:10.1016/j.canlet.2019.01.025
12. Vahdat V, Ryan KE, Keating PL, et al. Atomic-scale wear of amorphous hydrogenated carbon during intermittent contact: a combined study using experiment, simulation, and theory. *ACS Nano.* 2014;8(7):7027–7040. doi:10.1021/nn501896e
13. Clancy AJ, Bayazit MK, Hodge SA, et al. Charged carbon nanomaterials: redox chemistries of fullerenes, carbon nanotubes, and graphenes. *Chem Rev.* 2018;118(16):7363–7408. doi:10.1021/acs.chemrev.8b00128
14. Deline AR, Frank BP, Smith CL, et al. Influence of oxygen-containing functional groups on the environmental properties, transformations, and toxicity of carbon nanotubes. *Chem Rev.* 2020;120(20):11651–11697. doi:10.1021/acs.chemrev.0c00351
15. Zou M, Zhao W, Wu H, et al. Single carbon fibers with a macroscopic-thickness, 3D highly porous carbon nanotube coating. *Adv Mater.* 2018;30(13):e1704419. doi:10.1002/adma.201704419
16. Nekoueián K, Amiri M, Sillanpää M, et al. Carbon-based quantum particles: an electroanalytical and biomedical perspective. *Chem Soc Rev.* 2019;48(15):4281–4316. doi:10.1039/C8CS00445E
17. Tunuguntla RH, Henley RY, Yao YC, et al. Enhanced water permeability and tunable ion selectivity in subnanometer carbon nanotube porins. *Science.* 2017;357(6353):792–796. doi:10.1126/science.aan2438
18. Zhang S, Kang L, Wang X, et al. Arrays of horizontal carbon nanotubes of controlled chirality grown using designed catalysts. *Nature.* 2017;543(7644):234–238. doi:10.1038/nature21051
19. Chen S, Qiu L, Cheng HM. Carbon-based fibers for advanced electrochemical energy storage devices. *Chem Rev.* 2020;120(5):2811–2878. doi:10.1021/acs.chemrev.9b00466
20. Stergiou A, Tagmatarchis N. Interfacing carbon dots for charge-transfer processes. *Small.* 2021;e2006005. doi:10.1002/smll.202006005
21. Wang B, Yu J, Sui L, et al. Rational design of multi-color-emissive carbon dots in a single reaction system by hydrothermal. *Adv Sci (Weinh).* 2020;8(1):2001453. doi:10.1002/advs.202001453
22. Xu X, Ray R, Gu Y, et al. Electrophoretic analysis and purification of fluorescent single-walled carbon nanotube fragments. *J Am Chem Soc.* 2004;126(40):12736–12737. doi:10.1021/ja040082h
23. Cao L, Wang X, Mezziani MJ, et al. Carbon dots for multiphoton bioimaging. *J Am Chem Soc.* 2007;129(37):11318–11319. doi:10.1021/ja0735271
24. Lin Z, Xue W, Chen H, Lin JM. Classical oxidant induced chemiluminescence of fluorescent carbon dots. *Chem Commun (Camb).* 2012;48(7):1051–1053. doi:10.1039/C1CC15290D
25. Yuan F, Wang Z, Li X, et al. Bright multicolor bandgap fluorescent carbon quantum dots for electroluminescent light-emitting diodes. *Adv Mater.* 2017;29(3):1604436.
26. Guo Z, Li Q, Li Z, et al. Fabrication of efficient alginate composite beads embedded with N-doped carbon dots and their application for enhanced rare earth elements adsorption from aqueous solutions. *J Colloid Interface Sci.* 2020;562:224–234. doi:10.1016/j.jcis.2019.12.030
27. Tao Y, Lin J, Wang D, Wang Y. Na⁺-functionalized carbon dots with aggregation-induced and enhanced cyan emission. *J Colloid Interface Sci.* 2021;588:469–475. doi:10.1016/j.jcis.2020.12.104
28. Li H, He X, Kang Z, et al. Water-soluble fluorescent carbon quantum dots and photocatalyst design. *Angew Chem Int Ed Engl.* 2010;49(26):4430–4434. doi:10.1002/anie.200906154
29. Liu Y, Tian Y, Tian Y, et al. Carbon-dot-based nanosensors for the detection of intracellular redox state. *Adv Mater.* 2015;27(44):7156–7160. doi:10.1002/adma.201503662
30. Xue X, Fang T, Yin L. Multistage delivery of CDs-DOX/ICG-loaded liposome for highly penetration and effective chemo-photothermal combination therapy. *Drug Deliv.* 2018;25(1):1826–1839. doi:10.1080/10717544.2018.1482975
31. Xu B, Zhao C, Wei W, et al. Aptamer carbon nanodot sandwich used for fluorescent detection of protein. *Analyst.* 2012;137(23):5483–5486. doi:10.1039/c2an36174d
32. Wei JS, Ding C, Zhang P, et al. Robust negative electrode materials derived from carbon dots and porous hydrogels for high-performance hybrid supercapacitors. *Adv Mater.* 2019;31(5):e1806197.
33. Lim SY, Shen W, Gao Z. Carbon quantum dots and their applications. *Chem Soc Rev.* 2015;44(1):362–381. doi:10.1039/C4CS00269E
34. Yan C, Wang C, Hou T, et al. Lasting tracking and rapid discrimination of live gram-positive bacteria by peptidoglycan-targeting carbon quantum dots. *ACS Appl Mater Interfaces.* 2021;13(1):1277–1287. doi:10.1021/acsami.0c19651
35. Hua XW, Bao YW, Wu FG. Fluorescent carbon quantum dots with intrinsic nucleolus-targeting capability for nucleolus imaging and enhanced cytosolic and nuclear drug delivery. *ACS Appl Mater Interfaces.* 2018;10(13):10664–10677. doi:10.1021/acsami.7b19549
36. Mosquera J, Garcia I, Henriksen-Lacey M, et al. Reversible control of protein corona formation on gold nanoparticles using host-guest interactions. *ACS Nano.* 2020;14(8):10745–10746. doi:10.1021/acsnano.0c06355
37. Song Y, Wang H, Zhang L, et al. Protein Corona formation of human serum albumin with carbon quantum dots from roast salmon. *Food Funct.* 2020;11(3):2358–2367. doi:10.1039/C9FO02967B
38. Yan C, Guo L, Shao X, et al. Amino acid-functionalized carbon quantum dots for selective detection of Al³⁺ ions and fluorescence imaging in living cells. *Anal Bioanal Chem.* 2021;413(15):3965–3974. doi:10.1007/s00216-021-03348-x
39. He X, Luo Q, Zhang J, et al. Gadolinium-doped carbon dots as nano-theranostic agents for MR/FL diagnosis and gene delivery. *Nanoscale.* 2019;11(27):12973–12982. doi:10.1039/C9NR03988K
40. Zhang J, Zhang H, Jiang J, et al. Doxorubicin-loaded carbon dots lipid-coated calcium phosphate nanoparticles for visual targeted delivery and therapy of tumor. *Int J Nanomedicine.* 2020;15:433–444. doi:10.2147/IJN.S229154
41. Wu J. The enhanced permeability and retention (EPR) effect: the significance of the concept and methods to enhance its application. *J Pers Med.* 2021;11(8):771. doi:10.3390/jpm11080771
42. Nakamura Y, Mochida A, Choyke PL, Kobayashi H. Nanodrug delivery: is the enhanced permeability and retention effect sufficient for curing cancer? *Bioconjug Chem.* 2016;27(10):2225–2238. doi:10.1021/acs.bioconjchem.6b00437

43. Arcudi F, Đorđević L, Prato M. Design, synthesis, and functionalization strategies of tailored carbon nanodots. *Acc Chem Res.* 2019;52(8):2070–2079. doi:10.1021/acs.accounts.9b00249
44. Papaioannou N, Titirici MM, Sapelkin A. Investigating the effect of reaction time on carbon dot formation, structure, and optical properties. *ACS Omega.* 2019;4(26):21658–21665. doi:10.1021/acsomega.9b01798
45. Wu ZL, Liu ZX, Yuan YH. Carbon dots: materials, synthesis, properties and approaches to long-wavelength and multicolor emission. *J Mater Chem B.* 2017;5(21):3794–3809. doi:10.1039/C7TB00363C
46. Zhang M, Wang H, Wang B, et al. Maltase decorated by chiral carbon dots with inhibited enzyme activity for glucose level control. *Small.* 2019;15(48):e1901512. doi:10.1002/smll.201901512
47. Qiao ZA, Wang Y, Gao Y, et al. Commercially activated carbon as the source for producing multicolor photoluminescent carbon dots by chemical oxidation. *Chem Commun (Camb).* 2010;46(46):8812–8814. doi:10.1039/c0cc02724c
48. Yu H, Li X, Zeng X, Lu Y. Preparation of carbon dots by non-focusing pulsed laser irradiation in toluene. *Chem Commun (Camb).* 2016;52(4):819–822. doi:10.1039/C5CC08384B
49. Zahiri M, Shafiee Afarani M, Arabi AM. Combustion synthesis of ZnO/ZnS nanocomposite phosphors. *J Fluoresc.* 2019;29(5):1227–1239. doi:10.1007/s10895-019-02434-9
50. Ye X, Xiang Y, Wang Q, et al. A red emissive two-photon fluorescence probe based on carbon dots for intracellular pH detection. *Small.* 2019;15(48):e1901673. doi:10.1002/smll.201901673
51. Atchudan R, Edison TNJI, Aseer KR, et al. Highly fluorescent nitrogen-doped carbon dots derived from Phyllanthus acidus utilized as a fluorescent probe for label-free selective detection of Fe³⁺ ions, live cell imaging and fluorescent ink. *Biosens Bioelectron.* 2018;99:303–311. doi:10.1016/j.bios.2017.07.076
52. Jiang K, Gao X, Feng X, et al. Carbon dots with dual-emissive, robust, and aggregation-induced room-temperature phosphorescence characteristics. *Angew Chem Int Ed Engl.* 2020;59(3):1263–1269. doi:10.1002/anie.201911342
53. Zheng XT, Tan YN. Development of blood-cell-selective fluorescent biodots for lysis-free leukocyte imaging and differential counting in whole blood. *Small.* 2020;16(12):e1903328. doi:10.1002/smll.201903328
54. Zhang M, Ju H, Zhang L, et al. Engineering iodine-doped carbon dots as dual-modal probes for fluorescence and X-ray CT imaging. *Int J Nanomedicine.* 2015;10:6943–6953.
