



Draft Genome Sequence of a Novel *Methylobacterium brachiatum* Strain Isolated from Human Skin

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ABSTRACT *Methylobacterium brachiatum* MBRA is an aerobic alphaproteobacterium isolated from the human skin on methanol-containing minimal medium. The genome was sequenced using Illumina and Nanopore technology, and the genome was assembled using Unicycler. *M. brachiatum* MBRA possesses two *sox* genes, one gene pair, *mx*A and *mx*B, and a complete serine pathway.

Methanol is a by-product of plant (1) and healthy human metabolism (2). Facultative methylophilic bacteria from the genera *Methylobacterium* and *Methylobacterium* have been isolated from the plant phyllosphere (1), soil (3), and the human body (4–6). *Methylobacterium brachiatum* was first isolated from a water sample from a food processing plant in Japan (7). Here, we present a draft genome of *M. brachiatum* MBRA, which we isolated from human skin as part of a laboratory course.

Strain MBRA was isolated by placing a human thumb (Nijmegen area, The Netherlands) on solid mineral salt medium (MSM) containing (per liter) 15 g agar, 1.5 g KH₂PO₄, 7.9 g Na₂HPO₄·2H₂O, 0.8 g NH₄Cl, 0.1 g MgSO₄·7H₂O, 0.5% methanol, 1 ml 1,000× SL10 trace element solution (8), 0.1 ml 10 mM CeCl₃ solution, and 0.02 ml 50 mM nitrilotriacetic acid (NTA) solution (medium pH set at 6.8). After growing at room temperature for 1 week, a single colony was picked and restreaked onto an MSM plate. After 5 d at 25°C, biomass was scraped from the plate, and DNA was isolated using a DNeasy PowerSoil kit (Qiagen, Venlo, The Netherlands). Long-read sequencing was performed with the MinION R9 flow cell (FLO-MIN106; Nanopore, Oxford, UK), according to the manufacturer's protocol, using unfragmented DNA (no size selection) and NEBNext formalin-fixed, paraffin-embedded (FFPE) repair mix (M6630), a ligation sequencing kit (SQK-LSK109), the NEBNext end repair/dA-tailing module (E7546), a flow cell priming kit (EXP-FLP001), NEB Blunt/TA ligase master mix (M0367), the NEBNext quick ligation module E6056, and barcode kit EXPNBD104. MiSeq sequencing (Illumina, San Diego, CA, USA) was performed with the Nextera XT kit according to the manufacturer's protocol. All DNA was measured using a Qubit fluorometer (Thermo Fisher Scientific, Waltham, MA, USA). The quality of the 2,329,486 paired-end Illumina reads (average, 253 bp) was checked using CLC Genomics Workbench v12 (Qiagen Aarhus A/S, Denmark). Read error correction was performed using SPAdes v3.13 (9) in the Unicycler v0.4.4 (10) pipeline, so no trimming was completed prior to assembly. Nanopore reads were base called and demultiplexed using Guppy v3.0.7 (minimum length, 3,000 bp), leaving 129,787 reads with an average length of 8,234 bp for assembly.

Assembly using reads from both methods in Unicycler (10) resulted in 25 contigs with an *N*₅₀ value of 1,578,407 bp. Eight contigs were manually binned by GC content (GC content, ~69%) and Illumina read-based coverage (coverage, ~50×) (Table 1). Unicycler also identified five circular high-copy-number small contigs, most likely plasmids. The completeness of the genome was checked using CheckM v1.0.11, indicating 97.2% completeness at the *Rhizobiales* level and 0% contamination (11). Average

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TABLE 1 Characteristics of the *Methylobacterium brachiatum* MBRA genome sequence and plasmids

Characteristic	Data for:					
	Genome	Plasmid 1	Plasmid 2	Plasmid 3	Plasmid 4	Plasmid 5
Size (bp)	6,459,278	53,490	48,251	45,734	24,909	16,067
DNA GC content (%)	69.4	61.4	68.0	64.9	67.0	56.1
No. of contigs	8	1	1	1	1	1
N_{50} (bp)	1,573,792	53,490	48,135	45,734	24,910	16,067
Circular	No	Yes	Yes	Yes	Yes	Yes
Total no. of genes	6,236	57	60	64	25	23
Protein coding density (%)	84.5	85.6	88.9	85.7	84.5	84.9
No. of rRNA genes	15	0	0	0	0	0
No. of tRNA genes	67	0	0	0	0	1
Coverage (\times) ^a	50	107	117	149	194	296

^a Based on MiSeq reads.

nucleotide identity (ANI) analysis by JSpeciesWS v3.6.1 (12) placed the genome at 97.9% similarity to *M. brachiatum*. The genome was annotated using Prokka v1.12 (13) and manually curated. Default parameters for all programs were used except where otherwise noted.