55. Liu C, Zhang P, Zhai X, et al. Nano-carrier for gene delivery and bioimaging based on carbon dots with PEI-passivation enhanced fluorescence. *Biomaterials.* 2012;33(13):3604–3613. doi:10.1016/j.biomaterials.2012.01.052
56. Singh R, Kashayap S, Singh V, et al. QPRTase modified N-doped carbon quantum dots: a fluorescent bioprobe for selective detection of neurotoxin quinolinic acid in human serum. *Biosens Bioelectron.* 2018;101:103–109. doi:10.1016/j.bios.2017.10.017
57. Yang P, Zhu Z, Zhang T, et al. Orange-emissive carbon quantum dots: toward application in wound pH monitoring based on colorimetric and fluorescent changing. *Small.* 2019;15(44):e1902823. doi:10.1002/smll.201902823
58. Pierrat P, Wang R, Kereselidze D, et al. Efficient in vitro and in vivo pulmonary delivery of nucleic acid by carbon dot-based nanocarriers. *Biomaterials.* 2015;51:290–302. doi:10.1016/j.biomaterials.2015.02.017
59. Du FY, Jin X, Chen JH, et al. Nitrogen-doped carbon dots as multifunctional fluorescent probes. *J Nanoparticle Res.* 2014;16(11):2720. doi:10.1007/s11051-014-2720-8
60. Yang P, Zhang ZW, Zou GD, et al. Template thermolysis to create a carbon dots-embedded mesoporous titanium-oxo sulfate framework for visible-light photocatalytic applications. *Inorg Chem.* 2020;59(3):2062–2069. doi:10.1021/acs.inorgchem.9b03493
61. Yen YC, Lin CC, Chen PY, et al. Green synthesis of carbon quantum dots embedded onto titanium dioxide nanowires for enhancing photocurrent. *Royal Soc Open Sci.* 2017;4(5):161051–161060. doi:10.1098/rsos.161051
62. Hu C, Zhu Y, Zhao X. On-off-on nanosensors of carbon quantum dots derived from coal tar pitch for the detection of Cu²⁺, Fe³⁺, and L-ascorbic acid. *Spectrochim Acta A Mol Biomol Spectrosc.* 2021;250:119325. doi:10.1016/j.saa.2020.119325
63. Kang S, Kim KM, Jung K, et al. Graphene oxide quantum dots derived from coal for bioimaging: facile and green approach. *Sci Rep.* 2019;9(1):4101. doi:10.1038/s41598-018-37479-6
64. Han Y, Tang D, Yang Y, et al. Non-metal single/dual doped carbon quantum dots: a general flame synthetic method and electro-catalytic properties. *Nanoscale.* 2015;7(14):5955–5962. doi:10.1039/C4NR07116F
65. Zhang KY, Yu Q, Wei H, et al. Long-lived emissive probes for time-resolved photoluminescence bioimaging and biosensing. *Chem Rev.* 2018;118(4):1770–1839. doi:10.1021/acs.chemrev.7b00425
66. Chouksey S, Sankaranarayanan S, Pendem V, et al. Strong size dependency on the carrier and photon dynamics in InGaN/GaN single nanowalls determined using photoluminescence and ultrafast transient absorption spectroscopy. *Nano Lett.* 2017;17(8):4596–4603. doi:10.1021/acs.nanolett.7b00970
67. Yakunin S, Benin BM, Shynkarenko Y, et al. High-resolution remote thermometry and thermography using luminescent low-dimensional tin-halide perovskites. *Nat Mater.* 2019;18(8):846–852. doi:10.1038/s41563-019-0416-2
68. Nguyen HA, Srivastava I, Pan D, Gruebele M. Unraveling the fluorescence mechanism of carbon dots with sub-single-particle resolution. *ACS Nano.* 2020;14(5):6127–6137. doi:10.1021/acsnano.0c01924
69. Ehrat F, Bhattacharyya S, Schneider J, et al. Tracking the source of carbon dot photoluminescence: aromatic domains versus molecular fluorophores. *Nano Lett.* 2017;17(12):7710–7716. doi:10.1021/acs.nanolett.7b03863
70. Feng T, Zhu S, Zeng Q, et al. Supramolecular cross-link-regulated emission and related applications in polymer carbon dots. *ACS Appl Mater Interfaces.* 2018;10(15):12262–12277. doi:10.1021/acsaami.7b14857
71. Wang BB, Jin JC, Xu ZQ, et al. Single-step synthesis of highly photoluminescent carbon dots for rapid detection of Hg²⁺ with excellent sensitivity. *J Colloid Interface Sci.* 2019;551:101–110. doi:10.1016/j.jcis.2019.04.088
72. Bai J, Sun C, Jiang X. Carbon dots-decorated multiwalled carbon nanotubes nanocomposites as a high-performance electrochemical sensor for detection of H₂O₂ in living cells. *Anal Bioanal Chem.* 2016;408(17):4705–4714. doi:10.1007/s00216-016-9554-4
73. Tian XT, Yin XB. Carbon dots, unconventional preparation strategies, and applications beyond photoluminescence. *Small.* 2019;15(48):e1901803. doi:10.1002/smll.201901803
74. Wang B, Tan H, Zhang T, et al. Hydrothermal synthesis of N-doped carbon dots from an ethanalamine–ionic liquid gel to construct label-free multifunctional fluorescent probes for Hg²⁺, Cu²⁺ and S₂O₃²⁻. *Analyst.* 2019;144(9):3013–3022. doi:10.1039/C9AN00116F
75. Nasrin A, Hassan M, Gomes VG. Two-photon active nucleus-targeting carbon dots: enhanced ROS generation and photodynamic therapy for oral cancer. *Nanoscale.* 2020;12(40):20598–20603. doi:10.1039/D0NR05210H

76. Wu W, Zheng T, Tian Y. An enzyme-free amplification strategy based on two-photon fluorescent carbon dots for monitoring miR-9 in live neurons and brain tissues of Alzheimer's disease mice. *Chem Commun (Camb)*. 2020;56(58):8083–8086. doi:10.1039/D0CC01971B
77. Liu C, Xiao G, Yang M, et al. Mechanofluorochromic carbon nanodots: controllable pressure-triggered blue- and red-shifted photoluminescence. *Angew Chem Int Ed Engl*. 2018;57(7):1893–1897. doi:10.1002/anie.201711409
78. Ding H, Yu SB, Wei JS, Xiong HM. Full-color light-emitting carbon dots with a surface-state-controlled luminescence mechanism. *ACS Nano*. 2016;10(1):484–491. doi:10.1021/acsnano.5b05406
79. Zhang M, Zhao L, Du F, et al. Facile synthesis of cerium-doped carbon quantum dots as a highly efficient antioxidant for free radical scavenging. *Nanotechnology*. 2019;30(32):325101. doi:10.1088/1361-6528/ab12ef
80. Li L, Jin J, Liu J, et al. Accurate SERS monitoring of the plasmon mediated UV/visible/NIR photocatalytic and photothermal catalytic process involving Ag@carbon dots. *Nanoscale*. 2021;13(2):1006–1015. doi:10.1039/D0NR06293F
81. Tsurugaya T, Yoshida K, Yajima F, et al. Terahertz spectroscopy of individual carbon nanotube quantum dots. *Nano Lett*. 2019;19(1):242–246. doi:10.1021/acs.nanolett.8b03801
82. Jangra H, Chen Q, Fuks E, et al. Nucleophilicity and electrophilicity parameters for predicting absolute rate constants of highly asynchronous 1,3-dipolar cycloadditions of aryl diazomethanes. *J Am Chem Soc*. 2018;140(48):16758–16772. doi:10.1021/jacs.8b09995
83. Atchudan R, Edison TNJI, Lee YR. Nitrogen-doped carbon dots originating from unripe peach for fluorescent bioimaging and electrocatalytic oxygen reduction reaction. *J Colloid Interface Sci*. 2016;482:8–18. doi:10.1016/j.jcis.2016.07.058
84. Ansari F, Kahrizi D. Hydrothermal synthesis of highly fluorescent and non-toxic carbon dots using Stevia rebaudiana Bertoni. *Cell Mol Biol (Noisy-Le-Grand)*. 2018;64(12):32–36. doi:10.14715/cmb/2018.64.12.7
85. Li T, Wang ES, Wang J, Chen X. Regulating the properties of carbon dots via a solvent-involved molecule fusion strategy for improved sensing selectivity. *Anal Chim Acta*. 2019;1088:107–115. doi:10.1016/j.aca.2019.08.027
86. Qi H, Teng M, Liu M, et al. Biomass-derived nitrogen-doped carbon quantum dots: highly selective fluorescent probe for detecting Fe³⁺ ions and tetracyclines. *J Colloid Interface Sci*. 2019;539:332–341. doi:10.1016/j.jcis.2018.12.047
87. Liu H, He Z, Jiang LP, Zhu JJ. Microwave-assisted synthesis of wavelength-tunable photoluminescent carbon nanodots and their potential applications. *ACS Appl Mater Interfaces*. 2015;7(8):4913–4920. doi:10.1021/am508994w
88. Zhao DL, Chung TS. Applications of carbon quantum dots (CQDs) in membrane technologies: a review. *Water Res*. 2018;147:43–49. doi:10.1016/j.watres.2018.09.040
89. Biswas MC, Islam MT, Nandy PK, Hossain MM. Graphene Quantum Dots (GQDs) for bioimaging and drug delivery applications: a review. *ACS Mater Lett*. 2021;3(6):899–911. doi:10.1021/acsmaterialslett.0c00550
90. Chen WF, Li DJ, Tian L, et al. Synthesis of graphene quantum dots from natural polymer starch for cell imaging. *Green Chem*. 2018;20(19):4438–4442. doi:10.1039/C8GC02106F
91. Yan CR, Hu XL, Guan P, et al. Highly biocompatible graphene quantum dots: green synthesis, toxicity comparison and fluorescence imaging. *J Mater Sci*. 2020;55(3):1198–1215. doi:10.1007/s10853-019-04079-2
92. Hage FS, Radtke G, Kepaptsoglou DM, et al. Single-atom vibrational spectroscopy in the scanning transmission electron microscope. *Science*. 2020;367(6482):1124–1127. doi:10.1126/science.ab1136
93. Zhang L, Yang T, Du C, et al. Lithium whisker growth and stress generation in an in situ atomic force microscope-environmental transmission electron microscope set-up. *Nat Nanotechnol*. 2020;15(2):94–98. doi:10.1038/s41565-019-0604-x
94. Schorb M, Haberbosch I, Hagen WJH, et al. Software tools for automated transmission electron microscopy. *Nat Methods*. 2019;16(6):471–477. doi:10.1038/s41592-019-0396-9
95. Lim K, Bae Y, Jeon S, et al. A large-scale array of ordered graphene-sandwiched chambers for quantitative liquid-phase transmission electron microscopy. *Adv Mater*. 2020;32(39):e2002889. doi:10.1002/adma.202002889
96. Parent LR, Bakalis E, Proetto M, et al. Tackling the challenges of dynamic experiments using liquid-cell transmission electron microscopy. *Acc Chem Res*. 2018;51(1):3–11. doi:10.1021/acs.accounts.7b00331
97. Nakamura E. Atomic-resolution transmission electron microscopic movies for study of organic molecules, assemblies, and reactions: the first 10 years of development. *Acc Chem Res*. 2017;50(6):1281–1292. doi:10.1021/acs.accounts.7b00076
98. Hernandez-Cerdan P, Mansel BW, Leis A, et al. Structural analysis of polysaccharide networks by transmission electron microscopy: comparison with small-angle X-ray scattering. *Biomacromolecules*. 2018;19(3):989–995. doi:10.1021/acs.biomac.7b01773
99. Zhang J, Yuan Y, Liang G, Yu SH. Scale-up synthesis of fragrant nitrogen-doped carbon dots from bee pollens for bioimaging and catalysis. *Adv Sci (Weinh)*. 2015;2(4):1500002. doi:10.1002/advs.201500002
100. Li J, Li P, Wang D, Dong C. One-pot synthesis of aqueous soluble and organic soluble carbon dots and their multi-functional applications. *Talanta*. 2019;202:375–383. doi:10.1016/j.talanta.2019.05.019
101. Sun X, Brückner C, Lei Y. One-pot and ultrafast synthesis of nitrogen and phosphorus co-doped carbon dots possessing bright dual wavelength fluorescence emission. *Nanoscale*. 2015;7(41):17278–17282. doi:10.1039/C5NR05549K
102. Liu X, Li J, Huang Y, et al. Adsorption, Aggregation, and Deposition Behaviors of Carbon Dots on Minerals. *Environ Sci Technol*. 2017;51(11):6156–6164. doi:10.1021/acs.est.6b06558
103. Amarie S, Ganz T, Keilmann F. Mid-infrared near-field spectroscopy. *Opt Express*. 2009;17(24):21794–21801. doi:10.1364/OE.17.021794
104. Griffiths PR. The early days of commercial FT-IR spectrometry: a personal perspective. *Appl Spectrosc*. 2017;71(3):329–340. doi:10.1177/0003702816683529
105. Erfan M, Sabry YM, Sakr M, et al. On-chip micro-electromechanical system fourier transform infrared (MEMS FT-IR) spectrometer-based gas sensing. *Appl Spectrosc*. 2016;70(5):897–904. doi:10.1177/0003702816638295
106. Andreeva AB, Le KN, Chen L, et al. Soft mode metal-linker dynamics in carboxylate MOFs evidenced by variable-temperature infrared spectroscopy. *J Am Chem Soc*. 2020;142(45):19291–19299. doi:10.1021/jacs.0c09499
107. Wu Y, Xu L, Qian J, et al. Methotrexate-Mn²⁺ based nanoscale coordination polymers as a theranostic nanoplatform for MRI guided chemotherapy. *Biomater Sci*. 2020;8(2):712–719. doi:10.1039/C9BM01584A
108. Zhang M, Yao Q, Lu C, et al. Layered double hydroxide-carbon dot composite: high-performance adsorbent for removal of anionic organic dye. *ACS Appl Mater Interfaces*. 2014;6(22):20225–20233. doi:10.1021/am505765e
109. Nguyen L, Tao FF, Tang Y, et al. Understanding catalyst surfaces during catalysis through near ambient pressure X-ray photoelectron spectroscopy. *Chem Rev*. 2019;119(12):6822–6905. doi:10.1021/acs.chemrev.8b00114
110. Borgwardt M, Mahl J, Roth F, et al. Photoinduced charge carrier dynamics and electron injection efficiencies in au nanoparticle-sensitized TiO₂ determined with picosecond time-resolved X-ray photoelectron spectroscopy. *J Phys Chem Lett*. 2020;11(14):5476–5481. doi:10.1021/acs.jpcclett.0c00825