The genome sequence of strain MBRA reflects its facultative methylotrophic life-style. For growth on methanol, the genome encodes two lanthanide-dependent XoxF-type methanol dehydrogenases and a calcium-dependent MxaFI type (14), together with the recently discovered *lanM* gene (15) and the full serine pathway for carbon assimilation (1). Strain MBRA harbors a glycolysis pathway and a near-complete tricarboxylic acid (TCA) cycle. In addition, the genes for both dissimilatory and assimilatory nitrate reduction were present.

Data availability. This whole-genome shotgun sequencing project has been deposited in ENA under accession number [PRJEB35543](https://www.ebi.ac.uk/ena/record/PRJEB35543). The assembled genome (13 contigs) is deposited under accession numbers [CACTHX01000001](https://www.ebi.ac.uk/ena/record/CACTHX01000001) through [CACTHX01000013](https://www.ebi.ac.uk/ena/record/CACTHX01000013). The versions described in this paper are the first versions. The raw reads are available under SRA accession numbers [ERX3765083](https://www.ncbi.nlm.nih.gov/sra/ERX3765083) and [ERX3765085](https://www.ncbi.nlm.nih.gov/sra/ERX3765085).

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REFERENCES

- Vorholt JA. 2012. Microbial life in the phyllosphere. *Nat Rev Microbiol* 10:828–840. <https://doi.org/10.1038/nrmicro2910>.
- Dorokhov YL, Shindyapina AV, Sheshukova EV, Komarova TV. 2015. Metabolic methanol: molecular pathways and physiological roles. *Physiol Rev* 95:603–644. <https://doi.org/10.1152/physrev.00034.2014>.
- Kumar M, Tomar RS, Lade H, Paul D. 2016. Methylotrophic bacteria in sustainable agriculture. *World J Microbiol Biotechnol* 32:120. <https://doi.org/10.1007/s11274-016-2074-8>.
- Anesti V, McDonald IR, Ramaswamy M, Wade WG, Kelly DP, Wood AP. 2005. Isolation and molecular detection of methylotrophic bacteria occurring in the human mouth. *Environ Microbiol* 7:1227–1238. <https://doi.org/10.1111/j.1462-2920.2005.00805.x>.
- Anesti V, Vohra J, Goonetilleka S, McDonald IR, Sträubler B, Stackebrandt E, Kelly DP, Wood AP. 2004. Molecular detection and isolation of facultatively methylotrophic bacteria, including *Methylobacterium podarium* sp. nov., from the human foot microflora. *Environ Microbiol* 6:820–830. <https://doi.org/10.1111/j.1462-2920.2004.00623.x>.
- Carvajal TM, Tan RL, Lee AC. 2011. *Methylobacterium zatmanii*, a pink pigmented facultative methylotrophic (PPFM) bacterium isolated from the human oral cavity. *Philippine J Syst Biol* V:1–9.
- Kato Y, Asahara M, Goto K, Kasai H, Yokota A. 2008. *Methylobacterium persicinum* sp. nov., *Methylobacterium komagatae* sp. nov., *Methylobacterium brachiatum* sp. nov., *Methylobacterium tardum* sp. nov. and *Methylobacterium gregans* sp. nov., isolated from freshwater. *Int J Syst Evol Microbiol* 58:1134–1141. <https://doi.org/10.1099/ijs.0.65583-0>.
- Widdel F, Kohring G-W, Mayer F. 1983. Studies on dissimilatory sulfate-reducing bacteria that decompose fatty-acids: III. Characterization of the filamentous gliding *Desulfonema limicola* gen. nov. sp. nov., and *Desulfonema magnum* sp. nov. *Arch Microbiol* 134:286–294. <https://doi.org/10.1007/BF00407804>.
- Nurk S, Bankevich A, Antipov D, Gurevich A, Korobeynikov A, Lapidus A, Pribelsky A, Pyshkin A, Sirotkin A, Sirotkin Y, Stepanauskas R, McLean J, Lasken R, Clingenpeel SR, Woyke T, Tesler G, Alekseyev MA, Pevzner PA. 2013. Assembling genomes and mini-metagenomes from highly chimeric reads, p 158–170. In Deng M, Jiang R, Sun F, Zhang X (ed), *Research in computational molecular biology. RECOMB 2013. Lecture notes in computer science*, vol 7821. Springer, Berlin, Germany.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
- Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a Web server for prokaryotic species circumscription based on pairwise

- genome comparison. *Bioinformatics* 32:929–931. <https://doi.org/10.1093/bioinformatics/btv681>.
13. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
14. Keltjens JT, Pol A, Reimann J, Op den Camp HJM. 2014. PQQ-dependent methanol dehydrogenases: rare-earth elements make a difference. *Appl Microbiol Biotechnol* 98:6163–6183. <https://doi.org/10.1007/s00253-014-5766-8>.
15. Cotruvo JA, Jr, Featherston ER, Mattocks JA, Ho JV, Laremore TN. 2018. Lanmodulin: a highly selective lanthanide-binding protein from a lanthanide-utilizing bacterium. *J Am Chem Soc* 140:15056–15061. <https://doi.org/10.1021/jacs.8b09842>.