111. Zhang Y, Tamijani AA, Taylor ME, et al. Molecular surface functionalization of carbon materials via radical-induced grafting of terminal alkenes. *J Am Chem Soc.* 2019;141(20):8277–8288. doi:10.1021/jacs.9b02369
112. Takagi Y, Uruga T, Tada M, et al. Ambient pressure hard X-ray photoelectron spectroscopy for functional material systems as fuel cells under working conditions. *Acc Chem Res.* 2018;51(3):719–727. doi:10.1021/acs.accounts.7b00563
113. Mahmood A, Shi G, Wang Z, et al. Carbon quantum dots-TiO₂ nanocomposite as an efficient photocatalyst for the photodegradation of aromatic ring-containing mixed VOCs: an experimental and DFT studies of adsorption and electronic structure of the interface. *J Hazard Mater.* 2021;401:123402. doi:10.1016/j.jhazmat.2020.123402
114. Ghreghlou M, Esmaeili AA, Darroudi M. Green synthesis of fluorescent carbon dots from *elaegnus angustifolia* and its application as tartrazine sensor. *J Fluoresc.* 2021;31(1):185–193. doi:10.1007/s10895-020-02645-5
115. Wu Y, Song X, Wang N, et al. Carbon dots from roasted chicken accumulate in lysosomes and induce lysosome-dependent cell death. *Food Funct.* 2020;11(11):10105–10113. doi:10.1039/D0FO02144J
116. Liu Y, Wu P, Wu X, et al. Nitrogen and copper (II) co-doped carbon dots for applications in ascorbic acid determination by non-oxidation reduction strategy and cellular imaging. *Talanta.* 2020;210:120649. doi:10.1016/j.talanta.2019.120649
117. Drake JM, Paull EO, Graham NA, et al. Phosphoproteome integration reveals patient-specific networks in prostate cancer. *Cell.* 2016;166(4):1041–1054. doi:10.1016/j.cell.2016.07.007
118. Klaeger S, Heinzlmeir S, Wilhelm M, et al. The target landscape of clinical kinase drugs. *Science.* 2017;358(6367):eaan4368. doi:10.1126/science.aan4368
119. Coleman RL, Fleming GF, Brady MF, et al. Veliparib with first-line chemotherapy and as maintenance therapy in ovarian cancer. *N Engl J Med.* 2019;381(25):2403–2415. doi:10.1056/NEJMoa1909707
120. Barton MK. Encouraging long-term outcomes reported in patients with stage I non-small cell lung cancer treated with stereotactic ablative radiotherapy. *CA Cancer J Clin.* 2017;67(5):349–350. doi:10.3322/caac.21375
121. Hu Z, Li Z, Ma Z, Curtis C. Multi-cancer analysis of clonality and the timing of systemic spread in paired primary tumors and metastases. *Nat Genet.* 2020;52(7):701–708. doi:10.1038/s41588-020-0628-z
122. Liu HW, Chen L, Xu C, et al. Recent progresses in small-molecule enzymatic fluorescent probes for cancer imaging. *Chem Soc Rev.* 2018;47(18):7140–7180.
123. Li D, Jing P, Sun L, et al. Near-infrared excitation/emission and multiphoton-induced fluorescence of carbon dots. *Adv Mater.* 2018;30(13):e1705913. doi:10.1002/adma.201705913
124. Du J, Xu N, Fan J, et al. Carbon dots for in vivo bioimaging and theranostics. *Small.* 2019;15(32):e1805087. doi:10.1002/smll.201805087
125. Lesani P, Singh G, Viray CM, et al. Two-photon dual-emissive carbon dot-based probe: deep-tissue imaging and ultrasensitive sensing of intracellular ferric ions. *ACS Appl Mater Interfaces.* 2020;12(16):18395–18406. doi:10.1021/acsami.0c05217
126. Lu W, Jiao Y, Gao Y, et al. Bright yellow fluorescent carbon dots as a multifunctional sensing platform for the label-free detection of fluoroquinolones and histidine. *ACS Appl Mater Interfaces.* 2018;10(49):42915–42924. doi:10.1021/acsami.8b16710
127. Wu L, Huang C, Emery BP, et al. Förster resonance energy transfer (FRET)-based small-molecule sensors and imaging agents. *Chem Soc Rev.* 2020;49(15):5110–5139. doi:10.1039/C9CS00318E
128. Hou S, Chen Y, Lu D, et al. A self-assembled plasmonic substrate for enhanced fluorescence resonance energy transfer. *Adv Mater.* 2020;32(8):e1906475. doi:10.1002/adma.201906475
129. Wypijewska Del Nogal A, Fuchtbauer AF, Bood M, et al. Getting DNA and RNA out of the dark with 2CNqA: a bright adenine analogue and interbase FRET donor. *Nucleic Acids Res.* 2020;48(14):7640–7652. doi:10.1093/nar/gkaa525
130. Baibakov M, Patra S, Claude JB, et al. Extending single-molecule Förster resonance energy transfer (FRET) range beyond 10 nanometers in zero-mode waveguides. *ACS Nano.* 2019;13(7):8469–8480. doi:10.1021/acsnano.9b04378
131. Zheng W, Borgia A, Buholzer K, et al. Probing the action of chemical denaturant on an intrinsically disordered protein by simulation and experiment. *J Am Chem Soc.* 2016;138(36):11702–11713. doi:10.1021/jacs.6b05443
132. Kudr J, Richtera L, Xhaxhiu K, et al. Carbon dots based FRET for the detection of DNA damage. *Biosens Bioelectron.* 2017;92:133–139. doi:10.1016/j.bios.2017.01.067
133. Kim YJ, Guo P, Schaller RD. Aqueous carbon quantum dot-embedded PC60-PC61BM nanospheres for ecological fluorescent printing: contrasting fluorescence resonance energy-transfer signals between watermelon-like and random morphologies. *J Phys Chem Lett.* 2019;10(21):6525–6535. doi:10.1021/acs.jpcclett.9b02426
134. Wang Y, Meng H, Jia M, et al. Intraparticle FRET of Mn(II)-doped carbon dots and its application in discrimination of volatile organic compounds. *Nanoscale.* 2016;8(39):17190–17195. doi:10.1039/C6NR05927A
135. Hamd-Ghadareh S, Salimi A, Fathi F, Bahrami S. An amplified comparative fluorescence resonance energy transfer immunosensing of CA125 tumor marker and ovarian cancer cells using green and economic carbon dots for bio-applications in labeling, imaging and sensing. *Biosens Bioelectron.* 2017;96:308–316. doi:10.1016/j.bios.2017.05.003
136. Lee S, Lee HJ, Ji Y, et al. Electrochemiluminescent transistors: a new strategy toward light-emitting switching devices. *Adv Mater.* 2021;33(5):e2005456. doi:10.1002/adma.202005456
137. Zhao Y, Yu J, Xu G, et al. Photoinduced electrochemiluminescence at silicon electrodes in water. *J Am Chem Soc.* 2019;141(33):13013–13016. doi:10.1021/jacs.9b06743
138. Zanut A, Palomba F, Rossi Scota M, et al. Dye-doped silica nanoparticles for enhanced ECL-based immunoassay analytical performance. *Angew Chem Int Ed Engl.* 2020;59(49):21858–21863. doi:10.1002/anie.202009544
139. Liu G, Chen Z, Jin BK, Jiang LP. A ratiometric electrochemiluminescent cytosensor based on polyaniline hydrogel electrodes in spatially separated electrochemiluminescent systems. *Analyst.* 2021;146(6):1835–1838. doi:10.1039/D0AN02408B
140. Liu Q, Ma C, Liu XP, et al. A novel electrochemiluminescence biosensor for the detection of microRNAs based on a DNA functionalized nitrogen doped carbon quantum dots as signal enhancers. *Biosens Bioelectron.* 2017;92:273–279. doi:10.1016/j.bios.2017.02.027
141. Qin D, Jiang X, Mo G, et al. Electrochemiluminescence immunoassay of human chorionic gonadotropin using silver carbon quantum dots and functionalized polymer nanospheres. *Mikrochim Acta.* 2020;187(8):482. doi:10.1007/s00604-020-04450-0
142. Qiu Y, Zhou B, Yang X, et al. Novel single-cell analysis platform based on a solid-state zinc-coadsorbed carbon quantum dots electrochemiluminescence probe for the evaluation of CD44 expression on breast cancer cells. *ACS Appl Mater Interfaces.* 2017;9(20):16848–16856. doi:10.1021/acsami.7b02793
143. Chen A, Liang W, Wang H, et al. Anodic electrochemiluminescence of carbon dots promoted by nitrogen doping and application to rapid cancer cell detection. *Anal Chem.* 2020;92(1):1379–1385. doi:10.1021/acs.analchem.9b04537
144. Gu C, Guo C, Li Z, et al. Bimetallic ZrHf-based metal-organic framework embedded with carbon dots: ultra-sensitive platform for early diagnosis of HER2 and HER2-overexpressed living cancer cells. *Biosens Bioelectron.* 2019;134:8–15. doi:10.1016/j.bios.2019.03.043

145. Won HJ, Ryplida B, Kim SG, et al. Diselenide-bridged carbon-dot-mediated self-healing, conductive, and adhesive wireless hydrogel sensors for label-free breast cancer detection. *ACS Nano*. 2020;14(7):8409–8420. doi:10.1021/acsnano.0c02517
146. Kalytchuk S, Zdražil L, Bađura Z, et al. Carbon dots detect water-to-ice phase transition and act as alcohol sensors via fluorescence turn-off/on mechanism. *ACS Nano*. 2021;15(4):6582–6593. doi:10.1021/acsnano.0c09781
147. Li L, Shi L, Jia J, et al. Dual photoluminescence emission carbon dots for ratiometric fluorescent GSH sensing and cancer cell recognition. *ACS Appl Mater Interfaces*. 2020;12(16):18250–18257. doi:10.1021/acsnano.0c00283
148. Mohammadi S, Salimi A. Fluorometric determination of microRNA-155 in cancer cells based on carbon dots and MnO₂ nanosheets as a donor-acceptor pair. *Mikrochim Acta*. 2018;185(8):372. doi:10.1007/s00604-018-2868-5
149. Tang C, Zhou J, Qian Z, et al. A universal fluorometric assay strategy for glycosidases based on functional carbon quantum dots: β -galactosidase activity detection in vitro and in living cells. *J Mater Chem B*. 2017;5(10):1971–1979. doi:10.1039/C6TB03361J
150. Sidhu JS, Singh A, Garg N, Singh N. Carbon dot based, naphthalimide coupled FRET pair for highly selective ratiometric detection of thioredoxin reductase and cancer screening. *ACS Appl Mater Interfaces*. 2017;9(31):25847–25856. doi:10.1021/acsnano.7b07046
151. Liu Q, Xu S, Niu C, et al. Distinguish cancer cells based on targeting turn-on fluorescence imaging by folate functionalized green emitting carbon dots. *Biosens Bioelectron*. 2015;64:119–125. doi:10.1016/j.bios.2014.08.052
152. Wu L, Wang J, Ren J, et al. Highly sensitive electrochemiluminescent cytosensing using carbon nanodot@Ag hybrid material and graphene for dual signal amplification. *Chem Commun (Camb)*. 2013;49(50):5675–5677. doi:10.1039/c3cc2637h
153. Xie X, Peng Z, Wang Z, et al. Monitoring biothiols dynamics in living cells by ratiometric fluorescent gold carbon dots. *Talanta*. 2020;218:121214. doi:10.1016/j.talanta.2020.121214
154. Zheng M, Ruan S, Liu S, et al. Self-targeting fluorescent carbon dots for diagnosis of brain cancer cells. *ACS Nano*. 2015;9(11):11455–11461. doi:10.1021/acsnano.5b05575
155. Ji X, Lv H, Sun X, Ding C. Green-emitting carbon dot loaded silica nanoparticles coated with DNA-cross-linked hydrogels for sensitive carcinoembryonic antigen detection and effective targeted cancer therapy. *Chem Commun (Camb)*. 2019;55(100):15101–15104. doi:10.1039/C9CC07831B
156. Chramiec A, Teles D, Yeager K, et al. Integrated human organ-on-a-chip model for predictive studies of anti-tumor drug efficacy and cardiac safety. *Lab Chip*. 2020;20(23):4357–4372. doi:10.1039/D0LC00424C
157. Wu S, Zhou R, Chen H, et al. Highly efficient oxygen photosensitization of carbon dots: the role of nitrogen doping. *Nanoscale*. 2020;12(9):5543–5553. doi:10.1039/C9NR10986B
158. Li RS, Gao PF, Zhang HZ, et al. Chiral nanoprobe for targeting and long-term imaging of the Golgi apparatus. *Chem Sci*. 2017;8(10):6829–6835. doi:10.1039/C7SC01316G
159. Broxterman HJ, Gotink KJ, Verheul HM. Understanding the causes of multidrug resistance in cancer: a comparison of doxorubicin and sunitinib. *Drug Resist Updat*. 2009;12(4–5):114–126. doi:10.1016/j.drug.2009.07.001
160. Han D, Wang Y, Wang Y, et al. The tumor-suppressive human circular RNA CircITCH sponges miR-330-5p to ameliorate doxorubicin-induced cardiotoxicity through upregulating SIRT6, survivin, and SERCA2a. *Circ Res*. 2020;127(4):e108–e125. doi:10.1161/CIRCRESAHA.119.316061
161. Gyöngyösi M, Lukovic D, Zlabinger K, et al. Liposomal doxorubicin attenuates cardiotoxicity via induction of interferon-related DNA damage resistance. *Cardiovasc Res*. 2020;116(5):970–982.
162. Harris LN, Broadwater G, Abu-Khalaf M, et al. Topoisomerase II {alpha} amplification does not predict benefit from dose-intense cyclophosphamide, doxorubicin, and fluorouracil therapy in HER2-amplified early breast cancer: results of CALGB 8541/150013. *J Clin Oncol*. 2009;27(21):3430–3436. doi:10.1200/JCO.2008.18.4085
163. Fornari FA, Randolph JK, Yalowich JC, et al. Interference by doxorubicin with DNA unwinding in MCF-7 breast tumor cells. *Mol Pharmacol*. 1994;45(4):649–656.
164. Xiang S, Dauchy RT, Hauch A, et al. Doxorubicin resistance in breast cancer is driven by light at night-induced disruption of the circadian melatonin signal. *J Pineal Res*. 2015;59(1):60–69. doi:10.1111/jpi.12239
165. Koukourakis MI, Koukouraki S, Giatromanolaki A, et al. Liposomal doxorubicin and conventionally fractionated radiotherapy in the treatment of locally advanced non-small-cell lung cancer and head and neck cancer. *J Clin Oncol*. 1999;17(11):3512–3521. doi:10.1200/JCO.1999.17.11.3512
166. Wallin JJ, Guan J, Prior WW, et al. Nuclear phospho-Akt increase predicts synergy of PI3K inhibition and doxorubicin in breast and ovarian cancer. *Sci Transl Med*. 2010;2(48):48ra66. doi:10.1126/scitranslmed.3000630
167. Pan XQ, Zheng X, Shi G, et al. Strategy for the treatment of acute myelogenous leukemia based on folate receptor beta-targeted liposomal doxorubicin combined with receptor induction using all-trans retinoic acid. *Blood*. 2002;100(2):594–602. doi:10.1182/blood.V100.2.594
168. Di Lauro L, Belli F, Arena MG, et al. Epirubicin, cisplatin and docetaxel combination therapy for metastatic gastric cancer. *Ann Oncol*. 2005;16(9):1498–1502. doi:10.1093/annonc/mdi281
169. Gong X, Zhang Q, Gao Y, et al. Phosphorus and nitrogen dual-doped hollow carbon dot as a nanocarrier for doxorubicin delivery and biological imaging. *ACS Appl Mater Interfaces*. 2016;8(18):11288–11297. doi:10.1021/acsnano.6b01577
170. Li S, Amat D, Peng Z, et al. Transferrin conjugated nontoxic carbon dots for doxorubicin delivery to target pediatric brain tumor cells. *Nanoscale*. 2016;8(37):16662–16669. doi:10.1039/C6NR05055G
171. Su W, Guo R, Yuan F, et al. Red-emissive carbon quantum dots for nuclear drug delivery in cancer stem cells. *J Phys Chem Lett*. 2020;11(4):1357–1363. doi:10.1021/acs.jpclett.9b03891
172. Hou L, Chen D, Wang R, et al. Transformable honeycomb-like nanoassemblies of carbon dots for regulated multisite delivery and enhanced antitumor chemimmunotherapy. *Angew Chem Int Ed Engl*. 2021;60(12):6581–6592. doi:10.1002/anie.202014397
173. Kang MS, Singh RK, Kim TH, et al. Optical imaging and anticancer chemotherapy through carbon dot created hollow mesoporous silica nanoparticles. *Acta Biomater*. 2017;55:466–480. doi:10.1016/j.actbio.2017.03.054
174. Türk S, Altunsoy I, Gç E, et al. A novel multifunctional NCQDs-based injectable self-crosslinking and in situ forming hydrogel as an innovative stimuli responsive smart drug delivery system for cancer therapy. *Mater Sci Eng C Mater Biol Appl*. 2021;121:111829. doi:10.1016/j.msec.2020.111829
175. Sutton EC, McDevitt CE, Prochnau JY, et al. Nucleolar stress induction by oxaliplatin and derivatives. *J Am Chem Soc*. 2019;141(46):18411–18415. doi:10.1021/jacs.9b10319
176. Hamfjord J, Guren TK, Dajani O, et al. Total circulating cell-free DNA as a prognostic biomarker in metastatic colorectal cancer before first-line oxaliplatin-based chemotherapy. *Ann Oncol*. 2019;30(7):1088–1095. doi:10.1093/annonc/mdz139
177. Singhal C, Pundir CS, Narang J. A genosensor for detection of consensus DNA sequence of dengue virus using ZnO/Pt-Pd nanocomposites. *Biosens Bioelectron*. 2017;97:75–82. doi:10.1016/j.bios.2017.05.047

178. Zheng M, Liu S, Li J, et al. Integrating oxaliplatin with highly luminescent carbon dots: an unprecedented theranostic agent for personalized medicine. *Adv Mater*. 2014;26(21):3554–3560. doi:10.1002/adma.201306192
179. Feng T, Ai X, An G, et al. Charge-convertible carbon dots for imaging-guided drug delivery with enhanced in vivo cancer therapeutic efficiency. *ACS Nano*. 2016;10(4):4410–4420. doi:10.1021/acsnano.6b00043
180. Feng T, Ai X, Ong H, Zhao Y. Dual-responsive carbon dots for tumor extracellular microenvironment triggered targeting and enhanced anticancer drug delivery. *ACS Appl Mater Interfaces*. 2016;8(29):18732–18740. doi:10.1021/acsnano.6b06695
181. High KA, Roncarolo MG. Gene therapy. *N Engl J Med*. 2019;381(5):455–464. doi:10.1056/NEJMr1706910
182. Kuzmin DA, Shutova MV, Johnston NR, et al. The clinical landscape for AAV gene therapies. *Nat Rev Drug Discov*. 2021;20(3):173–174. doi:10.1038/d41573-021-00017-7
183. Sun J, Roy S. Gene-based therapies for neurodegenerative diseases. *Nat Neurosci*. 2021;24(3):297–311. doi:10.1038/s41593-020-00778-1
184. Liu X, Bao X, Hu M, et al. Inhibition of PCSK9 potentiates immune checkpoint therapy for cancer. *Nature*. 2020;588(7839):693–698. doi:10.1038/s41586-020-2911-7
185. Yang X, Wang Y, Shen X, et al. One-step synthesis of photoluminescent carbon dots with excitation-independent emission for selective bioimaging and gene delivery. *J Colloid Interface Sci*. 2017;492:1–7. doi:10.1016/j.jcis.2016.12.057
186. Devi P, Saini S, Kim KH. The advanced role of carbon quantum dots in nanomedical applications. *Biosens Bioelectron*. 2019;141:111158. doi:10.1016/j.bios.2019.02.059
187. Ju E, Li T, Liu Z, et al. Specific inhibition of viral MicroRNAs by carbon dots-mediated delivery of locked nucleic acids for therapy of virus-induced cancer. *ACS Nano*. 2020;14(1):476–487. doi:10.1021/acsnano.9b06333
188. Wu D, Li BL, Zhao Q, et al. Assembling defined DNA nanostructure with nitrogen-enriched carbon dots for theranostic cancer applications. *Small*. 2020;16(19):e1906975. doi:10.1002/sml.201906975
189. Li S, Su W, Wu H, et al. Targeted tumour theranostics in mice via carbon quantum dots structurally mimicking large amino acids. *Nat Biomed Eng*. 2020;4(7):704–716. doi:10.1038/s41551-020-0540-y
190. Zhou Q, Gong N, Zhang D, et al. Mannose-derived carbon dots amplify microwave ablation-induced antitumor immune responses by capturing and transferring “danger signals” to dendritic cells. *ACS Nano*. 2021;15(2):2920–2932. doi:10.1021/acsnano.0c09120
191. Ghosal K, Ghosh S, Ghosh D, Sarkar K. Natural polysaccharide derived carbon dot based in situ facile green synthesis of silver nanoparticles: synergistic effect on breast cancer. *Int J Biol Macromol*. 2020;162:1605–1615.
192. Kalaiyaran G, Veerapandian M, JebaMercy G, et al. Amygdalin-functionalized carbon quantum dots for probing β -glucosidase activity for cancer diagnosis and therapeutics. *ACS Biomater Sci Eng*. 2019;5(6):3089–3099. doi:10.1021/acsbomaterials.9b00394
193. Wang HJ, He X, Luo TY, et al. Amphiphilic carbon dots as versatile vectors for nucleic acid and drug delivery. *Nanoscale*. 2017;9(18):5935–5947. doi:10.1039/C7NR01029J
194. Prasad R, Aiyer S, Chauhan DS, et al. Bioresponsive carbon nano-gated multifunctional mesoporous silica for cancer theranostics. *Nanoscale*. 2016;8(8):4537–4546. doi:10.1039/C5NR06756A
195. Hailing Y, Xiufang L, Lili W, et al. Doxorubicin-loaded fluorescent carbon dots with PEI passivation as a drug delivery system for cancer therapy. *Nanoscale*. 2020;12(33):17222–17237. doi:10.1039/D0NR01236J
196. Li WQ, Wang Z, Hao S, et al. Mitochondria-based aircraft carrier enhances in vivo imaging of carbon quantum dots and delivery of anticancer drug. *Nanoscale*. 2018;10(8):3744–3752. doi:10.1039/C7NR08816G
197. Zhang Y, Zhang C, Chen J, et al. Trackable mitochondria-targeting nanomicellar loaded with doxorubicin for overcoming drug resistance. *ACS Appl Mater Interfaces*. 2017;9(30):25152–25163. doi:10.1021/acsnano.7b07219
198. Ma J, Kang K, Zhang Y, et al. Detachable polyzwitterion-coated ternary nanoparticles based on peptide dendritic carbon dots for efficient drug delivery in cancer therapy. *ACS Appl Mater Interfaces*. 2018;10(50):43923–43935. doi:10.1021/acsnano.8b17041
199. Hettiarachchi SD, Graham RM, Mintz KJ, et al. Triple conjugated carbon dots as a nano-drug delivery model for glioblastoma brain tumors. *Nanoscale*. 2019;11(13):6192–6205. doi:10.1039/C8NR08970A
200. Li W, Liu Q, Zhang P, Liu L. Zwitterionic nanogels crosslinked by fluorescent carbon dots for targeted drug delivery and simultaneous bioimaging. *Acta Biomater*. 2016;40:254–262. doi:10.1016/j.actbio.2016.04.006
201. Gao N, Yang W, Nie H, et al. Turn-on theranostic fluorescent nanoprobe by electrostatic self-assembly of carbon dots with doxorubicin for targeted cancer cell imaging, in vivo hyaluronidase analysis, and targeted drug delivery. *Biosens Bioelectron*. 2017;96:300–307. doi:10.1016/j.bios.2017.05.019
202. Feng T, Chua HJ, Zhao Y. Reduction-responsive carbon dots for real-time ratiometric monitoring of anticancer prodrug activation in living cells. *ACS Biomater Sci Eng*. 2017;3(8):1535–1541. doi:10.1021/acsbomaterials.7b00264
203. Li X, Hu S, Lin Z, et al. Dual-responsive mesoporous silica nanoparticles coated with carbon dots and polymers for drug encapsulation and delivery. *Nanomedicine (Lond)*. 2020;15(25):2447–2458. doi:10.2217/nnm-2019-0440
204. Li X, Lovell JF, Yoon J, Chen X. Clinical development and potential of photothermal and photodynamic therapies for cancer. *Nat Rev Clin Oncol*. 2020;17(11):657–674. doi:10.1038/s41571-020-0410-2
205. Zhou C, Zhang L, Sun T, et al. Activatable NIR-II plasmonic nanotheranostics for efficient photoacoustic imaging and photothermal cancer therapy. *Adv Mater*. 2021;33(3):e2006532. doi:10.1002/adma.202006532
206. Li L, Lu Y, Jiang C, et al. Actively targeted deep tissue imaging and photothermal-chemo therapy of breast cancer by antibody-functionalized drug-loaded x-ray-responsive bismuth sulfide@mesoporous silica core-shell nanoparticles. *Adv Funct Mater*. 2018;28(5):1704623. doi:10.1002/adfm.201704623
207. Xu C, Pu K. Second near-infrared photothermal materials for combinational nanotheranostics. *Chem Soc Rev*. 2021;50(2):1111–1137. doi:10.1039/D0CS00664E
208. Zhang J, Yang C, Zhang R, et al. Biocompatible D-A semiconducting polymer nanoparticle with light-harvesting unit for highly effective photoacoustic imaging guided photothermal therapy. *Adv Funct Mater*. 2017;27(13):1605094. doi:10.1002/adfm.201605094
209. Liu X, Wang C, Wang X, et al. A dual-targeting Fe₃O₄@C/ZnO-DOX-FA nanoplatfrom with pH-responsive drug release and synergetic chemo-photothermal antitumor in vitro and in vivo. *Mater Sci Eng C Mater Biol Appl*. 2021;118:111455. doi:10.1016/j.msec.2020.111455
210. Diroll BT, Brumberg A, Leonard AA, et al. Photothermal behaviour of titanium nitride nanoparticles evaluated by transient X-ray diffraction. *Nanoscale*. 2021;13(4):2658–2664. doi:10.1039/D0NR08202C
211. Espinosa A, Curcio A, Cabana S, et al. Intracellular biodegradation of Ag nanoparticles, storage in ferritin, and protection by a Au shell for enhanced photothermal therapy. *ACS Nano*. 2018;12(7):6523–6535. doi:10.1021/acsnano.8b00482

212. Zhou S, Shang L, Zhao Y, et al. Pd single-atom catalysts on nitrogen-doped graphene for the highly selective photothermal hydrogenation of acetylene to ethylene. *Adv Mater.* 2019;31(18):e1900509. doi:10.1002/adma.201900509
213. Neelgund GM, Oki A, Bandara S, Carson L. Photothermal effect and cytotoxicity of CuS nanoflowers deposited over folic acid conjugated nanographene oxide. *J Mater Chem B.* 2021;9(7):1792–1803. doi:10.1039/D0TB02366C
214. Xue K, Wei F, Lin J, et al. Tumor acidity-responsive carrier-free nanodrugs based on targeting activation via ICG-templated assembly for NIR-II imaging-guided photothermal-chemotherapy. *Biomater Sci.* 2021;9(3):1008–1019. doi:10.1039/D0BM01864C
215. Yu Y, Song M, Chen C, et al. Bortezomib-encapsulated CuS/carbon dot nanocomposites for enhanced photothermal therapy via stabilization of polyubiquitinated substrates in the proteasomal degradation pathway. *ACS Nano.* 2020;14(8):10688–10703. doi:10.1021/acsnano.0c05332
216. Wang H, Mukherjee S, Yi J, et al. Biocompatible chitosan-carbon dot hybrid nanogels for NIR-imaging-guided synergistic photothermal-chemo therapy. *ACS Appl Mater Interfaces.* 2017;9(22):18639–18649. doi:10.1021/acsami.7b06062
217. Qian M, Chen L, Du Y, et al. Biodegradable mesoporous silica achieved via carbon nanodots-incorporated framework swelling for debris-mediated photothermal synergistic immunotherapy. *Nano Lett.* 2019;19(12):8409–8417. doi:10.1021/acs.nanolett.9b02448
218. Lo PC, Rodríguez-Morgade MS, Pandey RK, et al. The unique features and promises of phthalocyanines as advanced photosensitizers for photodynamic therapy of cancer. *Chem Soc Rev.* 2020;49(4):1041–1056. doi:10.1039/C9CS00129H
219. Li M, Sun W, Tian R, et al. Smart J-aggregate of cyanine photosensitizer with the ability to target tumor and enhance photodynamic therapy efficacy. *Biomaterials.* 2021;269:120532. doi:10.1016/j.biomaterials.2020.120532
220. Wu Y, Li F, Zhang X, et al. Tumor microenvironment-responsive PEGylated heparin-pyropheophorbide-a nanoconjugates for photodynamic therapy. *Carbohydr Polym.* 2021;255:117490. doi:10.1016/j.carbpol.2020.117490
221. Zhao X, Liu J, Fan J, et al. Recent progress in photosensitizers for overcoming the challenges of photodynamic therapy: from molecular design to application. *Chem Soc Rev.* 2021;50(6):4185–4219. doi:10.1039/D0CS00173B
222. Zhu T, Shi L, Ma C, et al. Fluorinated chitosan-mediated intracellular catalase delivery for enhanced photodynamic therapy of oral cancer. *Biomater Sci.* 2021;9(3):658–662. doi:10.1039/D0BM01898H
223. Li Y, Sui H, Jiang C, et al. Dihydroartemisinin increases the sensitivity of photodynamic therapy Via NF- κ B/HIF-1 α /VEGF pathway in esophageal cancer cell in vitro and in vivo. *Cell Physiol Biochem.* 2018;48(5):2035–2045. doi:10.1159/000492541
224. Liu B, Qiao G, Han Y, et al. Targeted theranostics of lung cancer: PD-L1-guided delivery of gold nanoprisms with chlorin e6 for enhanced imaging and photothermal/photodynamic therapy. *Acta Biomater.* 2020;117:361–373. doi:10.1016/j.actbio.2020.09.040
225. Tham HP, Xu K, Lim WQ, et al. Microneedle-assisted topical delivery of photodynamically active mesoporous formulation for combination therapy of deep-seated melanoma. *ACS Nano.* 2018;12(12):11936–11948. doi:10.1021/acsnano.8b03007
226. Dos Santos AF, Inague A, Arini GS, et al. Distinct photo-oxidation-induced cell death pathways lead to selective killing of human breast cancer cells. *Cell Death Dis.* 2020;11(12):1070. doi:10.1038/s41419-020-03275-2
227. Cho HJ, Park SJ, Jung WH, et al. Injectable single-component peptide depot: autonomously rechargeable tumor photosensitization for repeated photodynamic therapy. *ACS Nano.* 2020;14(11):15793–15805. doi:10.1021/acsnano.0c06881
228. Nash GT, Luo T, Lan G, et al. Nanoscale metal-organic layer isolates phthalocyanines for efficient mitochondria-targeted photodynamic therapy. *J Am Chem Soc.* 2021;143(5):2194–2199. doi:10.1021/jacs.0c12330
229. Huang Z, Xu Z, Mahboub M, et al. Enhanced near-infrared-to-visible upconversion by synthetic control of PbS nanocrystal triplet photosensitizers. *J Am Chem Soc.* 2019;141(25):9769–9772. doi:10.1021/jacs.9b03385
230. Jia Q, Ge J, Liu W, et al. A magnetofluorescent carbon dot assembly as an acidic H₂O₂-driven oxygenerator to regulate tumor hypoxia for simultaneous bimodal imaging and enhanced photodynamic therapy. *Adv Mater.* 2018;30(13):e1706090. doi:10.1002/adma.201706090
231. Zheng DW, Li B, Li CX, et al. Carbon-dot-decorated carbon nitride nanoparticles for enhanced photodynamic therapy against hypoxic tumor via water splitting. *ACS Nano.* 2016;10(9):8715–8722. doi:10.1021/acsnano.6b04156
232. Teh DBL, Bansal A, Chai C, et al. A flexi-PEGDA upconversion implant for wireless brain photodynamic therapy. *Adv Mater.* 2020;32(29):e2001459. doi:10.1002/adma.202001459
233. Chen G, Jaskula-Sztul R, Esquibel CR, et al. Neuroendocrine tumor-targeted upconversion nanoparticle-based micelles for simultaneous NIR-controlled combination chemotherapy and photodynamic therapy, and fluorescence imaging. *Adv Funct Mater.* 2017;27(8):1604671. doi:10.1002/adfm.201604671
234. Chan MH, Pan YT, Lee IJ, et al. Minimizing the heat effect of photodynamic therapy based on inorganic nanocomposites mediated by 808 nm near-infrared light. *Small.* 2017;13(21):1700038.
235. Wang Y, Luo S, Wu Y, et al. Highly penetrable and on-demand oxygen release with tumor activity composite nanosystem for photothermal/photodynamic synergetic therapy. *ACS Nano.* 2020;14(12):17046–17062. doi:10.1021/acsnano.0c06415
236. Chen J, Wen K, Chen H, et al. Achieving high-performance photothermal and photodynamic effects upon combining D-A structure and nonplanar conformation. *Small.* 2020;16(17):e2000909. doi:10.1002/smll.202000909
237. Huang J, He B, Zhang Z, et al. Aggregation-induced emission luminogens married to 2D black phosphorus nanosheets for highly efficient multimodal theranostics. *Adv Mater.* 2020;32(37):e2003382. doi:10.1002/adma.202003382
238. Lv R, Yang P, He F, et al. A yolk-like multifunctional platform for multimodal imaging and synergistic therapy triggered by a single near-infrared light. *ACS Nano.* 2015;9(2):1630–1647. doi:10.1021/nn5063613
239. Sun S, Chen Q, Tang Z, et al. Tumor microenvironment stimuli-responsive fluorescence imaging and synergistic cancer therapy by Carbon-Dot-Cu²⁺ nanoassemblies. *Angew Chem Int Ed Engl.* 2020;59(47):21041–21048. doi:10.1002/anie.202007786
240. Hua XW, Bao YW, Zeng J, Wu FG. Ultrasmall all-in-one nanodots formed via carbon dot-mediated and albumin-based synthesis: multimodal imaging-guided and mild laser-enhanced cancer therapy. *ACS Appl Mater Interfaces.* 2018;10(49):42077–42087. doi:10.1021/acsami.8b16065
241. Zhang M, Wang W, Wu F, et al. Biodegradable Poly(γ -glutamic acid)@glucose oxidase@carbon dot nanoparticles for simultaneous multimodal imaging and synergetic cancer therapy. *Biomaterials.* 2020;252:120106. doi:10.1016/j.biomaterials.2020.120106
242. Lu Y, Li L, Li M, et al. Zero-dimensional carbon dots enhance bone regeneration, osteosarcoma ablation, and clinical bacterial eradication. *Bioconj Chem.* 2018;29(9):2982–2993. doi:10.1021/acs.bioconjchem.8b00400
243. Geng B, Qin H, Zheng F, et al. Carbon dot-sensitized MoS₂ nanosheet heterojunctions as highly efficient NIR photothermal agents for complete tumor ablation at an ultralow laser exposure. *Nanoscale.* 2019;11(15):7209–7220. doi:10.1039/C8NR10445J

244. Geng B, Shen W, Li P, et al. Carbon dot-passivated black phosphorus nanosheet hybrids for synergistic cancer therapy in the NIR-II window. *ACS Appl Mater Interfaces*. 2019;11(48):44949–44960. doi:10.1021/acsami.9b15569
245. Li Y, Bai G, Zeng S, Hao J. Theranostic carbon dots with innovative NIR-II emission for in vivo renal-excreted optical imaging and photothermal therapy. *ACS Appl Mater Interfaces*. 2019;11(5):4737–4744. doi:10.1021/acsami.8b14877
246. Wang X, Li X, Mao Y, et al. Multi-stimuli responsive nanosystem modified by tumor-targeted carbon dots for chemophototherapy synergistic therapy. *J Colloid Interface Sci*. 2019;552:639–650. doi:10.1016/j.jcis.2019.05.085
247. Xue M, Zhao J, Zhan Z, et al. Dual functionalized natural biomass carbon dots from lychee exocarp for cancer cell targetable near-infrared fluorescence imaging and photodynamic therapy. *Nanoscale*. 2018;10(38):18124–18130. doi:10.1039/C8NR05017A
248. Wang J, Zhang Z, Zha S, et al. Carbon nanodots featuring efficient FRET for two-photon photodynamic cancer therapy with a low fs laser power density. *Biomaterials*. 2014;35(34):9372–9381. doi:10.1016/j.biomaterials.2014.07.063
249. Zhang M, Wang W, Cui Y, et al. Near-infrared light-mediated photodynamic/photothermal therapy nanoplatform by the assembly of Fe₃O₄ carbon dots with graphitic black phosphorus quantum dots. *Int J Nanomedicine*. 2018;13:2803–2819. doi:10.2147/IJN.S156434
250. Phan LMT, Gul AR, Le TN, et al. One-pot synthesis of carbon dots with intrinsic folic acid for synergistic imaging-guided photothermal therapy of prostate cancer cells. *Biomater Sci*. 2019;7(12):5187–5196. doi:10.1039/C9BM01228A
251. Jia Q, Zheng X, Ge J, et al. Synthesis of carbon dots from *Hypocrella bambusae* for bimodal fluorescence/photoacoustic imaging-guided synergistic photodynamic/photothermal therapy of cancer. *J Colloid Interface Sci*. 2018;526:302–311. doi:10.1016/j.jcis.2018.05.005
252. Jia Q, Ge J, Liu W, et al. Gold nanorod@silica-carbon dots as multifunctional phototheranostics for fluorescence and photoacoustic imaging-guided synergistic photodynamic/photothermal therapy. *Nanoscale*. 2016;8(26):13067–13077. doi:10.1039/C6NR03459D
253. Bajpai VK, Khan I, Shukla S, et al. Multifunctional N-P-doped carbon dots for regulation of apoptosis and autophagy in B16F10 melanoma cancer cells and in vitro imaging applications. *Theranostics*. 2020;10(17):7841–7856. doi:10.7150/thno.42291
254. Yao H, Li J, Song Y, et al. Synthesis of ginsenoside re-based carbon dots applied for bioimaging and effective inhibition of cancer cells. *Int J Nanomedicine*. 2018;13:6249–6264. doi:10.2147/IJN.S176176
255. Emam HE, Ahmed HB. Antitumor/antiviral carbon quantum dots based on carrageenan and pullulan. *Int J Biol Macromol*. 2021;170:688–700. doi:10.1016/j.ijbiomac.2020.12.151

International Journal of Nanomedicine

Dovepress

Publish your work in this journal

The International Journal of Nanomedicine is an international, peer-reviewed journal focusing on the application of nanotechnology in diagnostics, therapeutics, and drug delivery systems throughout the biomedical field. This journal is indexed on PubMed Central, MedLine, CAS, SciSearch®, Current Contents®/Clinical Medicine,

Journal Citation Reports/Science Edition, EMBase, Scopus and the Elsevier Bibliographic databases. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/international-journal-of-nanomedicine-journal>